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### A Genome-Wide Association Meta-Analysis of Circulating Sex Hormone-Binding Globulin Reveals Multiple Loci Implicated in Sex Steroid Hormone Regulation

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DOI: 10.1371/journal.pgen.1002805

2012

#### Link to publication

Citation for published version (APA):

Coviello, A. D., Haring, R., Wellons, M., Vaidya, D., Lehtimaki, T., Keildson, S., Lunetta, K. L., He, C., Fornage, M., Lagou, V., Mangino, M., Onland-Moret, N. C., Chen, B., Eriksson, J., Garcia, M., Mei, Y., Koster, A., Lohman, K., Lyytikainen, L.-P., ... Perry, J. R. B. (2012). A Genome-Wide Association Meta-Analysis of Circulating Sex Hormone-Binding Globulin Reveals Multiple Loci Implicated in Sex Steroid Hormone Regulation. PLoS Genetics, 8(7). https://doi.org/10.1371/journal.pgen.1002805

Total number of authors: 99

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## A Genome-Wide Association Meta-Analysis of Circulating Sex Hormone–Binding Globulin Reveals Multiple Loci Implicated in Sex Steroid Hormone Regulation

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#### Abstract

Sex hormone-binding globulin (SHBG) is a glycoprotein responsible for the transport and biologic availability of sex steroid hormones, primarily testosterone and estradiol. SHBG has been associated with chronic diseases including type 2 diabetes (T2D) and with hormone-sensitive cancers such as breast and prostate cancer. We performed a genome-wide association study (GWAS) meta-analysis of 21,791 individuals from 10 epidemiologic studies and validated these findings in 7,046 individuals in an additional six studies. We identified twelve genomic regions (SNPs) associated with circulating SHBG concentrations. Loci near the identified SNPs included SHBG (rs12150660, 17p13.1,  $p = 1.8 \times 10^{-106}$ ), PRMT6 (rs17496332, 1p13.3,  $p = 1.4 \times 10^{-11}$ ), GCKR (rs780093, 2p23.3,  $p = 2.2 \times 10^{-16}$ ), ZBTB10 (rs440837, 8q21.13,  $p = 3.4 \times 10^{-09}$ ), JMJD1C (rs7910927, 10q21.3,  $p = 6.1 \times 10^{-35}$ ), SLCO1B1 (rs4149056, 12p12.1,  $p = 1.9 \times 10^{-08}$ ), NR2F2 (rs8023580, 15q26.2,  $p = 8.3 \times 10^{-12}$ ), ZNF652 (rs2411984, 17q21.32,  $p = 3.5 \times 10^{-14}$ ), TDGF3 (rs1573036, Xq22.3,  $p = 4.1 \times 10^{-14}$ ), LHCGR (rs10454142, 2p16.3,  $p = 1.3 \times 10^{-07}$ ), BAIAP2L1  $(rs3779195, 7q21.3, p = 2.7 \times 10^{-08})$ , and UGT2B15  $(rs293428, 4q13.2, p = 5.5 \times 10^{-06})$ . These genes encompass multiple biologic pathways, including hepatic function, lipid metabolism, carbohydrate metabolism and T2D, androgen and estrogen receptor function, epigenetic effects, and the biology of sex steroid hormone-responsive cancers including breast and prostate cancer. We found evidence of sex-differentiated genetic influences on SHBG. In a sex-specific GWAS, the loci 4q13.2-*UGT2B15* was significant in men only (men  $p = 2.5 \times 10^{-08}$ , women p = 0.66, heterogeneity p = 0.003). Additionally, three loci showed strong sex-differentiated effects: 17p13.1-SHBG and Xq22.3-TDGF3 were stronger in men, whereas 8q21.12-ZBTB10 was stronger in women. Conditional analyses identified additional signals at the SHBG gene that together almost double the proportion of variance explained at the locus. Using an independent study of 1,129 individuals, all SNPs identified in the overall or sexdifferentiated or conditional analyses explained ~15.6% and ~8.4% of the genetic variation of SHBG concentrations in men and women, respectively. The evidence for sex-differentiated effects and allelic heterogeneity highlight the importance of considering these features when estimating complex trait variance.

Citation: Coviello AD, Haring R, Wellons M, Vaidya D, Lehtimäki T, et al. (2012) A Genome-Wide Association Meta-Analysis of Circulating Sex Hormone–Binding Globulin Reveals Multiple Loci Implicated in Sex Steroid Hormone Regulation. PLoS Genet 8(7): e1002805. doi:10.1371/journal.pgen.1002805

