



# LUND UNIVERSITY

## Systematics, phylogeography and polyploid evolution in the *Dactylorhiza maculata* complex (Orchidaceae)

Ståhlberg, David

2007

[Link to publication](#)

*Citation for published version (APA):*

Ståhlberg, D. (2007). *Systematics, phylogeography and polyploid evolution in the *Dactylorhiza maculata* complex (Orchidaceae)*. [Doctoral Thesis (compilation), Department of Biology]. Department of Ecology, Lund University.

*Total number of authors:*

1

### General rights

Unless other specific re-use rights are stated the following general rights apply:

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

Read more about Creative commons licenses: <https://creativecommons.org/licenses/>

### Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

LUND UNIVERSITY

PO Box 117  
221 00 Lund  
+46 46-222 00 00



# **Systematics, phylogeography and polyploid evolution in the *Dactylorhiza maculata* complex (Orchidaceae)**

**David Ståhlberg**

## **AKADEMISK AVHANDLING**

som för avläggande av filosofie doktorsexamen vid naturvetenskapliga fakulteten, Lunds universitet, kommer att offentligen försvaras vid Ekologiska institutionen, Avdelningen för växtekologi och systematik, fredagen den 27 april 2007 kl 10.00.

Plats: Röda rummet, Ekologihuset, Sölvegatan 37.

Fakultetsopponent: Professor Hilde Nybom, Sveriges lantbruksuniversitet, Balsgård.

Lund 2007

A doctoral thesis at a university in Sweden is produced either as a monograph or as a collection of papers. In the latter case, the introductory part constitutes the formal thesis, which summarizes the accompanying papers. These have either already been published or are manuscripts at various stages (in press, submitted or in ms).

Cover: Satyrium Basilicum alterum (vel femina)

[*Dactylorhiza maculata* ssp. *maculata*]

Picture from Rembert Dodoens (1583): *Stirpium historiae pemptades sex. Antuerpiae.*

ISBN: 978-91-7105-254-4

SE-LUNBDS/NBBE-07/1084+189pp

© 2007 David Ståhlberg

Printed in Sweden by Media-Tryck, Lund, 2007

This thesis is based on the following papers, which are referred to by their Roman numerals:

- I Hedrén M., Nordström S., Ståhlberg D. Plastid DNA variation and the origin of allopolyploid *Dactylorhiza* (Orchidaceae) in Scandinavia. Submitted.
- II Ståhlberg D., Hedrén M. Systematics and phylogeography of *Dactylorhiza maculata* s.l. (Orchidaceae) in Scandinavia: insights from cytological, morphological and molecular data. Manuscript.
- III Ståhlberg D. Habitat differentiation, hybridization and gene flow patterns in mixed populations of diploid and autotetraploid *Dactylorhiza maculata* s.l. (Orchidaceae) in Scandinavia. Manuscript.
- IV Ståhlberg D., Hedrén M. Phylogeography of the *Dactylorhiza maculata* polyploid complex (Orchidaceae): high genetic diversity in the north and indications of plastid DNA recombination. Manuscript.



# Systematics, phylogeography and polyploid evolution in the *Dactylorhiza maculata* complex (Orchidaceae)

## Introduction and background

### *Influence of Quaternary climatic changes on speciation*

Quaternary climatic changes have had a profound impact on speciation, structuring of genetic diversity and the shaping of the present-day distributions of plant and animal taxa (Vuilleumier, 1971; Hewitt, 1996, 2000, 2004; Avise, 2000). In Europe, the repeated cycles of glacials and interglacials during the Pleistocene (c. 2 Ma until 10 000 BP) have caused massive fluctuations in the distributions of taxa. Fragmentation and isolation of populations during the long-lasting glacials and expansion during the short interglacials have resulted in marked differences among regions in intraspecific diversity. Oscillations of population sizes, bottle necks, founder events and other population historical events associated with climatic shifts have further contributed to differentiation among regional population groups. As a combined effect of range shifts and population differentiation, divergent lineages have occasionally formed contact zones, leading to reticulate speciation by means of hybridization and polyploidization (Grant, 1981; Stebbins, 1984; Hewitt, 1988, 2001). A high frequency of polyploid taxa may be a particularly important feature of regions harbouring young floras and faunas, including Fennoscandia and adjacent areas of northern Europe that were covered by thick ice sheets during the maximum of the last (Weichselian) glaciation (LGM; c. 22 000 to 18 000 BP), as compared to more climatologically stable regions (Löve and Löve 1974; Stebbins, 1971; Hewitt, 1988; Soltis et al., 2003).

Analysis of macrofossil and pollen data together with organellar markers have shown that populations of many temperate species in the European flora and fauna survived the LGM in various southern refugia in the Mediterranean region (the Iberian, Apennine and Balkan peninsulas), and in the Caspian/Caucasian region (Huntley and Birks, 1983; Petit et al., 1993; Demesure et al., 1996; Hewitt, 2004). Similarly, patterns of postglacial migration have been reconstructed for many temperate species, and a general picture of high intraspecific genetic diversity in refugial areas in the south and low diversity in previously glaciated areas in the north has been established (Ferris et al., 1999). However, this picture may be too simple (cf. Widmer and Lexer, 2001). Increasing evidence suggests that the southern refugia for temperate species were supplemented by more northern refugia during the LGM (reviewed by Stewart and Lister, 2001), which clearly would have resulted in more complex patterns of the distribution of intraspecific genetic diversity. Areas of sheltered topography in mountainous parts of Central Europe may have provided suitable stable microclimates for thermophilous plant and animal species (Litynska-Zajac, 1995; Willis and van Andel, 2004; Magri et al., 2006; Ursenbacher et al., 2006; Sommer and Nadachowski, 2006). Populations of more cold tolerant species may as well have survived close to the southern and eastern edges of the Fennoscandian ice sheet together with arctic-alpine species (Rendell and Ennos, 2002; Palmé et al., 2003; Alsos et al., 2005; Skrede et al., 2006).

Increased intraspecific genetic diversity also occurs in contact zones where divergent populations from separate refugia meet (Petit et al., 2003). Such zones of secondary contact have been demonstrated for both plants and animals at intermediate latitudes in Central Europe (Petit et al. 2003). Polyploidization appears to be common at these latitudes (Stebbins, 1984). Several studies have indicated that central-northern Scandinavia may be another area of secondary contact between divergent populations immigrating from the northeast and the south (Jaarola and Tegelström, 1995; Fredga, 1996; Nyberg-Berglund and Westerbergh, 2001).

## *Polyplloid evolution*

Polyplloid speciation has long been recognized as an important process in plant evolution (Müntzing, 1936; Stebbins 1950; Grant, 1981). Recent genomic studies have made it clear that angiosperms possess genomes with considerable gene redundancy, indicating that “most (if not all) plants have undergone one or more episodes of polyplloidization” (Soltis et al., 2003). Many taxa are ancient polyploids that have become secondarily diploidized due to gene silencing and other genomic processes (e.g. Lynch and Conery, 2000). Functional polyploids in the traditional sense refer to polyploids that are integral parts of polyplloid complexes consisting of closely related taxa at various ploidy levels (cf. Grant, 1981; Soltis et al., 2003). Polyplloidization is probably the most common mechanism of sympatric speciation (Otto and Whitton, 2000), and it is widely accepted that a single taxon may arise several times by independent polyplloidization events, via both allopolyploidization and autopolyplloidization (Soltis and Soltis, 1993, 1999; Soltis et al., 2003).

Chromosome doubling is an instantaneous mode of speciation that results in effective postzygotic barriers between new polyploids and their diploid progenitors (Ramsey and Schemske, 1998). On the other hand, new polyploid plants are subjected to competitive constraints because they are also exposed to pollen from diploid plants, which results in triploid offspring that have low fitness (minority cytotype disadvantage; Levin, 1975). Adaptive strategies are needed for establishment and persistence of new polyploids. Such strategies include apomixis, self-pollination, pollinator shift and habitat differentiation (Soltis et al., 2003). However, even though the literature on polyploidy is comprehensive there are relatively few studies that focus on the ecological aspects of polyplloidization.

## *Variation in Dactylorhiza*

*Dactylorhiza* Necker ex Nevski is one of the most taxonomically investigated genera in the orchid family (e.g. Klinge, 1898; Vermeulen, 1947; Soó, 1960; Senghas, 1968; Nelson, 1976; Averyanov, 1990; Pedersen, 1998). The genus is widespread (Eurasia, Northern Africa, Alaska), and consists of a confusing variety of forms that are difficult to sort into discrete taxa. Consequently, the number of species varies strongly among authors, ranging from 12 (Klinge, 1898) to 75 (Averyanov, 1990). Part of the taxonomic complexity could be explained by the frequent interpretation of aberrant populations and specimens as separate taxa (Bateman and Denholm, 2003), but more important explanations may be innate factors such as phenotypic plasticity, or that many taxa are young and have not yet acquired good separating characters. Since long, hybridization and polyplloidization have been recognized as critical factors for the understanding of the diversification in *Dactylorhiza* (Hagerup, 1938; Heslop-Harrison, 1957). During the last decade, molecular tools have provided deeper insights into the evolutionary history of *Dactylorhiza* and the intricate patterns of speciation that characterize the genus (Hedrén, 1996, 2001, 2003; Hedrén et al., 2001, 2007; Pridgeon et al., 1997; Bateman et al., 1997; Pedersen, 1998, 2004, 2006; Bullini et al., 2001; Devos et al., 2003, 2005, 2006a, b; Shipunov et al., 2004, 2005; Pillon et al., 2006, 2007).

Most species of *Dactylorhiza* form a polyplloid complex that has undergone extensive reticulate evolution (summarized by Hedrén, 2002). In Europe, a large number of allotetraploid taxa (i.e. taxa belonging to the *D. majalis* [Rchb.] P.F. Hunt & Summerh. group) have evolved repeatedly by hybridization between two broadly defined parental lineages: the *D. incarnata* (L.) Soó s.l. lineage and the *D. maculata* (L.) Soó s.l. lineage. Extensive studies of these two parental lineages are necessary to achieve a detailed comprehension of polyplloid evolution in *Dactylorhiza*. A better understanding of the

variation patterns in *D. incarnata* s.l. and *D. maculata* s.l. is urgent also for proper decisions about conservation priorities, since many allotetraploid taxa in Europe are threatened by habitat loss (e.g. Janečková et al., 2006; Pillon et al., 2006).

It appears that more information will be gained by studying the *D. maculata* s.l. parental lineage, rather than the *D. incarnata* s.l. parental lineage. Nearly all investigated allotetraploids with *D. incarnata* s.l. x *D. maculata* s.l. origin have inherited their plastid genomes from the *D. maculata* s.l. parent (Hedrén, 2003; Hedrén et al., 2007; Devos et al., 2003, 2006a; Shipunov et al., 2004, 2005; Pillon et al., 2007). It is therefore of particular interest to investigate *D. maculata* s.l. for plastid DNA variation. Also at nuclear marker loci it appears more profitable to analyze *D. maculata* s.l. rather than *D. incarnata* s.l. *Dactylorhiza maculata* s.l. is considerably more variable than *D. incarnata* s.l. at all nuclear marker loci investigated so far: allozymes (Hedrén, 1996, 2001; Pedersen, 1998, 2004, 2006), amplified fragment length polymorphisms (AFLPs; Hedrén et al. 2001, 2007) and internal transcribed spacers of nuclear ribosomal DNA (ITS nrDNA; Shipunov et al., 2004, 2005; Devos et al., 2005, 2006a; Pillon et al., 2007). Since *D. maculata* s.l. is morphologically very variable and occupies a wide range of habitats, molecular genetic studies should ideally be combined with morphometric and ecological studies.

