



# LUND UNIVERSITY

## Diversity of Skin Infections

Ekström, Johanna

2011

[Link to publication](#)

*Citation for published version (APA):*

Ekström, J. (2011). *Diversity of Skin Infections*. [Doctoral Thesis (compilation), Clinical Microbiology, Malmö]. Avd för medicinsk mikrobiologi.

*Total number of authors:*

1

### General rights

Unless other specific re-use rights are stated the following general rights apply:

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

Read more about Creative commons licenses: <https://creativecommons.org/licenses/>

### Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

LUND UNIVERSITY

PO Box 117  
221 00 Lund  
+46 46-222 00 00

# DIVERSITY OF SKIN INFECTIONS

Johanna Ekström

Avdelningen för medicinsk mikrobiologi  
Skånes universitetssjukhus  
Lunds universitet



**LUNDS**  
UNIVERSITET

## AKADEMISK AVHANDLING

som med vedebörligt tillstånd av Medicinska Fakulteten vid Lunds universitet för avläggande av doktorsexamen i medicinsk vetenskap kommer att offentligen försvaras i Patologiska Institutionens föreläsningssal, ingång 78, Skånes universitetssjukhus, Malmö, torsdagen den 17 februari 2011 kl. 09.00

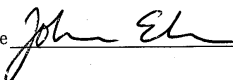
Fakultetsopponent:  
Professor Jan Albert  
Avdelningen för mikrobiologi, tumör och cellbiologi  
Karolinska institutet

Organization <b>LUND UNIVERSITY</b>  Division of Medical Microbiology, Department of Laboratory Medicine, Skåne University Hospital, Malmö, Sweden	Document name <b>DOCTORAL DISSERTATION</b>	
	Date of issue February 17, 2011	
	Sponsoring organization	
Author(s) Johanna Ekström		
Title and subtitle Diversity of skin infections		
Abstract <p>The identification of infectious agents in cancer has been one of the most rewarding endeavors in cancer research. Currently about 20% of the global cancer burden is linked to an infection. A common characteristic of virus-induced cancer is an increased incidence in immunosuppressed patients, presumably because of impaired host control of virus. Yet non-melanoma skin cancer (NMSC), the cancer that increases most among the immunosuppressed, does not have an established link to infection. NMSC, including squamous cell carcinoma (SCC) and basal cell carcinoma, is the most common cancer among Caucasians. Ultraviolet radiation is an established risk factor.</p> <p>Human papillomaviruses (HPVs) have been established as the major cause of cervical cancer. Many NMSCs contain one or several cutaneous types of HPV. Exploration of a possible infectious etiology of NMSC requires an unbiased and comprehensive approach for detection of as many infections as possible in the tumor. We examined NMSCs and other presumably HPV-associated lesions for the presence of unidentified HPV types or other microorganisms, using a combination of multiple displacement amplification (MDA), which amplifies all DNA in a sample without any requirement of prior knowledge of the nucleotide sequence, degenerate "general HPV primers" PCR and high-throughput sequencing. The most common microbial DNA in NMSC was Staphylococcus aureus (S. aureus). We also identified sequences from at least 40 previously not described putative HPV types, of which three novel types (HPV 109, 112 and 114) and an HPV 88 isolate were cloned and completely sequenced. Prevalences and viral loads were investigated in skin and genital samples from different patient groups. S. aureus DNA was more commonly detected in SCC compared to healthy skin (odds ratio, 6.23; 95% confidence interval, 3.10 – 12.53). However, the study design could not determine the causality of the association. HPV 88, 109 and 112 were almost only found in their index patients, whereas HPV114 was found in 1.7% of the female genital samples.</p> <p>In summary, we find that there is a wide diversity of HPV types in the skin. The association of S. aureus with SCC raises the possibility of general susceptibility to infection in SCC. An association of NMSC with a specific infection remains to be found.</p>		
Key words: Human papillomavirus, non-melanoma skin cancer, Staphylococcus aureus, multiple displacement amplification, high-throughput sequencing		
Classification system and/or index terms (if any):		
Supplementary bibliographical information:		Language English
ISSN and key title: 1652-8220		ISBN 978-91-86671-62-4
Recipient's notes	Number of pages 104	Price
	Security classification	

Distribution by (name and address)

I, the undersigned, being the copyright owner of the abstract of the above-mentioned dissertation, hereby grant to all reference sources permission to publish and disseminate the abstract of the above-mentioned dissertation.

Signature



Date 2011-01-11

# DIVERSITY OF SKIN INFECTIONS

Johanna Ekström

Doctoral Thesis



LUND UNIVERSITY

Malmö 2011

Department of Laboratory Medicine, Medical Microbiology,  
Lund University, Skåne University Hospital, Malmö, Sweden

ISBN 978-91-86671-62-4

ISSN 1652-8220

Copyright © Johanna Ekström

Lund University, Faculty of Medicine Doctoral Dissertation Series 2011:13

Printed by Media Tryck, Lund University, Sweden

*FOR MY FAMILY*



# Table of Contents

<b>SUMMARY</b> .....	<b>7</b>
<b>POPULÄRVETENSKAPLIG SAMMANFATTNING</b> .....	<b>8</b>
<b>LIST OF PAPERS</b> .....	<b>10</b>
<b>ABBREVIATIONS</b> .....	<b>11</b>
<b>INTRODUCTION</b> .....	<b>12</b>
INFECTIONS AND CANCER .....	12
NON-MELANOMA SKIN CANCER .....	15
<i>The skin</i> .....	15
<i>Skin cancer</i> .....	16
<i>Non-melanoma skin cancer</i> .....	16
Risk factors for non-melanoma skin cancer .....	17
PAPILLOMAVIRUSES .....	19
<i>Generalities</i> .....	19
<i>Evolution</i> .....	19
<i>Classification</i> .....	20
<i>Genomic organization</i> .....	23
E1 .....	23
E2 .....	24
E4 .....	24
E5 .....	24
E6 and E7.....	25
L1and L2.....	25
Non coding region.....	26
<i>The papillomavirus life cycle</i> .....	26
<i>HPV and mucosal infections</i> .....	27
Genital infections .....	27
Oral infections.....	28
<i>HPV and cutaneous infections</i> .....	28
Warts.....	28
Psoriasis .....	28
Epidermodysplasia verruciformis.....	29
Non-melanoma skin cancer .....	29
<i>Identification of novel human papillomavirus types</i> .....	30
STAPHYLOCOCCUS AUREUS .....	31
<i>Generalities</i> .....	31
<i>Pathogenesis and virulence factors</i> .....	31
<i>Staphylococcus aureus associated diseases</i> .....	32



METHODS FOR DETECTION OF POTENTIAL PATHOGENS.....	33
<i>Multiple displacement amplification</i> .....	33
Applications of MDA .....	34
<i>High-throughput sequencing</i> .....	34
High-throughput sequencing using GS FLX .....	35
Applications of GS FLX.....	36
<b>SUMMARY OF PAPERS.....</b>	<b>37</b>
AIMS .....	37
<i>Paper I</i> .....	37
<i>Paper II</i> .....	37
<i>Paper III</i> .....	37
<i>Paper IV</i> .....	37
MATERIALS AND METHODS.....	38
<i>Human papillomavirus-modified multiple displacement amplification</i> .....	38
<i>Real-time PCR</i> .....	38
<i>Paper I</i> .....	38
<i>Paper II</i> .....	39
<i>Paper III</i> .....	40
<i>Paper IV</i> .....	41
RESULTS AND DISCUSSION .....	42
<i>Paper I</i> .....	42
<i>Paper II</i> .....	43
<i>Paper III</i> .....	43
<i>Paper IV</i> .....	44
CONCLUDING REMARKS AND FUTURE PERSPECTIVES .....	46
<b>ACKNOWLEDGEMENTS.....</b>	<b>48</b>
<b>REFERENCES .....</b>	<b>50</b>

# Summary

The identification of infectious agents in cancer has been one of the most rewarding endeavors in cancer research. Currently about 20% of the global cancer burden is linked to an infection. A common characteristic of virus-induced cancer is an increased incidence in immunosuppressed patients, presumably because of impaired host control of virus. Yet non-melanoma skin cancer (NMSC), the cancer that increases most among the immunosuppressed, does not have an established link to infection. NMSC, including squamous cell carcinoma (SCC) and basal cell carcinoma, is the most common cancer among Caucasians. Ultraviolet radiation is an established risk factor.

Human papillomaviruses (HPVs) have been established as the major cause of cervical cancer. Many NMSCs contain one or several cutaneous types of HPV. Exploration of a possible infectious etiology of NMSC requires an unbiased and comprehensive approach for detection of as many infections as possible in the tumor.

We examined NMSCs and other presumably HPV-associated lesions for the presence of unidentified HPV types or other microorganisms, using a combination of multiple displacement amplification (MDA), which amplifies all DNA in a sample without any requirement of prior knowledge of the nucleotide sequence, degenerate “general HPV primers” PCR and high-throughput sequencing. The most common microbial DNA in NMSC was *Staphylococcus aureus* (*S. aureus*). We also identified sequences from at least 40 previously not described putative HPV types, of which three novel types (HPV 109, 112 and 114) and an HPV 88 isolate were cloned and completely sequenced. Prevalences and viral loads were investigated in skin and genital samples from different patient groups. *S. aureus* DNA was more commonly detected in SCC compared to healthy skin (odds ratio, 6.23; 95% confidence interval, 3.10 – 12.53). However, the study design could not determine the causality of the association. HPV 88, 109 and 112 were almost only found in their index patients, whereas HPV114 was found in 1.7% of the female genital samples.

In summary, we find that there is a wide diversity of HPV types in the skin. The association of *S. aureus* with SCC raises the possibility of general susceptibility to infection in SCC. An association of NMSC with a specific infection remains to be found.

# Populärvetenskaplig sammenfattning

Det är väl känt att infektioner kan orsaka eller medverka till utveckling av vissa former av cancer hos människan. Cirka 20% av den globala cancerincidensen orsakas av infektioner. Patienter med nedsatt immunförsvar har en ökad förekomst av ett flertal virus-orsakade cancerformer, t.ex. livmoderhalscancer som orsakas av humant papillomvirus (HPV) och levercancer som orsakas av Hepatit B och C. Upptäckten av dessa samband har möjliggjort att man numera kan vaccineras mot dessa sjukdomar. Även för icke-melanom hudcancer (NMSC) kan man påvisa en kraftig ökning hos dessa patienter. Hittills har inget samband mellan infektion och NMSC kunnat säkerställas. NMSC, som huvudsakligen består av diagnoserna skivepitelcancer och basalcellscancer, är den vanligast förekommande cancer bland vithyade, och ultraviolett ljus är en känd riskfaktor.

HPV orsakar ett flertal sjukdomar; utöver livmoderhalscancer t.ex. även kondylom och cancer i munhålan. I NMSC hittas vanligen flera olika HPV-typer, men de påträffas även i frisk hud. För att kunna undersöka en möjlig association mellan en infektion och NMSC krävs därför en mångsidig objektiv metod som kan upptäcka maximalt antal patogener i en tumör.

Vi undersökte NMSC och andra möjliga HPV-relaterade lesioner efter förekomst av nya HPV-typer och andra mikroorganismer med en metod som amplifierar allt DNA i ett prov, samt amplifiering med en teknik som kan påvisa många olika HPV-typer följt av sekvensering med en effektiv sekvenseringsteknik. Vi identifierade sekvenser från minst 40 tidigare ej kända HPV typer. Av dessa klonades och helgenomssekvenserades tre typer, HPV 109, 112 och 114, samt den sedan tidigare kända typen HPV 88. HPV 88 och 109 hittades bägge i skivepitelcancer, HPV 112 upptäcktes i ett kondylom och HPV114 i en lätt cellförändring i livmoderhalsen. När vi undersökte skivepitelcancer för förekomst

av nya HPV-typer och andra mikroorganismer fann vi även flera sekvenser som tillhörde bakterien *Staphylococcus aureus* (*S. aureus*).

Vidare undersöktes förekomsten av *S. aureus* samt HPV 88, 109, 112 och 114 i olika hud- samt genitala prover. Vi fann att DNA från *S. aureus* var betydligt vanligare i skivepitelcancer än i frisk hud. HPV 88, 109 och 112 är sällsynta virus, medan HPV 114 återfanns i 1.7% av de genitala proven från kvinnor.

Sammanfattningsvis visar dessa resultat på att mångfalden av HPV i huden är mycket stor. Det faktum att *S. aureus* var associerat med skivepitelcancer kan visa på en allmän mottaglighet för infektioner av skivepitelcancer och att ett samband mellan skivepitelcancer och en specifik infektion återstår att finna.

# List of papers

This thesis is based on the following papers:

- I. Cutaneous human papillomavirus 88: remarkable differences in viral load.  
**Kullander J**, Handisurya A, Forslund O, Geusau A, Kirnbauer R, Dillner J.  
Int J Cancer. 2008 Jan 15;122(2):477-80.
  
- II. *Staphylococcus aureus* and squamous cell carcinoma of the skin.  
**Kullander J**, Forslund O, Dillner J.  
Cancer Epidemiol Biomarkers Prev. 2009 Feb;18(2):472-8. Epub 2009 Jan 20.
  
- III. Three novel papillomaviruses (HPV 109, HPV 112 and HPV 114) and their presence in cutaneous and mucosal samples.  
**Ekström J**, Forslund O, Dillner J.  
Virology. 2010 Feb 20;397(2):331-6. Epub 2009 Dec 6.
  
- IV. High-throughput sequencing reveals diversity of Human Papillomaviruses in cutaneous lesions.  
**Ekström J**, Bzhalava D, Svenback D, Forslund O and Dillner J.  
Manuscript.

Paper I and II were published in the maiden name Kullander.

# Abbreviations

AIDS	Acquired immune deficiency syndrome
AK	Actinic keratosis
ASCUS	Atypical cell of undetermined significance
ATP	Adenosine triphosphate
BCC	Basal cell carcinoma
bp	Base pair
CCD	Colony collapse disorder
CI	Confidence interval
CIN	Cervical intraepithelial neoplasia
DNA	Deoxyribonucleic acid
EBV	Epstein Barr virus
EV	Epidermodysplasia verruciformis
GS FLX	Genome sequencer FLX
HCC	Hepatocellular carcinoma
HHV	Human herpes virus
HIV	Human immunodeficiency virus
HPV	Human papillomavirus
KA	Keratoachantoma
kb	Kilo base
MCC	Merkel cell carcinoma
MCV	Merkel cell poylomavirus
MDA	Multiple displacement amplification
NF- $\kappa$ B	Nuclear factor kappa B
NMSC	Non-melanoma skin cancer
OR	Odds ratio
ORF	Open reading frame
PCR	Polymerase chain reaction
pRb	Retinoblastoma protein
PV	Papillomavirus
RDA	Representational differences analysis
RNA	Ribonucleic acid
RRP	Recurrent respiratory papillomatosis
SCC	Squamous cell carcinoma
SISPA	Sequence-independent, single primer amplification
SK	Seborrhoeic keratosis
UV	Ultraviolet

# Introduction

## Infections and cancer

Identification of unknown pathogens is an urgent task of importance for outbreak preparedness as well as for studies of chronic diseases of unknown etiology (11, 61, 201). Currently, about 20 % of the global cancer incidence can be linked to infectious agents (270).

The search to identify infectious agents as causative factors for human cancers is difficult due to of several reasons as summarized in the Nobel lecture 2008, by Harald zur Hausen (270):

- 1) *No human cancer arises as the acute consequence of infection. The latency period between primary infection and development of cancer is usually in the range of 15 to 40 years (270).*

This increases the importance of longitudinal epidemiological studies that follow patients over time, preferably over many decades. Infected healthy subjects may not develop cancer until many years later. Also, it is possible that a causative infection could have disappeared long before the cancer has developed (so called hit-and-run mechanism) (10, 229).

- 2) *No synthesis of the infectious agents occurs in the cells, besides some exceptions (270).*

Nowadays, virus detection by isolation in tissue culture is not commonplace, but indeed detection of the infectious agent by other molecular methods could also be complicated when there is no ongoing virus production. E.g., immunohistochemistry might be false negative if it is using antibodies to a protein that is only expressed in the productive phase of the viral life cycle or a polymerase chain reaction (PCR) might be

false negative if it uses primers targeting a piece of deoxyribonucleic acid (DNA) that could be lost if the genome of a virus is integrated.

- 3) *Most of the infections linked to human cancers are common in the whole human population, while only a proportion develops cancer (270).*
- 4) *Mutations in host cell genes or within the viral genome are mandatory for malignant conversion (270).*
- 5) *Mutations caused by chemical and physical carcinogens act synergistically with carcinogenic infectious agents (270).*
- 6) *Some infectious agents act as indirect carcinogens, without persistence of their genes within their respective cancer cells (270).*

E. g. *Helicobacter Pylori* that presumably induces cancer by causing a chronic inflammation (164), and is not present in all parts of the carcinogenic tissue. This indirect carcinogenic activity is more difficult to link to cancer than case for HPV where part of the viral DNA is always present in malignant cells and production of oncogenic proteins E6 and E7 occurs (67).

An important issue in investigations of cancer causes is the direction of causality, one of nine criteria required to establish a causal relationship between an exposure and a disease, as formulated by Bradford Hill (123), If an infection is more common in cancers than in controls what came first? Does a particular lesion attract (or activate) a microbe or does the microbe contribute to the development of the lesion?

With appropriate molecular and epidemiological approaches to the issue, several pathogens have been identified to be implicated in different types of cancer. The first human tumor virus, Epstein-Barr virus (EBV), was 1965 linked to Burkitt's lymphoma (77, 122). Another herpesvirus associated to a human cancer, Kaposi's sarcoma, is human herpes virus (HHV) -8, also called Kaposi sarcoma virus, discovered by Yang Chang and Patrick Moore in 1994 (45). Chang and Moore also discovered a new polyomavirus in Merkel cell carcinoma, Merkel cell polyomavirus (85). The revelation of a relationship between hepatitis B and hepatocellular carcinoma (HCC) in 1975 (34) led to the development of the first



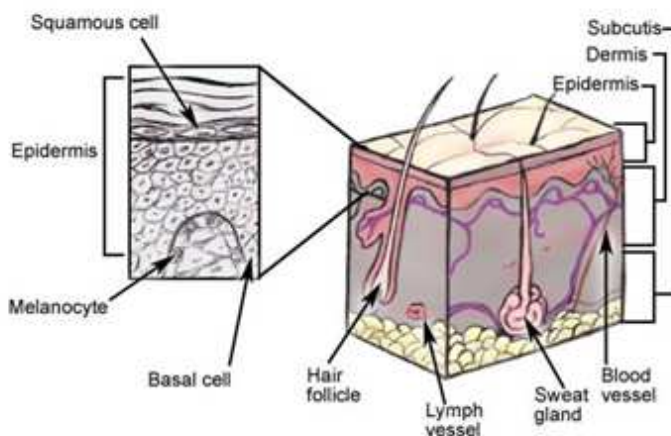
vaccine shown to be effective in preventing a human cancer. Later also hepatitis C was linked to HCC (9, 146). The second human cancer vaccine, that became available in 2006, prevents the majority of cervical cancer cases caused by human papillomavirus (HPV) (254, 269) The discovery of the oncogenic anogenital HPVs that cause cervical cancer was made by Harald zur Hausen and co-workers and was honoured with the Nobel Prize in Physiology or Medicine 2008 (254, 269). Besides viruses, also other pathogens have been identified as causes of cancer, such as the bacterium, *Helicobacter Pylori*, the major cause of gastric cancer (164), and parasitic infections, e.g. *Schistosoma hematobium* associated with bladder cancer (35).