Editor: Greg Gibson, Georgia Institute of Technology, United States of America

Received February 20, 2012; Accepted May 19, 2012; Published July 19, 2012

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Funding: This meta-analysis is a collaborative effort involving data from many individual studies and many sources of funding. The details of funding sources for each study are detailed in Text S1 as well as below. Framingham Heart Study (FHS): The phenotype-genotype association analyses were funded through grants from the NIA R21AG032598 (JM Murabito, KL Lunetta), R01HL094755 (AD Coviello, RS Vasan, S Bandinelli), and R01AG31206 (RS Vasan, S Bandinelli), R01 AR/AG 41398 (DP Kiel). This research was conducted in part using data and resources from the Framingham Heart Study of the National Heart, Lung, and Blood Institute of the National Institutes of Health and Boston University School of Medicine. The analyses reflect intellectual input and resource development from the Framingham Heart Study investigators participating in the SNP Health Association Resource (SHARe) project. This work was partially supported by the National Heart, Lung, and Blood Institute's Framingham Heart Study (Contract No. N01-HC-25195) and its contract with Affymetrix for genotyping services (Contract No. N02-HL-6-4278). A portion of this research utilized the Linux Cluster for Genetic Analysis (LinGA-II), funded by the Robert Dawson Evans Endowment of the Department of Medicine at Boston University School of Medicine and Boston Medical Center. Gothenburg Osteoporosis and Obesity Determinants (GOOD) Study: Financial support was received from the Swedish Research Council (K2010-54X-09894-19-3, 2006-3832, and K2010-52X-20229-05-3), the Swedish Foundation for Strategic Research, the ALF/LUA research grant in Gothenburg, the Lundberg Foundation, the Torsten and Ragnar Söderberg's Foundation, Petrus and Augusta Hedlunds Foundation, the Västra Götaland Foundation, the Göteborg Medical Society, the Novo Nordisk foundation, and the European Commission grant HEALTH-F2-2008-201865-GEFOS. We would like to acknowledge Maria Nethander at the genomics core facility at University of Gothenburg for statistical analyses. We would also like to thank Dr. Tobias A. Knoch, Luc V. de Zeeuw, Anis Abuseiris, and Rob de Graaf as well as their institutions the Erasmus Computing Grid, Rotterdam, The Netherlands, and especially the national German MediGRID and Services@MediGRID part of the German D-Grid, both funded by the German Bundesministerium fuer Forschung und Technology under grants #01 AK 803 A-H and #01 IG 07015 G for access to their grid resources. We would also like to thank Karol Estrada, Department ofInternal Medicine, Erasmus MC, Rotterdam, The Netherlands, for advice regarding