### The *Dactylorhiza maculata* complex

*Dactylorhiza maculata* s.l. is a morphologically and genetically variable polyploid complex consisting of diploid ( $2n = 40$ ) and tetraploid ( $2n = 80$ ) cytotypes (Averyanov, 1990; Hedrén, 1996; Hedrén et al., 2001; Tyteca, 2001; Bateman and Denholm, 2003; Shipunov et al., 2004; Devos et al., 2005). At least 30 taxa at various taxonomic levels have been described (Soó, 1960; Delforge, 1995), but most contemporary authors distinguish between three or four morphologically and largely cytologically defined taxa: (1) *D. maculata* ssp. *fuchsii* (Druce) Hyl., a predominantly diploid taxon that typically grows in semi-open woodlands on fertile soils throughout most of northwestern Eurasia (absent or rare in southern and southeastern Europe); (2) *D. maculata* ssp. *saccifera* (Brongn.) Diklic, a diploid taxon that gradually replaces ssp. *fuchsii* on the Apennine peninsula and in southeastern Europe; (3) *D. maculata* ssp. *maculata*, a tetraploid taxon that characteristically is found in more open habitats such as grasslands, coastal moorlands and boreal-subarctic peatlands in western and northern Eurasia (absent in southeastern Europe); (4) *D. foliosa* (Sol. ex Lowe) Soó, a geographically isolated Madeiran diploid. It should be observed that tetraploid populations of *D. maculata* ssp. *fuchsii* are common in the mountain areas of Central Europe (e.g. Groll, 1965; Vöth and Greilhuber, 1980), and that ssp. *maculata* is rare or absent in the same region (e.g. Klein and Kerschbaumsteiner, 1996).

Chromosome counts and ploidy level determinations of almost 400 populations of *D. maculata* s.l. from throughout all of the distribution range have been reported in the literature (Table 1). The distributions of diploid and tetraploid populations with respect to geography and taxonomy are shown in Figure 1. This compilation shows that there is a clear correlation between ploidy level and taxonomy, except for populations of ssp. *fuchsii*. Of more than 150 cytologically investigated populations of ssp. *fuchsii* from the Alps, northern Apennines, western Carpathians and adjacent mountain areas in eastern Germany and the Czech Republic, 80 % have turned out to consist of tetraploid plants; in the Alps the proportion is even higher. Outside Central Europe, populations of ssp. *fuchsii* consist of diploid plants.

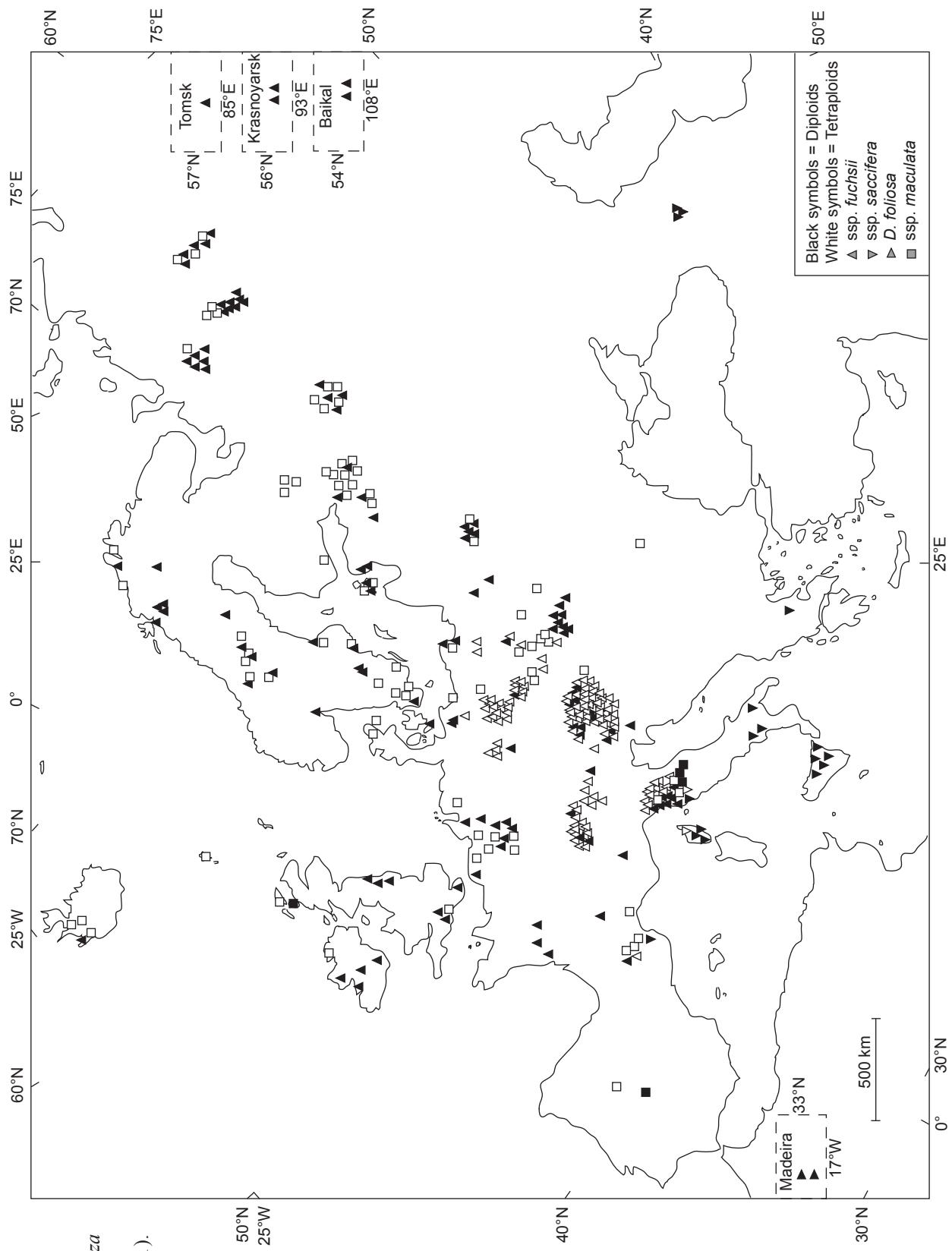
Morphometric studies have shown that ssp. *fuchsii* and ssp. *maculata* are relatively distinct in southern and western Europe (Heslop-Harrison, 1951; Bateman and Denholm, 1989, Dufrêne et al., 1991; Tyteca and Gathoye, 2004). In contrast, other studies have indicated that

the morphological differentiation decreases towards the north (Averyanov, 1990; Shipunov et al., 2004, 2005). In general, hybridization between ssp. *fuchsii* and ssp. *maculata* is considered to be common (e.g. Delforge, 1995), and triploid hybrids are occasionally reported (Table 1). So far, no taxonomic studies of the *D. maculata* complex have incorporated detailed ecological data.

Using allozyme markers it has been shown that tetraploid populations of *D. maculata* s.l. have originated by autopolyploidization (Hedrén, 1996). Regionally focused studies based on plastid DNA and/or nrDNA markers (e.g. Devos et al., 2003, 2005, 2006a; Shipunov et al., 2004; Hedrén, 2003; Pillon et al., 2007), as well as on morphometry and/or cytometry (e.g. Heslop-Harrison, 1951; Vöth, 1978; Reinhard, 1985; Jagiełło, 1986-1987; Bateman and Denholm, 1989; Dufrêne et al., 1991; Tyteca and Gathoye, 2004) suggest, when considered together, that tetraploid populations of *D. maculata* s.l. may include at least three separate autotetraploid lineages: (i) *D. maculata* ssp. *maculata* from southern and western Europe, (ii) *D. maculata* ssp. *maculata* from northern and eastern Europe and (iii) *D. maculata* ssp. *fuchsii* from the mountain areas of Central Europe. According to more or less obvious morphological differences between ssp. *maculata* and present-day diploids, the first two lineages may be relatively ancient. Moreover, there are differences in chromosome size (Jagiełło and Lankosz-Mróz, 1986-1987) and in AFLP banding patterns (Hedrén et al., 2001) between ssp. *maculata* and present-day diploids. In contrast, the third lineage may be relatively young since diploid and tetraploid populations of ssp. *fuchsii* are morphologically indistinguishable (Groll, 1965; Vaucher, 1966; Scharfenberg, 1977; Vöth, 1978; Vöth and Greilhuber, 1980; Jagiełło, 1986-1987; Jagiełło and Lankosz-Mróz, 1986-1987; Reinhardt, 1988; Gölz and Reinhard, 1997; Bertolini et al., 2000).

## Aims of this thesis

The association between taxonomic complexity and universal biological issues such as Quaternary migration history and polyploid evolution makes the foundation for this thesis. The objective was to investigate and describe different aspects of variation in the *Dactylorhiza maculata* complex and relate the variation patterns to underlying biological processes. Four separate studies are included in the thesis. In the first study (**Paper I**) focus was on the origin of allopolyploid taxa in Scandinavia. This study was also a methodological study in which plastid DNA markers with appropriate variation were developed and examined. The next study (**Paper II**) had a more taxonomic approach. Molecular markers from both the plastid and nuclear genomes were combined with cytological and morphological data in order to unravel patterns of differentiation in the *D. maculata* complex in Scandinavia. The third study (**Paper III**) was a detailed investigation of habitat differentiation, hybridization and gene flow patterns in mixed populations of diploid ssp. *fuchsii* and tetraploid ssp. *maculata*. Plastid and nuclear DNA markers, as well as cytological, morphological and ecological data were used for this investigation. The last study (**Paper IV**) was a large-scale overview of population genetic structure, postglacial migration and polyploidization in the *D. maculata* complex. This study was based on both plastid and nuclear DNA markers.



**Figure 1.** Distribution of diploid and tetraploid populations of *Dactylorhiza maculata* s.l. based on literature data (see Table 1).

## Material and methods

### *Sampling*

The variation patterns in the *Dactylorhiza maculata* complex were investigated at various geographical levels.

For the study of allotetraploid speciation (Paper I) plant material was mainly collected from northern Europe, but some reference material from other parts of Europe was included as well. More than 1 000 individuals from c. 150 populations of *D. majalis* s.l. (allotetraploids) and *D. maculata* s.l. (the maternal lineage) were sampled for this study. In the taxonomically focused study of the *D. maculata* complex in Scandinavia (Paper II) we investigated about 30 pure populations of either diploid ssp. *fuchsii* or tetraploid ssp. *maculata*. The populations were chosen to cover as much as possible of the biogeographic variation in Scandinavia. One population from the adjacent Kola Peninsula was included to improve the representation of morphologically controversial northern populations. Between 5 and 20 individuals from each population were analysed. In the third study (Paper III), which had an ecological perspective, I investigated two mixed populations of diploid ssp. *fuchsii* and tetraploid ssp. *maculata*. The populations were located in the coastal lowland of central Sweden (Sjösa nature reserve; 58°45'N, 17°07'E) and the central part of the Scandinavian mountain ridge (Hamra nature reserve; 62°34'N, 12°15'E). I chose these two sites since they represent contrasting biogeographic regions with different environmental conditions. About 50 samples/sample plots at each site were analysed. The fourth study (Paper IV) had a broad geographic perspective. Almost 2 000 samples of *D. maculata* s.l. from c. 300 populations from all parts of Europe were analysed.

### *Ploidy level determination*

In two of the studies (Papers II-III) the relative ploidy level of each sampled plant was assessed by flow cytometry. The analyses were performed by Gerard Geenen, Plant Cytometry Services (Schijndel, The Netherlands).