A few cancers occur at an increased incidence in immunosuppressed patients compared to the general population (76, 113). Most of them have an established infectious etiology, e.g. cervical cancer and HPV (254, 269), Burkitts lymphoma and EBV (77, 122) and Kaposi's sarcoma and HHV-8 (45). Non-melanoma skin cancer is the cancer form that increases most after immunosuppression, but still does not have an established infectious etiology. Interestingly, several additional cancer forms in addition to the above mentioned have also been reported to have some increased incidence in immunosuppression, e.g. cancer of the kidney, leukemia and colon cancer (113), warranting a search for infections also in these cancer forms.

# Non-melanoma skin cancer

## The skin

The skin is the largest organ in the body. It protects the body against foreign microorganisms and prevents loss of too much water. The skin consists of three layers; epidermis, dermis and the hypodermis (the subcutaneous fatty tissue) (Figure 1). The purpose of the hypodermis is to attach the skin to underlying bone and muscles as well as supply it with blood vessels and nerves. The dermis contains hair follicles, sweat glands, lymphatic vessels and blood vessels. The blood vessels in the dermis provide nourishment and waste removal from its own cells as well as from the epidermis. The epidermis consists of three layers with the bottom layer made up of basal cells. The basal cells divide to form keratinocytes. As new keratinocytes are formed the old ones move up the epithelia, change shape, die and form the outermost layer (Figure 1). Also melanocytes, Langerhans cells and Merkel cells are found in the epidermis.



**Figure 1.** Schematic picture of a cross section of the skin. Image reprinted with permission from Medscape.com, 2010.

## Skin cancer

There are several different types of cancer originating in the skin. Merkel cell carcinoma (MCC), from Merkel cells, is very rare but has the highest mortality rates among skin tumors (37, 196). Risk factors for MCC are ultraviolet (UV) radiation and immunosuppression (169). Since the discovery of Merkel cell polyoma virus (MCV) in MCC (85) several studies have investigated if MCV is associated with MCC (14, 42, 85, 190, 261). MCV DNA is more common in MCC than in healthy controls or other skin lesions (14, 85, 261). Antibodies to MCV are common in the general population, but MCC patients have higher antibody levels (42, 190). Another aggressive skin tumor is malignant melanoma that develops from the melanocytes. It is curable at early stages, but can spread to other parts of the body and be highly mortal (37). Non-melanoma skin cancer (NMSC) is the most common cancer among Caucasians and is described in more detail below (158, 235). Another common neoplasm is keratoachantoma (KA), a skin tumor characterized by a rapid onset most often followed by spontaneous regression within a few months (137, 217). The histological pattern is often difficult to distinguish from squamous cell carcinoma (SCC), a form of NMSC.

### Non-melanoma skin cancer

The predominating NMSCs are SCC and basal cell carcinoma (BCC). BCC is more common than SCC, with a ratio of approximately 4:1 (102). The increased incidence of NMSC in Sweden from 1970 to 2008 can be seen in Figure 2. The mortality rate is fortunately very low, but the lesions can cause disfigurement and treatment costs are high (197). Prevention and early detection would reduce both morbidity and cost.

BCC is a slowly growing tumor that has a low degree of malignancy (208, 244). However, BCC can be locally invasive and can cause massive tissue damage (244). The lesion is most frequently found on areas exposed to the sun as the head and neck, followed by the trunk, arms and legs (208). Treatment can be both surgical, e.g. excision of the lesion, cryosurgery or electrodesiccation, and nonsurgical, e.g. radiotherapy (208).

SCC is the second most common skin cancer and, as for BCC, the tumors are most common on sun-exposed areas such as the head (37). In contrast to BCC, which is

believed to arise de novo (i.e. it has no known precursor), a small proportion of SCCs are suggested to arise from actinic keratosis (AK) even though the rate of malignant conversion varies greatly in different studies (148, 162, 163). SCCs are generally slow growing but have the capability to metastasize (49, 66). Nevertheless, most patients have an excellent prognosis and most tumors can be eliminated by electrodesiccation, curettage, excision or cryosurgery, with a low risk of metastasis (7).

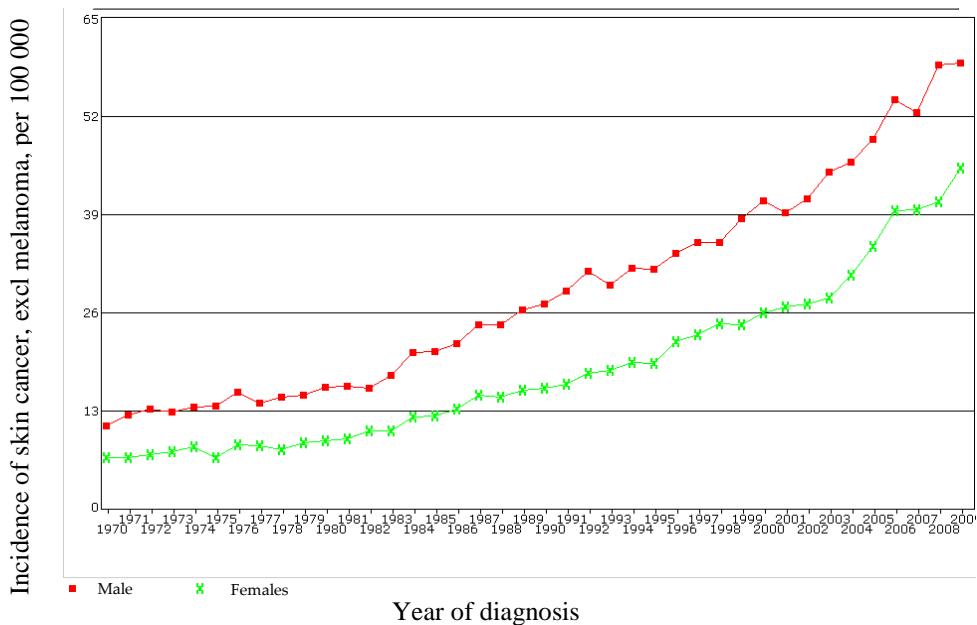


Figure 2. Incidence of non-melanoma skin cancer per 100 000 for males and females. Adapted from Cancer incidence in Sweden 2008, published by Socialstyrelsen.

### *Risk factors for non-melanoma skin cancer*

The main risk factor for developing NMSC is exposure to ultraviolet (UV) light (159, 198). In the case of SCC, it appears that long-term exposure to UV light is responsible for the progression, but for BCC short term burning episodes seem more important (37, 159). Other risk factors include older age, male sex, fair skin that tan poorly, red, blond or light-brown hair, blue or light-colored eyes and a

number of inherited genetic skin conditions, e.g. epidermodysplasia verruciformis (197).

There are several lines of evidence suggesting that there may also be other preventable risk factors for this cancer. There is a greatly increased incidence of NMSCs, varying from 10- to 250-fold, in patients receiving immunosuppressive therapy because of organ transplantation, (28, 117, 132, 154). The BCC:SCC ratio is reversed in immunocompromised patients, with SCC being the most common (78, 117). Among these patients, the impaired immune surveillance against viral antigens results in increased incidence of skin warts as well as several virus-associated cancers (113) such as Kaposi's sarcoma, Epstein-Barr virus associated lymphoma and HPV-associated cancers (32, 40, 232). An association of NMSC with HPV has been investigated (reviewed in 118, 177) and will be discussed further in the section "HPV and cutaneous infections".

# Papillomaviruses

## Generalities

Papillomaviruses (PVs) are small, nonenveloped viruses that belong to the family *Papillomaviridae*. Their genome consists of double-stranded circular DNA, about 8 kilo bases (kb), surrounded by an icosahedral capsid. The capsid consists of 72 capsomers, resembling a golf ball when viewed by an electron microscope (27). The first PV was identified by Richard Shope in the 1930s in wild cottontail rabbits (221). Soon thereafter Rous and Beard showed that cottontail rabbit papillomavirus caused skin cancer in domestic rabbits (206, 207). In 1949 the first human PVs (HPVs) were visualized in skin warts by electron microscope (238) and in 1972 Jablonska et al. demonstrated that HPV is responsible for cutaneous lesions in patients with the disease epidermodysplasia verruciformis (129). The association of genital HPVs with cervical cancer was proposed by Harald zur Hausen in the late 1970s (269) and the first major oncogenic HPV (HPV 16) was cloned in 1983 (72). In 2006, after sufficient amounts of epidemiological data had accumulated, the US Food and Drug Administration approved the first vaccine for prevention of cervical cancer (168).

## Evolution

The PV genome is very stable and as they utilize the host cell replication machinery they evolve at the same rate as the human genome, mainly through point mutations (31). Papillomaviruses have been detected worldwide and in a wide variety of different species (16, 199, 200, 236). The extensive genomic diversity of *Papillomaviridae* has taken millions of years to arise. By comparison Human Immunodeficiency Virus (HIV) can diverge to a similar extent during a 10-year infection (31). This difference is due to the high-fidelity proof-reading capacity of the DNA-dependent DNA polymerases used by HPV.

The current hypothesis is that PVs are ancient and existed already at the evolutionary origin of humans. This theory is based on the wide geographic distribution of PVs that cannot be explained by airborne transmission as close physical mucosal or cutaneous contact is required for transmission of PV types (31, 160). Studies of different HPV 16 and 18 isolates indicates co-evolution with humans since the origin of man-kind (Ho, Ong, Chan).

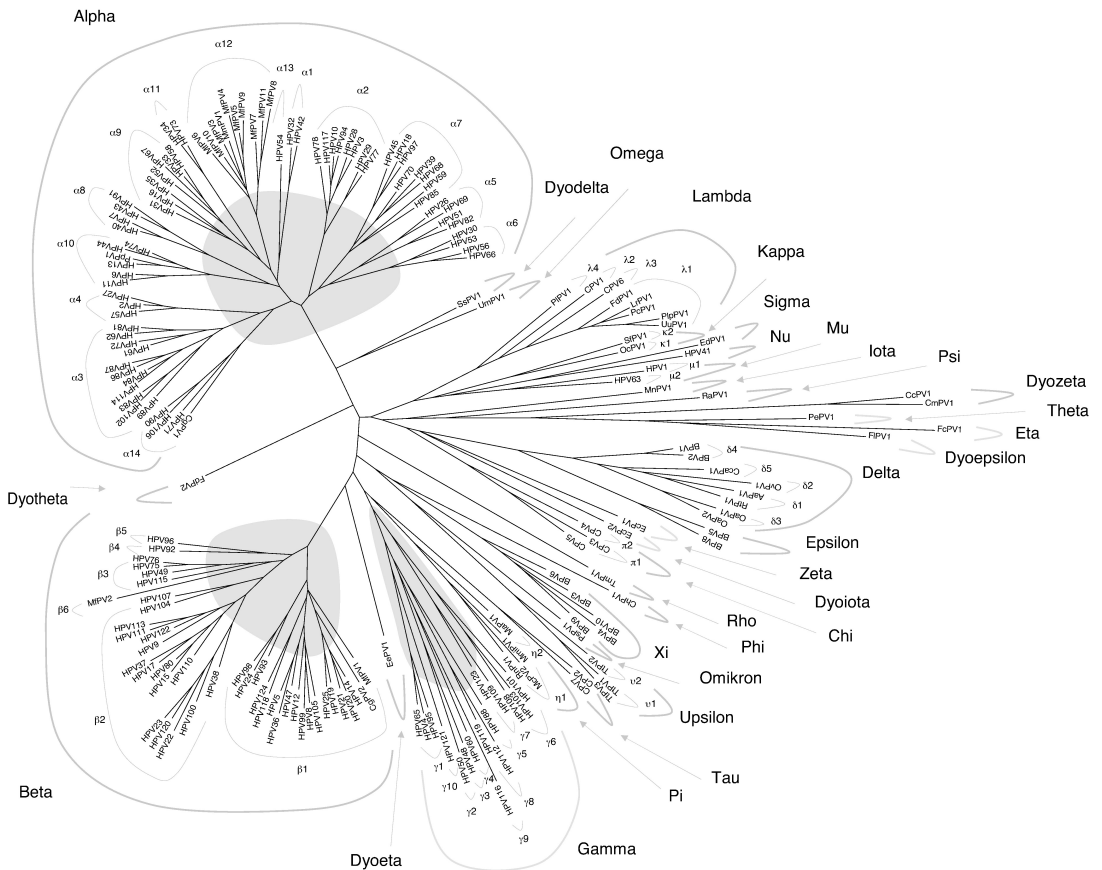
## Classification

PVs comprise a large group of viruses with at least 200 completely characterized types divided into 29 genera infecting a wide variety of mammals and birds, including e.g. cows, parrots and dolphins in addition to humans (30). Classification of PVs is based on the sequence of the capsid protein L1 (58). PV types within a genus show less than 60% similarity in the L1 gene to PV types of other genera and different viral species within a genus share between 60 and 70 % similarity. A novel PV types has less than 90% similarity to any known type. Subtypes and variants differ 2-10% and maximum 2% respectively from any PV type. The PV types infecting humans are found in five different genera: Alpha-, Beta-, Gamma-, Mu- and Nu-PVs (Figure 3).

The majority of the at least 74 types found in genus Alpha-PV infect the human mucosa, but a few types that infect animals are also found, e.g. Rhesus Macaque and Colobus monkey, in addition to a small number of types infecting human skin (Figure 3). The PV types with mainly cutaneous tissue tropism are found in species 2 (HPV 3, 10, 28, 29, 77, 78, 94 and 117), 4 (HPV 2, 27 and 57) and 8 (HPV 7, 40, 43 and 91) (58). The mucosal types are further divided into high- and low-risk types depending on their ability to cause cancer (173). Most of the high-risk types cluster within species 7 and 9, e.g. HPV 16 and 18 (51, 173), which are the two most common types in cervical cancer (173). In species 10 we find the low-risk types 6 and 11 which are the main etiological agents of condyloma acuminata (genital warts) (112). The two different HPV vaccines approved, Gardasil<sup>®</sup> (Merck and Co) and Cervarix<sup>®</sup> (GlaxoSmithKline) both protect against HPV16 and 18, while Gardasil also protects against HPV 6 and 11 (12).

Genus Beta-PV contains six different species, with at least 42 different types, mainly causing cutaneous lesions, with the exception of a few animal types (Figure 3). Patients with the rare disease epidermodysplasia verruciformis are often infected with HPV types from this genus, especially HPV 5 and 8 from species 1 (183). Different studies have found an association of SCC with HPV type within genus beta species 2 (21, 94).

In the classification study by Bernard et al. (published in May 2010), genus Gamma-PV consists of 16 types divided into 10 different species (Figure 3). Since then at least 10 novel types have been deposited in GenBank (Table 1). In addition



**Figure 3.** Phylogenetic tree based on the L1 sequences from 189 papillomaviruses. Reprinted from Bernard et al., Classification of papillomaviruses (PVs) based on 189 PV types and proposal of taxonomic amendments, in *Virology* 2010; **401**:70-9 with permission from Elsevier.

a phylogenetic study clustered subgenomic sequences of 97 putative HPV types into this genus (90) suggesting a very large diversity of gamma types. Subgenomic sequences used in that study were generated using the FAP59 and FAP64 primers (92), producing an amplicon of approximately 450 base pairs (bp) in L1. Although, most of the HPV types in genus gamma PV are found in cutaneous infections, 5 types are found in various mucosal samples (Table 1). HPV 101, 103 and 108 are all cloned from cervicovaginal cells (48, 179), HPV 112 from a condyloma acuminata (75) and HPV 116 from a rectal swab (152).



The genera Mu- and Nu-PVs contain only three different types (HPV 1, 63 and 41) detected in cutaneous lesions (Figure 3).

**Table 1.** Characteristics of HPV types within the genus Gamma.

Species	Types	Original lesion	Accession nr	Reference
1	4	Cutaneous wart	X70827	Pfister and Gissman, 1978 (192)
	65	Cutaneous wart	X70829	Egawa et al., 2005 (73)
	95	Cutaneous wart	AJ620210	Egawa et al., 1993 (74)
2	48	SCC* of the skin	NC_001690	Müller et al., 1989 (172)
	131**	Cutaneous wart	GU117631	Nindle et al. unpubl.
3	50	AK* from an EV patient	U31790	Favre et al., 1989 (81)
4	60	Plantar cyst	U31792	Matsukura et al, 1992 (165)
5	88	NA***	NC_010329	Egawa et al., unpubl.
6	101	CIN III*	NC_008189	Chen et al., 2007 (48)
	103	Normal cervicovaginal cells	NC_008188	Chen et al., 2007 (48)
	108	Low-grade cervical lesion	NC_012213	Nobre et al., 2009 (179)
7	109	SCC of the skin	EU541441	Paper III (75)
	123	NA	GQ845445	Chen et al., unpubl.
	134**	Cutaneous wart	GU117634	Nindle et al., unpubl.
	149**	Cutaneous wart	GU117629	Nindle et al., unpubl.
8	112	Condyloma accuminata	EU541442	Paper II (75)
	119	NA	GQ845441	Chen et al., unpubl.
9	116	Rectal swab	FJ804072	Li et al., 2009 (152)
	129**	Cutaneous wart	GU233853	Nindle et al. unpubl.
10	121	NA	GQ845443	Chen et al., unpubl.
	130**	Cutaneous wart	GU117630	Nindle et al. unpubl.
	133**	Cutaneous wart	GU117633	Nindle et al. unpubl.
X****	128**	Cutaneous wart	GU225708	Nindle et al. unpubl.
Y****	127**	Healthy skin	HM011570	Schwalter et al., 2010 (216)
	132**	Cutaneous wart	GU117632	Nindle et al., unpubl.
	148**	Cutaneous wart	GU129016	Nindle et al., unpubl.

\*Abbreviations: SCC= squamous cell carcinoma, AK= actinic keratosis, EV= epidermodysplasia verruciformis, CIN III= cervical intraepithelial neoplasia grade III.

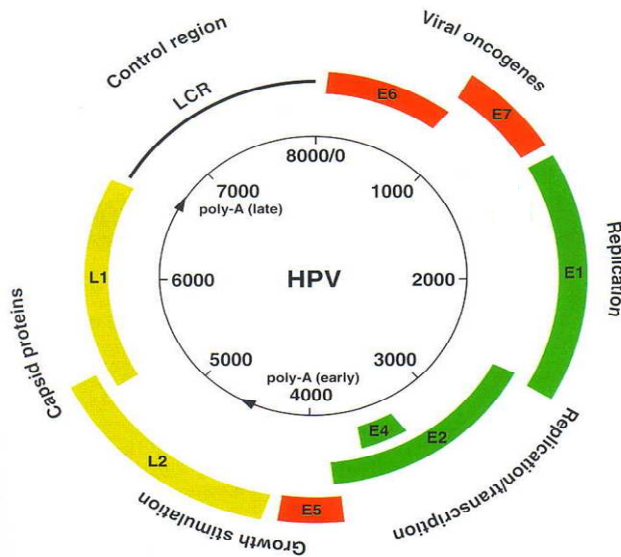
\*\*HPV types not included in the phylogenetic tree in Figure 3, was found in GenBank and classification was determined by comparing their L1 sequences to the L1 sequences of other gamma types.

\*\*\*NA= not available as there is no article describing the origin of the lesion.

\*\*\*\*These types have not yet been designated a species.

## Genomic organization

All HPV types have a similar genomic organization. It is divided into three regions, a noncoding, an early and a late region (Figure 4). The non-coding region regulates transcription of the open reading frames (ORFs) and the early region, E1, E2, E4, E5, E6 and E7, encodes the proteins required for viral DNA replication and cellular transformation. The capsid proteins, L1 and L2, are encoded by the late region.