the grid resources. Health, Aging, and Body Composition (Health ABC) Study: This Health ABC Study was supported by NIA contracts N01AG62101, N01AG62103, and N01AG62106. The genome-wide association study was funded by NIA grant 1R01AG032098-01A1 to Wake Forest University Health Sciences and genotyping services were provided by the Center for Inherited Disease Research (CIDR). CIDR is fully funded through a federal contract from the National Institutes of Health to The Johns Hopkins University, contract number HHSN268200782096C. This research was also supported in part by the Intramural Research Program of the National Institute on Aging, NIH, Bethesda, Maryland. Invecchiare in Chianti (InCHIANTI): The InCHIANTI study baseline (1998-2000) was supported as a "targeted project" (ICS110.1/RF97.71) by the Italian Ministry of Health and in part by the U.S. National Institute on Aging (Contracts: 263 MD 9164 and 263 MD 821336); the InCHIANTI Follow-up 1 (2001–2003) was funded by the U.S. National Institute on Aging (Contracts: N.1-AG-1-1 and N.1-AG-1-2111); the InCHIANTI Follow-ups 2 and 3 studies (2004–2010) were financed by the U.S. National Institute on Aging (Contract: N01-AG-5-0002), supported in part by the Intramural research program of the National Institute on Aging, National Institutes of Health, Baltimore, Maryland. JRB Perry is a Sir Henry Wellcome Postdoctoral Research Fellow (092447/Z/10/Z). Cooperative Health Research in the Region of Augsburg (KORA): The KORA research platform was initiated and financed by the Helmholtz Zentrum Munich, German Research Center for Environmental Health, which is funded by the German Federal Ministry of Education and Research (BMBF) and by the State of Bavaria. Part of this work was financed by the German National Genome Research Network (NGFN-2 and NGFNPlus: 01G50823). Our research was supported within the Munich Center of Health Sciences (MC Health) as part of LMUinnovativ. Multi-Ethnic Study of Atherosclerosis (MESA): MESA and the MESA SHARe project are conducted and supported by the National Heart, Lung, and Blood Institute (NHLBI) in collaboration with MESA investigators. Support is provided by grants and contracts N01 HC-95159, N01-HC-95160, N01-HC-95161, N01-HC-95162, N01-HC-95163, N01-HC-95164, N01-HC-95165, N01-HC-95166, N01-HC-95167, N01-HC-95168, N01-HC-95169, and RR-024156. Funding support for the sex hormone dataset was provided by grants HL074406 and HL074338. Funding for SHARe genotyping was provided by NHLBI Contract N02-HL-6-4278. Genotyping was performed at the Broad Institute of Harvard and MIT (Boston, Massachusetts, USA) and at Affymetrix (Santa Clara, California, USA) using the Affymetric Genome-Wide Human SNP Array 6.0. Northern Finland Birth Cohort 1966 Study (NFBC-66): NFBC1966 received financial support from the Academy of Finland (project grants 104781, 120315, 129269, 1114194, Center of Excellence in Complex Disease Genetics, and SALVE), University Hospital Oulu, Biocenter, University of Oulu, Finland (75617), the European Commission (EURO-BLCS, Framework 5 award QLG1-CT-2000-01643), NHLBI grant 5R01HL087679-02 through the STAMPEED program (1RL1MH083268-01), NIH/NIMH (5R01MH63706:02), ENGAGE project and grant agreement HEALTH-F4-2007-201413, the Medical Research Council UK (G0500539, G0600705, PrevMetSyn/SALVE), and the Wellcome Trust (project grant GR069224). We acknowledge the support of U.S. National Heart, Lung, and Blood Institute grant HL087679 through the STAMPEED program; grants MH083268, GM053275-14, and U54 RR020278 from the U.S. National Institutes of Health; grant DMS-0239427 from the National Science Foundation; the Medical Research Council of the UK, EURO-BLCS, QLG1-CT-2000-01643 and the European Community's Seventh Framework Programme (FP7/2007–2013); ENGAGE project and grant agreement HEALTH-F4-2007-201413. The authors would like to thank the Center of Excellence in Common Disease Genetics of the Academy of Finland and Nordic Center of Excellence in Disease Genetics, the Sydantautisaatio (Finnish Foundation of Heart Diseases), the Broad Genotyping Center, D. Mirel, H. Hobbs, J. DeYoung, P. Rantakallio, M. Koiranen, and M. Isohanni for advice and assistance. Rotterdam study (RS1): The generation and management of GWAS genotype data for the Rotterdam Study are supported by the Netherlands Organisation of Scientific Research NWO Investments (nr. 175.010.2005.011, 911-03-012). This study is funded by the Research Institute for Diseases in the Elderly (014-93-015; RIDE2), the Netherlands Genomics Initiative (NGI)/Netherlands Organisation for Scientific Research (NWO) project nr. 050-060-810, and funding from the European Commision (HEALTH-F2-2008-201865, GEFOS; HEALTH-F2-2008-35627, TREAT-OA). The Rotterdam Study is funded by Erasmus Medical Center and Erasmus University, Rotterdam; Netherlands Organization for the Health Research and Development (ZonMw); the Research Institute for Diseases in the Elderly (RIDE); the Ministry of Education, Culture, and Science; the Ministry for Health, Welfare, and Sports; the European Commission (DG XII); and the Municipality of Rotterdam. We thank Pascal Arp, Mila Jhamai, Dr. Michael Moorhouse, Marijn Verkerk, and Sander Bervoets for their help in creating the GWAS database. The authors are grateful to the study participants, the staff from the Rotterdam Study and the participating general practitioners and pharmacists. We would like to thank Dr. Tobias A. Knoch, Karol Estrada, Luc V. de Zeeuw, Anis Abuseiris, and Rob de Graaf as well as their institutions the Erasmus Computing Grid, Rotterdam, The Netherlands, and especially the national German MediGRID and Services@MediGRID part of the German D-Grid, both funded by the German Bundesministerium fuer Forschung und Technology under grants #01 AK 803 A-H and # 01 IG 07015 Gfor access to their grid resources. Study of Health in Pomerania (SHIP): SHIP is part of the Community Medicine Research Net of the University of Greifswald, Germany, which is funded by the Federal Ministry of Education and