### *Molecular methods*

Two categories of molecular markers were used: plastid DNA and ITS alleles from the nuclear genome. Both marker systems are supposed to be selectively neutral and are standard tools for population genetic investigations. They have previously been used in studies of *Dactylorhiza* (see background). Plastid markers are generally maternally inherited in angiosperms and are particularly useful in phylogeographic studies since they are expected to provide a more simplified reflection of migration patterns than biparentally inherited markers (Ferris et al., 1999). In this thesis, plastid markers were used in all the studies. In the first study (Paper I) we found appropriate variation at seven microsatellite loci and three loci with indel variation. These ten loci were then used in the following studies. ITS markers (six different alleles) were used in three of the studies (Papers II-III).

### *Morphometry*

Morphological data were used in two studies (Papers II-III). In these studies a total of 35 quantitative and qualitative characters were measured on all plants. Both floral and vegetative characters were taken into account. The methodology was slightly modified from Bateman and Denholm (1985).

### *Ecological data*

Ecological data was used in one study (Paper II). In this study the niche of each sampled plant was characterized by data of presence and cover of associated plant species (herbaceous plants, dwarf shrubs and saplings) in a 40 x 40 cm quadrat centred on the target *Dactylorhiza* individual (cf. Du Rietz, 1921). Eight environmental variables were recorded in each quadrat in addition to the vegetation data: (1) shading, (2) cover of exposed soil, (3) cover of litter, (4) cover of *Sphagnum*, (5) cover of mosses other than *Sphagnum*, (6) pH on the top 10 cm of soil, (7) moisture and (8) grass sward density.

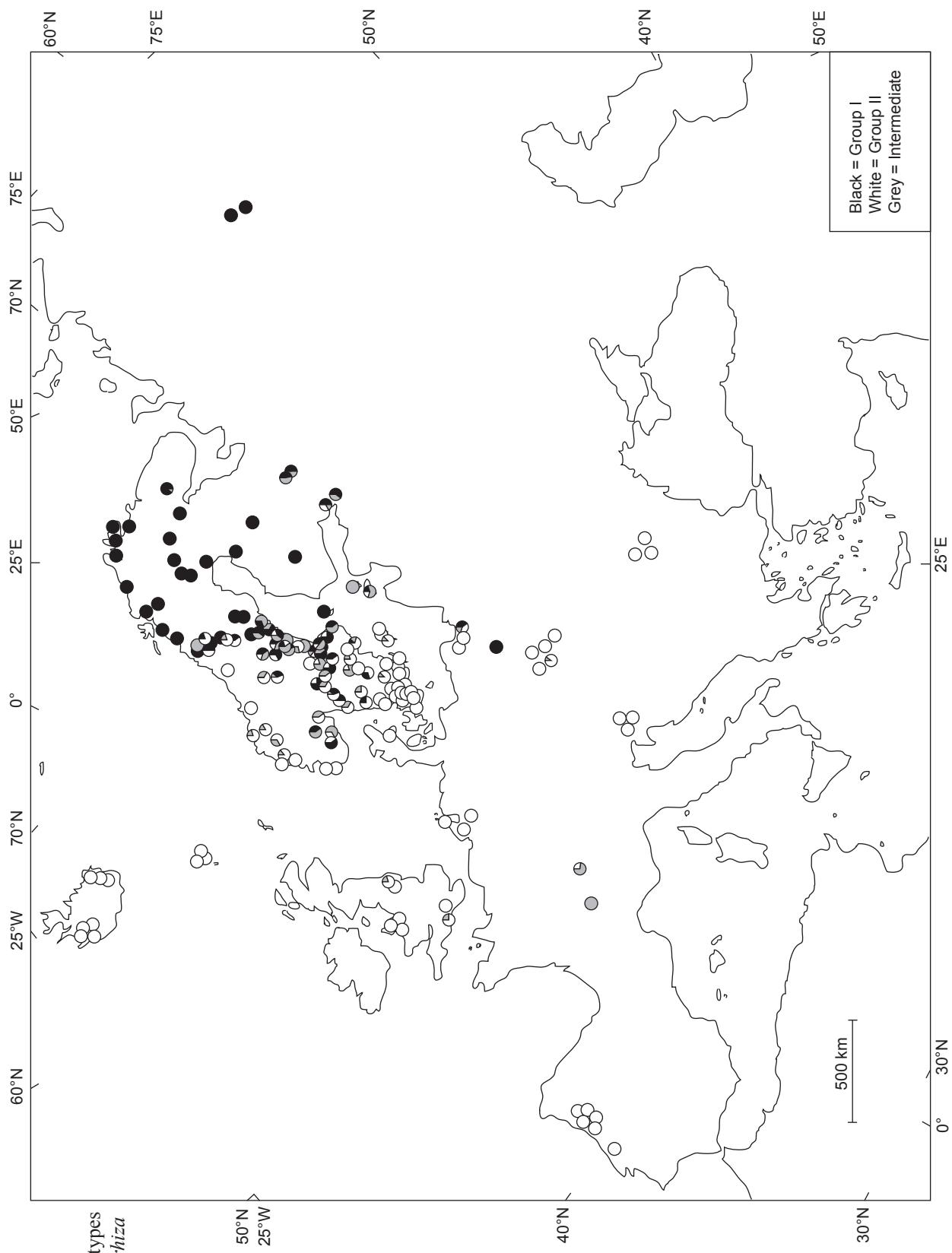
### *Data analysis*

Variation patterns in different data sets were summarized by means of appropriate multivariate techniques (see Papers I-IV for details): canonical correspondence analysis (CCA), canonical variates analysis (CVA), detrended correspondence analysis (DCA), discriminant analysis, multidimensional scaling (MDS), principal coordinates analysis (PCO) and principal components analysis (PCA). Relationships between plastid haplotypes were visualized in minimum spanning networks. Analysis of molecular variance (AMOVA) was used to describe the partitioning of genetic diversity among various spatial levels. Mantel tests were performed to unravel large-scale geographic patterns. Geographic patterns were also described by means of distribution maps of plastid haplotypes and ITS alleles. Genetic diversity at various spatial levels was calculated according to the indices of gene diversity ( $H$ ) and average gene diversity over loci ( $\pi$ ) (Excoffier et al., 2005).

## **Results and conclusions**

### *Plastid haplotypes (Papers I-IV)*

We identified almost 300 plastid haplotypes in the total material of *D. maculata* s.l. and allotetraploid taxa that was investigated. The haplotypes were differentiated into three distinct groups. Group I haplotypes were dominating in populations of ssp. *fuchsii* (including ssp. *saccifera*) and in populations of ssp. *maculata* from northern and northeastern Europe (“northern/eastern ssp. *maculata*”). Group II haplotypes were dominating in populations of ssp. *maculata* from southern, central and western Europe (“southern/western ssp. *maculata*”). Intermediate haplotypes were found in contact zones between the two lineages of ssp. *maculata*. The geographic distribution of haplotypes in populations of ssp. *maculata* is shown in Figure 2. Both haplotype groups were represented in allotetraploid taxa in Scandinavia. The third group of haplotypes was rare and almost completely restricted to allotetraploid material, but must have originated from *D. incarnata* s.l. (see Introduction).



**Figure 2.** Distribution of Group I, Group II and intermediate plastid haplotypes in populations of *Dactylorhiza maculata* ssp. *maculata*.

### *Origin of allotetraploids* (Paper I, complemented by data from Paper IV)

We found several distinct plastid haplotypes in widespread allotetraploid taxa (entities belonging to the *Dactylorhiza majalis* complex), which shows that the allotetraploid group must have multiple origins. Similar (mostly identical) haplotypes were found in both *D. majalis* s.l. and in *D. maculata* s.l. (the maternal lineage). However, some haplotypes are common in *D. majalis* s.l. but rare in *D. maculata* s.l., which suggests a pre-Holocene origin of most allotetraploid lineages. Populations of *D. majalis* s.l. in Scandinavia are invariably associated with widespread allotetraploid lineages and do not seem to have arisen via recent polyploidization events *in situ*. Morphologically distinct populations should be considered as segregates of already existing allotetraploid lineages. Our data indicate further that back-crossing between allotetraploids and their parental lineages is relatively common, and this may be an alternative pathway for the formation of new allopolyploid lineages. Hybridization between independently derived allotetraploids may also occur.

### *Taxonomy of the Dactylorhiza maculata complex* (Papers II-IV)

Both plastid and nuclear markers support the current view of four major taxa in the *Dactylorhiza maculata* complex: *D. maculata* ssp. *fuchsii*, *D. maculata* ssp. *saccifera*, *D. maculata* ssp. *maculata* and *D. foliosa* (cf. Devos et al., 2005, 2006a). It should however be observed that ssp. *fuchsii* and ssp. *saccifera* grade into each other. In Scandinavia, we found that ssp. *fuchsii* and ssp. *maculata* have partly overlapping morphological variation patterns, but that the two subspecies could be kept separate if morphology is considered together with habitat features. Based on ITS allele frequencies and morphology, populations of ssp. *maculata* from northernmost Europe can be recognized as var. *kolaënsis*. Morphologically more or less distinct groups of populations from other parts of Europe are sometimes recognized as independent taxonomic entities as well (e.g. Delforge, 1995). However, apart from var. *kolaënsis*, other taxa separated at subspecies or variety level are not supported by molecular markers. Furthermore, some taxa previously recognized on basis of ecology and distribution, including “*elodes*”, “*ericetorum*” and “*psychrophila*”, were also poorly separated in morphology, based on Scandinavian material.

### *Niche differentiation between diploids and autotetraploids* (Paper III, complemented by data from Paper IV)

Mixed populations of diploid and autotetraploid *Dactylorhiza maculata* s.l. in Scandinavia represent secondary contact zones between diploid ssp. *fuchsii* and tetraploid ssp. *maculata*. I found no patterns of recent and local (*in situ*) autoploidization. Based on both molecular markers and morphology it must be concluded that diploids and tetraploids from mixed populations are no less differentiated than diploids and tetraploids from cytologically homogeneous populations. Furthermore, diploid ssp. *fuchsii* and tetraploid ssp. *maculata* are separated on a microhabitat level in mixed populations. Both taxa appear to have wider ecological amplitude in pure populations. The ecological constraints may thus be strengthened in mixed populations, which should contribute to the maintenance of hybrid zones. I found unexpectedly few triploid hybrids. Most of them grew in intermediate habitats between diploids and tetraploids. Introgressive gene flow between ploidy levels was also limited, especially from tetraploid to diploid level (cf. Stebbins, 1971). However, I observed that hybridization and introgression seem to be slightly more common in the Scandinavian

mountains than in the lowland, which may be related to differences in disturbance regimes (cf. Anderson, 1948; Arnold, 1997).

The general conclusion that introgression is restricted between ssp. *fuchsii* and ssp. *maculata* is strongly supported by data from particularly southern/western Europe. Of several hundred investigated individuals of ssp. *maculata* from this area, none contained ssp. *fuchsii* plastid haplotypes. However, a local influence of introgression (past or present) was observed in Central Europe, where most populations of ssp. *fuchsii* consist of tetraploid plants. Reproductive barriers between taxa on the same ploidy level should be less efficient than barriers between taxa on different ploidy levels (Grant, 1981).