**Figure 4.** Schematic picture of the genomic organization of HPV.

### *E1*

Only two viral proteins are needed for efficient replication, E1 and E2, as the host cell is providing all other proteins and factors. E1 is the largest of the PV coded proteins and the only one with enzymatic activities; it is an adenosine triphosphate (ATP) dependent DNA helicase (265). The E1 protein binds to the origin of replication as a dihexamer, unwinds the DNA, recruits DNA polymerase  $\alpha$ /primase and the single stranded DNA binding protein replication protein A to initiate replication (52, 115, 153, 218). E1 is required both for initiation and elongation of viral DNA synthesis (157).

## *E2*

E2 is a multifunctional DNA binding protein that regulates both replication and transcription. It is divided into three distinct regions; a DNA binding domain in the carboxyl terminal end, transcriptional activity in the amino terminal end and in between a flexible hinge region (103). It binds to the DNA as a dimer at E2 protein binding sites in the replication origin and enhances replication by interaction with E1 (68, 147, 166). E2 comes in many different forms depending on alternative ribonucleic acid (RNA) splicing and alternative promoter usage. The full length protein is required for the interaction with E1 (5). The shorter forms compete with the full length protein to regulate transcription and replication (50, 157, 240). This modulation is abrogated in most cervical cancers as the HPV genome is integrated into the human genome at sites that disrupts the E2 gene (237).

E2 also has a role in viral DNA attachment to chromosomes, ensuring episomal maintenance within replicating cells. Segregation of viral episomes depends on tethering to viral chromosomes (150, 225), an event mediated by the cellular protein bromodomain-containing protein 4 (267). It has also been shown that, for at least some HPV types, the E2 protein associates with spindle fibers rather than the chromosome during mitosis (251).

## *E4*

The most abundant HPV protein is the fusion protein E1<sup>E4</sup> and different functions has been suggested; e.g. promoting genome amplification and S phase maintenance (263, 264) but results have not been consistent (80), suggesting that HPV type-specific differences exist.

## *E5*

The E5 protein is the main transforming protein in bovine PVs (64, 212). For human PVs, the E5 gene is expressed in high-risk HPV types, but many low-risk genital types and cutaneous types lack an E5 ORF or a translation start codon for E5 (98, 211). E5 may play a role in HPV infection (46, 65, 83), but is frequently missing in cervical cancer, due to integration, (167) and is probably not necessary for maintenance of the transformed phenotype.

### *E6 and E7*

E6 and E7 are the major oncogenes in humans and they are regularly expressed in HPV associated lesions and cancers. Both genes code for growth-stimulating proteins and especially in high-risk HPV types this can lead to malignant growth.

The most well-known mechanism of the high risk E6 protein is its ability to bind to and degrade the p53 tumor suppressor protein (210, 259). The E6 protein of low risk types do not share this property, but some ability to repress p53 dependent transcription regulatory functions have been shown (246). The E6 proteins of high risk types all have a PDZ binding motif (245). The PDZ motifs are found in many different proteins and the biological consequence of the interaction with E6 is currently investigated. The oncogenic HPV E6 proteins disrupt cellular tight junctions through the degradation of MAGI-1 (143), a protein possessing PDZ motifs.

Interestingly there exists three HPV types, HPV 101, 103 and 108, lacking an E6 gene (48, 179). They were all isolated from cervicovaginal cells, despite being phylogenetically clustered with HPV types normally infecting the skin in the genus Gamma-PVs.

The function of E7 is to maintain viral replication by promoting S-phase entry in otherwise non dividing differentiating cells in the epithelia (108). The tumor suppressor protein pRb, together with relative p130, inhibit cell cycle progression until a cell is ready to divide. Interaction of E7 with pRb disrupts the growth-suppressive pRb-E2F complex, thereby promoting S-phase replication (108). The ability to bind and degrade pRb differs between the HPV types, with the high risk types being more efficient (97). The E7 protein from both high and low risk types bind and degrade p130 thereby inducing viral replication (100).

### *L1 and L2*

The two late genes, L1 and L2, code for the structural proteins. The L1 ORF is the most conserved region of HPVs and as mentioned previously used for classification (58). Five units of L1 form the pentameric capsomers of which 72 constitute most of the capsid, while one L2 molecule is found beneath each of the pentamers (41). The L1 proteins can self-assemble into virus-like particles (140), which is the basis for the HPV vaccine (12). L2 is not required for capsid formation, but may facilitate the assembly (87, 114). L2 has also been proposed to

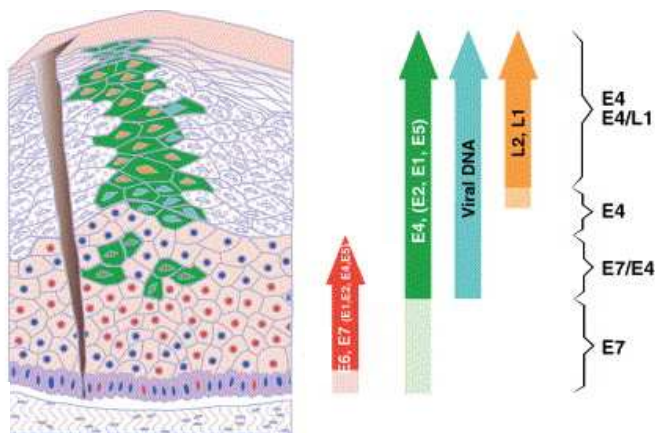
have a role in e.g. the capsid assembly by introducing HPV DNA to the virus particles (268) and to facilitate penetration of the viral genome from endosomes (136).

### *Non coding region*

The noncoding region, also called the upstream regulatory region or the long control region is the least conserved region among PVs (58). The non coding region contains the replication origin as well as binding sites to the regulatory protein E2 and also to various cellular transcriptional regulatory factors (20, 44, 107).

## The papillomavirus life cycle

PVs infect epithelial cells in the skin or the mucosa and they are dependent on the epithelial differentiation for viral replication (Figure 5). Little is known about the initial steps of infection and it has long been thought that the entry occurs in the basal epithelial cells at sites of microlesions (67), but a recent study showed that HPV pseudovirions bind to the basement membrane and become cell-associated at a later time-point (139, 203). The receptor necessary for uptake of the virion is not



**Figure 5.** Overview of the papillomavirus life cycle. After infection, the early proteins E6 and E7 are produced to enable S-phase entry. In the higher epithelial layers production of proteins required for replication is increased. In the upper layers of the epithelium the late capsid proteins L1 and L2 are expressed and the viral capsid is released. Adapted from Doorbar, The papillomavirus life cycle in *Journal of Clinical Virology* 32S (2005) S7–S15 with permission from Elsevier.

known, but the widely expressed heparin sulphate proteoglycan mediates the initial attachment (104, 135, 139) probably also involving a co-receptor (219). Internalization is not completely understood, and both the clathrin and caveolar mediated endocytic pathways have been described to be involved (38, 54). Inside the cell, the virus starts to replicate: first in a non-productive way with on average one replication round per cell cycle with the viral genome maintained as an episome (125). When an uninfected cell differentiates it leaves the cell cycle, losing the ability to replicate (241). In order to continue replication in differentiating cells the virus stimulates G1 to S-phase progression utilising the transforming proteins E6 and E7 (108). During differentiation the expression of E6 and E7 is upregulated (67), viral DNA is amplified at a high copy number and in the higher epithelial layers the viral capsid proteins, L1 and L2, are synthesized and finally the virions are assembled and released (67, 187).

## HPV and mucosal infections

### *Genital infections*

The most important malignancy associated with HPV is cervical cancer, representing one of the most common cancers in women worldwide (86), with more than half a million new cases every year and a yearly mortality of more than 250 000 deaths. The prevalence of HPV in cervical tumors is nearly 100% (254), and the most common high risk HPV types are HPV 16 followed by 18, 31, 33 and 35 (51, 173). An infection with a sexually transmitted HPV in the genital area is highly prevalent among women, especially adolescents (57), but most infections are transient (79, 195). Persistence of infection (124, 213) and high viral load (171, 214, 266) are risk factors for malignant conversion. Other co-factors are smoking (19), multi-parity (174), use of hormonal contraceptives (227) and other sexually transmitted diseases (226, 228).

Condyloma acuminata (genital warts) is a very common sexually transmitted disease caused by low-risk HPV types, mainly HPV 6 and 11 (112). Condyloma is a benign lesion that may regress spontaneously or after treatment (71).

HPV also causes several other genital diseases, notably vulvar, vaginal, anal and penile cancer (4).

### *Oral infections*

Although consumption of alcohol and tobacco are risk factors for oral and pharyngeal cancers (189), there is an increased incidence in young people with no smoking and drinking history (176). Epidemiological studies suggest a strong association of HPV infection and oral cancer development, with HPV 16 being the most prevalent type (144). Globally, HPV is estimated to attribute to 3% of oral cancers and 12 % of oropharynx (189).

Recurrent respiratory papillomatosis (RRP) is a rare disease with an estimated incidence of 2 per 100 000 in adults and 4 per 100 000 in children (109). RRP is primarily caused by HPV 6 and 11 (234). Although benign, there is a significant morbidity due to repeated treatment caused by recurrent lesions (109). Extension of the growths into the lower airways indicates a poorer prognosis.

### HPV and cutaneous infections

Infections of HPV on the skin are very common (12, 17, 32) and acquisition appears to occur already shortly after birth (18, 110, 126, 256). A large spectrum of cutaneous HPV is commonly detected both on healthy skin (16, 17), in plucked eyebrow samples (39, 55, 194) as well as in different skin lesions (193).

### *Warts*

Common manifestations of cutaneous HPV are skin warts, a benign skin disorder, considered to be no more than a cosmetic nuisance (130). The warts can occur on almost every location on the skin and generally, they resolve spontaneously. Cutaneous HPV types that are frequently associated with warts are type 1, 2, 3, 4, 7, 10, 41 and 57 (105, 130, 184).

### *Psoriasis*

HPV DNA, especially from HPV 5 and 8, can be detected in around 90 % of psoriatic lesions (82, 257). There is a significantly higher prevalence of antibodies to HPV 5 and 8 in patients with psoriasis compared to healthy donors (82, 233). A role of HPV in the development of psoriasis is uncertain. A recent study proposed that the presence of the virus is due to the immunosuppression induced in patients receiving phototherapy (209).

### *Epidermodysplasia verruciformis*

The association of HPV with skin cancer was as previously mentioned first demonstrated in patients with epidermodysplasia verruciformis (EV) (129). Patients suffering from the rare hereditary disease EV develop skin lesion in early infancy. The EV lesions are refractory to conventional wart treatment and by the fourth decade, more than half of the patients develop precancerous lesions and invasive NMSC, especially SCC (reviewed in 131, 183). HPV types within the genus Beta PV are commonly found in EV lesions, with HPV 5 being the most prevalent type (182). HPV 5 is also the type that has the strongest association with the malignant conversion to SCC in these patients (185).

### *Non-melanoma skin cancer*

Many studies have investigated the presence of HPV in NMSCs with prevalence rates up to 90 % and generally a little higher in immunosuppressed patients compared to immunocompetent individuals (reviewed in 118, 177). Many cutaneous HPV types are also common on healthy skin (17, 22, 39). Detection of DNA in tumors does not necessarily mean an infection, as it may merely be a viral contamination of the skin surface. Forslund with colleagues showed that cleansing of the skin with tape before taking punch biopsies clearly reduces the proportion of HPV positive samples (95): An HPV prevalence of 69% in swab samples taken on top of the lesions from SCC, BCC, AK and seborrheic keratosis (SK) was reduced to 12% in the cleansed biopsies (94). The viral load of known HPV types is typically very low in NMSCs (91, 105, 120, 184, 186, 258), normally with less than 1 copy per cell indicating that they are not involved in the growth of the lesion. Genital cancers require continued presence of HPV for continued growth of the cells and they have at least 1 copy per cell of the causative HPV type (171, 214).

Nevertheless, recent studies have found an association of Beta-PV species 2 DNA in biopsies from SCC of the skin compared to adjacent healthy skin (odds ratio (OR), 4.0; 95% confidence interval (CI), 1.3–12.0) (21) and OR, 4.40; 95% CI, 1.92–10.06) (94) in immunocompetent individuals. An association of Beta-PV and has also been found in the Netherlands (OR, 2.8; 95% CI 1.3–5.8) using hair bulbs from plucked eye brows for detection of HPV (26). Hair bulbs are thought to be a reservoir of PV (39), and eyebrow hairs harbor persistent beta PV (55, 194). Persistence of HPV has also been observed in forehead swab samples (121).



Studies analyzing prevalence of antibodies to HPV have also found an association of Beta-PVs with SCC (26, 84, 138, 239, 255). In two of the studies, an association of HPV and BCC was investigated, but with contradictory results (84, 138). One study also included Gamma-PVs and found an association with SCC (255). There are also serological studies showing no association (13, 43), implicating that additional investigations are required to clarify the possible role of HPV in NMSC.

## Identification of novel human papillomavirus types

In the 1970s, the identification of novel PV types was a difficult process as conventional cell-culture systems do not allow reproduction of PVs (58). The first HPV types were purified using cesium-chloride gradient centrifugation followed by extraction of DNA, restriction enzyme digestion and gel electrophoresis (105, 106). Closely related types could be detected by Southern blot hybridization (186). These techniques require large amounts of viral DNA. During the last decades there has been a rapid increase in the number of putative HPV types identified by the use of polymerase chain reaction (PCR) and degenerate primers within the conserved L1 ORF (29, 56, 92, 111, 119, 220). At least 150 HPV types have been completely sequenced (30) and new types are continuously found (47, 48, 179, 252, 253). Unfortunately, this technique is limited to detection of viruses with similarity of the L1 gene to previously characterized HPV types and only a small part of the genome is obtained. Despite amplification with PCR cloning and sequencing of complete genomes can be troublesome by the low amounts of virus available in many samples.

# Staphylococcus Aureus

## Generalities

*S. aureus* is a gram-positive, nonmotile, catalase-positive coccus. The name *staphylococcus*, from the Greek *staphylē* (=a bunch of grapes), was suggested by Sir Alexander Ogston (180) more than 100 years ago. *S. aureus* appears as golden-yellow colonies when grown on blood agar plates, thereby the name aureus (gold in Latin).

The bacterium may occur as a commensal on human skin and in the nose. The carriage rate varies depending on body site, with the highest prevalence of approximately 40 % in the nose (141, 262). The prevalence on skin varies between 10 to 20% (141). There are three different types of carriers; persistent carriers, persistent non carriers and intermittent carriers (262). Approximately 10 to 20% are consistently *S. aureus* negative in swabs from the nose, another 10 to 20% are positive at every testing. The intermittent carriers can be positive for several weeks and then negative for comparable periods.

The genome of *S. aureus* consists of a single circular chromosome of 2.8 Mbp and the majority of *S. aureus* strains also carry one or more plasmids ranging from 1 to 60 kbp (170).

## Pathogenesis and virulence factors

*S. aureus* is normally not able to cause an infection in an immunocompetent person unless normal barriers have been breached by surgery or a splinter wound (248). Once inside the blood stream access and adherence to host tissue is mediated by surface receptors or adhesions. *S. aureus* produces many different surface proteins involved in cell binding e.g fibronectin-binding proteins (134) and collagen adhesions (243). *S. aureus* also produces different toxins facilitating colonization, one example being  $\alpha$ -toxin also called  $\alpha$ -hemolysin.  $\alpha$ -toxin is a pore-forming toxin that induces a wide array of cellular events in the infected epithelial cells and also in neighboring cells by diffusion of toxins (128, 223). This includes activation of nuclear factor kappa B (NF- $\kappa$ B) and up regulation of various inflammatory cytokines (69, 205). Other virulence factors are involved in the immune response to the bacterium, e.g. protein A (89, 191). Protein A facilitates the survival of *S. aureus* by inhibiting opsonization and thereby phagocytosis.

## Staphylococcus aureus associated diseases

As mentioned previously many humans are carriers of *S. aureus* and disease is not developed until *S. aureus* is spread into the blood stream and to other tissues and organs. The most common infections caused by *S. aureus* are different skin infections such as impetigo and carbuncles (70). More severe complications include septic arthritis, and staphylococcal endocarditis (infection of the heart valves) and pneumonia (248). During surgery or dialysis, carriage of *S. aureus* has been identified as a risk factor for the development of infections (141). Other complications are food poisoning and toxic shock syndrome in women (248). A tobacco tar-resistant strain of *S. aureus* has also been suggested to have a carcinogenic potential in the buccal cavity (96).

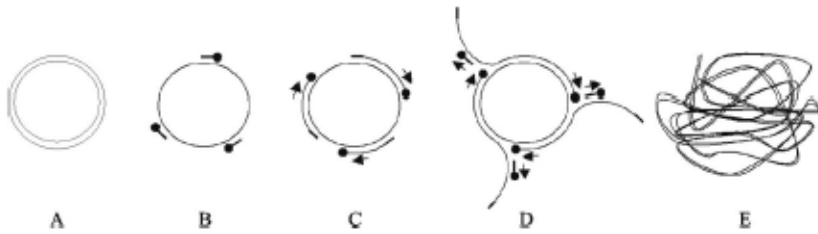
## Methods for detection of potential pathogens

Identification of previously unrecognized pathogenic agents is of great medical interest. PCR using “general” or “degenerate” primers, has been extensively used to discover HPV types, but this method limits the discovery to pathogens with partly conserved DNA. Representational differences analysis (RDA) (156) require no prior knowledge of the nucleic acid sequence. RDA is based on a sequence-independent PCR amplification with subtractive hybridization and is used to identify sequences that is present in cases but not in controls. Both human herpes virus type 8 (45) and torque teno virus (178) were discovered by RDA. Sequence-independent, single primer amplification (SISPA) is another technique enabling nucleic acids of unknown sequence to be amplified, by ligation of primers to blunt end DNA (202). SISPA has been used to discover e.g. hepatitis G virus (155) and two bovine parvovirus’s (8).

Among the techniques that have been used in this thesis two are worth attention, namely multiple displacement amplification (MDA) and high throughput sequencing.

### Multiple displacement amplification

The MDA technique is based on rolling circle amplification (88) and has been developed to amplify all DNA in a sample from a very small amount of starting material (60). MDA was initially used to amplify circular DNA (60) but has been extended to work for linear DNA as well (59, 149). The method makes use of a strand displacing DNA polymerase originating from bacteriophage phi29 (33, 99). The strand displacing activity of the enzyme gives it advantages compared to other polymerases, because when the polymerase reaches a primer it displaces it and continues to synthesize DNA (Figure 6). This activity makes the processivity high, the average product length in every primer-extension event being >10 kb (59). The enzyme is very stable and synthesis can carry on for many hours (60).



**Figure 6.** Overview of the multiple displacement amplification reaction: (A) Double stranded circular DNA. (B) Denatured single stranded DNA with annealed primers (-). Phi29 (•) binds to the primers and (C) amplification can start. (D) When Phi29 reaches a downstream primer strand displacement occurs and new primers can anneal to the displaced product. (E) The end product is double stranded repeated copies of the genome. Adapted from Rector et al., A sequence-independent strategy for detection and cloning of circular DNA virus genomes by using multiply primed rolling-circle amplification in *Journal of Virology* 2004, 78(10):4993-8 with permission from American Society for Microbiology.

### *Applications of MDA*

The MDA technique has been used for several different applications, e.g. whole genome amplification of human DNA (25, 59). The ability to increase the quantity of genomic DNA can eliminate technical problems in situations where only limited amounts of DNA is available, e.g. in forensic studies (101), even though the starting DNA has to be of good quality (24). Archival plasma and serum samples for genetic epidemiological studies can also be successfully amplified using MDA (224). MDA has been applied to amplify circular viral genomes and several novel PV types (199, 200, 236, 247, 250) as well as two polyoma viruses (133) have been detected.