Research (grants no. 01ZZ9603, 01ZZ0103, and 01ZZ0403), and the Ministry of Cultural Affairs, as well as the Social Ministry of the Federal State of Mecklenburg, West Pomerania. Genome-wide data have been supported by the Federal Ministry of Education and Research (grant no. 03ZIK012) and a joint grant from Siemens Healthcare, Erlangen, Germany, and the Federal State of Mecklenburg, West Pomerania. The University of Greifswald is a member of the "Center of Knowledge Interchange" program of the Siemens AG. This work is also part of the research project Greifswald Approach to Individualized Medicine (GANI\_MED). The GANI\_MED consortium is funded by the Federal Ministry of Education and Research and the Ministry of Cultural Affairs of the Federal State of Mecklenburg, West Pomerania (03IS2061A). The SHBG reagents used were sponsored by Siemens Healthcare Diagnostics, Eschborn, formerly DPC Biermann GmbH, Bad Nauheim, Germany, Novo Nordisk provided partial grant support for the determination of serum samples and data analysis. R Haring received honorarium for lectures by Bayer Pharma AG. H Wallaschofski has received research grants from Novo Nordisk and Pfizer for research unrelated to the contents of this manuscript and honorarium for lectures by Bayer Pharma AG. TWINS UK: The study was funded by the Wellcome Trust, European Community's Seventh Framework Programme (FP7/2007–2013) grant agreement HEALTH-F2-2008-201865-GEFOS and (FP7/2007–2013), ENGAGE project grant agreement HEALTH-F4-2007-201413, and the FP-5 GenomEUtwin Project (QLG2-CT-2002-01254). The study also receives support from the Department of Health via the National Institute for Health Research (NIHR) comprehensive Biomedical Research Centre award to Guy's and St. Thomas' NHS Foundation Trust in partnership with King's College London. TD Spector is an NIHR senior Investigator. The project also received support from a Biotechnology and Biological Sciences Research Council (BBSRC) project grant (G20234). The authors acknowledge the funding and support of the National Eye Institute via an NIH/CIDR genotyping project (PI: Terri Young). We thank the staff from the Genotyping Facilities at the Wellcome Trust Sanger Institute for sample preparation, quality control, and genotyping led by Leena Peltonen and Panos Deloukas; Le Centre National de Génotypage, France, led by Mark Lathrop, for genotyping; Duke University, North Carolina, USA, led by David Goldstein, for genotyping; and the Finnish Institute of Molecular Medicine, Finnish Genome Center, University of Helsinki, led by Aarno Palotie. Genotyping was also performed by CIDR as part of an NEI/NIH project grant. The Cardiovascular Risk in Young Finns Study (YFS): The Young Finns Study has been financially supported by the Academy of Finland: grants 134309 (Eye), 126925, 121584, 124282, 129378 (Salve), 117787 (Gendi), and 41071 (Skidi); the Social Insurance Institution of Finland; Kuopio, Tampere, and Turku University Hospital Medical Funds (grant 9M048 for 9N035 for TeLeht); Juho Vainio Foundation; Paavo Nurmi Foundation; Finnish Foundation of Cardiovascular Research and Finnish Cultural Foundation; Tampere Tuberculosis Foundation; and Emil Aaltonen Foundation (T Lehtimäki). The expert technical assistance in the statistical analyses by Irina Lisinen and Ville Aalto are gratefully acknowledged. Women's Health Initiative (WHI): Genotyping was performed at the Broad Institute (Cambridge, MA) through the NHGRI-funded Genomics and Randomized Clinical Network (U01 HG005152) or GARNET. The WHI program is funded by the National Heart, Lung, and Blood Institute, National Institutes of Health, U.S. Department of Health and Human Services through contracts N01WH22110, 24152, 32100-2, 32105-6, 32108-9, 32111-13, 32115, 32118-32119, 32122, 42107-26, 42129–32, and 44221. The authors thank the WHI investigators and staff for their dedication, and the study participants for making the program possible. A listing of WHI investigators can be found at http://www.whiscience.org/publications/WHI\_investigators\_shortlist.pdf. Coronary Artery Risk Development in Young Adults (CARDIA) Women's Study: The CARDIA study is funded by contracts N01-HC-95095, N01-HC-48047, N01-HC-48048, N01-HC-48049, N01-HC-48050, N01-HC-45134, N01-HC-05187, N01-HC-48049, N01 HC-45205, and N01-HC-45204 and by the CARDIA Women's study by R01-HL065611 from the National Heart, Lung, and Blood Institute to the CARDIA investigators Genotyping of the CARDIA participants was supported by grants U01-HG-004729, U01-HG-004446, and U01-HG-004424 from the National Human Genome Research Institute. Statistical analyses were supported by grants U01-HG-004729 and R01-HL-084099 to M Fornage. M Wellons is supported by the Career Development Award 5-K23-HL087114. European Prospective Investigation into Cancer and Nutrition (Prospect-EPIC): The Prospect-EPIC study was funded by "Europe against Cancer" Programme of the European Commission (SANCO); the Dutch Ministry of Health, Welfare, and Sports (VWS); and ZONMw. Osteoporotic fractures in men (MrOS) study Sweden: Financial support was received from the Swedish Research Council (2006-3832), the Swedish Foundation for Strategic Research, the ALF/LUA research grant in Gothenburg, the Lundberg Foundation, the Torsten and Ragnar Söderberg's Foundation, Petrus and Augusta Hedlunds Foundation, the Västra Götaland Foundation, the Göteborg Medical Society, the Novo Nordisk Foundation, and the European Commission grant HEALTH-F2-2008-201865-GEFOS. Nurses' Health Study (NHS): The NHS breast cancer GWAS was performed as part of the Cancer Genetic Markers of Susceptibility (CGEMS) initiative of the NCI. We particularly acknowledge the contributions of R. Hoover, A. Hutchinson, K. Jacobs and G. Thomas. The current research is supported by CA87969, CA49449, CA40356, CA128034, and U01-CA98233 from the National Cancer Institute. We acknowledge the study participants in the NHS for their contribution in making this study possible. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