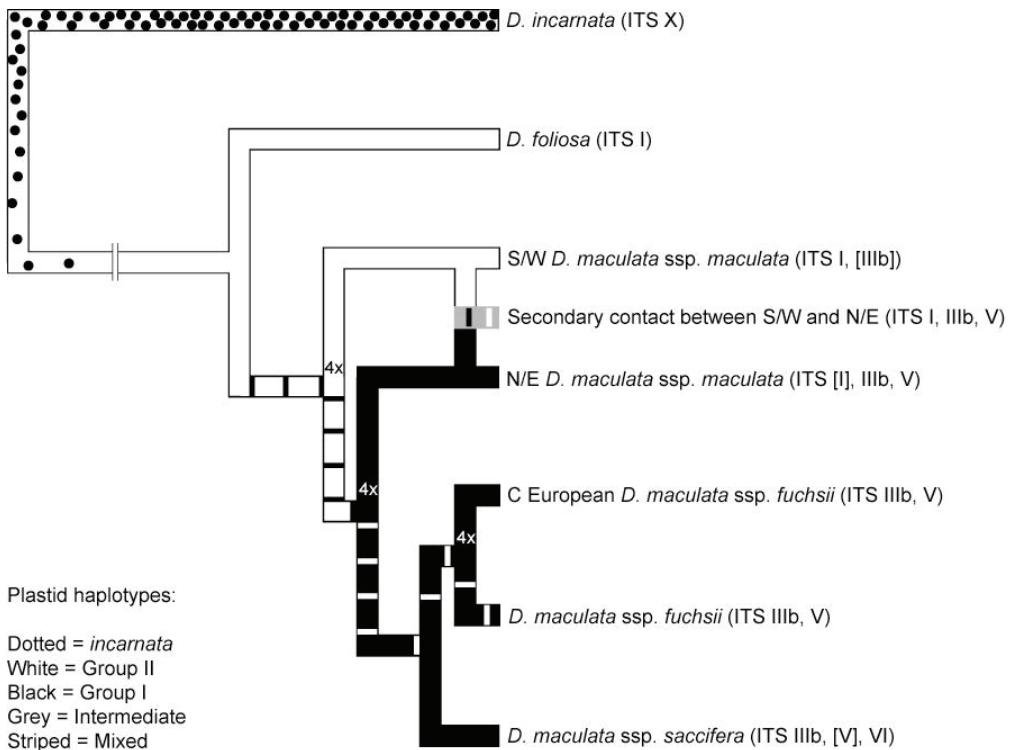
#### *Polyplloid evolution* (Paper IV, supported by data from Paper II)

A model of the evolutionary history of the *Dactylorhiza maculata* complex is given in Figure 3. Based on both plastid and nuclear markers we identified three distinct autotetraploid lineages: a southern/western lineage of ssp. *maculata*, a northern/eastern lineage of ssp. *maculata* and a Central European lineage of ssp. *fuchsii*. Given the level of differentiation between major plastid haplotype groups, and the present-day distribution of haplotypes belonging to these groups, we conclude that both the southern/western and northern/eastern lineages of ssp. *maculata* must have arisen before the Holocene. The tetraploid lineage of ssp. *fuchsii* is genetically and morphologically indistinguishable from diploid ssp. *fuchsii* and is most likely of postglacial origin (cf. references in Introduction). The southern/western lineage of ssp. *maculata* has probably arisen from diploid ancestors common to this lineage and to *D. foliosa*. The northern/eastern lineage of ssp. *maculata* has probably arisen from diploid ancestors common to this lineage and to ssp. *fuchsii*. It should be observed that populations belonging to ssp. *maculata* are found in two different genetically defined lineages, and in each of these ssp. *maculata* is connected to a diploid taxon that is different in morphology. This pattern indicates restricted morphological evolution in the tetraploid lineages, which may thus have preserved some characters that have been modified in the diploids. We found that southern/western populations of ssp. *maculata* are morphologically somewhat different from northern/eastern populations, but there is no distinct morphological limit between the two lineages.

#### *Glacial refugia and postglacial recolonization* (Paper IV)

The separation of *Dactylorhiza maculata* ssp. *maculata* in two distinct lineages evidently indicates postglacial recolonization from two separate refugial areas. However, the phylogeographic signal within each lineage is weak. Both lineages are dominated by a few widespread plastid haplotypes and by a large number of rare and geographically restricted haplotypes. The weak phylogeographic signal within the two ssp. *maculata* lineages may reflect effective seed dispersal and a propensity for long distance gene flow, as have been suggested for other plant taxa that exhibit similar patterns of plastid haplotype distribution (e.g. *Betula*; Palmé et al, 2003). Orchid seeds are dust-like and wind-borne, which implies a potential for long-distance dispersal. Based on the present-day distribution of genetic diversity, we suggest that source areas for postglacial migration of ssp. *maculata* may have been Central Europe and parts of central Russia located between the Fennoscandian ice sheet and the Urals. Populations of ssp. *fuchsii* were also characterized by a few widespread plastid haplotypes and by many local haplotypes. During the LGM, areas of sheltered topography in Central Europe may have provided suitable habitats for ssp. *fuchsii*, which is a more

thermophilous taxon than ssp. *maculata*. The Mediterranean region and the Caucasus have not contributed to northward migration, neither for ssp. *fuchsii*, nor for ssp. *maculata*.



**Figure 3.** Evolutionary history of the *Dactylorhiza maculata* polyploid complex as indicated by plastid and nrDNA markers. Autopolyploidization events are marked by “4x.” Note changes in relative frequencies of Group II and Group I haplotypes. ITS alleles that occur in low frequencies are placed between square brackets.

#### Secondary contact between northern/eastern and southern/western lineages (Paper IV)

Both plastid and nuclear markers conclusively show that the northern/eastern and southern/western lineages of ssp. *maculata* meet in central Scandinavia (Fig. 2). The main route of immigration for the northern/eastern lineage is via northern Finland, but it can also be assumed that some immigration has taken place via the Åland Archipelago in the Baltic Sea. A second contact zone involving the same two lineages seems to occur in the eastern European lowland, between Poland and Lake Ladoga. For Scandinavia, contrasting immigration routes from the northeast and the south has previously been suggested for e.g. brown bear (*Ursus arctos*; Taberlet and Bouvet, 1994), field vole (*Microtus agrestis*; Jaarola and Tegelström, 1995), common shrew (*Sorex araneus*; Fredga, 1996) and some vascular plants (*Festuca ovina*, Bengtsson et al., 1995; *Viola rupestris*, Nordal and Jonsell, 1998; *Cerastium alpinum*; Nyberg-Berglund and Westerbergh (2001); *Silene dioica*, Malm and Prentice, 2005; *Arabidopsis thaliana*, Jakobsson et al., 2007).

We found that the contact zone between the northern/eastern and southern/western lineages of ssp. *maculata* has an extensive distribution in central Scandinavia. The centre is located to the provinces of Hälsingland, Medelpad and Ångermanland along the Bothnian Sea. Jaarola and Tegelström (1995) and Fredga (1996) localized hybrid zones for field vole and common shrew to the same Bothnian region. Nyberg-Berglund and Westerbergh (2001) also suggested that northeastern and southern lineages of *Cerastium alpinum* form a contact zone in this area. Such a pattern of coinciding contact zones could be explained by the deglaciation history of the Weichselian ice sheet. The centre of the ice sheet during the LGM

was located to the Ångermanland area, and the deglaciation of southern Ångermanland took place only c. 9300 BP (c. 10 500 cal. yrs. BP) when southern and northeastern Fennoscandia was already ice free (Berglund, 2004). Many species of plants and animals may thus have accumulated in the bordering areas left by the retreating ice. When the ice had finally melted away, the Bothnian region may have become quickly colonized from both the north and the south, which should explain the coincidence of contact zones.

For plastid data, we observed that the genetic diversity is markedly higher in the contact zone in central Scandinavia than in adjacent areas to the north and the south. This is reflected by all measures of genetic diversity. The average gene diversity over loci, which considers divergence between haplotypes, reveals that populations in central Scandinavia, together with populations from the putative contact zone in the eastern European lowland, are more diverse than any other European populations of ssp. *maculata*.

#### *Plastid DNA recombination (Paper IV)*

Intermediate plastid haplotypes between northern/eastern Group I haplotypes and southern/western Group II haplotypes are conspicuously common in the contact zone in central Scandinavia (Fig. 2). A quarter of the individuals of ssp. *maculata* in central Scandinavia have intermediate haplotypes. We also observed a high frequency of intermediate haplotypes in the putative contact zone in the eastern European lowland. These remarkable results strongly suggest that recombination takes place in the plastid genome. So far, only a few cases of suspected plastid DNA recombination have been reported (viz. *Pinus contorta*, Marshall et al., 2001; *Cycas taitungensis*, Huang et al., 2001). Our study is the first to provide clear phylogeographic evidence. As a comparison, there is an increasing amount of evidence for mitochondrial genome recombination (e.g. Bergthorsson et al., 2003; Barr et al., 2005; Tsiaousis et al., 2005). Plastid DNA markers are standard tools for population genetic and phylogenetic analysis. It is obvious that recombination can be problematic for phylogenetic inference at the species level.

#### **Main conclusions and perspectives**

This thesis has resulted in several interesting and important findings:

- Gene flow between diploid and autotetraploid cytotypes of *Dactylorhiza maculata* s.l. is restricted. In hybrid zones between diploid ssp. *fuchsii* and tetraploid ssp. *maculata*, the differentiation between taxa is reinforced by niche separation at the microhabitat level.
- For *Dactylorhiza maculata* s.l., and many other temperate organisms, postglacial recolonization from source populations at intermediate latitudes in Central and Eastern Europe has played a much more important role than previously thought. In contrast, populations of *D. maculata* s.l. in traditional refugial areas in southernmost Europe have not contributed to northward migration.
- Hybrid zones between southern and northern/eastern immigrant lineages may be a common phenomenon in central Scandinavia, which should have profound consequences for the structuring of genetic diversity.
- The paradigm of the non-recombinant plastid genome is questioned.

**Table 1.** Chromosome counts and ploidy level determinations of *Dactylorhiza maculata* s.l. *N* is number of populations. Taxon refers to taxa listed in the Introduction.

Country, region(s)	<i>N</i>	Taxon	<i>2n</i>	Comments	Reference
Austria: All parts	10	<i>fuchsii</i>	40		Vöth and Greilhuber, 1980
Austria: Kärnten	1	<i>fuchsii</i>	60	Together with 2x and 4x.	Vöth and Greilhuber, 1980
Austria: All parts	53	<i>fuchsii</i>	80		Vöth and Greilhuber, 1980
Austria: Niederösterreich	1	<i>fuchsii</i>	40		Groll, 1965
Austria: Niederösterreich	1	<i>fuchsii</i>	40		Titz, 1965
Austria: Niederösterreich	1	<i>fuchsii</i>	40		Vöth, 1978
Austria: Niederösterreich	1	<i>fuchsii</i>	60	Together with 2x and 4x.	Groll, 1965
Austria: Niederösterreich	15	<i>fuchsii</i>	80		Groll, 1965
Austria: Niederösterreich	1	<i>fuchsii</i>	80		Titz, 1965
Austria: Niederösterreich	1	<i>fuchsii</i>	80		Vöth, 1978
Austria: Tirol	n.d.	<i>fuchsii</i>	80		Vermeulen, 1968
Belarus: Minsk Area	5	<i>fuchsii</i>	40		Semerenko, 1989
Belarus: Minsk Area	2	<i>maculata</i>	80		Semerenko, 1989
Belgium: All parts	5	<i>maculata</i>	80		Gathoye and Tyteca, 1989
Belgium: Luxembourg	1	<i>fuchsii</i> × <i>maculata</i>	60	Together with 2x and 4x.	Gathoye and Tyteca, 1989
Belgium: Wallonia	6	<i>fuchsii</i>	40		Gathoye and Tyteca, 1989
Czech Republik: Sudeten Mts.	1	<i>fuchsii</i>	80		Krahulcová, 2003
Denmark: Jylland	n.d.	<i>maculata</i>	80		Hagerup, 1944
Denmark: Læsø	n.d.	<i>maculata</i>	80		Holmén and Kaad, 1956
Denmark: Møn	n.d.	<i>fuchsii</i>	40		Hagerup, 1944
England (?)	n.d.	<i>maculata</i>	79		M. M. Richardson unpubl. in Maude, 1939
England (?)	≥ 2	<i>maculata</i>	80		Lord and Richards, 1977
England: Durham	1	<i>fuchsii</i>	40		Heslop-Harrison, 1951
England: Hampshire	1	<i>maculata</i>	80		Heslop-Harrison, 1951
England: Hampshire, Durham	2	<i>fuchsii</i>	40		Heslop-Harrison, 1953
England: Hampshire, Kent	2	<i>fuchsii</i>	40		Montgomery et al., 1997
England: Yorkshire	1	<i>fuchsii</i>	40		Jagielo et al., 1989
Estonia: Saaremaa	2	<i>maculata</i>	80		Jagielo et al., 1989
Estonia: Saaremaa, W Mainland	4	<i>fuchsii</i>	40		Hagerup, 1944
Faeroe Islands	n.d.	<i>maculata</i>	80		Heslop-Harrison, 1951
Finland: Lappland	n.d.	<i>fuchsii</i>	40		

Table 1. Continued.