### High-throughput sequencing

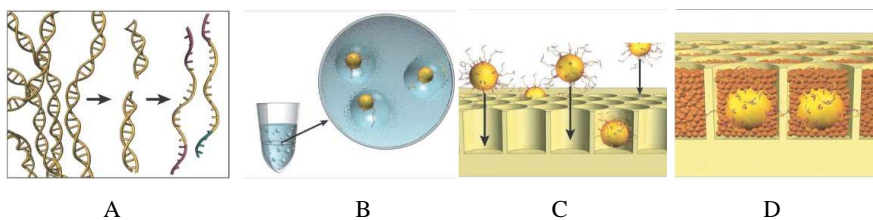
The high demand on low-cost and fast sequencing has driven the development of high-throughput sequencing forward and there are now several alternatives available e.g. Applied Biosystems SOLiD system (2), Illumina Genome Analyzer (3) and Genome sequencer (GS) FLX developed by Roche (1). Their sequence methodologies differ and have various advantages. Both SOLiD and Genome Analyzer have relatively short read lengths, 75 and 150 bp respectively, but have high throughputs of up to 20-30 Gb per day for SOLiD and 6 Gb for Genome Analyzer. The GS FLX has the longest read length with an average of 200 to 300

bp, and for the Titanium update of GS FLX 400 to 600 bp can be achieved. Even though the throughput is lower for GS FLX than for the other technologies, approximately 1 billion bases per day, the long read length makes this technology more suitable for de novo sequencing. The GX FLX has been used in this thesis and is described in more detail below.

### *High-throughput sequencing using GS FLX*

The GS FLX method is based on emulsion-based amplification and pyrosequencing. The hallmark of the technology is the PicoTiterPlate™, which allows a single instrument to produce millions of nucleotide bases per run (161).

Genomic DNA is isolated and fragmented by nebulisation, ligated to adaptors and separated into single strands. The sheared DNA strands are then blunt-ended to allow ligation of adaptors and fragments are captured on beads under conditions that favour one fragment per bead. The beads are captured within the droplets of a PCR-reaction-mixture-in-oil emulsion and PCR amplification occurs within each droplet, resulting in beads each carrying ten million copies of a unique DNA template. Emulsion is broken, DNA strands are denatured, and beads carrying single-stranded DNA clones are deposited into wells of a fibre-optic slide, the PicoTiterPlate™, containing hundreds of thousands of wells, just deep and wide enough for one DNA-containing bead. Smaller beads carrying immobilized enzymes required for pyrophosphate sequencing are deposited in each well. The DNA is sequenced using a technology known as pyrosequencing.



**Figure 7.** Overview of high-throughput sequencing using GS FLX: (A) Fragmentation of DNA and ligation of adaptors. (B) Emulsion-based PCR amplification. (C) PicoTiterplate with one DNA-containing bead per well. (D) Addition of smaller beads with enzymes required for sequencing. Adapted from Margulies et al., Genome sequencing in microfabricated high-density picolitre reactors in *Nature* 2005, 415;376-380 with permission from nature Publishing group.

### *Applications of GS FLX*

The GS FLX has been used in a variety of applications, e.g. sequencing of the human genome in less than two months (260), exploring the structural variations in humans by mapping DNA from two individuals against a reference genome (142) and sequencing of the complete genome for several large bacteria (23, 62).

In addition, this high-throughput sequencing technology has been used for several metagenomic studies (15, 53, 151, 188, 204, 231, 249). As the process does not require cloning or preamplification before sample preparation, previously unknown and unculturable organisms can be easily detected. Studies have shown an enormous diversity of microbes in the marine environment (15, 231). Most sequences found in these environmental samples were not present in the current GenBank database, predicting that the composition of microbial communities is greater than the estimation of a few thousand distinct microbes per litre of seawater. The technique has also been used to investigate the microbiome of soil (151, 204).

Using metagenomic analysis with the Genome Sequencer FLX system, researchers can quickly discover known and unknown organisms in outbreaks of infectious diseases, which could have major implications in health care. E.g. three women in Australia who received organ transplants from a single donor died of unknown causes shortly after the transplantation (188). RNA was evaluated from the transplanted organs and high-throughput sequencing identified a novel Arenavirus as the cause. Further, a novel Ebola virus was linked to an outbreak of hemorrhagic fever in Uganda (249). As the virus diverged substantially from other known ebolaviruses the study have important implications for design of effective diagnostics and vaccines. Another important discovery is that of Israeli acute paralysis virus, which is strongly correlated to colony collapse disorder (CCD) in honeybees (53). CCD is responsible for the loss of the adult bee population in colonies (181) and a decrease in healthy honey bee colonies can have severe impact in agricultural commodities that depend on insect pollination. Metagenomic sequences from healthy colonies compared to CCD hives detected candidate pathogens (53). Screening of samples collected from various sites during three years then found the association with Israeli acute paralysis virus.

# Summary of papers

## Aims

### Paper I

#### **Cutaneous human papillomavirus 88: remarkable differences in viral load.**

To analyze skin biopsies with paired controls for the presence and viral load of HPV 88 that was found at extreme high viral loads in a patient infected with HIV.

### Paper II

#### ***Staphylococcus aureus* and squamous cell carcinoma of the skin.**

To investigate if squamous cell carcinomas of the skin contain as yet unidentified HPV types or other microorganisms. As most of the detected sequences were from *Staphylococcus aureus*, different skin lesions and controls were investigated for its presence.

### Paper III

#### **Three novel papillomaviruses (HPV 109, HPV 112 and HPV 114) and their presence in cutaneous and mucosal samples.**

To characterize the complete genomes of three novel HPV types, HPV 109, 112 and 114 and analyze for presence and viral load in skin and genital samples.

### Paper IV

#### **High-throughput sequencing reveals diversity of Human Papillomaviruses in cutaneous lesions.**

To use high-throughput sequencing to determine the presence of known and previously unknown HPV types in skin lesions preamplified by degenerate PCR.



## Materials and methods

### Human papillomavirus-modified multiple displacement amplification

An in-house multiple displacement amplification (MDA) (see the section "Multiple displacement amplification"), that preferentially should amplify human papillomaviruses (HPVs), was developed. In addition to random hexamer primers, primers generic to any HPV were included. Design of the HPV primers was based on an alignment of 72 HPV types, belonging to the genera Alpha-, Beta-, Gamma-, Mu- and Nu-PVs, from the HPV database (Los Alamos, 1997). Two undecamers were chosen in the relatively conserved regions L1 and E1. The random hexamer primers and the "HPV-general" undecamer primers were thiophosphate-modified to be protected from degradation of the 3'-5' exonuclease proofreading activity of the  $\phi$ 29 DNA polymerase.

### Real-time PCR

In Paper I to III a single- or triplex real time PCR was designed to analyze presence and viral load of different HPV types. Primers and probes were designed, to only amplify the HPV type of interest. A standard curve was included in each PCR and at least 5 copies were detected in each approved run. To calculate viral load per cell, human DNA was analyzed with real time PCR using primers and probe for the  $\beta$ -globin gene.

### Paper I

HPV 88 was isolated, by the use of the in-house HPV-generic MDA, from a human immunodeficiency virus (HIV)-infected man with several extensive squamous cell carcinomas (SCCs) on fingers and on one toe. The MDA product was digested with Bam HI, separated using gel electrophoresis and cloned. HPV 88 was also visualized without preamplification on an ethidium bromide stained gel. The complete genome was sequenced using primer walking.

Patients used for screening of HPV 88 were from three different studies. Skin biopsies were collected from 362 immunocompetent patients attending a hospitalized based case control study in Swedish and Austrian hospitals (94); including 84 SCCs, 147 basal cell carcinomas (BCCs), 58 actinic keratosis (AKs) and 73 seborrhoeic keratosis (SKs). To ensure that viruses detected are from the

lesion and not from surface contamination the skin was stripped by using a tape that was attached and removed five times, followed by the same procedure with a new tape. After tape stripping a biopsy was taken from both the lesion and from adjacent healthy skin. The second series of samples were immunosuppressed patients from Austria (AK, n=5, SCC, n=21). The last patient group of both immunosuppressed (n= 38) and immunocompromised (n=21) were attending a clinic in Australia. They were diagnosed with SCC (n= 21), BCC (n=22) and AK (n= 16). All samples were extracted and DNA quality checked by real-time PCR for the  $\beta$ -globin gene.

Prevalence and copy number of HPV 88 was investigated in eight of the fingers from the HIV patient and in the three different studies using real time PCR. As the lesions also were shown to be positive for HPV 26 (116), viral loads for HPV 26 were investigated.

## Paper II

Patient samples from the same hospitalized based case control study as in Paper I were used also in Paper II with a few modifications: 82 SCCs, 142 BCCs, 57 AKs and 72 SKs were included. In addition to biopsies from the lesions and healthy skin, swab samples from the top of the lesions and from healthy skin were also used. Swab samples from the top of the lesions were collected using a cotton-tipped swab drawn back and forth over the lesion. From healthy adjacent skin the swab was drawn back and forth 15 times within an area of 5 x 5 cm. The swab was thereafter suspended in 1 ml saline.

The HPV-generic MDA was performed on 83 SCC biopsies. MDA treated samples were digested with Hinc II and analyzed by agarose gel. DNA fragments were excised, cloned and sequenced. To increase the amount of DNA in the biopsy samples, MDA using only random hexamer primers was performed. Ordinary PCR with primers against the nuc gene was used to analyze for presence of *S. aureus* in biopsies and swab samples. SYBR Green was added and a dissociation curve was carried out in a Gene Amp 5700 SDS. Samples containing a detectable product of the same melting point as the positive control *S. aureus* were confirmed by gel electrophoresis.

Odds ratio (OR) and 95% confidence interval (CI) was calculated using LogXact, version 8, and the statistical software R, version 2.7.2.

### Paper III

Three novel HPV types were cloned and sequenced in this paper:

HPV 109 was isolated from an SCC of the skin using MDA with primers generic to HPV. The MDA product was digested using Hinc II and run on a gel. A band of approximately 4 kb was excised, purified and cloned. The remainder of the genome was amplified using long PCR with primers designed within the MDA fragment. The complete genome was sequenced using primer walking. HPV 112 was isolated from a condyloma acuminata brush sample using the HPV modified MDA followed by amplification with FAP-PCR. The third type, HPV 114 was found in a CIN I lesion using a modified HPV general priming system, MGP (230), followed by typing with Luminex (215). Luminex is a bead-based multiplex genotyping method that includes type-specific probes and a universal probe. In the case of the HPV 114 index sample, only the universal probe but none of the probes for known HPV types was positive and the amplicon was sequenced. Specific primers were designed for both HPV 112 and 114 and complete genomes were amplified with long PCR. PCR products were separated on E-gel iBase Power System and cloned. The genomes were sequenced using transposons where the transposon EZ-Tn5 <TET-1> was randomly inserted into the clones and the clones were subsequently sequenced bidirectional using primers in the transposon.

Skin samples included were biopsies from different lesions and paired healthy skin from the hospitalized-based study described in Paper I: 52 SCCs, 118 BCCs, 52 AKs and 47 SKs. Genital samples from four different studies were included: From Mozambique 312 cervical cancer biopsies and 271 brush samples from controls were analyzed (175). A second series of samples came from Latvia; 431 brush samples from cervical cancer and 234 brush samples from population based controls were included (222). Also included were 1581 samples from women with atypical cells of undetermined significance (ASCUS) or cervical intraepithelial neoplasia (CIN) I attending the Swedish cervical screening program in Stockholm (63). Finally 27 brush samples from patients with condyloma acuminata were included (242). These samples were previously found to be negative for HPV with PCR using GP5/6+ primers. In total 2856 samples from the genital area and 538

cutaneous samples were analyzed. All patient samples were extracted and checked for presence of human DNA using  $\beta$ -globin PCR.

A triplex real-time PCR with minor groove binding probes was used to investigate presence and viral load of the three viruses in the patient panels.

A phylogenetic tree using MEGA 3.1 was based on an alignment of the L1 sequences of HPV 109, 112 and 114 and representative relatives.

## Paper IV

Also in this paper the cutaneous samples from the study from Sweden and Austria described in Paper I was analyzed in addition to keratoacantoma (KA) biopsies from both immunosuppressed and immunocompromised patients that were collected in Norway (93).

The samples were amplified (one by one) with the general primer pair FAP, and then mixed into three pools: 1) fresh frozen biopsies from 37 SCC lesions and 36 AK lesions, 2) fresh frozen biopsies from 92 KA lesions and 3) swab samples from the top of lesion from 86 SCCs and 92 AKs. The samples were run on a gel, purified and thereafter sent for sequencing on the GS FLX platform at KTH, Stockholm, Sweden (see the section “High-throughput sequencing using GS FLX”).

Sequences from the GS FLX platform were filtered to remove human DNA and remaining sequences were assembled and compared to GenBank to classify them as non-HPV or HPV-related.

The program Mr Bayes was used to construct a phylogenetic tree of putative new HPV types with relatives.

# Results and discussion

## Paper I

The complete genome of HPV 88 was 7 326 bp with a genomic organization typical for PVs. HPV 88 belongs to species 5 within the genus Gamma, with the closest relative being HPV 60 with a similarity of 61 %.

All eight SCCs of the HIV infected male were positive for HPV 88, with the highest viral load in the left hand fingers. The SCC that HPV 88 was isolated from contained an exceptionally high viral load of  $1.3 \times 10^6$  copies per cell, but also the other fingers of the left hand had high copy numbers varying from approximately 30 to 16 000 copies per cell. The four tumors of the right hand were also positive for HPV 88, but with lower copy number ranging from 0.1 to 1.6 copies per cell. By contrast, HPV 26 were found at high copy numbers on the right hand SCCs (56 to 44 000 copies per cell) and at lower copy number on the left hand SCCs (0.4 to 1.2 copies per cell). Comparing to cervical cancers, which normally carry at least 1 viral genome per cell (171, 214), the HPV viral load of known types in NMSCs are with a few exceptions very low (91, 105, 120, 184, 186, 258). Screening of 809 skin samples, both immunocompetent and immunosuppressed, detected only seven HPV 88 positive specimens. Five of these (SCC =4 and AK =1) were from immunosuppressed patients visiting the same Austrian clinic as the index patient. This result could indicate an association of HPV 88 with SCC in immunosuppressed patients, but as we also had a series of HPV 88 negative immunosuppressed patients from Australia it appears that HPV 88 infection is not generally associated with immunosuppression or with SCC. A possible explanation for the HPV 88 positive patients in the Austrian clinic is that the index patient could have contaminated the waiting room. The same explanation may also be applied to his own hands; HPV 88 of the left hand could have contaminated the right hand and vice versa for HPV 26. The high viral loads of HPV 88 in the SCCs of the left hand and of HPV 26 in the SCCs of right hand suggest that these viruses could be the causal agents for these SCCs.

HPV 88 was cloned using HPV generic MDA, but with the knowledge of its high copy number and the fact that it was possible to visualize HPV 88 directly on an ethidium stained gel without prior amplification, cloning without any amplification would probably have been possible.

## Paper II

Eighty-three SCC biopsies were analyzed using the HPV generic MDA for presence of HPV types and other microorganisms. Strong bands were visualized in 22 of the 83 biopsies, and 28 bands were cloned and sequenced. Half of the sequences (14 of 28) contained sequences from the human genome, another 12 were found to be different plasmids belonging to *S. aureus*. Only one sequence was a new HPV type, and the last one matched with *Escherichia coli*.

As so many of the sequences belonged to *S. aureus* we decided to investigate different skin lesions and swab samples for its presence. The highest prevalence of *S. aureus* was in biopsies from SCC lesions, 29.3%, compared to 1.4 % in SK, 12.3% in AK, 7.7% in BCC and 5.7% in biopsies from healthy skin. Presence *S. aureus* DNA in biopsies was strongly associated with SCC (OR, 6.23; 95% CI, 1.47 – 4.83) when using biopsies from healthy skin as the reference. There is a possibility that the bacterium could merely adhere to protruding growth of the skin so the same analysis was made using the benign protruding growth SK as the reference. These calculations gave an even stronger association of *S. aureus* DNA with SCC (OR, 23.84; 95% CI, 3.69-1004) and also a weak association for AK (OR, 10.01; 95% CI, 1.37-∞). As AK is considered a precursor to SCC (36, 163), this finding is of interest as it suggests an increased colonization of *S. aureus* early in the carcinogenic process. Also when considering the swab samples, the highest prevalence of *S. aureus* was on top of the SCC lesions (31.7 %) (OR, 2.67; 95% CI, 1.47-4.83). *S. aureus* is a commensal with reported prevalences on healthy skin varying from 10 to 20 % (262) which was confirmed in this study with 15 % positive swab samples taken on healthy skin. Some humans appear to be persistent carriers of *S. aureus* and thus we wanted to investigate if SCC subjects could be more susceptible to *S. aureus* infection by comparing the *S. aureus* positivity of SCC lesions to biopsies from healthy skin from the same patient with the result that *S. aureus* was more frequently detected in SCC biopsies (OR, 7.26; 95% CI, 1.38-76.55) than in the matched healthy skin biopsies. Consequently, the association cannot be explained by genetic or disease-induced overall susceptibility to bacterial colonization.

## Paper III

The complete genomes of three novel HPV types were characterized; HPV 109-7346 bp, HPV 112 – 7227 bp and HPV 114 – 8069 bp. All three types had a

genomic organization typical for HPV types except that HPV 114 lacks an E5 ORF which is normally present in genital types. Phylogenetic analysis suggests that HPV 112 constitutes a new species in genus Gamma-PVs and it was most closely related to HPV 65 with 64 % similarity. HPV 114 belongs to species alpha-3, with 84 % sequence similarity to HPV 84. The classification of HPV 109 is difficult as the bootstrap values in the phylogenetic analysis indicate low reliability of the tree. It clusters with the recently identified HPV types lacking an E6 ORF: HPV 101, 103 and 108, but the closest relative based on similarity in the L1 gene is the Gamma-type HPV 4, with a 65 % identity in the L1 gene.

The prevalences of HPV 109 and 112 were very low. HPV 109 was positive in only three specimens; the index SCC, an SK and as a co-infection with HPV 114 in a CIN I lesion. The viral loads in the SK and CIN lesion were low ( $4 \times 10^{-2}$  and  $5 \times 10^{-6}$  respectively) as for most cutaneous samples (91, 105, 120, 184, 186, 258), but the index SCC had a relatively high viral load of 11 copies per cell. HPV 112 is a rare virus and was only positive in the index condyloma, despite 2856 samples from the genital area and 538 cutaneous samples analyzed. The high viral load of 842 copies per cell in the index condyloma indicates that its presence in this lesion is probably most likely not a contamination.

HPV 114 was found in 48 of the 2856 genital samples with a viral load ranging from  $5 \times 10^{-4}$  to 240 copies per cell (mean = 11 copies per cell). The highest prevalence of HPV 114, 2.7% (42/1581), was among women with ASCUS and CIN I lesions (ASCUS, 1.8% (10/554) and CIN I, 3.1% (32/1027)). HPV 114 existed as a co-infection in 37 of 42 samples, with up to five different HPV types. The MGP-Luminex missed only one of the HPV 114 single infections indicating adequate sensitivity of detecting unknown HPV types as long as the universal probe can hybridize to the sequence.