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#### Author Summary

Sex hormone-binding globulin (SHBG) is the key protein responsible for binding and transporting the sex steroid hormones, testosterone and estradiol, in the circulatory system. SHBG regulates their bioavailability and therefore their effects in the body. SHBG has been linked to chronic diseases including type 2 diabetes and to hormonesensitive cancers such as breast and prostate cancer. SHBG concentrations are approximately 50% heritable in family studies, suggesting SHBG concentrations are under significant genetic control; yet, little is known about the specific genes that influence SHBG. We conducted a large study of the association of SHBG concentrations with markers in the human genome in  $\sim$ 22,000 white men and women to determine which loci influence SHBG concentrations. Genes near the identified genomic markers in addition to the SHBG protein coding gene included PRMT6, GCKR, ZBTB10, JMJD1C, SLCO1B1, NR2F2, ZNF652, TDGF3, LHCGR, BAIAP2L1, and UGT2B15. These genes represent a wide range of biologic pathways that may relate to SHBG function and sex steroid hormone biology, including liver function, lipid metabolism, carbohydrate metabolism and type 2 diabetes, and the development and progression of sex steroid hormone-responsive cancers.

#### Introduction

Sex hormone-binding globulin (SHBG) is a protein secreted mainly by the liver that binds to the sex steroids, testosterone, dihydrotestosterone, and estradiol, transports them in the circulation, and influences their action in target tissues by regulating their bioavailability. SHBG thereby influences the expression of sex hormone sensitive phenotypes including sexual characteristics and reproductive function in men and women [1]. In addition to regulating sex steroid hormone effects, SHBG may exert independent effects through its own receptor [2]. Variation in SHBG concentration has also been associated with various chronic diseases including cancers [3], polycystic ovary syndrome (PCOS) [4,5] and type 2 diabetes (T2D) [6,7]. Although SHBG is estimated to have a heritable component ( $\sim 50\%$ ) [8], little is known about the genetic regulation of SHBG. Polymorphisms at the SHBG gene locus have been associated with SHBG concentrations [9,10], but much remains unknown about specific genetic variants that may determine circulating SHBG concentrations. Identifying genetic factors that influence SHBG may provide insights into the biology of sex steroid hormone regulation, metabolism and tissue effects that underlie their relationship with chronic diseases such as T2D as well as hormone-sensitive cancers such as breast and prostate cancer.

#### Results

We identified nine loci associated with SHBG concentrations at the genome-wide significance threshold of  $p = 5 \times 10^{-8}$  (Table 1 and Figure 1) in a genome-wide association study (GWAS) metaanalysis of circulating SHBG concentrations in 21,791 men and women from 10 studies (Table S1). All nine lead SNPs at these loci had effects in the same direction (seven with p<0.05) in the validation dataset of 7,046 men and women from six additional studies (Table S2). The strongest association was within the *SHBG* locus (rs12150660,  $p = 2 \times 10^{-106}$ ). Together, these nine lead SNPs explained 7.2% of the genetic variance (assuming 50% heritability) in SHBG concentrations.

We next performed a series of additional analyses to explain more of the phenotypic variance (Figure 2). First, we hypothesized that genetic effects may be different in men and women, as SHBG concentrations are >50% higher in females than males, and may be differentially regulated between sexes. In a sex stratified analysis, three of the nine loci showed evidence of sex-differentiated effects at p < 0.02 when we would not expect any signals to have reached this level of significance by chance. The associations at the 17p13.1-SHBG and Xq22.3 loci were stronger in males whereas the association at the 8q21.13 locus was stronger in females. To investigate the apparent differential sex effect for the X chromosome further we ran a recessive regression model for the X chromosome SNP rs1573036 in women in the Framingham Heart Study and found no association with SHBG suggesting the sex-differentiated effect is not the result of a recessive inheritance pattern. Sex stratified GWAS identified one novel signal in men, which showed no association in women (4q13.2: men  $p = 2.5 \times 10^{-8}$ , women p = 0.66, heterogeneity p = 0.003).

A series of conditional analyses were performed to identify statistically independent signals. At the SHBG locus, three apparently independent additional signals separate from the main index SNP were observed, based on low  $(r^2 < 0.05)$  pairwise correlations in HapMap (rs6258  $p = 2.7 \times 10^{-46}$ , rs1625895  $p = 1.2 \times 10^{-14}$  and rs3853894  $p = 2.5 \times 10^{-11}$ ). A series of iterative conditional analyses (Table 2) involving SNPs at the SHBG locus generated a final regression model including six statistically independent SHBG SNPs. Four of these SNPs (#1-4 Table 2) retained GWS when conditioned against the other five, and two were nominally associated (SNP#5 p = 0.0001, SNP#6 p = 0.01). Re-running the GWAS meta-analysis adjusting for these six SNPs revealed evidence for three additional statistically independent (pairwise HapMap  $r^2 < 0.01$ ) signals at the SHBG locus (SNP#7  $p = 1.5 \times 10^{-7}$ , SNP#8  $p = 4.6 \times 10^{-5}$ , SNP#9  $p = 9.9 \times 10^{-6}$ ) (Figure 3). There were also two additional trans signals located at 2p16.3 and 7q21.3 (Table 1). Although the 2p16.3 signal dropped below GWS when combined with follow-up samples ( $p = 1 \times 10^{-7}$ ), the index SNP at 2p16.3 is  $\sim$  300 kb away from a strong candidate gene, the luteinizing hormone receptor gene (LHCGR).

The majority of pair-wise correlations for the nine SHBG locus SNPs highlighted by our conditional analyses showed very low HapMap  $r^2$  values. However, the pairwise D' values are often high (Table S3) indicating that no or few recombination events have occurred between some SNPs, and that combinations of SNPs may be tagging un-typed variants on a common haplotype. To investigate this possibility, we performed more extensive analyses in a single study (NFBC1966, n = 4467). We used a denser set of SNPs imputed from the June 2011 version of the 1000 Genomes data and performed model selection analyses. Model selection identifies a set of SNPs that best explain phenotypic variation, while simultaneously penalizing each SNP included in this set, and therefore correlated SNPs tend to be excluded from the final model. These analyses consistently included at least seven SNPs in the model, although it is hard to estimate the false-negative rate of using the reduced sample size. While we are underpowered to accurately pinpoint the exact number of independent signals, these analyses support the results of the conditional analysis and suggest that multiple variants at the SHBG locus are independently associated with SHBG concentrations.