Country, region(s)	N	Taxon	2n	Comments	Reference
Finland: Uusimaa	1	<i>maculata</i>	80		Sorsa, 1962
France: Ardennes, Gard	2	<i>maculata</i>	80		Gathoye and Tyteca, 1989
France: Corsica	3	<i>saccifera</i>	40		Gathoye and Tyteca, 1989
France: Corsica	n.d.	<i>saccifera</i>	80		Vermeulen, 1947
France: E Pyrenees	1	<i>fuchsii</i>	40		Cauwet-Marc and Balayer, 1984
France: E Pyrenees	1	<i>fuchsii</i>	80		Cauwet-Marc and Balayer, 1984
France: E Pyrenees	3	<i>maculata</i>	80		Cauwet-Marc and Balayer, 1984
France: E Pyrenees	1	<i>saccifera</i>	40		Cauwet-Marc and Balayer, 1984
France: E Pyrenees	1	<i>fuchsii</i>	60	Together with 2x.	Gathoye and Tyteca, 1989
France: Hautes-Alpes	3	<i>fuchsii</i>	40		Gathoye and Tyteca, 1989
France: Indre, Averyon, Hautes-Alpes	n.d.	<i>fuchsii</i>	40		Van Loon and de Jong, 1978
France: Pas-de-Calais	2	<i>fuchsii</i>	40		Vermeulen, 1947
France: Poitou-Charentes	3	<i>maculata</i>	40 <sup>1</sup>		D. Ståhlberg, unpublished data
Georgia: E Tbilisi	16	<i>fuchsii</i>	80		Scharfenberg, 1977
Germany: Brandenburg	1	<i>maculata</i>	80		Wegener, 1966
Germany: Mecklenburg-Vorpommern	2	<i>fuchsii</i>	40		Wegener, 1966
Germany: Mecklenburg-Vorpommern, Sachsen-Anhalt	1	<i>fuchsii</i>	80		Wegener, 1966
Germany: Sachsen-Anhalt	≥ 3	<i>fuchsii</i>	80		Reinhardt, 1988
Germany: Thüringen	1	<i>saccifera</i>	42		Van Loon and Oudemans, 1982
Greece: Thessalia	n.d.	<i>maculata</i>	80		Hagerup, 1944
Iceland	≥ 2	<i>maculata</i>	80		Löve and Löve, 1956
Iceland: SW	n.d.	<i>fuchsii</i>	40		Löve and Löve, 1956
Ireland	n.d.	<i>fuchsii</i>	40		Vermeulen, 1947
Ireland: Clare	1	<i>fuchsii</i>	40		Heslop-Harrison, 1951
Ireland: Donegal	1	<i>maculata</i>	80		Heslop-Harrison, 1951
Ireland: Mayo, Wexford	2	<i>fuchsii</i>	40		Heslop-Harrison, 1953
Italy: Foggia, Salerno, Sicily	3	<i>saccifera</i>	40		D'Emerico et al., 2002
Italy: Roma, Umbria/Marche	2	<i>maculata</i>	40		Del Prete et al., 1980
Italy: Sicily	4	<i>saccifera</i>	40		Mazzolla et al., 1981
Italy: Toscana	1	<i>fuchsii</i>	40		Vöth and Grellhuber, 1980
Italy: Toscana	1	<i>fuchsii</i>	60	Together with 2x.	Vöth and Grellhuber, 1980

Table 1. Continued.

Country, region(s)	N	Taxon	2n	Comments	Reference
Italy: Toscana	6	<i>fuchsii</i>	80		Vöth and Greilhuber, 1980
Italy: Toscana	3	<i>maculata</i>	80		Bertolini et al., 2000
Italy: Toscana	n.d.	<i>saccifera</i>	80		Vermeulen, 1947
Italy: Toscana, Campania	2	<i>saccifera</i>	40		Gathoye and Tyteca, 1989
Italy: Toscana, Emilia-Romagna	ca 5	<i>fuchsii</i>	40		Bertolini et al., 2000
Italy: Toscana, Emilia-Romagna	ca 10	<i>fuchsii</i>	80		Bertolini et al., 2000
Italy: Toscana/Emilia-Romagna	1	<i>maculata</i>	40		Bertolini et al., 2000
Netherlands	n.d.	<i>fuchsii</i>	40		Kliphuis, 1963
Netherlands	n.d.	<i>maculata</i>	80		Kliphuis, 1963
Netherlands (?)	n.d.	<i>fuchsii</i>	40	Cult.	Vermeulen, 1938
Netherlands: Limburg	n.d.	<i>fuchsii</i>	40		Vermeulen, 1947
Norway: Akershus, Nordland	2	<i>fuchsii</i>	40		Knaben and Engelskjön, 1967
Norway: Finmark	1	<i>fuchsii</i>	40 <sup>1</sup>		D. Ståhlberg and M. Hedrén, in prep.
Norway: Sør-Trøndelag	1	<i>fuchsii</i>	40 <sup>1</sup>		Aagaard et al. 2005
Norway: Sør-Trøndelag	1	<i>maculata</i>	80 <sup>1</sup>		D. Ståhlberg and M. Hedrén, in prep.
Norway: Troms, Finnmark	2	<i>maculata</i>	80 <sup>1</sup>		Jagiello and Lankosz-Mróz, 1986-1987
Poland: All parts	10	<i>fuchsii</i>	40		Jagiello and Lankosz-Mróz, 1986-1987
Poland: All parts	11	<i>maculata</i>	80		Jagiello and Lankosz-Mróz, 1986-1987
Poland: Central Area, Sudeten Mts.	13	<i>fuchsii</i>	80		Jagiello and Lankosz-Mróz, 1986-1987
Poland: NE	1	<i>fuchsii</i>	60	Together with 2x.	Jagiello and Lankosz-Mróz, 1986-1987
Poland: Tatra Mts -W Carpathians	4	<i>fuchsii</i>	40		Pegan and Wcislo, 1957
Portugal: Madeira	n.d.	<i>foliosa</i>	40	Cult.	Vermeulen, 1938
Portugal: Madeira	n.d.	<i>foliosa</i>	40		Vermeulen, 1947
Romania: Transylvania	1	<i>saccifera</i>	80 <sup>1</sup>		D. Ståhlberg, unpublished data <sup>1</sup>
Russia: Baikal Area	2	<i>fuchsii</i>	40		Belaeva and Siplivinsky, 1975
Russia: Komi	8	<i>fuchsii</i>	40		Averyanov et al., 1982b
Russia: Komi	2	<i>fuchsii</i>	41	Together with 2x.	Averyanov et al., 1982b
Russia: Komi	2	<i>fuchsii</i> (?)	60	Together with 2x.	Averyanov et al., 1982b
Russia: Krasnoyarsk Area	3	<i>maculata</i>	80		Stepanov, 1994
Russia: Krasnoyarsk Area	3	<i>fuchsii</i>	20		Stepanov, 1994
	2	<i>fuchsii</i>	40		Stepanov, 1994

Table 1. Continued.

Country, region(s)	N	Taxon	2n	Comments	Reference
Russia: N Ural Mts.	5	<i>fuchsii</i>	40		Averyanov et al., 1982a
Russia: N Ural Mts.	1	<i>fuchsii</i>	60	Together with 2x.	Averyanov et al., 1982a
Russia: N Ural Mts.	1	<i>maculata</i>	78	Together with 4x.	Averyanov et al., 1982a
Russia: N Ural Mts.	3	<i>maculata</i>	80		Averyanov et al., 1982a
Russia: Novgorod Area	2	<i>maculata</i> (?)	60	Together with 4x.	Averyanov, 1979
Russia: St. Petersburg Area	1	<i>maculata</i> (?)	100	Together with 4x.	Averyanov, 1979
Russia: St. Petersburg Area, Novgorod Area, Karelia	8	<i>maculata</i>	80		Averyanov, 1979
Russia: St. Petersburg Area, Novgorod Area	3	<i>fuchsii</i>	40		Averyanov, 1979
Russia: St. Petersburg Area, Vologda Area	5	<i>fuchsii</i>	40		Averyanov, 1977
Russia: St. Petersburg Area, Vologda Area	11	<i>maculata</i>	80		Averyanov, 1977
Russia: Tomsk Area	1	<i>fuchsii</i>	40		Kartashova et al., 1974
Russia: Tyman	6	<i>fuchsii</i>	40		Averyanov et al., 1980
Russia: Vologda Area	3	<i>maculata</i>	80		Averyanov et al., 1980
Scotland: Inner Hebrides	1	<i>fuchsii</i> × <i>maculata</i>	60	Together with 2x and 4x.	Averyanov, 1977
Scotland: Outer Hebrides	n.d.	<i>maculata</i>	40		Heslop-Harrison, 1948
Slovakia: Bratislava Area	1	<i>maculata</i>	80		Heslop-Harrison, 1951
Slovakia: Tatra Mts.	1	<i>fuchsii</i>	40		Uhriková, 1976
Slovenia: Bevke	1	<i>fuchsii</i>	40		Uhriková, 1978
Spain: Ávila	1	<i>maculata</i>	80		Lovka et al., 1971
Spain: Castilla-La Mancha	1	<i>maculata</i>	40		Bernardos et al., 2004
Sweden: All parts	7	<i>fuchsii</i>	40 <sup>1</sup>		Fernández Casas et al., 1979
Sweden: Jämtland, Småland, Åsele lappmark	5	<i>maculata</i>	80		D. Ståhlberg and M. Hedrén, in prep.
Sweden: Skåne	1	<i>maculata</i>	120 <sup>1</sup>	Together with 4x.	Heslop-Harrison, 1951
Sweden: Skåne, Uppland	4	<i>maculata</i>	80 <sup>1</sup>		D. Ståhlberg and M. Hedrén, in prep.
Sweden: Södermanland, Häradedalen	2	<i>fuchsii</i> × <i>maculata</i>	60 <sup>1</sup>	Together with 2x and 4x.	D. Ståhlberg, in prep.
Sweden: Torne lappmark	1	<i>fuchsii</i>	40		Löve and Löve, 1944
Sweden: Uppland	1	<i>fuchsii</i>	40		Afzelius, 1958
Sweden: Östergötland, Jämtland, Torne lappmark	3	<i>fuchsii</i>	40		Heslop-Harrison, 1951
Switzerland: Neuchâtel	1	<i>fuchsii</i>	60		Vaucher, 1966
Switzerland: Neuchâtel, Bern	13	<i>fuchsii</i>	80		Vaucher, 1966

Table 1. Continued.