## Paper IV

In total 19 436 reads were obtained from the three pools. The non human sequences (n=17 022) were assembled to 3898 reads of which 2196 reads, 56%, were identified as HPV related. An explanation for the non-HPV sequences found could be that the FAP primers are degenerated and also contains two inosines. In total 43 known HPV types and 67 previously described putative HPV types were detected. Putative types are subgenomic sequences of HPVs, e.g. FA- isolates

amplified using the FAP primers. Most of the HPV types were found in the pool of SCC- and AK-swab samples, 35 types and 48 putative types, compared to 29 types and 19 putative types in the pool of SCC- and AK-biopsies and 26 types and 38 putative types in the pool of KA-biopsies. The majority of the HPV-sequences found were from the genera Beta-PVs (n= 52) and Gamma-PVs (n= 55) which contain the HPV types that mostly are found in cutaneous lesions (58). However three types, HPV 3, 16 and 77, from species alpha were found as well. HPV types within the genus Alpha-PVs are normally found in the mucosa, but HPV types within alpha species 2 and 4, where HPV 3 and 77 are found, more frequently cause cutaneous than mucosal lesions (58). HPV 16 and other mucosal types have also occasionally been found in skin infections (6, 127, 145).

The SCC-, AK- and KA- biopsies had previously been tested using conventional cloning and sequencing after PCR, but still additional sequences from 60 novel putative new types were detected with the use of high-throughput sequencing. The read length for the GS FLX is 200-300 bp and consequently a complete FA-fragment cannot be sequenced from one read. For 11 types more than 400 bp was obtained and for the remaining 49 a sequence from either the 5' - (n= 26) or the 3' -end (n=23) was obtained. Most likely many of them belong to the same type but as the length of the partial sequences varied from 84 to 258 bp there were no or too short overlaps to make larger contigs. Most of the novel putative new types were found in this pool (4 with > 400 bp and 30 with a shorter sequence). In the pool with SCC-and AK-biopsies 4 novel putative types with more than 400 bp and 15 additional shorter sequences were detected and in the KA-pool 4 sequences larger than 400bp and 11 with a shorter sequence were found.

A phylogenetic tree based on 34 sequences (>200 bp from the 5'-end) clustered 23 of the novel putative types within the genus Gamma-PVs and 11 within the genus Beta-PVs.



## Concluding remarks and future perspectives

The large spectrum of different HPV types found on human skin continuously grows and in this thesis the complete genomes of three novel types, HPV 109, 112 and 114, were characterized. In addition, sequences from at least 37 novel putative types were detected. Many known HPV types have been discovered using degenerate PCR-systems, which require some similarity to previously known types. In paper I to III we tried to overcome this problem by using MDA with HPV generic primers. The HPV generic primers were designed in regions relatively conserved in 72 HPV types from genera Alpha-, Beta-, Gamma-, Mu- and Nu-PVs, and with the addition of random hexamer primers the MDA should allow amplification of any HPV type present at a sufficient amount. In paper I we identified HPV 88 in digital SCCs of an HIV infected man using this HPV modified MDA. HPV 88 is only distantly related to known HPV types and would be difficult to amplify using general HPV detection systems. A disadvantage with the MDA is that the virus probably has to be present at a relatively high copy number to be able to visualize on a gel after amplification. Both HPV 88 and HPV 109, cloned after MDA amplification, had high viral loads.

The characterization of HPV 114 expands the genus Alpha-PV and this infection was not uncommon in a large material of genital lesions. HPV 88 and 112 were classified into the genus Gamma-PV and later a phylogenetic study also clustered HPV 109 within the same genus (30). Genus Gamma-PV appears to be the genus most rapidly growing with at least 26 completely characterized types, including HPV 88, 109 and 112 from paper I and III, 97 putative types, clustered into the genus Gamma-PV in a phylogenetic study (90), and at least 23 of the novel putative HPV sequences found in paper IV. Many of the partially sequenced novel putative types were detected in samples previously tested for HPV, indicating a diversity of HPV greater than what has been revealed with conventional methods.

For future studies we believe that unbiased MDA amplification, avoiding preamplification by PCR, followed by high-throughput sequencing with the GS FLX system could increase the rate at which new HPV types are discovered. With the new high-throughput sequencing techniques it would be possible to sequence a sufficient number of cases and controls separately to provide sufficiently large amounts of epidemiological data to allow rapid establishment of whether some of these new viruses are associated with a major human disease such as NMSC.

*S. aureus* is a commensal on human skin and in the nose, but is also present in many skin infections. The association of *S. aureus* DNA with SCC of the skin found in paper II is stronger than what has been found for HPV. However the study design cannot determine the causality of the association. It would be of interest to investigate if the high prevalence of *S.aureus* in SCC is due to the ulcerating growth of the SCC or whether the bacterium could be involved in the development of cancer. A possible mechanism for *S. aureus* to contribute to the cancer development is by production of a chronic inflammation, as in the case of *Helicobacter Pylori* causing gastric cancer and Hepatitis B and C causing liver cancer. A well known tumor promoter, produced in Staphylococcal inflammation, is nuclear factor- $\kappa$ B. *S. aureus* has also been suggested to have a carcinogenic potential in the buccal cavity.

Taken together, the work in this thesis has expanded our knowledge of the wide genomic diversity of human papillomaviruses on the skin and has discovered that SCCs of the skin are also associated with *S. aureus*. Future studies aiming to elucidate a possible infectious etiology of SCC will obviously need to include an unbiased metagenomic sequencing of both viral and bacterial genomes.

# Acknowledgements

I wish to express my sincere gratitude to everyone who has been involved in this work and to family and friends who have supported me during these years.

My supervisors,

Joakim Dillner, for showing me the wonderful world of research that I love and for making me feel inspired after almost every Friday meeting.

Ola Forslund, for always taking time for discussions, helping me with laboratory work as well as writing issues.

My co-authors for fruitful collaborations.

Special thanks to:

All the past and present members of our group, especially: Agustin, Anna, Annika, Carina, for helping me with this and that, knowing so much about almost everything, Cecilia, Christina, for all help with laboratory work, Davit, co-author and sequence guru, Hanna, Helena, co-author and for all help with ultracentrifugation, Herman, Janka, Kia, for being helpful in the lab, Kristin, Malin, Natasa, for being such a good travel companion, Vesna and Zoltan. You all make life at work fun, with cake fridays/thursdays and interesting discussions at lunch-time.

My students, Carly, Caroline, Carl and Martin.

Maria Anderberg, for all fun times, in and outside work, for all help with e.g. excel-problems.

Sophia Harlid, my new roommate, for many discussions both regarding science and the life outside.

Helena Person and Anna Olofsson Franzoia, for all administrative help and for always being so kind.

Margareta, Anki, Nasida and Lars-Göran for always being so helpful with preparing laboratory stuff and ordering supplies.

Members of the “tjejmiddag” Dr Tina Hazard, for helping when I came to the lab knowing nothing and for all fun we have had during the years, both at the lab, conferences and outside work, Jenny and Margareta for all delicious food and fun conversations.

My friends scattered all over Sweden (and Germany) for all fun times.

My parents, Lotta och Kjell, for supporting and believing in me and for always being there for me.

My sister and best friend, Malin, for all fun we have together and for being the best god-mother for Isak.

My little family,

Isak, my wonderful son, for making me so happy every day.

Mats, the love of my life, for making me believe in myself, without you I would not be where I am today!

This work was supported by the Swedish Cancer Society and the Swedish Research Council, the European Union FP5 project VIRASKIN and the Swedish Center for Metagenomic Sequencing.

# References

1. Roche, 454. <http://454.com/products-solutions/454-sequencing-system-portfolio.asp>, accessed December 29th, 2010.
2. Applied Biosystems. <http://www.appliedbiosystems.com/absite/us/en/home/applications-technologies/solid-next-generation-sequencing/next-generation-systems.html>, Accessed December 29, 2010.
3. Illumina. [http://www.illumina.com/systems/genome\\_analyzer\\_iix.ilmn](http://www.illumina.com/systems/genome_analyzer_iix.ilmn), Accessed December 29th, 2010.
4. 1995. Human Papillomaviruses. IARC monographs on assessment of carcinogenic risk to humans; IARC, Lyon, France **64**.
5. **Abbate, E. A., J. M. Berger, and M. R. Botchan.** 2004. The X-ray structure of the papillomavirus helicase in complex with its molecular matchmaker E2. *Genes Dev* **18**:1981-96.
6. **Alam, M., J. B. Caldwell, and Y. D. Eliezri.** 2003. Human papillomavirus-associated digital squamous cell carcinoma: literature review and report of 21 new cases. *J Am Acad Dermatol* **48**:385-93.
7. **Alam, M., and D. Ratner.** 2001. Cutaneous squamous-cell carcinoma. *N Engl J Med* **344**:975-83.
8. **Allander, T., S. U. Emerson, R. E. Engle, R. H. Purcell, and J. Bukh.** 2001. A virus discovery method incorporating DNase treatment and its application to the identification of two bovine parvovirus species. *Proc Natl Acad Sci U S A* **98**:11609-14.
9. **Alter, H. J., R. H. Purcell, J. W. Shih, J. C. Melpolder, M. Houghton, Q. L. Choo, and G. Kuo.** 1989. Detection of antibody to hepatitis C virus in prospectively followed transfusion recipients with acute and chronic non-A, non-B hepatitis. *N Engl J Med* **321**:1494-500.
10. **Ambinder, R. F.** 2000. Gammaherpesviruses and "Hit-and-Run" oncogenesis. *Am J Pathol* **156**:1-3.
11. **Ambrose, H. E., and J. P. Clewley.** 2006. Virus discovery by sequence-independent genome amplification. *Rev Med Virol* **16**:365-83.
12. **Ames, A., and P. Gravitt.** 2007. Human papillomavirus vaccine update. *Curr Infect Dis Rep* **9**:151-8.
13. **Andersson, K., T. Waterboer, R. Kirnbauer, K. Slupetzky, T. Iftner, E. M. de Villiers, O. Forslund, M. Pawlita, and J. Dillner.** 2008. Seroreactivity to cutaneous human papillomaviruses among patients with nonmelanoma skin cancer or benign skin lesions. *Cancer Epidemiol Biomarkers Prev* **17**:189-95.

14. **Andres, C., B. Belloni, U. Puchta, C. A. Sander, and M. J. Flaig.** 2009. Prevalence of MCPyV in Merkel cell carcinoma and non-MCC tumors. *J Cutan Pathol*.
15. **Angly, F. E., B. Felts, M. Breitbart, P. Salamon, R. A. Edwards, C. Carlson, A. M. Chan, M. Haynes, S. Kelley, H. Liu, J. M. Mahaffy, J. E. Mueller, J. Nulton, R. Olson, R. Parsons, S. Rayhawk, C. A. Suttle, and F. Rohwer.** 2006. The marine viromes of four oceanic regions. *PLoS Biol* **4**:e368.
16. **Antonsson, A., C. Erfurt, K. Hazard, V. Holmgren, M. Simon, A. Kataoka, S. Hossain, C. Hakangard, and B. G. Hansson.** 2003. Prevalence and type spectrum of human papillomaviruses in healthy skin samples collected in three continents. *J Gen Virol* **84**:1881-6.
17. **Antonsson, A., O. Forslund, H. Ekberg, G. Sterner, and B. Hansson.** 2000. The ubiquity and impressive genomic diversity of human skin papillomaviruses suggest a commensalic nature of these viruses. *Journal of Virology* **74**:11636-11641.
18. **Antonsson, A., S. Karanfilovska, P. G. Lindqvist, and B. G. Hansson.** 2003. General acquisition of human papillomavirus infections of skin occurs in early infancy. *J Clin Microbiol* **41**:2509-14.
19. **Appleby, P., V. Beral, A. Berrington de Gonzalez, D. Colin, S. Franceschi, A. Goodill, J. Green, J. Peto, M. Plummer, and S. Sweetland.** 2006. Carcinoma of the cervix and tobacco smoking: collaborative reanalysis of individual data on 13,541 women with carcinoma of the cervix and 23,017 women without carcinoma of the cervix from 23 epidemiological studies. *Int J Cancer* **118**:1481-95.
20. **Apt, D., T. Chong, Y. Liu, and H. U. Bernard.** 1993. Nuclear factor I and epithelial cell-specific transcription of human papillomavirus type 16. *J Virol* **67**:4455-63.
21. **Asgari, M. M., N. B. Kiviat, C. W. Critchlow, J. E. Stern, Z. B. Argenyi, G. J. Raugi, D. Berg, P. B. Odland, S. E. Hawes, and E. M. de Villiers.** 2008. Detection of human papillomavirus DNA in cutaneous squamous cell carcinoma among immunocompetent individuals. *J Invest Dermatol* **128**:1409-17.
22. **Astori, G., D. Lavergne, C. Benton, B. Hockmayr, K. Egawa, C. Garbe, and E. M. de Villiers.** 1998. Human papillomaviruses are commonly found in normal skin of immunocompetent hosts. *J Invest Dermatol* **110**:752-5.
23. **Baltrus, D. A., M. R. Amieva, A. Covacci, T. M. Lowe, D. S. Merrell, K. M. Ottemann, M. Stein, N. R. Salama, and K. Guillemin.** 2009. The complete genome sequence of *Helicobacter pylori* strain G27. *J Bacteriol* **191**:447-8.
24. **Barber, A. L., and D. R. Foran.** 2006. The utility of whole genome amplification for typing compromised forensic samples. *J Forensic Sci* **51**:1344-9.
25. **Barker, D. L., M. S. Hansen, A. F. Faruqi, D. Giannola, O. R. Irsula, R. S. Lasken, M. Latterich, V. Makarov, A. Oliphant, J. H. Pinter, R. Shen, I. Sleptsova, W. Ziehler, and E. Lai.** 2004. Two methods of whole-genome amplification enable accurate genotyping across a 2320-SNP linkage panel. *Genome Res* **14**:901-7.
26. **Bavinck, J. N., R. E. Neale, D. Abeni, S. Euvrard, A. C. Green, C. A. Harwood, M. N. de Koning, L. Naldi, I. Nindl, M. Pawlita, H. Pfister, C. M.**

- Proby, W. G. Quint, J. Ter Schegget, T. Waterboer, S. Weissenborn, and M. C. O. B. Feltkamp.** Multicenter Study of the Association between Betapapillomavirus Infection and Cutaneous Squamous Cell Carcinoma. *Cancer Res* **70**:9777-9786.
27. **Belnap, D. M., N. H. Olson, N. M. Cladel, W. W. Newcomb, J. C. Brown, J. W. Kreider, N. D. Christensen, and T. S. Baker.** 1996. Conserved features in papillomavirus and polyomavirus capsids. *J Mol Biol* **259**:249-63.
28. **Berg, D., and C. C. Otley.** 2002. Skin cancer in organ transplant recipients: Epidemiology, pathogenesis, and management. *J Am Acad Dermatol* **47**:1-17; quiz 18-20.
29. **Berkhout, R. J., L. M. Tieben, H. L. Smits, J. N. Bavinck, B. J. Vermeer, and J. ter Schegget.** 1995. Nested PCR approach for detection and typing of epidermodysplasia verruciformis-associated human papillomavirus types in cutaneous cancers from renal transplant recipients. *J. Clin. Microbiol.* **33**:690-695.
30. **Bernard, H. U., R. D. Burk, Z. Chen, K. van Doorslaer, H. Hausen, and E. M. de Villiers.** Classification of papillomaviruses (PVs) based on 189 PV types and proposal of taxonomic amendments. *Virology* **401**:70-9.
31. **Bernard, H. U., I. E. Calleja-Macias, and S. T. Dunn.** 2006. Genome variation of human papillomavirus types: phylogenetic and medical implications. *Int J Cancer* **118**:1071-6.
32. **Birkeland, S. A., H. H. Storm, L. U. Lamm, L. Barlow, I. Blohme, B. Forsberg, B. Eklund, O. Fjeldborg, M. Friedberg, L. Frodin, and et al.** 1995. Cancer risk after renal transplantation in the Nordic countries, 1964-1986. *Int J Cancer* **60**:183-9.
33. **Blanco, L., A. Bernad, J. M. Lazaro, G. Martin, C. Garmendia, and M. Salas.** 1989. Highly efficient DNA synthesis by the phage phi 29 DNA polymerase. Symmetrical mode of DNA replication. *J Biol Chem* **264**:8935-40.
34. **Blumberg, B. S., B. Larouze, W. T. London, B. Werner, J. E. Hesser, I. Millman, G. Saimot, and M. Payet.** 1975. The relation of infection with the hepatitis B agent to primary hepatic carcinoma. *Am J Pathol* **81**:669-82.
35. **Botelho, M. C., J. C. Machado, and J. M. da Costa.** Schistosoma haematobium and bladder cancer: what lies beneath? *Virulence* **1**:84-7.
36. **Boukamp, P.** 2005. Non-melanoma skin cancer: what drives tumor development and progression? *Carcinogenesis* **26**:1657-67.
37. **Boukamp, P.** 2005. UV-induced skin cancer: similarities--variations. *J Dtsch Dermatol Ges* **3**:493-503.
38. **Bousarghin, L., A. Touze, P. Y. Sizaret, and P. Coursaget.** 2003. Human papillomavirus types 16, 31, and 58 use different endocytosis pathways to enter cells. *J Virol* **77**:3846-50.
39. **Boxman, I. L., R. J. Berkhout, L. H. Mulder, M. C. Wolkers, J. N. Bouwes Bavinck, B. J. Vermeer, and J. ter Schegget.** 1997. Detection of human papillomavirus DNA in plucked hairs from renal transplant recipients and healthy volunteers. *J Invest Dermatol* **108**:712-5.