Data from an independent study, the InCHIANTI study, was used to calculate the proportion of genetic variance in SHBG concentrations explained when accounting for sex specific effects, the multiple signals of association at the *SHBG* locus, and the additional *trans* signals identified post conditional analysis. In men and women we explained ~15.6% and ~8.4% of the heritable

										Discovery Samples	amples		Discovery	Discovery+Follow-up	
Index SNP	Analysis	Region	Nr Gene	Chr	Position	Effect Allele	Other Allele		EAF B	Beta S	SE	٩	Beta	SE	٩
rs17496332	Main	1p13.3	PRMT6	-	107347898	a	б		0.67 –	-0.026 0	0.0046	1.0E-08	-0.028	0.0041	1.4E-11
rs780093	Main	2p23.3	GCKR	2	27596107	t	υ		0.40 -	-0.033 0	0.0043	5.8E-14	-0.032	0.0039	2.2E-16
rs440837	Main	8q21.13	ZBTB10	ø	81624529	a	D		0.78 -	-0.030 0	0.0052	6.7E-09	-0.028	0.0047	3.4E-09
rs7910927	Main	10q21.3	JMJD1C	10	64808916	t	g		0.51 -	-0.044 0	0.0043	7.4E-25	-0.048	0.0039	6.1E-35
rs4149056	Main	12p12.1	SLCO1B1	12	21222816	t	υ	-	0.82 0.	0.032 0	0.0057	1.5E-08	0.029	0.0052	1.9E-08
rs8023580	Main	15q26.2	NR2F2	15	94509295	t	υ		0.72	-0.029 0	0.0049	2.8E-09	-0.03	0.0044	8.3E-12
rs12150660	Main	17p13.1	SHBG	17	7462640	t	D		0.24 0.	0.100 0	0.0053	1.2E-79	0.103	0.0047	1.8E-106
rs2411984	Main	17q21.32	ZNF652	17	44800750	a	g		0.28 0.	0.034 0	0.0049	1.5E-12	0.033	0.0044	3.5E-14
rs1573036	Main	Xq22.3	TDGF3	23	109706724	t	υ		0.39 0.	0.031 0	0.0043	5.1E-13	0.028	0.0037	4.1E-14
rs10454142	Conditional	2p16.3	LHCGR	2	48499903	t	U		0.69 0.	0.026 0	0.0047	2.8E-08	0.023	0.0044	1.3E-07
rs3779195	Conditional	7q21.3	BAIAP2L1	7	97831298	a	t	-	0.17 -	-0.033 0	0.0057	1.2E-08	-0.028	0.0051	2.7E-08
rs293428	Sex-specific	4q13.2	UGT2B15	4	69626371	a	g		- 69.0	-0.023 0	0.0047	1.6E-06	-0.019	0.0042	5.5E-06
	Discovery Men	' Men		Discovery	ry+Follow-up Men		Discovery Women	Women		Di	scovery+	Discovery+Follow-up Women	Vomen	Heterogeneity	<b>jeneity</b>
Index SNP	Beta	SE	٩	Beta	SE	<u>م</u>	Beta	SE	٩	Be	Beta	SE	٩	٩	Sex
rs17496332	-0.027	0.0054	9.6E-07	-0.027	0.0051	1.5E-07	-0.02	0.008	0.003	0	-0.029	0.0067	1.8E-05	0.79	Females
rs780093	-0.029	0.0052	1.8E-08	-0.026	0.0049	7.0E-08	-0.04	0.0076	1.3E-07	-	-0.041	0.0063	8.6E-11	0.07	Females
rs440837	-0.021	0.0062	0.0009	-0.019	0.0058	0.001	-0.049	0.0093	1.2E-07	-	-0.042	0.0078	7.2E-08	0.02	Females
rs7910927	-0.049	0.0051	5.3E-22	-0.050	0.0048	1.2E-25	-0.038	0.0075	6.4E-07	-	-0.046	0.0063	1.7E-13	0.63	Males
rs4149056	0.028	0.0067	0.00003	0.027	0.0063	1.5E-05	0.049	0.0103	0.000002	2 0.037	37	0.0086	1.7E-05	0.36	Females
rs8023580	-0.024	0.0057	0.00002	-0.025	0.0054	5.1E-06	-0.038	0.0087	0.00001	-	-0.038	0.0071	7.8E-08	0.13	Females
rs12150660	0.106	0.0063	1.8E-63	0.110	0.0058	3.7E-80	0.085	0.0094	1.8E-19	0.087	87	0.0077	5.8E-30	0.02	Males
rs2411984	0.034	0.0058	5.9E-09	0.034	0.0054	2.3E-10 (	0.032	0.0084	0.0001	0.029	29	0.007	3.2E-05	0.54	Males
rs1573036	0.04	0.0048	9.1E-17	0.035	0.0043	2.8E-16 (	0.012	0.0083	0.15	0.016	16	0.007	0.02	0.02	Males
rs10454142															
rs3779195															
rs293428	-0.032	0.0056	1.5E-08	-0.029	0.0053	2.5E-08	-0.005	0.0085	0.57	0-	-0.003	0.0071	0.66	0.003	Males

Table 1. SNPs representing loci associated with circulating SHBG concentrations.