Country, region(s)	N	Taxon	2n	Comments	Reference
Switzerland: Vaud, Bern	2	<i>fuchsii</i>	40		Vaucher, 1966
Switzerland: Zürich	1	<i>fuchsii</i>	40		Heusser, 1938
Switzerland: Zürich, Graubünden, Tessin	7	<i>fuchsii</i>	80		Heusser, 1938

<sup>1</sup>Assessed by flow cytometry.

## References

Aagaard S. M. D., Såstad S. M., Greilhuber J., Moen A. (2005) A secondary hybrid zone between diploid *Dactylorhiza incarnata* ssp. *cruenta* and allotetraploid *D. laponica* (Orchidaceae). *Heredity* 94: 488-496.

Afzelius K. (1958) En egendomlig form av *Orchis maculata* L. sens. lat. *Svensk Bot. Tidskr.* 52: 18-22.

Alsos I. G., Engelskjøn T., Gielly L., Taberlet P., Brochmann C. (2005) Impact of ice ages on circumpolar molecular diversity: insights from an ecological key species. *Mol. Ecol.* 14: 2739-2753.

Anderson E. (1948) Hybridization of the habitat. *Evolution* 2: 1-9.

Arnold M. L. (1997) Natural hybridization and evolution. Oxford University Press, Oxford.

Averyanov L. V. (1977) Chromosome numbers of some species of the Orchidaceae family in the Leningrad and Vologda districts. *Bot. Zhur.* 62: 547-553 (in Russian).

Averyanov L. V. (1979) Chromosome numbers of some species of the Orchidaceae family from the northwestern part of the USSR. *Bot. Zhur.* 64: 863-877 (in Russian).

Averyanov L. V. (1990) A review of the genus *Dactylorhiza*. In: Arditti J. (ed.), *Orchid biology. Reviews and perspectives*. V. Timber Press, Portland (Oregon), pp. 159-206.

Averyanov L. V., Averyanova E. L., Lavrenko A. N. (1980) Caryosystematic characterization of the genus *Dactylorhiza* (Orchidaceae) in the Middle Tyman. *Bot. Zhur.* 65: 983-989 (in Russian).

Averyanov L. V., Averyanova E. L., Lavrenko A. N. (1982a) Caryosystematic study of orchids (Orchidaceae) of the Pechoro-Ilyshsky reservation. *Bot. Zh.* 67: 945-951 (in Russian).

Averyanov L. V., Averyanova E. L., Lavrenko A. N. (1982b) Caryosystematic study of orchids (Orchidaceae) on the territory of the Komi ASSR. *Bot. Zh.* 67: 1491-1499 (in Russian).

Avise J. C. (2000) *Phylogeography: The history and formation of species*. Harvard Univ. Press, Cambridge, Massachusetts.

Barr C. M., Neiman M., Taylor D. R. (2005) Inheritance and recombination of mitochondrial genomes in plants, fungi and animals. *New Phytol.* 168: 39-50.

Bateman R. M., Denholm I. (1985) A reappraisal of the British and Irish dactylochids, 2. The diploid marsh-orchids. *Watsonia* 15: 321-355.

Bateman R. M., Denholm I. (1989) A reappraisal of the British and Irish dactylochids, 3. The spotted-orchids. *Watsonia* 17: 319-349.

Bateman R. M., Denholm I. (2003) The Heath Spotted-orchid (*Dactylorhiza maculata* (L.) Soó) in the British Isles: a cautionary case-study in delimitating infraspecific taxa and inferring their evolutionary relationships.

Bateman R. M., Hollingsworth P. M., Preston J., Yi-Bo L., Pridgeon A. M., Chase M. W. (2003) Molecular phylogenetics and evolution of Orchidinae and selected Habenariinae (Orchidaceae). *Bot. J. Linn. Soc.* 142: 1-40.

Belaeva V. A., Siplivinsky V. N. (1975) Chromosome numbers and taxonomy of some species of the Baikal flora. *Bot. Zhur.* 60: 864-872 (in Russian).

Bengtsson B. O., Weibull P., Ghatnekar L. (1995) The loss of alleles by sampling: A study of the common outbreeding grass *Festuca ovina* over three geographic scales. *Hereditas* 122: 221-238.

Berglund M. (2004) Holocene shore displacement and chronology in Ångermanland, eastern Sweden, the Scandinavian glacio-isostatic uplift centre. *Boreas* 33: 48-60.

Bergthorsson U., Adams K. L., Thomason B., Palmer J. D. (2003) Widespread horizontal transfer of mitochondrial genes in flowering plants. *Nature* 424: 197-201.

Bernardos S., Tyteca D., Amich F. (2004) Cytotaxonomic study of some taxa of the subtribe Orchidinae (Orchidoideae, Orchidaceae) from the Iberian Peninsula. *Isr. J. Plant Sci.* 52: 161-170.

Bertolini V., Del Prete C., Garbari F. (2000) Karyological and biometrical studies on some species of the genus *Dactylorhiza* Necker ex Nevski, sect. *Dactylorhiza* (Orchidaceae) of Central-Northern Italy. *Portugaliae Acta Biol.* 19: 249-265.

Bullini L., Cianchi R., Arduino P., De Bonis L., Mosco M. C., Verardi A., Porretta D., Corrias B., Rossi W. (2001) Molecular evidence for allopolyploid speciation and a single origin of the western Mediterranean orchid *Dactylorhiza insularis* (Orchidaceae). *Bot. J. Linn. Soc.* 72: 193-201.

Cauwet-Marc A. M., Balayer M. (1984) Les genres *Orchis* L., *Dactylorhiza* Necker ex Newski, *Neotinea* Reichenb. et *Traunsteinera* Reichenb. : Caryologie et proposition de phylogénie et d'évolution. *Bot. Helv.* 94 : 391-406.

Delforge P. (1995) *Orchids of Britain and Europe*. Harper Collins Publishers, London.

Del Prete C., Gabari F., Giordani A. (1980) Numeri cromosomici per la Flora Italiana. *Inform. Bot. Ital.* 12 : 117-120.

D'Emerico S., Cozzolino S., Pellegrino G., Pignone D., Scrugli A. (2002) Karyotype structure, supernumerary chromosomes and heterochromatin distribution suggest a pathway of karyotype evolution in *Dactylorhiza* (Orchidaceae). *Bot. J. Linn. Soc.* 138: 85-91.

Demesure B., Comps B., Petit R. J. (1996) Chloroplast DNA phylogeography of the common beech (*Fagus sylvatica*) in Europe. *Evolution* 50: 2525-2520.

Devos N., Oh S.-H., Raspé O., Jacquemart A.-L., Manos P. S. (2005) Nuclear ribosomal DNA sequence variation and evolution of spotted marsh-orchids (*Dactylorhiza maculata* group). *Mol. Phylogenetic Evol.* 36: 568-580.

Devos N., Raspé O., Jacquemart A.-L., Tyteca D., (2006b) On the monophyly of *Dactylorhiza* Necker ex Nevski (Orchidaceae): is *Coeloglossum viride* (L.) Hartman a *Dactylorhiza*? *Bot. J. Linn. Soc.* 152: 261-269.

Devos N., Raspé O., Oh S.-H., Tyteca D., Jacquemart A.-L. (2006a) The evolution of *Dactylorhiza* (Orchidaceae) allotetraploid complex: Insights from nrDNA sequences and cpDNA PCR-RFLP data. *Mol. Phylogenetic Evol.* 38: 767-778.

Devos N., Tyteca D., Raspé O., Wesselingh R. A., Jacquemart A.-L. (2003) Patterns of chloroplast diversity among western European *Dactylorhiza* species (Orchidaceae). *Pl. Syst. Evol.* 243: 85-97.

Du Rietz G. E. (1921) Zur methodologischen Grundlage der modernen Pflanzensoziologie. Holzhausen, Wien.

Dufrêne M., Gathoye J. L., Tyteca D. (1991) Biostatistical studies on western European *Dactylorhiza* (Orchidaceae): the *D. maculata* group. *Pl. Syst. Evol.* 175: 55-72.

Excoffier L., Laval G., Schneider S. (2005) Arlequin version. 3.0: An integrated software package for population genetics data analysis. *Evol. Bioinf. Online* 1: 47-50.

Fernández Casas J., Pons-Sorolla A., Susanna A. (1979) Numeros chromosomaticos de plantas occidentales. *Anal. Jard. Bot. Madrid* 36 : 401-405.

Ferris C., King R. A., Hewitt G. M. (1999) Isolation within species and the history of glacial refugia. In: Hollingsworth R. M., Bateman R. M., Gornall R. J. (eds.), *Molecular systematics and plant evolution*. Taylor & Francis, London, pp. 20-34.

Fredga K. (1996) The chromosome races of *Sorex araneus* in Scandinavia. *Hereditas*: 125: 123-135.

Gathoye J.-L., Tyteca D. (1989) Contribution à l'étude cytotaxonomique des *Dactylorhiza* d'Europe occidentale. *Mém. Soc. Roy. Bot. Belg.* 11: 30-42Groll, 1965

Grant V. (1981) Plant speciation. Columbia Univ. Press, New York.

Groll M. (1965) Fruchtansatz, Bestäubung und Merkmalsanalyse bei diploiden und polyploiden Sippen von *Dactylorhizis (Orchis) maculata* und *Gymnadenia conopsea*. *Österr. Bot. Z.* 112: 657-700.

Götz P., Reinhard H. R. (1997) Über die Gattung *Dactylorhiza* – neue Erkenntnisse und neue Fragen. *J. Eur. Orchid.* 29: 585-640.

Hagerup O. (1938) Studies on the significance of polypliody. II. *Orchis*. *Hereditas* 24: 258-264.

Hagerup O. (1944) On fertilisation, polypliody and haploidy in *Orchis maculatus* L. sens. lat. *Dansk Bot. Ark.* 11: 1-26.

Hedrén M. (1996) Genetic differentiation, polyploidization and hybridization in Northern European *Dactylorhiza* (Orchidaceae): Evidence from allozyme markers. *Pl. Syst. Evol.* 201: 31-55.

Hedrén M. (2001) Systematics of the *Dactylorhiza euxina/incipitata/maculata* polyploid complex (Orchidaceae) in Turkey: evidence from allozyme data. *Pl. Syst. Evol.* 229: 23-44.

Hedrén M. (2002) Speciation patterns in the *Dactylorhiza incarnata/maculata* polyploid complex (Orchidaceae): evidence from molecular markers. *Jour. Eur. Orch.* 34: 707-731.

Hedrén M. (2003) Plastid DNA variation in the *Dactylorhiza incarnata/maculata* polyploid complex and the origin of allotetraploid *D. sphagnicola* (Orchidaceae). *Mol. Ecol.* 12: 2669-2680.

Hedrén M., Fay M. F., Chase M. W. (2001) Amplified fragment length polymorphisms (AFLP) reveal details of polyploid evolution in *Dactylorhiza* (Orchidaceae). *Am. J. Bot.* 88: 1868-1880.

Hedrén M., Nordström S., Persson Hovmalm H. A., Pedersen H. A., Hansson S. (2007) Patterns of polyploid evolution in Greek Marsh Orchids (*Dactylorhiza* Orchidaceae) as revealed by allozymes, AFLPs and plastid DNA data. *Am. J. Bot.*, in press.