40. **Boyle, J., R. M. MacKie, J. D. Briggs, B. J. Junor, and T. C. Aitchison.** 1984. Cancer, warts, and sunshine in renal transplant patients. A case-control study. *Lancet* **1**:702-5.
41. **Buck, C. B., N. Cheng, C. D. Thompson, D. R. Lowy, A. C. Steven, J. T. Schiller, and B. L. Trus.** 2008. Arrangement of L2 within the papillomavirus capsid. *J Virol* **82**:5190-7.
42. **Carter, J. J., K. G. Paulson, G. C. Wipf, D. Miranda, M. M. Madeleine, L. G. Johnson, B. D. Lemos, S. Lee, A. H. Warcola, J. G. Iyer, P. Nghiem, and D. A. Galloway.** 2009. Association of Merkel cell polyomavirus-specific antibodies with Merkel cell carcinoma. *J Natl Cancer Inst* **101**:1510-22.
43. **Casabonne, D., A. Lally, L. Mitchell, K. M. Michael, T. Waterboer, M. Pawlita, B. Imko-Walczuk, F. Wojnarowska, C. Proby, C. Harwood, and R. Newton.** 2009. A case-control study of cutaneous squamous cell carcinoma among Caucasian organ transplant recipients: the role of antibodies against human papillomavirus and other risk factors. *Int J Cancer* **125**:1935-45.
44. **Chan, W. K., T. Chong, H. U. Bernard, and G. Klock.** 1990. Transcription of the transforming genes of the oncogenic human papillomavirus-16 is stimulated by tumor promoters through AP1 binding sites. *Nucleic Acids Res* **18**:763-9.
45. **Chang, Y., E. Cesarman, M. S. Pessin, F. Lee, J. Culpepper, D. M. Knowles, and P. S. Moore.** 1994. Identification of herpesvirus-like DNA sequences in AIDS-associated Kaposi's sarcoma. *Science* **266**:1865-9.
46. **Chen, S. L., and P. Mounts.** 1990. Transforming activity of E5a protein of human papillomavirus type 6 in NIH 3T3 and C127 cells. *J Virol* **64**:3226-33.
47. **Chen, Z., M. Schiffman, R. Herrero, and R. D. Burk.** 2007. Identification and characterization of two novel human papillomaviruses (HPVs) by overlapping PCR: HPV102 and HPV106. *J Gen Virol* **88**:2952-5.
48. **Chen, Z., M. Schiffman, R. Herrero, R. Desalle, and R. D. Burk.** 2007. Human papillomavirus (HPV) types 101 and 103 isolated from cervicovaginal cells lack an E6 open reading frame (ORF) and are related to gamma-papillomaviruses. *Virology* **360**:447-53.
49. **Cherpelis, B. S., C. Marcusen, and P. G. Lang.** 2002. Prognostic factors for metastasis in squamous cell carcinoma of the skin. *Dermatol Surg* **28**:268-73.
50. **Chiang, C. M., T. R. Broker, and L. T. Chow.** 1991. An E1M--E2C fusion protein encoded by human papillomavirus type 11 is a sequence-specific transcription repressor. *J Virol* **65**:3317-29.
51. **Clifford, G. M., J. S. Smith, M. Plummer, N. Munoz, and S. Franceschi.** 2003. Human papillomavirus types in invasive cervical cancer worldwide: a meta-analysis. *Br J Cancer* **88**:63-73.
52. **Conger, K. L., J. S. Liu, S. R. Kuo, L. T. Chow, and T. S. Wang.** 1999. Human papillomavirus DNA replication. Interactions between the viral E1 protein and two subunits of human dna polymerase alpha/primase. *J Biol Chem* **274**:2696-705.
53. **Cox-Foster, D. L., S. Conlan, E. C. Holmes, G. Palacios, J. D. Evans, N. A. Moran, P. L. Quan, T. Briese, M. Hornig, D. M. Geiser, V. Martinson, D. vanEngelsdorp, A. L. Kalkstein, A. Drysdale, J. Hui, J. Zhai, L. Cui, S. K. Hutchison, J. F. Simons, M. Egholm, J. S. Pettis, and W. I. Lipkin.** 2007. A



- metagenomic survey of microbes in honey bee colony collapse disorder. *Science* **318**:283-7.
54. **Day, P. M., D. R. Lowy, and J. T. Schiller.** 2003. Papillomaviruses infect cells via a clathrin-dependent pathway. *Virology* **307**:1-11.
  55. **de Koning, M. N., L. Struijk, J. N. Bavinck, B. Kleter, J. ter Schegget, W. G. Quint, and M. C. Feltkamp.** 2007. Betapapillomaviruses frequently persist in the skin of healthy individuals. *J Gen Virol* **88**:1489-95.
  56. **de Roda Husman, A. M., J. M. Walboomers, A. J. van den Brule, C. J. Meijer, and P. J. Snijders.** 1995. The use of general primers GP5 and GP6 elongated at their 3' ends with adjacent highly conserved sequences improves human papillomavirus detection by PCR. *J Gen Virol* **76 (Pt 4)**:1057-62.
  57. **de Sanjose, S., M. Diaz, X. Castellsague, G. Clifford, L. Bruni, N. Munoz, and F. X. Bosch.** 2007. Worldwide prevalence and genotype distribution of cervical human papillomavirus DNA in women with normal cytology: a meta-analysis. *Lancet Infect Dis* **7**:453-9.
  58. **de Villiers, E. M., C. Fauquet, T. R. Broker, H. U. Bernard, and H. zur Hausen.** 2004. Classification of papillomaviruses. *Virology* **324**:17-27.
  59. **Dean, F. B., S. Hosono, L. Fang, X. Wu, A. F. Faruqi, P. Bray-Ward, Z. Sun, Q. Zong, Y. Du, J. Du, M. Driscoll, W. Song, S. F. Kingsmore, M. Egholm, and R. S. Lasken.** 2002. Comprehensive human genome amplification using multiple displacement amplification. *Proc Natl Acad Sci U S A* **99**:5261-6.
  60. **Dean, F. B., J. R. Nelson, T. L. Giesler, and R. S. Lasken.** 2001. Rapid amplification of plasmid and phage DNA using Phi 29 DNA polymerase and multiply-primed rolling circle amplification. *Genome Res* **11**:1095-9.
  61. **Delwart, E. L.** 2007. Viral metagenomics. *Rev Med Virol* **17**:115-31.
  62. **Di Bonaventura, M. P., R. DeSalle, M. Pop, N. Nagarajan, D. H. Figurski, D. H. Fine, J. B. Kaplan, and P. J. Planet.** 2009. Complete genome sequence of *Aggregatibacter (Haemophilus) aphrophilus* NJ8700. *J Bacteriol* **191**:4693-4.
  63. **Dillner, L., L. Kemetli, K. Elfgrén, G. Bogdanovic, P. Andersson, A. Carlsten-Thor, S. Andersson, E. Persson, E. Rylander, L. Grillner, J. Dillner, and S. Tornberg.** Randomized healthservices study of human papillomavirus-based management of low-grade cytological abnormalities. *Int J Cancer*.
  64. **DiMaio, D., D. Guralski, and J. T. Schiller.** 1986. Translation of open reading frame E5 of bovine papillomavirus is required for its transforming activity. *Proc Natl Acad Sci U S A* **83**:1797-801.
  65. **DiMaio, D., and D. Mattoon.** 2001. Mechanisms of cell transformation by papillomavirus E5 proteins. *Oncogene* **20**:7866-73.
  66. **Dinehart, S. M., and S. V. Pollack.** 1989. Metastases from squamous cell carcinoma of the skin and lip. An analysis of twenty-seven cases. *J Am Acad Dermatol* **21**:241-8.
  67. **Doorbar, J.** 2005. The papillomavirus life cycle. *J Clin Virol* **32 Suppl 1**:S7-15.
  68. **Dostatni, N., F. Thierry, and M. Yaniv.** 1988. A dimer of BPV-1 E2 containing a protease resistant core interacts with its DNA target. *Embo J* **7**:3807-16.
  69. **Dragneva, Y., C. D. Anuradha, A. Valeva, A. Hoffmann, S. Bhakdi, and M. Husmann.** 2001. Subcytotoxic attack by staphylococcal alpha-toxin activates NF-kappaB and induces interleukin-8 production. *Infect Immun* **69**:2630-5.

70. **Dryden, M. S.** 2009. Skin and soft tissue infection: microbiology and epidemiology. *Int J Antimicrob Agents* **34 Suppl 1**:S2-7.
71. **Dupin, N.** 2004. Genital warts. *Clin Dermatol* **22**:481-6.
72. **Durst, M., L. Gissmann, H. Ikenberg, and H. zur Hausen.** 1983. A papillomavirus DNA from a cervical carcinoma and its prevalence in cancer biopsy samples from different geographic regions. *Proc Natl Acad Sci U S A* **80**:3812-5.
73. **Egawa, K., H. Delius, T. Matsukura, M. Kawashima, and E. M. de Villiers.** 1993. Two novel types of human papillomavirus, HPV 63 and HPV 65: comparisons of their clinical and histological features and DNA sequences to other HPV types. *Virology* **194**:789-99.
74. **Egawa, K., R. Kimmel, and E. M. De Villiers.** 2005. A novel type of human papillomavirus (HPV 95): comparison with infections of closely related human papillomavirus types. *Br J Dermatol* **153**:688-9.
75. **Ekstrom, J., O. Forslund, and J. Dillner.** 2010. Three novel papillomaviruses (HPV109, HPV112 and HPV114) and their presence in cutaneous and mucosal samples. *Virology* **397**:331-6.
76. **Engels, E. A., R. J. Biggar, H. I. Hall, H. Cross, A. Crutchfield, J. L. Finch, R. Grigg, T. Hylton, K. S. Pawlish, T. S. McNeel, and J. J. Goedert.** 2008. Cancer risk in people infected with human immunodeficiency virus in the United States. *Int J Cancer* **123**:187-94.
77. **Epstein, M. A., G. Henle, B. G. Achong, and Y. M. Barr.** 1965. Morphological and Biological Studies on a Virus in Cultured Lymphoblasts from Burkitt's Lymphoma. *J Exp Med* **121**:761-70.
78. **Euvrard, S., J. Kanitakis, C. Pouteil-Noble, G. Dureau, J. L. Touraine, M. Faure, A. Claudy, and J. Thivolet.** 1995. Comparative epidemiologic study of premalignant and malignant epithelial cutaneous lesions developing after kidney and heart transplantation. *J Am Acad Dermatol* **33**:222-9.
79. **Evander, M., K. Edlund, A. Gustafsson, M. Jonsson, R. Karlsson, E. Rylander, and G. Wadell.** 1995. Human papillomavirus infection is transient in young women: a population-based cohort study. *Journal of Infectious Diseases* **171**:1026-1030.
80. **Fang, L., L. R. Budgeon, J. Doorbar, E. R. Briggs, and M. K. Howett.** 2006. The human papillomavirus type 11 E1/E4 protein is not essential for viral genome amplification. *Virology* **351**:271-9.
81. **Favre, M., S. Obalek, S. Jablonska, and G. Orth.** 1989. Human papillomavirus (HPV) type 50, a type associated with epidermodysplasia verruciformis (EV) and only weakly related to other EV-specific HPVs. *J Virol* **63**:4910.
82. **Favre, M., G. Orth, S. Majewski, S. Baloul, A. Pura, and S. Jablonska.** 1998. Psoriasis: A possible reservoir for human papillomavirus type 5, the virus associated with skin carcinomas of epidermodysplasia verruciformis. *J. Invest. Dermatol.* **110**:311-317.
83. **Fehrmann, F., D. J. Klumpp, and L. A. Laimins.** 2003. Human papillomavirus type 31 E5 protein supports cell cycle progression and activates late viral functions upon epithelial differentiation. *J Virol* **77**:2819-31.

84. **Feltkamp, M. C., R. Broer, F. M. di Summa, L. Struijk, E. van der Meijden, B. P. Verlaan, R. G. Westendorp, J. ter Schegget, W. J. Spaan, and J. N. Bouwes Bavinck.** 2003. Seroreactivity to epidermodysplasia verruciformis-related human papillomavirus types is associated with nonmelanoma skin cancer. *Cancer Res* **63**:2695-700.
85. **Feng, H., M. Shuda, Y. Chang, and P. S. Moore.** 2008. Clonal integration of a polyomavirus in human Merkel cell carcinoma. *Science* **319**:1096-100.
86. **Ferlay, J., H. R. Shin, F. Bray, D. Forman, C. Mathers, and D. M. Parkin.** Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int J Cancer*.
87. **Finnen, R. L., K. D. Erickson, X. S. Chen, and R. L. Garcea.** 2003. Interactions between papillomavirus L1 and L2 capsid proteins. *J Virol* **77**:4818-26.
88. **Fire, A., and S. Q. Xu.** 1995. Rolling replication of short DNA circles. *Proc Natl Acad Sci U S A* **92**:4641-5.
89. **Forsgren, A., and J. Sjoquist.** 1966. "Protein A" from *S. aureus*. I. Pseudo-immune reaction with human gamma-globulin. *J Immunol* **97**:822-7.
90. **Forslund, O.** 2007. Genetic diversity of cutaneous human papillomaviruses. *J Gen Virol* **88**:2662-9.
91. **Forslund, O., A. Antonsson, G. Higgins, H. Ly, H. Delius, A. Hunziker, and E. M. de Villiers.** 2003. Nucleotide sequence and phylogenetic classification of candidate human papilloma virus type 92. *Virology* **312**:255-60.
92. **Forslund, O., A. Antonsson, P. Nordin, B. Stenquist, and B. G. Hansson.** 1999. A broad range of human papillomavirus types detected with a general PCR method suitable for analysis of cutaneous tumours and normal skin. *J Gen Virol* **80 ( Pt 9)**:2437-43.
93. **Forslund, O., P. M. DeAngelis, M. Beigi, A. R. Schjolberg, and O. P. Clausen.** 2003. Identification of human papillomavirus in keratoacanthomas. *J Cutan Pathol* **30**:423-9.
94. **Forslund, O., T. Iftner, K. Andersson, B. Lindelöf, E. Hradil, P. Nordin, B. Stenquist, O. Kirnbauer, J. Dillner, and E.-M. deVilliers.** 2007. Cutaneous human papillomaviruses found in sun-exposed skin: Beta-papillomavirus species 2 predominates in squamous cell carcinoma. *Journal of Infectious Diseases* **196**:876-883.
95. **Forslund, O., B. Lindelof, E. Hradil, P. Nordin, B. Stenquist, R. Kirnbauer, K. Slupetzky, and J. Dillner.** 2004. High prevalence of cutaneous human papillomavirus DNA on the top of skin tumors but not in "Stripped" biopsies from the same tumors. *J Invest Dermatol* **123**:388-94.
96. **Fujiki, H., H. Takeuchi, N. Nishitani, H. Yamanaka, K. Suzuki, M. Kurusu, and M. Suganuma.** 2004. Carcinogenic potential of tobacco tar-resistant *Staphylococcus aureus* in buccal cavity. *J Cancer Res Clin Oncol* **130**:301-5.
97. **Gage, J. R., C. Meyers, and F. O. Wettstein.** 1990. The E7 proteins of the nononcogenic human papillomavirus type 6b (HPV-6b) and of the oncogenic HPV-16 differ in retinoblastoma protein binding and other properties. *J Virol* **64**:723-30.

98. **Garcia-Vallve, S., A. Alonso, and I. G. Bravo.** 2005. Papillomaviruses: different genes have different histories. *Trends Microbiol* **13**:514-21.
99. **Garmendia, C., A. Bernad, J. A. Esteban, L. Blanco, and M. Salas.** 1992. The bacteriophage phi 29 DNA polymerase, a proofreading enzyme. *J Biol Chem* **267**:2594-9.
100. **Genovese, N. J., N. S. Banerjee, T. R. Broker, and L. T. Chow.** 2008. Casein kinase II motif-dependent phosphorylation of human papillomavirus E7 protein promotes p130 degradation and S-phase induction in differentiated human keratinocytes. *J Virol* **82**:4862-73.
101. **Giardina, E., I. Pietrangeli, C. Martone, S. Zampatti, P. Marsala, L. Gabriele, O. Ricci, G. Solla, P. Asili, G. Arcudi, A. Spinella, and G. Novelli.** 2009. Whole genome amplification and real-time PCR in forensic casework. *BMC Genomics* **10**:159.
102. **Giles, G. G., R. Marks, and P. Foley.** 1988. Incidence of non-melanocytic skin cancer treated in Australia. *Br Med J (Clin Res Ed)* **296**:13-7.
103. **Giri, I., and M. Yaniv.** 1988. Structural and mutational analysis of E2 trans-activating proteins of papillomaviruses reveals three distinct functional domains. *Embo J* **7**:2823-9.
104. **Giroglou, T., L. Florin, F. Schafer, R. E. Streeck, and M. Sapp.** 2001. Human papillomavirus infection requires cell surface heparan sulfate. *J Virol* **75**:1565-70.
105. **Gissmann, L., H. Pfister, and H. Zur Hausen.** 1977. Human papilloma viruses (HPV): characterization of four different isolates. *Virology* **76**:569-80.
106. **Gissmann, L., and H. zur Hausen.** 1976. Human papilloma virus DNA: physical mapping and genetic heterogeneity. *Proc Natl Acad Sci U S A* **73**:1310-3.
107. **Gloss, B., and H. U. Bernard.** 1990. The E6/E7 promoter of human papillomavirus type 16 is activated in the absence of E2 proteins by a sequence-aberrant Sp1 distal element. *J Virol* **64**:5577-84.
108. **Gonzalez, S. L., M. Stremlau, X. He, J. R. Basile, and K. Munger.** 2001. Degradation of the retinoblastoma tumor suppressor by the human papillomavirus type 16 E7 oncoprotein is important for functional inactivation and is separable from proteasomal degradation of E7. *J Virol* **75**:7583-91.
109. **Goon, P., C. Sonnex, P. Jani, M. Stanley, and H. Sudhoff.** 2008. Recurrent respiratory papillomatosis: an overview of current thinking and treatment. *Eur Arch Otorhinolaryngol* **265**:147-51.
110. **Gottschling, M., M. Goker, A. Kohler, M. D. Lehmann, E. Stockfleth, and I. Nindl.** 2009. Cutaneotropic human beta-/gamma-papillomaviruses are rarely shared between family members. *J Invest Dermatol* **129**:2427-34.
111. **Gravitt, P. E., C. L. Peyton, T. Q. Alessi, C. M. Wheeler, F. Coutlee, A. Hildesheim, M. H. Schiffman, D. R. Scott, and R. J. Apple.** 2000. Improved amplification of genital human papillomaviruses. *J Clin Microbiol* **38**:357-61.
112. **Greer, C. E., C. M. Wheeler, M. B. Ladner, K. Beutner, M. Y. Coyne, H. Liang, A. Langenberg, T. S. Yen, and R. Ralston.** 1995. Human papillomavirus (HPV) type distribution and serological response to HPV type 6 virus-like particles in patients with genital warts. *J Clin Microbiol* **33**:2058-63.