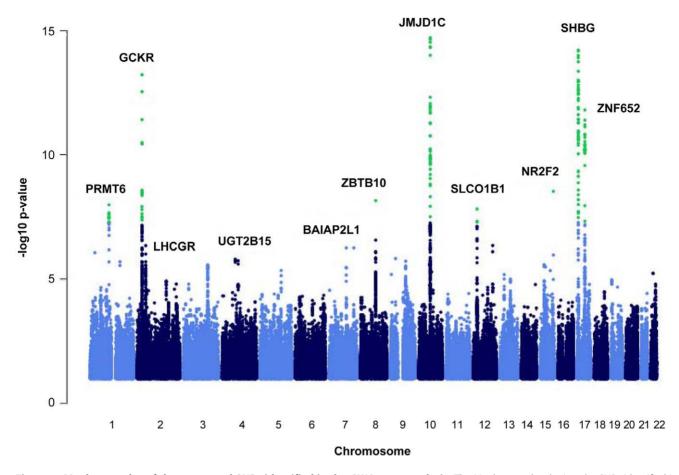


Figure 1. Manhattan plot of the autosomal SNPs identified in the GWA meta-analysis. The Manhattan plot depicts the SNPs identified in the GWAS analysis labeled with the nearest gene on the plot. The SNP identified on the X chromosome, rs1573036, at Xq22.3, is not included in this figure. doi:10.1371/journal.pgen.1002805.g001

component respectively. The *SHBG* locus accounted for  $\sim 10\%$  and  $\sim 6.6\%$  of the genetic variation in men and women respectively with the lead SNP in isolation accounting for  $\sim 7.8\%$  and  $\sim 3.3\%$  of the variation in men and women, respectively.

We identified genes near the associated SNPs and explored their biologic relevance to SHBG. The genes associated with identified SNPs included the *SHBG* locus (rs12150660, 17p13.1, p=1.8×10<sup>-106</sup>), *PRMT6* (rs17496332, 1p13.3, p=1.4×10<sup>-11</sup>), *GCKR* (rs780093, 2p23.3, p=2.2×10<sup>-16</sup>), *ZBTB10* (rs440837, 8q21.13, p=3.4×10<sup>-09</sup>), *JMJD1C* (rs7910927, 10q21.3, p=6.1×10<sup>-35</sup>), *SLCO1B1* (rs4149056, 12p12.1, p=1.9×10<sup>-08</sup>), *JR2F2* (rs8023580, 15q26.2, p=8.3×10<sup>-12</sup>), *ZNF652* (rs2411984, 17q21.32, p=3.5×10<sup>-14</sup>), *TDGF3* (rs1573036, Xq22.3, p=4.1×10<sup>-14</sup>), *LHCGR* (rs10454142, 2p16.3, p=1.3×10<sup>-07</sup>), *BAIAP2L1* (rs3779195, 7q21.3, p=2.7×10<sup>-08</sup>), and *UGT2B15* (rs293428, 4q13.2, p=5.5×10<sup>-06</sup>) (Figure 1).

We used the online tool STRING (www.string-db.org) to perform pathway analyses to explore possible interactions between the *SHBG* gene and the proteins encoded by the 11 most plausible genes nearest the 11 SNPs listed above. There was an interaction noted between *GCKR* and *JMJD1C* which were associated with the lipoprotein fractions VLDL and HDL, respectively [11]. In an expanded analysis, we assessed protein interactions among *SHBG* and 67 genes within 500 kb of our 11 identified SNPs and uncovered additional protein interaction pathways. An interaction between two proteins encoded by *GTF2A1L* and *STON1* was found; these proteins are co-expressed in testicular germ cells in the mouse [12]. An interaction between *LHCGR* and *BR13* encoded proteins that are associated with the G-protein coupled receptor complex in the human luteinizing hormone receptor was also identified [13]. Finally, an interaction between *LHCGR* and *LAPP* (amylin) proteins which are components of a ligand/G-protein receptor/G-protein alpha subunit complex was found (database: www.reactome.com).

Targeted analysis of two strong candidate genes, hepatocyte nuclear factor- $4\alpha$  (*HNF4* $\alpha$ ) and peroxisome-proliferating receptor  $\gamma$  (*PPAR* $\gamma$ ) did not identify any SNPs at *HNF4* $\alpha$  but did identify one SNP, rs2920502, at *PPAR* $\gamma$  that reached statistical significance (p = 9.9×10<sup>-5</sup>) and a second SNP at *PPAR* $\gamma$ , rs13081389, that reached nominal significance (p = 0.01).