Heslop-Harrison J. (1948) Field studies in *Orchis* L., I. The structure of dactylorhizid populations on certain islands in the Inner and Outer Hebrides. *Trans. Bot. Soc. Edinb.* 35: 26-66.

Heslop-Harrison J. (1951) A comparison of some Swedish and British forms of *Orchis maculata* L. sens. lat. *Svensk Bot. Tidskr.* 45: 608-635.

Heslop-Harrison J. (1953) Microsporogenesis in some triploid *Dactylorhiza* hybrids. *Ann. Bot.* 17: 539-549.

Heslop-Harrison J. (1957) On the hybridization of the common spotted orchid *Dactylorhizis fuchsii* (Druce) Vermln., with the marsh orchid *D. praetermissa* (Druce) Vermln., and *D. purpurella* (T. and T. A. Steph.) Vermln. *Proc. Linn. Soc. London* 167: 176-185.

Heusser C. (1938) Chromosomenverhältnisse bei schweizerischen bastionen Orchideen. *Ber. Schweiz. Ges.* 48: 562-605.

Hewitt G. M. (1988) Hybrid zones – natural laboratories for evolutionary studies. *Trends Ecol. Evol.* 3: 158-167.

Hewitt G. M. (1996) Some genetic consequences of ice ages, and their role in divergence and speciation. *Bot. J. Linn. Soc.* 58: 247-276.

Hewitt G. M. (2000) The genetic legacy of the Quaternary ice ages. *Nature* 405: 907-913.

Hewitt G. M. (2001) Speciation, hybrid zones and phylogeography – or seeing genes in space and time. *Mol. Ecol.* 10: 537-549.

Hewitt G. M. (2004) Genetic consequences of climatic oscillations in the Quaternary. *Philos. Trans. R Soc. Lond. Biol. Sci.* 359: 183-195.

Holmen K., Kaad P. (1956) Über *Dactylorhiza traunsteineri* auf der Insel Läsö. *Bot. Tidsskr.* 53: 35-48.

Huang S., Chiang Y. C., Schaal B. A., Chou C. H., Chiang T. Y. Organelle DNA phylogeography of *Cycas taitungensis*, a relict species in Taiwan. *Mol. Ecol.* 10: 2669-2681.

Huntley B., Birks H. J. B. (1983) An Atlas of past and present pollen maps for Europe: 0-13 000 years ago. Cambridge Univ. Press, Cambridge.

Jaarola M., Tegelström H. (1995) Colonization history of north European field voles (*Migrotus agrestis*) revealed by mitochondrial DNA. *Mol Ecol.* 4: 299-310.

Jagiełło M. (1986-1987) Analysis of population variability and distribution of species from the *Dactylorhiza maculata* group (Orchidaceae) in Poland. *Fragm. Florist. Geobot.* 31-32: 333-383.

Jagiełło M., Kuusk V., Lankosz-Mróz M. (1989) Karyological investigations on orchids of the Estonian SSR. Part I. Genus *Dactylorhiza* Necker ex Nevski (Orchidaceae). *Fragm. Florist. Geobot.* 34: 315-326.

Jagiełło M., Lankosz-Mróz M. (1986-1987) Cytotaxonomic studies in the *Dactylorhiza maculata* (L.) Soó group in Poland (Orchidaceae). *Fragm. Florist. Geobot.* 31-32: 385-394.

Jakobsson M., Säll T., Lind-Halldén C., Halldén C. (2007) The evolutionary history of the common chloroplast genome of *Arabidopsis thaliana* and *A. suecica*. *J. Evol. Biol.* 20: 104-121.

Janečková P., Wotavová K., Jersáková J., Kindlmann P. (2006) Relative effects of management and environmental conditions on performance and survival of populations of a terrestrial orchid, *Dactylorhiza majalis*. *Biol. Conserv.* 129: 40-49.

Kartashova N. N., Malakhova L. A., Kozlova A. A. (1974) Study of chromosomes of representatives of the Ob Region flora. I. Number of chromosomes of the Tomsk District. *Nauch. Dokl. Vyssh. Skh. Biol. Nauk.* 4: 114-119. *Nauchn. Dokl. Vyssh. Skh. Biol. Nauki* 4: 114-119 (in Russian).

Klein E., Kerschbaumsteiner H. (1996) Die Orchideen der Steiermark. Steiermärkisches Landesmuseum Joanneum, Graz.

Klinge J. (1898) Dactylorhids, orchids subgeneris, monographiae prodromus. *Acta Horti Petropol.* 17: 145-201.

Kliphuis E. (1963) Cytological observations in relation to the taxonomy of the orchids of the Netherlands. *Acta Bot. Neerland.* 12: 172-194.

Knaben G., Engelskjön T. (1967) Chromosome numbers of Scandinavian arctic-alpine plant species II. *Acta Borealia Scientia* 21: 9-10, 52-53.

Krahulcová A. (2003) Chromosome numbers in selected monocotyledons (Czech Republic, Hungary, and Slovakia). *Preslia* 75: 97-113.

Levin D. A. (1975) Minority cytotype exclusion in local plant populations. *Taxon* 24: 35-43.

Litynska-Zajac M. (1995) Anthracological analysis. In: Hromada J., Kozłowski J. (eds.), Complex of Upper Palaeolithic Sites Near Moravany, Western Slovakia. Jagiellonian Univ. Press, Krakow, pp. 74-79.

Lord R. M., Richards A. J. (1977) A hybrid swarm between the diploid *Dactylorhiza fuchsii* (Druce) Soó and the tetraploid *D. purpurella* (T. & T. A. Steph.) Soó in Durham. *Watsonia* 11: 205-210.

Lovka M., Sušnik F., Löve Á., Löve D. (1971) In: Löve Á. (ed.), IOPM Chromosome number reports XXXIV. *Taxon* 20: 785-797.

Lynch M., Conery J. S. (2000) The evolutionary fate of duplicated genes. *Science* 290: 1151-1154.

Löve Á., Löve D. (1944) Cyto-taxonomical studies on boreal plants. III. Some new chromosome numbers of Scandinavian plants. *Arkiv för Botanik* 31: 16.

Löve Á., Löve D. (1956) Conspectus of the Icelandic flora. *Acta Hort. Gothob.* 20: 65-291.

Löve Á., Löve D. (1974) Origin and the evolution of the arctic and alpine floras. In: Ives J. D., Barry R. G. (eds.), Arctic and alpine environments (eds.). Methuen, London, pp 571-603.

Magri D., Vendramin G.G., Comps B., Dupanloup I., Geburek T., Gömöry D., Lata'owa M., Litt T., Paule L., Roure J.M., Tantau I., van der Knaap W.O., Petit R.M., de Beaulieu J.-L. (2006). A new scenario for the Quaternary history of European beech populations: palaeobotanical evidence and genetic consequences. *New Phytol.* 171: 199–221.

Malm J. U., Prentice H. C. (2005) Chloroplast DNA haplotypes in Nordic *Silene dioica*: postglacial immigration from the east and the south. *Plant Syst. Evol.* 250: 27-38.

Marshall H. D. C. Newton, Ritland K. (2001) Sequence-repeat polymorphisms exhibit the signature of recombination in lodgepole pine chloroplast DNA. *Mol. Biol. Evol.* 18: 2136-2138.

Maude P. F. (1939) The Merton Catalogue. A list of chromosome numbers of species of British flowering plants. *New. Phytol.* 38: 1-31.

Mazolla P., Lidberg R., Raimondo F. M. (1981) Critical notes on the Sicilian flora: the genus *Dactylorhiza* Necker ex Nevski sect. *Dactylorhiza*. *Anales Jard. Bot. Madrid* 37: 661-676.

Müntzing A. (1936) The evolutionary significance of autoploidy. *Hereditas* 21: 263-378.

Montgomery L., Khalaf M., Bailey J. P., Gornall R. J. (1997) Contributions to a cytological catalogue of the British and Irish flora, 5. *Watsonia* 21: 365-368.

Nelson E. (1976) Monographie und Ikonographie der Orchidaceen-Gattung *Dactylorhiza*. Speich, Zürich.

Nordal I., Jonsell B. (1998) A phylogeographic analysis of *Viola rupestris* : three post-glacial immigration routes into the Nordic area? *Bot. J. Linn. Soc.* 128: 105-122.

Nyberg Berglund A.-B., Westerbergh A. (2001) Two postglacial lineages of the polyploid *Cerastium alpinum* (Caryophyllaceae). *Hereditas* 134: 171-183.

Otto S. P., Whitton J. (2000) Polyploid incidence and evolution. *Annu. Rev. Genet.* 34, 401-437.

Palmé A. E., Su Q., Rautenberg A., Manni F., Lascoux M. (2003) Postglacial recolonization and cpDNA variation of silver birch, *Betula pendula*. *Mol. Ecol.* 12 : 201-212.

Pedersen H. ÅE. (1998). Species concept and guidelines for infraspecific taxonomic ranking in *Dactylorhiza* (Orchidaceae). *Nord. J. Bot.* 18: 289-311.

Pedersen H. ÅE. (2004) *Dactylorhiza majalis* s.l. (Orchidaceae) in acid habitats: variation patterns, taxonomy and evolution. *Nord. J. Bot.* 22: 641-658.

Pedersen H. ÅE. (2006) Systematics and evolution of the *Dactylorhiza romana/sambucina* polyploid complex (Orchidaceae). *Bot. J. Linn. Soc.* 152: 405-434.

Petit R. J., Kremer A., Wagner D. B. (1993) Geographical structure of chloroplast DNA polymorphism in european oaks. *Theor. Appl. Genet.* 87: 122-128.

Petit R. J., Aguinagalde I., de Beaulieu J.-L., Bittkau C., Brewer S., Cheddadi R., Ennos R., Fineschi S., Grivet D., Lascoux M., Mohanty A., Müller-Starck G., Demesure-Musch B., Palmé A., Martín J. P., Rendell S., Vendramin G. G. (2003) Glacial refugia: hotspots but not melting pots of genetic diversity. *Science* 300: 1563-1565.

Pillon Y., Fay M. F., Hedrén M., Devey D., Shipunov A., van der Bank M., Bateman R. M., Chase M. W. (2007) Insights into the evolution and biogeography of Western European species complexes in *Dactylorhiza* (Orchidaceae). *Taxon*, in press.

Pillon Y., Fay M. F., Shipunov A. B., Chase M. W. (2006) Species diversity versus phylogenetic diversity: a practical study in the taxonomically difficult genus *Dactylorhiza* (Orchidaceae). *Biol. Conserv.* 129: 4-13.

Pogan E., Wcisło, H. (1957) In: Skalińska M., et al. , Further studies in chromosome numbers of Polish angiosperms. *Acta Soc. Bot. Pol.* 26: 231-245.

Pridgeon A. M., Bateman R. M., Cox A. V., Hapeman J. R., Chase M. W. (1997) Phylogenetics of subtribe orchidinae (Orchidoideae, Orchidaceae) based on nuclear ITS sequences. 1. Intergeneric relationships and polyphyly of *Orchis* *sensu lato*. *Lindleyana* 12: 89-109.

Ramsey J., Schemske D. W. (1998) Pathways, mechanisms, and rates of polyploid formation in flowering plants. *Ann. Rev. Ecol. Syst.* 29: 467-501.

Reinhard H. R. (1985) Skandinavische und alpine *Dactylorhiza*-arten (Orchidaceae). Ergebnisse populationsstatistischer Untersuchungen. *Mitt. Bl. Arbeitskr. Heim. Orch. Baden-Württ.* 17: 321-416.