113. **Grulich, A. E., M. T. van Leeuwen, M. O. Falster, and C. M. Vajdic.** 2007. Incidence of cancers in people with HIV/AIDS compared with immunosuppressed transplant recipients: a meta-analysis. *Lancet* **370**:59-67.
114. **Hagensee, M. E., N. Yaegashi, and D. A. Galloway.** 1993. Self-assembly of human papillomavirus type 1 capsids by expression of the L1 protein alone or by coexpression of the L1 and L2 capsid proteins. *J Virol* **67**:315-22.
115. **Han, Y., Y. M. Loo, K. T. Militello, and T. Melendy.** 1999. Interactions of the papovavirus DNA replication initiator proteins, bovine papillomavirus type 1 E1 and simian virus 40 large T antigen, with human replication protein A. *J Virol* **73**:4899-907.
116. **Handisurya, A., A. Rieger, A. Bankier, A. Koller, A. Salat, G. Stingl, and R. Kirnbauer.** 2007. Human papillomavirus type 26 infection causing multiple invasive squamous cell carcinomas of the fingernails in an AIDS patient under highly active antiretroviral therapy. *Br J Dermatol* **157**:788-94.
117. **Hartvelt, M. M., J. N. Bavinck, A. M. Kootte, B. J. Vermeer, and J. P. Vandenbroucke.** 1990. Incidence of skin cancer after renal transplantation in The Netherlands. *Transplantation* **49**:506-9.
118. **Harwood, C. A., and C. M. Proby.** 2002. Human papillomaviruses and non-melanoma skin cancer. *Curr Opin Infect Dis* **15**:101-14.
119. **Harwood, C. A., P. J. Spink, T. Suretheran, I. M. Leigh, E. M. de Villiers, J. M. McGregor, C. M. Proby, and J. Breuer.** 1999. Degenerate and nested PCR: a highly sensitive and specific method for detection of human papillomavirus infection in cutaneous warts. *J Clin Microbiol* **37**:3545-55.
120. **Hazard, K., L. Eliasson, J. Dillner, and O. Forslund.** 2006. Subtype HPV38b[FA125] demonstrates heterogeneity of human papillomavirus type 38. *Int J Cancer* **119**:1073-7.
121. **Hazard, K., A. Karlsson, K. Andersson, H. Ekberg, J. Dillner, and O. Forslund.** 2007. Cutaneous Human Papillomaviruses persist on healthy skin. *Journal of Investigative Dermatology*.
122. **Henle, G., and W. Henle.** 1966. Immunofluorescence in cells derived from Burkitt's lymphoma. *J Bacteriol* **91**:1248-56.
123. **Hill, A. B.** 1965. The Environment and Disease: Association or Causation? *Proc R Soc Med* **58**:295-300.
124. **Ho, G. Y., R. D. Burk, S. Klein, A. S. Kadish, C. J. Chang, P. Palan, J. Basu, R. Tachezy, R. Lewis, and S. Romney.** 1995. Persistent genital human papillomavirus infection as a risk factor for persistent cervical dysplasia. *J. Natl. Cancer Inst.* **87**:1365-1371.
125. **Hoffmann, R., B. Hirt, V. Bechtold, P. Beard, and K. Raj.** 2006. Different modes of human papillomavirus DNA replication during maintenance. *J Virol* **80**:4431-9.
126. **Hsu, J. Y., A. C. Chen, A. Keleher, N. A. McMillan, and A. Antonsson.** 2009. Shared and persistent asymptomatic cutaneous human papillomavirus infections in healthy skin. *J Med Virol* **81**:1444-9.
127. **Iftner, A., S. J. Klug, C. Garbe, A. Blum, A. Stancu, S. P. Wilczynski, and T. Iftner.** 2003. The prevalence of human papillomavirus genotypes in

- nonmelanoma skin cancers of nonimmunosuppressed individuals identifies high-risk genital types as possible risk factors. *Cancer Res* **63**:7515-9.
128. **Iwatsuki, K., O. Yamasaki, S. Morizane, and T. Oono.** 2006. Staphylococcal cutaneous infections: invasion, evasion and aggression. *J Dermatol Sci* **42**:203-14.
  129. **Jablonska, S., J. Dabrowski, and K. Jakubowicz.** 1972. Epidermodysplasia verruciformis as a model in studies on the role of papovaviruses in oncogenesis. *Cancer Res* **32**:583-9.
  130. **Jablonska, S., S. Majewski, S. Obalek, and G. Orth.** 1997. Cutaneous warts. *Clin Dermatol* **15**:309-19.
  131. **Jablonska, S., and G. Orth.** 1985. Epidermodysplasia verruciformis. *Clin Dermatol* **3**:83-96.
  132. **Jensen, P., S. Hansen, B. Moller, T. Leivestad, P. Pfeffer, O. Geiran, P. Fauchald, and S. Simonsen.** 1999. Skin cancer in kidney and heart transplant recipients and different long-term immunosuppressive therapy regimens. *J Am Acad Dermatol* **40**:177-86.
  133. **Johne, R., W. Wittig, D. Fernandez-de-Luco, U. Hofle, and H. Muller.** 2006. Characterization of two novel polyomaviruses of birds by using multiply primed rolling-circle amplification of their genomes. *J Virol* **80**:3523-31.
  134. **Jonsson, K., C. Signas, H. P. Muller, and M. Lindberg.** 1991. Two different genes encode fibronectin binding proteins in *Staphylococcus aureus*. The complete nucleotide sequence and characterization of the second gene. *Eur J Biochem* **202**:1041-8.
  135. **Joyce, J. G., J. S. Tung, C. T. Przysiecki, J. C. Cook, E. D. Lehman, J. A. Sands, K. U. Jansen, and P. M. Keller.** 1999. The L1 major capsid protein of human papillomavirus type 11 recombinant virus-like particles interacts with heparin and cell-surface glycosaminoglycans on human keratinocytes. *J Biol Chem* **274**:5810-22.
  136. **Kamper, N., P. M. Day, T. Nowak, H. C. Selinka, L. Florin, J. Bolscher, L. Hilbig, J. T. Schiller, and M. Sapp.** 2006. A membrane-destabilizing peptide in capsid protein L2 is required for egress of papillomavirus genomes from endosomes. *J Virol* **80**:759-68.
  137. **Karaa, A., and A. Khachemoune.** 2007. Keratoacanthoma: a tumor in search of a classification. *Int J Dermatol* **46**:671-8.
  138. **Karagas, M. R., H. H. Nelson, P. Sehr, T. Waterboer, T. A. Stukel, A. Andrew, A. C. Green, J. N. Bavinck, A. Perry, S. Spencer, J. R. Rees, L. A. Mott, and M. Pawlita.** 2006. Human papillomavirus infection and incidence of squamous cell and basal cell carcinomas of the skin. *J Natl Cancer Inst* **98**:389-95.
  139. **Kines, R. C., C. D. Thompson, D. R. Lowy, J. T. Schiller, and P. M. Day.** 2009. The initial steps leading to papillomavirus infection occur on the basement membrane prior to cell surface binding. *Proc Natl Acad Sci U S A* **106**:20458-63.
  140. **Kirnbauer, R., F. Booy, N. Cheng, D. R. Lowy, and A. T. Schiller.** 1992. Papillomavirus L1 major capsid protein self-assembles into virus-like particles that are highly immunogenic. *Proceedings of the National Academy of Sciences USA* **89**:12180-12184.

141. **Kluytmans, J., A. van Belkum, and H. Verbrugh.** 1997. Nasal carriage of *Staphylococcus aureus*: epidemiology, underlying mechanisms, and associated risks. *Clin Microbiol Rev* **10**:505-20.
142. **Korbel, J. O., A. E. Urban, J. P. Affourtit, B. Godwin, F. Grubert, J. F. Simons, P. M. Kim, D. Palejev, N. J. Carriero, L. Du, B. E. Taillon, Z. Chen, A. Tanzer, A. C. Saunders, J. Chi, F. Yang, N. P. Carter, M. E. Hurles, S. M. Weissman, T. T. Harkins, M. B. Gerstein, M. Egholm, and M. Snyder.** 2007. Paired-end mapping reveals extensive structural variation in the human genome. *Science* **318**:420-6.
143. **Kranjec, C., and L. Banks.** A systematic analysis of HPV E6 PDZ substrates identifies MAGI-1 as a major target of HPV-16 and HPV-18 whose loss accompanies disruption of Tight Junctions. *J Virol*.
144. **Kreimer, A. R., G. M. Clifford, P. Boyle, and S. Franceschi.** 2005. Human papillomavirus types in head and neck squamous cell carcinomas worldwide: a systematic review. *Cancer Epidemiol Biomarkers Prev* **14**:467-75.
145. **Kreuter, A., T. Gambichler, H. Pfister, and U. Wieland.** 2009. Diversity of human papillomavirus types in periungual squamous cell carcinoma. *Br J Dermatol* **161**:1262-9.
146. **Kuo, G., Q. L. Choo, H. J. Alter, G. L. Gitnick, A. G. Redeker, R. H. Purcell, T. Miyamura, J. L. Dienstag, M. J. Alter, C. E. Stevens, and et al.** 1989. An assay for circulating antibodies to a major etiologic virus of human non-A, non-B hepatitis. *Science* **244**:362-4.
147. **Kuo, S. R., J. S. Liu, T. R. Broker, and L. T. Chow.** 1994. Cell-free replication of the human papillomavirus DNA with homologous viral E1 and E2 proteins and human cell extracts. *J Biol Chem* **269**:24058-65.
148. **Kwa, R. E., K. Campana, and R. L. Moy.** 1992. Biology of cutaneous squamous cell carcinoma. *J Am Acad Dermatol* **26**:1-26.
149. **Lage, J. M., J. H. Leamon, T. Pejovic, S. Hamann, M. Lacey, D. Dillon, R. Seagraves, B. Vossbrinck, A. Gonzalez, D. Pinkel, D. G. Albertson, J. Costa, and P. M. Lizardi.** 2003. Whole genome analysis of genetic alterations in small DNA samples using hyperbranched strand displacement amplification and array-CGH. *Genome Res* **13**:294-307.
150. **Lehman, C. W., and M. R. Botchan.** 1998. Segregation of viral plasmids depends on tethering to chromosomes and is regulated by phosphorylation. *Proc Natl Acad Sci U S A* **95**:4338-43.
151. **Leininger, S., T. Urich, M. Schloter, L. Schwark, J. Qi, G. W. Nicol, J. I. Prosser, S. C. Schuster, and C. Schleper.** 2006. Archaea predominate among ammonia-oxidizing prokaryotes in soils. *Nature* **442**:806-9.
152. **Li, L., P. Barry, E. Yeh, C. Glaser, D. Schnurr, and E. Delwart.** 2009. Identification of a novel human gammapapillomavirus species. *J Gen Virol* **90**:2413-7.
153. **Lin, B. Y., A. M. Makhov, J. D. Griffith, T. R. Broker, and L. T. Chow.** 2002. Chaperone proteins abrogate inhibition of the human papillomavirus (HPV) E1 replicative helicase by the HPV E2 protein. *Mol Cell Biol* **22**:6592-604.

154. **Lindelof, B., B. Sigurgeirsson, H. Gabel, and R. S. Stern.** 2000. Incidence of skin cancer in 5356 patients following organ transplantation. *Br J Dermatol* **143**:513-9.
155. **Linnen, J., J. Wages, Jr., Z. Y. Zhang-Keck, K. E. Fry, K. Z. Krawczynski, H. Alter, E. Koonin, M. Gallagher, M. Alter, S. Hadziyannis, P. Karayiannis, K. Fung, Y. Nakatsuji, J. W. Shih, L. Young, M. Piatak, Jr., C. Hoover, J. Fernandez, S. Chen, J. C. Zou, T. Morris, K. C. Hyams, S. Ismay, J. D. Lifson, G. Hess, S. K. Fong, H. Thomas, D. Bradley, H. Margolis, and J. P. Kim.** 1996. Molecular cloning and disease association of hepatitis G virus: a transfusion-transmissible agent. *Science* **271**:505-8.
156. **Lisitsyn, N., and M. Wigler.** 1993. Cloning the differences between two complex genomes. *Science* **259**:946-51.
157. **Liu, J. S., S. R. Kuo, T. R. Broker, and L. T. Chow.** 1995. The functions of human papillomavirus type 11 E1, E2, and E2C proteins in cell-free DNA replication. *J Biol Chem* **270**:27283-91.
158. **MacKie, R. M.** 2004. Awareness, knowledge and attitudes to basal cell carcinoma and actinic keratoses among the general public within Europe. *J Eur Acad Dermatol Venereol* **18**:552-5.
159. **MacKie, R. M.** 2006. Long-term health risk to the skin of ultraviolet radiation. *Prog Biophys Mol Biol* **92**:92-6.
160. **Marc Van Ranst, J. B. K., Johan P. Sundberg and Robert D Burk.** 1995. Molecular evolution of papillomaviruses. *Molecular basis of viral evolution*:455-476.
161. **Margulies, M., M. Egholm, W. E. Altman, S. Attiya, J. S. Bader, L. A. Bembien, J. Berka, M. S. Braverman, Y. J. Chen, Z. Chen, S. B. Dewell, L. Du, J. M. Fierro, X. V. Gomes, B. C. Godwin, W. He, S. Helgesen, C. H. Ho, G. P. Irzyk, S. C. Jando, M. L. Alenquer, T. P. Jarvie, K. B. Jirage, J. B. Kim, J. R. Knight, J. R. Lanza, J. H. Leamon, S. M. Lefkowitz, M. Lei, J. Li, K. L. Lohman, H. Lu, V. B. Makhijani, K. E. McDade, M. P. McKenna, E. W. Myers, E. Nickerson, J. R. Nobile, R. Plant, B. P. Puc, M. T. Ronan, G. T. Roth, G. J. Sarkis, J. F. Simons, J. W. Simpson, M. Srinivasan, K. R. Tartaro, A. Tomasz, K. A. Vogt, G. A. Volkmer, S. H. Wang, Y. Wang, M. P. Weiner, P. Yu, R. F. Begley, and J. M. Rothberg.** 2005. Genome sequencing in microfabricated high-density picolitre reactors. *Nature* **437**:376-80.
162. **Marks, R., P. Foley, G. Goodman, B. H. Hage, and T. S. Selwood.** 1986. Spontaneous remission of solar keratoses: the case for conservative management. *Br J Dermatol* **115**:649-55.
163. **Marks, R., G. Rennie, and T. S. Selwood.** 1988. Malignant transformation of solar keratoses to squamous cell carcinoma. *Lancet* **1**:795-7.
164. **Marx, J.** 2004. Cancer research. Inflammation and cancer: the link grows stronger. *Science* **306**:966-8.
165. **Matsukura, T., T. Iwasaki, and M. Kawashima.** 1992. Molecular cloning of a novel human papillomavirus (type 60) from a plantar cyst with characteristic pathological changes. *Virology* **190**:561-4.
166. **McBride, A. A., J. C. Byrne, and P. M. Howley.** 1989. E2 polypeptides encoded by bovine papillomavirus type 1 form dimers through the common



- carboxyl-terminal domain: transactivation is mediated by the conserved amino-terminal domain. *Proc Natl Acad Sci U S A* **86**:510-4.
167. **McLaughlin-Drubin, M. E., and K. Munger.** 2009. Oncogenic activities of human papillomaviruses. *Virus Res* **143**:195-208.
  168. **McLemore, M. R.** 2006. Gardasil: Introducing the new human papillomavirus vaccine. *Clin J Oncol Nurs* **10**:559-60.
  169. **Miller, R. W., and C. S. Rabkin.** 1999. Merkel cell carcinoma and melanoma: etiological similarities and differences. *Cancer Epidemiol Biomarkers Prev* **8**:153-8.
  170. **Mlynarczyk, A., G. Mlynarczyk, and J. Jeljaszewicz.** 1998. The genome of *Staphylococcus aureus*: a review. *Zentralbl Bakteriol* **287**:277-314.
  171. **Moberg, M., I. Gustavsson, E. Wilander, and U. Gyllensten.** 2005. High viral loads of human papillomavirus predict risk of invasive cervical carcinoma. *Br J Cancer* **92**:891-4.
  172. **Muller, M., G. Kelly, M. Fiedler, and L. Gissmann.** 1989. Human papillomavirus type 48. *J Virol* **63**:4907-8.
  173. **Munoz, N., F. X. Bosch, S. de Sanjose, R. Herrero, X. Castellsague, K. V. Shah, P. J. Snijders, and C. J. Meijer.** 2003. Epidemiologic classification of human papillomavirus types associated with cervical cancer. *N Engl J Med* **348**:518-27.
  174. **Munoz, N., S. Franceschi, C. Bosetti, V. Moreno, R. Herrero, J. S. Smith, K. V. Shah, C. J. Meijer, and F. X. Bosch.** 2002. Role of parity and human papillomavirus in cervical cancer: the IARC multicentric case-control study. *Lancet* **359**:1093-101.
  175. **Naucler, P., F. M. Da Costa, O. Ljungberg, A. Bugalho, and J. Dillner.** 2004. Human papillomavirus genotypes in cervical cancers in Mozambique. *J Gen Virol* **85**:2189-90.
  176. **Nguyen, N. P., A. Chi, L. M. Nguyen, B. H. Ly, U. Karlsson, and V. Vinh-Hung.** Human papillomavirus-associated oropharyngeal cancer: a new clinical entity. *QJM* **103**:229-36.
  177. **Nindl, I., M. Gottschling, and E. Stockfleth.** 2007. Human papillomaviruses and non-melanoma skin cancer: basic virology and clinical manifestations. *Dis Markers* **23**:247-59.
  178. **Nishizawa, T., H. Okamoto, K. Konishi, H. Yoshizawa, Y. Miyakawa, and M. Mayumi.** 1997. A novel DNA virus (TTV) associated with elevated transaminase levels in posttransfusion hepatitis of unknown etiology. *Biochem Biophys Res Commun* **241**:92-7.
  179. **Nobre, R. J., E. Herraes-Hernandez, J. W. Fei, L. Langbein, S. Kaden, H. J. Grone, and E. M. de Villiers.** 2009. E7 oncoprotein of novel human papillomavirus type 108 lacking the E6 gene induces dysplasia in organotypic keratinocyte cultures. *J Virol* **83**:2907-16.
  180. **Ogston, A.** 1882. Micrococcus Poisoning. *J Anat Physiol* **17**:24-58.
  181. **Oldroyd, B. P.** 2007. What's killing American honey bees? *PLoS Biol* **5**:e168.
  182. **Orth, G.** 2006. Genetics of epidermodysplasia verruciformis: Insights into host defense against papillomaviruses. *Semin Immunol* **18**:362-74.

183. **Orth, G.** 2008. Host defenses against human papillomaviruses: lessons from epidermodysplasia verruciformis. *Curr Top Microbiol Immunol* **321**:59-83.
184. **Orth, G., M. Favre, and O. Croissant.** 1977. Characterization of a new type of human papillomavirus that causes skin warts. *J Virol* **24**:108-20.
185. **Orth, G., M. Favre, S. Majewski, and S. Jablonska.** 2001. Epidermodysplasia verruciformis defines a subset of cutaneous human papillomaviruses. *J Virol* **75**:4952-3.
186. **Ostrow, R. S., M. Bender, M. Niimura, T. Seki, M. Kawashima, F. Pass, and A. J. Faras.** 1982. Human papillomavirus DNA in cutaneous primary and metastasized squamous cell carcinomas from patients with epidermodysplasia verruciformis. *Proc Natl Acad Sci U S A* **79**:1634-8.
187. **Ozbun, M. A., and C. Meyers.** 1998. Human papillomavirus type 31b E1 and E2 transcript expression correlates with vegetative viral genome amplification. *Virology* **248**:218-30.
188. **Palacios, G., J. Druce, L. Du, T. Tran, C. Birch, T. Briese, S. Conlan, P. L. Quan, J. Hui, J. Marshall, J. F. Simons, M. Egholm, C. D. Paddock, W. J. Shieh, C. S. Goldsmith, S. R. Zaki, M. Catton, and W. I. Lipkin.** 2008. A new arenavirus in a cluster of fatal transplant-associated diseases. *N Engl J Med* **358**:991-8.
189. **Parkin, D., and F. Bray.** 2006. Chapter 2: The burden of HPV-related cancers. *Vaccine* **24 suppl 3**:S11-S25.
190. **Pastrana, D. V., Y. L. Tolstov, J. C. Becker, P. S. Moore, Y. Chang, and C. B. Buck.** 2009. Quantitation of human seroresponsiveness to Merkel cell polyomavirus. *PLoS Pathog* **5**:e1000578.
191. **Peterson, P. K., J. Verhoef, L. D. Sabath, and P. G. Quie.** 1977. Effect of protein A on staphylococcal opsonization. *Infect Immun* **15**:760-4.
192. **Pfister, H., and L. Gissmann.** 1978. Heterogeneity of human papilloma viruses. *Bull Cancer* **65**:165-7.
193. **Pfister, H., and J. Ter Schegget.** 1997. Role of HPV in cutaneous premalignant and malignant tumors. *Clin Dermatol* **15**:335-47.
194. **Plasmeijer, E. I., R. E. Neale, M. N. de Koning, W. G. Quint, P. McBride, M. C. Feltkamp, and A. C. Green.** 2009. Persistence of betapapillomavirus infections as a risk factor for actinic keratoses, precursor to cutaneous squamous cell carcinoma. *Cancer Res* **69**:8926-31.
195. **Plummer, M., M. Schiffman, P. E. Castle, D. Maucort-Boulch, and C. M. Wheeler.** 2007. A 2-year prospective study of human papillomavirus persistence among women with a cytological diagnosis of atypical squamous cells of undetermined significance or low-grade squamous intraepithelial lesion. *J Infect Dis* **195**:1582-9.
196. **Poulsen, M.** 2004. Merkel-cell carcinoma of the skin. *Lancet Oncol* **5**:593-9.
197. **Preston, D. S., and R. S. Stern.** 1992. Nonmelanoma cancers of the skin. *N Engl J Med* **327**:1649-62.
198. **Ramos, J., J. Villa, A. Ruiz, R. Armstrong, and J. Matta.** 2004. UV dose determines key characteristics of nonmelanoma skin cancer. *Cancer Epidemiol Biomarkers Prev* **13**:2006-11.