Reinhardt J. (1988) Zur Zytotaxonomie einiger *Dactylorhiza fuchsii* (Druce) Soó-Sippen im Eichsfeld. *Mitt. Bl. Arbeitskr. Heim. Orch. Baden-Württ.* 17: 14-18.

Rendell S., Ennos R. A. (2002) Chloroplast DNA diversity in *Calluna vulgaris* (heather) populations in Europe. *Mol. Ecol.* 11: 69-78.

Scharfenberg K. (1977) Beiträge zur Kenntnis der Sippenstruktur der Gattung *Dactylorhiza* Necker ex Nevski in den Bezirken Cottbus, Potsdam, Frankfurt (Oder) und Neubrandenburg. *Gleditschia* 5: 65-127.

Semerenko L. V. (1989) Chromosome numbers in some members of the families Asteraceae, Fabaceae, Orchidaceae and Poaceae from the Berezinsky biosphere reservation (Byelorussia) flora. *Bot. Zhur.* 74: 1671-1673 (in Russian).

Senghas K. (1968) Taxonomische Übersicht der Gattung *Dactylorhiza* Necker ex Nevski. In: Senghas K., Sundermann H (eds.), Probleme der Orchideengattung *Dactylorhiza*. *Jahresber. Naturwiss. Vereins Wuppertal* 21-22: 32-67.

Shipunov A. B., Fay M. F., Pillon Y., Bateman R. M., Chase M. W. (2004) *Dactylorhiza* (Orchidaceae) in European Russia: combined molecular and morphological analysis. *Am. J. Bot.* 91: 1419-1426.

Shipunov A. B., Fay M. F., Chase M. W. (2005) Evolution of *Dactylorhiza baltica* (Orchidaceae) in European Russia: evidence from molecular markers and morphology.

Skrede I., Bronken Eidesen P., Piñeiro Portela R., Brochmann C. (2006) Refugia, differentiation and postglacial migration in arctic-alpine Eurasia, exemplified by the mountain avens (*Dryas octopetala* L.)

Soltis D. E., Soltis P. S. (1993) Molecular data and the dynamic nature of polyploidy. *Crit. Rev. Plant Sci.* 12: 243-273.

Soltis D. E., Soltis P. S. (1999) Polyploidy: Recurrent formation and genome evolution. *Trends Ecol. Evol.* 14: 348-352.

Soltis D. E., Soltis P. S., Tate J. A. (2003) Advances in the study of polyploidy since *Plant speciation*. New Phytol. 161:173-191.

Sommer R. S., Nadachowski A. (2006) Glacial refugia of mammals in Europe: evidence from fossil records Mammal Rev. 36: 251–265.

Sóó R. (1960) Synopsis generis *Dactylorhiza* (*Dactylorhizis*). Ann. Univ. Sci. Budapest Biol. 3: 335-357.

Sorsa V. (1962) Chromosomenzahlen finnischer Kormophyten I. Ann. Acad. Scient. Fennicæ 58: 1-14.

Stebbins G. L. (1950) Variation and evolution in plants. Columbia Univ. Press

Stebbins G. L. (1971) The morphological, physiological, and cytogenetic significance of polyploidy. In: Barrington E. J., Willis A. J. (eds.), Chromosomal evolution in higher plants. Arnold E., London, pp. 124-154.

Stebbins G. L. (1984) Polyploidy and the distribution of the arctic-alpine flora: new evidence and a new approach. Bot. Helv. 94: 1-13.

Stepanov N. V. (1994) Chromosome numbers in some nemoral species of the West Sayan (Krasnoyarsk region). Bot. Zhur. 79: 125-128 (in Russian).

Stewart J. R., Lister A. M. (2001) Cryptic northern refugia and the origins of the modern biota. Trends Ecol. Evol. 16: 608-613.

Taberlet P., Bouvet J. (1994). Mitochondrial DNA polymorphism, phylogeography, and conservation genetics of the brown bear *Ursus arctos* in Europe. Proc. R. Soc. London 255: 195-200.

Titz W. (1965) Vergleichende Untersuchungen über den Grad der somatischen Polyploidie an nahe verwandten diploiden und polyploiden Sippen einschließlich der Cytologie von Antipoden. Österr. Bot. Z. 112: 101-173.

Tsaousis A. D., Martin D. P., Ladoukakis E. D., Posada D., Zouros E. (2005) Widespread recombination in published animal mtDNA sequences. Mol. Biol. Evol. 22: 925-933.

Tyteca D., Gathoye J.-L. (2004) Morphometric analyses of the *Dactylorhiza maculata* (L.) Soó group in western Europe. Ber. Arbeitskr. Heim. Orchid. 21: 4-35.

Uhriková A. (1976) In: Májovský J., et al., Index of chromosome numbers of Slovakian flora (Part 5). Acta Fac. Rerum Nat. Univ. Comen., Bot. 25: 1-18.

Uhriková A. (1978) In: Májovský J., et al., Index of chromosome numbers of Slovakian flora (Part 6). Acta Fac. Rerum Nat. Univ. Comen., Bot. 26: 1-42.

Ursenbacher S., Carlsson M., Helfer V., Tegelström H., Fumagalli L. (2006) Phylogeography and Pleistocene refugia of the adder (*Vipera berus*) as inferred from mitochondrial DNA sequence data. Mol. Ecol. 15: 3425-3437.

Van Loon J. C., de Jong H. (1978) In: Löve Á (ed.) IOPB chromosome number reports LIX. Taxon 27: 53-61.

Van Loon J. C., Oudemans J. J. M. H. (1982) In: Á. Löve (ed.), IOPB chromosome number reports LXXV. Taxon 31: 342-368.

Vaucher C. (1966) Contribution a l'étude cytologique du genre *Dactylorhiza* (Klinge) Vermeulen. Bull. Soc. Neuchat. Sci. Nat. 89: 75-85.

Vermeulen P. (1938) Chromosomes in *Orchis*. Chron. Bot. 4: 107-108.

Vermeulen P. (1947) Studies on dactylorhids. Schotanus & Jens, Utrecht.

Vermeulen P. (1968) *Dactylorhiza maculata* und ihre Formen. Jahresber. Naturwiss. Vereins Wuppertal 21-22: 68-76.

Vuilleumier B. S. (1971) Pleistocene changes in the fauna and flora of South America. Science 173: 771-780.

Vöth W. (1978) Biometrische Untersuchungen an *Dactylorhiza maculata* s.l. – Sippen in Niederösterreich. Linzer Biol. Beitr. 10: 179-215.

Vöth W., Greilhuber J. (1980) Zur Karyosystematik von *Dactylorhiza maculata* s.l. und ihrer Verbreitung, insbesondere in Niederösterreich. Linzer Biol. Beitr. 12: 415-468.

Wegener K.-A. von (1966) Ein Beitrag zur Zytologie von Orchideen aus dem Gebiet der DDR. Wiss. Z. Univ. Greifswald (Math.-nat. R.) 15: 1-7.

Widmer A., Lexer C. (2001) Glacial refugia: sanctuaries for allelic richness, but not for gene diversity. Trends Ecol. Evol. 16: 267- 269.

Willis K. J., van Andel T. H. (2004). Trees or no trees? The environments of central and eastern Europe during the last glaciation.

## Acknowledgements

Financial support was given by Anna och Svante Murbecks minnesfond, CFO Nordstedts fond, Elly Olssons fond, Kungliga Fysiografiska Sällskapet, Lunds botaniska förening, Margit Stiernwärds fond för miljövärdfsrsksning and Per Westlings minnesfond.

Many people have contributed to this thesis and I am grateful to all of you.

I was introduced to these studies by Mikael Hedrén. I thank you for many years of supervision, friendly support and cooperation, and for help with numerous things.

The *Dactylorhiza* group was established when Sofie Nordström arrived. Thank you for cooperation in the field and the lab, and for a lot of encouragement throughout the years.

I would not have been able to finish this work without the help from Louise Hathaway. Thank you for valuable discussions about methods and results, and thank you for reading and commenting on the manuscripts. I am also grateful to you for correcting and improving the language of some of the manuscripts.

There is no doubt that I would have given up already after a few hours without the help and support from my sister Ingela Ståhlberg. Thank you for invaluable assistance in the field.

Kerstin Isaksson is the best room mate I could imagine. I miss you already.

Maarten Ellmer was an excellent companion during the field trip to Transylvania. I am particularly grateful to you for saving me from the brown bears.

Nils Cronberg provided a lot of help during the last intensive weeks. Thank you for reading and commenting on the ill-disposed last manuscript, and thank you for the encouragement.

Stefan Andersson has always shown an interest in my work. Thank you for always being open to questions.

Galina Garkavia has helped me with thousands of administrative problems. I am also grateful to you for translating Russian texts.

Lina Steinke and Frida Rosengren helped me in the lab. Thank you for an eminent job.

During the last months I suffered together with Eva Månsby, Pernilla Göransson and Frida Andreasson. I think that we helped each other. I am most grateful to you.

I cannot mention each and every person at the Section of Plant Ecology and Systematics. Thanks to all of you for support in various ways.

I am also grateful to people at other sections in the Ecology Building. I would especially like to thank Ullrika Sahlin for intellectual input.

Many thanks to Ulrika Norberg for proofreading the summary.

I am debtful to Gerard Geenen, Plant Cytometry Services, for ploidy level determinations and for answering my questions.

I thank many friends and colleagues from various parts of Europe for providing information about *Dactylorhiza* localities or for sending me material. I am particularly grateful to Maia Akhalkatsi, Sven Birkedal, Ilona Blinova, Joanna Bloch-Orłowska, József Pál Frink, Sven Hansson and Crina Mocan for practical help in the field. Special thanks must also go to Rayna Natcheva, Maria Olsrud and Torbjörn Tyler. Other people include:

Sunniva Aagaard, Richard M. Bateman, P. Batoušek, Anders Bertilsson, Agust H. Bjarnason, Per Bjurulf, Andrew Bystrushkin, Åse Bøilestad, Mark W. Chase, Elzbieta Cieslak, Salvatore Cozzolino, Brone Dohnar, R. Dundr, Stefan Eriksson, Michael F. Fay, Lars Fröberg, Tore Frøland, Kjell Georgson, Trond Grøstad, Mikael Gustafsson, Bjørn Erik Halvorsen, Roger Halvorsen, Jan-Erik Hederås, Olle Hedvall, Svein Imsland, Jana Jersáková, Janne Johansson, John Inge Johnsen, Stig Johnsson, Bengt Jonsell, Zygmunt Kacki, Arne Kildebo, Erich Klein, Karel Kreutz, Pella Larsson, Rolf Lidberg(†), Marej Liposevic, Marje Loide, Richard Lorenz, Tor Melseth, Julita Minasiewicz, Kalle Mälson, Henrik Nilsson, Maria Noskova, Kjell-Arne Olsson, Dag Olav Ovstdal, B. Paszko, Wojciech Paul, Henrik Æ. Pedersen, Guiseppe Pellegrino, Tarmo Pikner, Mari Reitalu, Paula J. Rudall, Ingvald Røsberg, Carna Sigvardsson Lööv, Steinar Skrede, Magda Szczepaniak, Kjell Thowsen, Taavi Tuulik, Kai Vahtra, Ruud van der Meijden, B. Vreeken, Krister Wahlström, Bo Wallén, Erik Westberg, Åke Widgren, Ł. Wilk and Finn Wischmann.

A huge thanks to my friends outside the *Dactylorhiza* world.

And finally, most of all I would like to thank my family!