199. **Rector, A., H. Stevens, G. Lacave, P. Lemey, S. Mostmans, A. Salbany, M. Vos, K. Van Doorslaer, S. J. Ghim, M. Rehtanz, G. D. Bossart, A. B. Jenson, and M. Van Ranst.** 2008. Genomic characterization of novel dolphin papillomaviruses provides indications for recombination within the Papillomaviridae. *Virology* **378**:151-61.
200. **Rector, A., R. Tachezy, K. Van Doorslaer, T. MacNamara, R. D. Burk, J. P. Sundberg, and M. Van Ranst.** 2005. Isolation and cloning of a papillomavirus from a North American porcupine by using multiply primed rolling-circle amplification: the *Erethizon dorsatum* papillomavirus type 1. *Virology* **331**:449-56.
201. **Relman, D. A.** 1999. The search for unrecognized pathogens. *Science* **284**:1308-10.
202. **Reyes, G. R., and J. P. Kim.** 1991. Sequence-independent, single-primer amplification (SISPA) of complex DNA populations. *Mol Cell Probes* **5**:473-81.
203. **Roberts, J. N., C. B. Buck, C. D. Thompson, R. Kines, M. Bernardo, P. L. Choyke, D. R. Lowy, and J. T. Schiller.** 2007. Genital transmission of HPV in a mouse model is potentiated by nonoxynol-9 and inhibited by carrageenan. *Nat Med* **13**:857-61.
204. **Roesch, L. F., R. R. Fulthorpe, A. Riva, G. Casella, A. K. Hadwin, A. D. Kent, S. H. Daroub, F. A. Camargo, W. G. Farmerie, and E. W. Triplett.** 2007. Pyrosequencing enumerates and contrasts soil microbial diversity. *ISME J* **1**:283-90.
205. **Rose, F., G. Dahlem, B. Guthmann, F. Grimminger, U. Maus, J. Hanze, N. Duemmer, U. Grandel, W. Seeger, and H. A. Ghofrani.** 2002. Mediator generation and signaling events in alveolar epithelial cells attacked by *S. aureus* alpha-toxin. *Am J Physiol Lung Cell Mol Physiol* **282**:L207-14.
206. **Rous, P., and J. W. Beard.** 1935. The Progression to Carcinoma of Virus-Induced Rabbit Papillomas (Shope). *J Exp Med* **62**:523-48.
207. **Rous, P., and J. W. Beard.** 1934. A Virus-Induced Mammalian Growth with the Characters of a Tumor (the Shope Rabbit Papilloma) : I. The Growth on Implantation within Favorable Hosts. *J Exp Med* **60**:701-22.
208. **Rubin, A. I., E. H. Chen, and D. Ratner.** 2005. Basal-cell carcinoma. *N Engl J Med* **353**:2262-9.
209. **Salem, S. A., N. M. Zuel-Fakkar, G. Fathi, S. M. Abd El-Reheem, A. Abd Elmonem El-Tabakh, and D. M. Ragab.** Comparative study of human papilloma virus in untreated and ultraviolet-treated psoriatic patients. *Photodermatol Photoimmunol Photomed* **26**:78-82.
210. **Scheffner, M., B. A. Werness, J. M. Huibregtse, A. J. Levine, and P. M. Howley.** 1990. The E6 oncoprotein encoded by human papillomavirus types 16 and 18 promotes the degradation of p53. *Cell* **63**:1129-36.
211. **Schiffman, M., R. Herrero, R. Desalle, A. Hildesheim, S. Wacholder, A. Rodriguez, M. Bratti, M. Sherman, J. Morales, D. Guillen, M. Alfaro, M. Hutchinson, T. Wright, D. Solomon, Z. Chen, J. Schussler, P. Castle, and R. Burk.** 2005. The carcinogenicity of human papillomavirus types reflects viral evolution. *Virology* **337**:76-84.

212. **Schiller, J. T., W. C. Vass, K. H. Vousden, and D. R. Lowy.** 1986. E5 open reading frame of bovine papillomavirus type 1 encodes a transforming gene. *J Virol* **57**:1-6.
213. **Schlecht, N., S. Kulaga, J. Robitaille, S. Ferreira, M. Santos, R. Miyamura, E. Duarte-Franco, T. Rohan, A. Ferenczy, L. Villa, and E. Franco.** 2001. Persistent human papillomavirus infection as a predictor of cervical intraepithelial neoplasia. *JAMA* **286**:3106-3114.
214. **Schlecht, N. F., A. Trevisan, E. Duarte-Franco, T. E. Rohan, A. Ferenczy, L. L. Villa, and E. L. Franco.** 2003. Viral load as a predictor of the risk of cervical intraepithelial neoplasia. *Int J Cancer* **103**:519-24.
215. **Schmitt, M., I. G. Bravo, P. J. Snijders, L. Gissmann, M. Pawlita, and T. Waterboer.** 2006. Bead-based multiplex genotyping of human papillomaviruses. *J Clin Microbiol* **44**:504-12.
216. **Schwalter, R. M., D. V. Pastrana, K. A. Pumphrey, A. L. Moyer, and C. B. Buck.** Merkel cell polyomavirus and two previously unknown polyomaviruses are chronically shed from human skin. *Cell Host Microbe* **7**:509-15.
217. **Schwartz, R. A.** 2004. Keratoacanthoma: a clinico-pathologic enigma. *Dermatol Surg* **30**:326-33; discussion 333.
218. **Sedman, J., and A. Stenlund.** 1998. The papillomavirus E1 protein forms a DNA-dependent hexameric complex with ATPase and DNA helicase activities. *J Virol* **72**:6893-7.
219. **Selinka, H. C., L. Florin, H. D. Patel, K. Freitag, M. Schmidtke, V. A. Makarov, and M. Sapp.** 2007. Inhibition of transfer to secondary receptors by heparan sulfate-binding drug or antibody induces noninfectious uptake of human papillomavirus. *J Virol* **81**:10970-80.
220. **Shamanin, V., H. Delius, and E. M. de Villiers.** 1994. Development of a broad spectrum PCR assay for papillomaviruses and its application in screening lung cancer biopsies. *J Gen Virol* **75 ( Pt 5)**:1149-56.
221. **Shope, R. E., and E. W. Hurst.** 1933. Infectious Papillomatosis of Rabbits : With a Note on the Histopathology. *J Exp Med* **58**:607-24.
222. **Silins, I., X. Wang, A. Tadesse, K. U. Jansen, J. T. Schiller, E. Avall-Lundqvist, B. Frankendal, and J. Dillner.** 2004. A population-based study of cervical carcinoma and HPV infection in Latvia. *Gynecol Oncol* **93**:484-92.
223. **Sinha, B., and M. Fraunholz.** Staphylococcus aureus host cell invasion and post-invasion events. *Int J Med Microbiol* **300**:170-5.
224. **Sjöholm, M. I. L., J. Dillner, and J. Carlson.** 2007. Assessing quality and functionality of DNA from fresh and archival dried blood spots and recommendations for quality control guidelines. *Clin Chem* **53**:1401-1407.
225. **Skiadopoulos, M. H., and A. A. McBride.** 1998. Bovine papillomavirus type 1 genomes and the E2 transactivator protein are closely associated with mitotic chromatin. *J Virol* **72**:2079-88.
226. **Smith, J., C. Bosetti, N. Munoz, R. Herrero, F. Bosch, J. Eluf-Neto, C. Meijer, A. V. d. Brule, S. Franceschi, and R. Peeling.** 2004. Chlamydia trachomatis and invasive cervical cancer: a pooled analysis of the IARC multicentric case-control study. *Int J Cancer* **111**:431-439.

227. **Smith, J., J. Green, A. Berrington de Gonzalez, P. Appleby, J. Peto, M. Plummer, S. Franceschi, and V. Beral.** 2003. Cervical cancer and use of hormonal contraceptives: a systematic review. *Lancet* **361**:1159-1167.
228. **Smith, J. S., R. Herrero, C. Bosetti, N. Munoz, F. X. Bosch, J. Eluf-Neto, X. Castellsague, C. J. Meijer, A. J. Van den Brule, S. Franceschi, and R. Ashley.** 2002. Herpes simplex virus-2 as a human papillomavirus cofactor in the etiology of invasive cervical cancer. *J Natl Cancer Inst* **94**:1604-13.
229. **Smith, K. T., and M. S. Campq.** 1988. "Hit and run" transformation of mouse C127 cells by bovine papillomavirus type 4: the viral DNA is required for the initiation but not for maintenance of the transformed phenotype. *Virology* **164**:39-47.
230. **Soderlund-Strand, A., J. Carlson, and J. Dillner.** 2009. Modified general primer PCR system for sensitive detection of multiple types of oncogenic human papillomavirus. *J Clin Microbiol* **47**:541-6.
231. **Sogin, M. L., H. G. Morrison, J. A. Huber, D. Mark Welch, S. M. Huse, P. R. Neal, J. M. Arrieta, and G. J. Herndl.** 2006. Microbial diversity in the deep sea and the underexplored "rare biosphere". *Proc Natl Acad Sci U S A* **103**:12115-20.
232. **Stark, L. A., M. J. Arends, K. M. McLaren, E. C. Benton, H. Shahidullah, J. A. Hunter, and C. C. Bird.** 1994. Prevalence of human papillomavirus DNA in cutaneous neoplasms from renal allograft recipients supports a possible viral role in tumour promotion. *Br J Cancer* **69**:222-9.
233. **Stark, S., A. K. Petridis, S. J. Ghim, A. B. Jenson, J. N. Bouwes Bavinck, G. Gross, E. Stockfleth, P. Fuchs, and H. Pfister.** 1998. Prevalence of antibodies against virus-like particles of Epidermodysplasia verruciformis-associated HPV 8 in patients at risk of skin cancer. *J. Invest. Dermatol.* **111**:696-701.
234. **Steinbrook, R.** 2006. The potential of human papillomavirus vaccines. *N Engl J Med* **354**:1109-12.
235. **Stern, R. S.** 1999. The mysteries of geographic variability in nonmelanoma skin cancer incidence. *Arch Dermatol* **135**:843-4.
236. **Stevens, H., A. Rector, M. F. Bertelsen, P. S. Leifsson, and M. Van Ranst.** 2008. Novel papillomavirus isolated from the oral mucosa of a polar bear does not cluster with other papillomaviruses of carnivores. *Vet Microbiol* **129**:108-16.
237. **Stoler, M. H., C. R. Rhodes, A. Whitbeck, S. M. Wolinsky, L. T. Chow, and T. R. Broker.** 1992. Human papillomavirus type 16 and 18 gene expression in cervical neoplasias. *Hum Pathol* **23**:117-28.
238. **Strauss, M. J., E. W. Shaw, and et al.** 1949. Crystalline virus-like particles from skin papillomas characterized by intranuclear inclusion bodies. *Proc Soc Exp Biol Med* **72**:46-50.
239. **Struijk, L., L. Hall, E. van der Meijden, P. Wanningen, J. N. Bavinck, R. Neale, A. C. Green, J. Ter Schegget, and M. C. Feltkamp.** 2006. Markers of cutaneous human papillomavirus infection in individuals with tumor-free skin, actinic keratoses, and squamous cell carcinoma. *Cancer Epidemiol Biomarkers Prev* **15**:529-35.
240. **Stubenrauch, F., M. Hummel, T. Iftner, and L. A. Laimins.** 2000. The E8E2C protein, a negative regulator of viral transcription and replication, is required for

- extrachromosomal maintenance of human papillomavirus type 31 in keratinocytes. *J Virol* **74**:1178-86.
241. **Stubenrauch, F., and L. A. Laimins.** 1999. Human papillomavirus life cycle: active and latent phases. *Semin Cancer Biol* **9**:379-86.
242. **Sturegard, E., A. Johnsson, E. Gustafsson, and J. Dillner.** 2008. [Condyloma typing important for follow up of HPV vaccination. A condyloma reporting project]. *Lakartidningen* **105**:3648-50.
243. **Switalski, L. M., P. Speziale, and M. Hook.** 1989. Isolation and characterization of a putative collagen receptor from *Staphylococcus aureus* strain Cowan 1. *J Biol Chem* **264**:21080-6.
244. **Telfer, N. R., G. B. Colver, and C. A. Morton.** 2008. Guidelines for the management of basal cell carcinoma. *Br J Dermatol* **159**:35-48.
245. **Thomas, M., N. Narayan, D. Pim, V. Tomaic, P. Massimi, K. Nagasaka, C. Kranjec, N. Gammoh, and L. Banks.** 2008. Human papillomaviruses, cervical cancer and cell polarity. *Oncogene* **27**:7018-30.
246. **Thomas, M. C., and C. M. Chiang.** 2005. E6 oncoprotein represses p53-dependent gene activation via inhibition of protein acetylation independently of inducing p53 degradation. *Mol Cell* **17**:251-64.
247. **Tobler, K., C. Favrot, G. Nespeca, and M. Ackermann.** 2006. Detection of the prototype of a potential novel genus in the family Papillomaviridae in association with canine epidermodysplasia verruciformis. *J Gen Virol* **87**:3551-7.
248. **Todd, J. K.** 2005. Staphylococcal infections. *Pediatr Rev* **26**:444-50.
249. **Towner, J. S., T. K. Sealy, M. L. Khristova, C. G. Albarino, S. Conlan, S. A. Reeder, P. L. Quan, W. I. Lipkin, R. Downing, J. W. Tappero, S. Okware, J. Lutwama, B. Bakamutumaho, J. Kayiwa, J. A. Comer, P. E. Rollin, T. G. Ksiazek, and S. T. Nichol.** 2008. Newly discovered ebola virus associated with hemorrhagic fever outbreak in Uganda. *PLoS Pathog* **4**:e1000212.
250. **Van Doorslaer, K., A. Rector, P. Vos, and M. Van Ranst.** 2006. Genetic characterization of the *Capra hircus* papillomavirus: a novel close-to-root artiodactyl papillomavirus. *Virus Res* **118**:164-9.
251. **Van Tine, B. A., L. D. Dao, S. Y. Wu, T. M. Sonbuchner, B. Y. Lin, N. Zou, C. M. Chiang, T. R. Broker, and L. T. Chow.** 2004. Human papillomavirus (HPV) origin-binding protein associates with mitotic spindles to enable viral DNA partitioning. *Proc Natl Acad Sci U S A* **101**:4030-5.
252. **Vasiljevic, N., K. Hazard, J. Dillner, and O. Forslund.** 2008. Four novel human betapapillomaviruses of species 2 preferentially found in actinic keratosis. *Journal of General Virology* **89**:2467-2474.
253. **Vasiljevic, N., K. Hazard, L. Eliasson, H. Ly, A. Hunziker, E.-M. d. Villiers, B. Norrild, J. Dillner, and O. Forslund.** 2007. Characterization of two novel cutaneous Human Papillomaviruses, HPV 93 and HPV 96. *J Gen Virol* **88**:1479-1483.
254. **Walboomers, J. M., M. V. Jacobs, M. M. Manos, F. X. Bosch, J. A. Kummer, K. V. Shah, P. J. Snijders, J. Peto, C. J. Meijer, and N. Munoz.** 1999. Human papillomavirus is a necessary cause of invasive cervical cancer worldwide. *J Pathol* **189**:12-9.

255. **Waterboer, T., D. Abeni, F. Sampogna, A. Rother, C. Masini, P. Sehr, K. M. Michael, and M. Pawlita.** 2008. Serological association of beta and gamma human papillomaviruses with squamous cell carcinoma of the skin. *Br J Dermatol* **159**:457-9.
256. **Weissenborn, S. J., M. N. De Koning, U. Wieland, W. G. Quint, and H. J. Pfister.** 2009. Intrafamilial transmission and family-specific spectra of cutaneous betapapillomaviruses. *J Virol* **83**:811-6.
257. **Weissenborn, S. J., R. Hopfl, F. Weber, H. Smola, H. J. Pfister, and P. G. Fuchs.** 1999. High prevalence of a variety of epidermodysplasia verruciformis-associated human papillomaviruses in psoriatic skin of patients treated or not treated with PUVA. *J Invest Dermatol* **113**:122-6.
258. **Weissenborn, S. J., I. Nindl, K. Purdie, C. Harwood, C. Proby, J. Breuer, S. Majewski, H. Pfister, and U. Wieland.** 2005. Human papillomavirus-DNA loads in actinic keratoses exceed those in non-melanoma skin cancers. *J Invest Dermatol* **125**:93-7.
259. **Werness, B. A., A. J. Levine, and P. M. Howley.** 1990. Association of human papillomavirus types 16 and 18 E6 proteins with p53. *Science* **248**:76-9.
260. **Wheeler, D. A., M. Srinivasan, M. Egholm, Y. Shen, L. Chen, A. McGuire, W. He, Y. J. Chen, V. Makhijani, G. T. Roth, X. Gomes, K. Tartaro, F. Niazi, C. L. Turcotte, G. P. Irzyk, J. R. Lupski, C. Chinault, X. Z. Song, Y. Liu, Y. Yuan, L. Nazareth, X. Qin, D. M. Muzny, M. Margulies, G. M. Weinstock, R. A. Gibbs, and J. M. Rothberg.** 2008. The complete genome of an individual by massively parallel DNA sequencing. *Nature* **452**:872-6.
261. **Wieland, U., C. Mauch, A. Kreuter, T. Krieg, and H. Pfister.** 2009. Merkel cell polyomavirus DNA in persons without merkel cell carcinoma. *Emerg Infect Dis* **15**:1496-8.
262. **Williams, R. E.** 1963. Healthy carriage of *Staphylococcus aureus*: its prevalence and importance. *Bacteriol Rev* **27**:56-71.
263. **Wilson, R., F. Fehrmann, and L. A. Laimins.** 2005. Role of the E1-E4 protein in the differentiation-dependent life cycle of human papillomavirus type 31. *J Virol* **79**:6732-40.
264. **Wilson, R., G. B. Ryan, G. L. Knight, L. A. Laimins, and S. Roberts.** 2007. The full-length E1E4 protein of human papillomavirus type 18 modulates differentiation-dependent viral DNA amplification and late gene expression. *Virology* **362**:453-60.
265. **Yang, L., I. Mohr, E. Fouts, D. A. Lim, M. Nohaile, and M. Botchan.** 1993. The E1 protein of bovine papilloma virus 1 is an ATP-dependent DNA helicase. *Proc Natl Acad Sci U S A* **90**:5086-90.
266. **Ylitalo, N., P. Sorensen, A. M. Josefsson, P. K. Magnusson, P. K. Andersen, J. Ponten, H. O. Adami, U. B. Gyllensten, and M. Melbye.** 2000. Consistent high viral load of human papillomavirus 16 and risk of cervical carcinoma in situ: a nested case-control study. *Lancet* **355**:2194-8.
267. **You, J., J. L. Croyle, A. Nishimura, K. Ozato, and P. M. Howley.** 2004. Interaction of the bovine papillomavirus E2 protein with Brd4 tethers the viral DNA to host mitotic chromosomes. *Cell* **117**:349-60.

268. **Zhou, J., X. Y. Sun, K. Louis, and I. H. Frazer.** 1994. Interaction of human papillomavirus (HPV) type 16 capsid proteins with HPV DNA requires an intact L2 N-terminal sequence. *J Virol* **68**:619-25.
269. **zur Hausen, H.** 1976. Condylomata acuminata and human genital cancer. *Cancer Res* **36**:794.
270. **zur Hausen, H.** 2009. The search for infectious causes of human cancers: where and why (Nobel lecture). *Angew Chem Int Ed Eng* **48**:5798-5808.