#### Discussion

In total, we identified 12 genomic regions associated with circulating SHBG concentrations, including extensive allelic heterogeneity at the *SHBG* locus itself. Conditional meta-analyses carried out at the *SHBG* locus, identified nine genome-wide significant SNPs with low correlation ( $r^2 < 0.01$ ) between them. Two of these signals (rs6258 [10] and rs6259) are missense variants and two are low frequency variants (MAF ~2%). Furthermore, rs12150660 is highly correlated ( $r^2 > 0.95$ ) [10] with a pentanu-



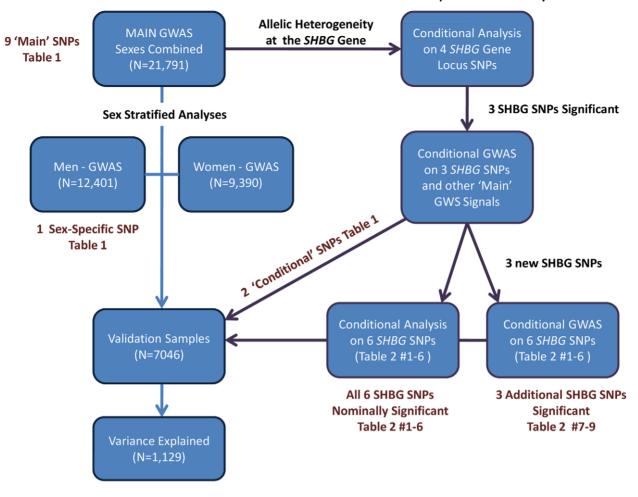


Figure 2. Summary of the analytic plan. doi:10.1371/journal.pgen.1002805.g002

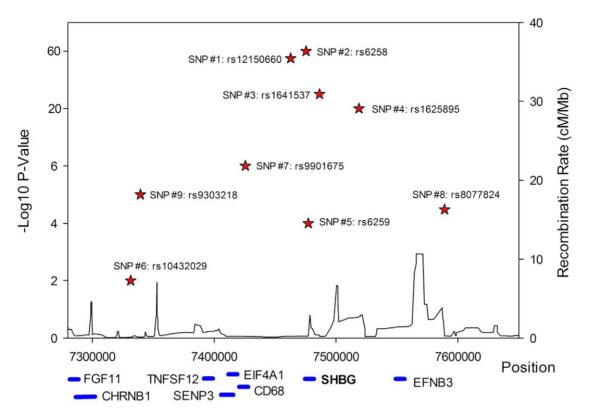
cleotide repeat, which affects *SHBG* expression *in-vitro* [14]. To our knowledge, the magnitude of secondary signals observed at this locus are the largest seen for any complex trait.

The proportion of genetic variance in SHBG serum concentrations explained when accounting for sex specific effects, the multiple signals of association at the *SHBG* locus, and the

SNP #	Model	Conditioned On SNP $\#$	SNP	Position	Effect Allele	Other Allele	EAF	Beta	SE	p-value	Discovery p-value	Discovery Beta
1	Full model	2–6	rs12150660	7462640	t	g	0.24	0.082	0.005	1.89E-55	1.19E-79	0.10
2	Full model	1,3–6	rs6258	7475403	t	с	0.02	-0.272	0.017	1.03E-60	2.69E-46	-0.2613
3	Full model	1–2,4–6	rs1641537	7486446	t	с	0.14	-0.064	0.006	1.20E-24	8.19E-39	-0.0814
4	Full model	1–3,5–6	rs1625895	7518840	t	с	0.12	-0.06	0.006	1.75E-21	1.17E-14	-0.052
5	Full model	1–4,6	rs6259	7477252	а	g	0.11	0.026	0.007	0.0001	1.46E-07	0.0372
6	Full model	1–5	rs10432029	7331393	а	g	0.79	0.0136	0.006	0.01	7.52E-16	0.0446
7	Conditional	1–6	rs9901675	7425536	а	g	0.05	-0.057	0.01	1.46E-07	5.2E-12	-0.07
8	Conditional	1–6	rs8077824	7588951	а	g	0.02	0.075	0.018	4.58E-05	0.01	0.0451
9	Conditional	1–6	rs9303218	7339386	t	с	0.77	0.026	0.006	9,89E-06	1.21E-11	0.0344

Table 2. Statistically independent signals at the SHBG gene locus.

All SNPs are on the+strand and positions are based on build 36. EAF = 'effect allele frequency'. Beta units are per-allele effect estimates in natural log transformed nmol/ L. 'Full model' SNPs were all included in a single regression model, where the effect estimates for each SNP are adjusted for the effect of the others in the model. 'Conditional' SNPs are SNPs with low pair-wise LD (HapMap  $r^2 < 0.01$ ) that were identified after conditioning on the full model SNPs. doi:10.1371/journal.pgen.1002805.t002



**Figure 3. Allelic heterogeneity at the** *SHBG* **gene locus.** There was significant allelic heterogeneity at the SHBG gene locus. The nine independent signals identified in the SHBG gene are shown in relation to their position within the gene. All positions based on build 36. Not all genes are shown.

doi:10.1371/journal.pgen.1002805.g003

additional *trans* signals identified post conditional analysis was  $\sim 15.6\%$  in men and  $\sim 8.4\%$  in women. The *SHBG* locus accounted for  $\sim 10\%$  and  $\sim 6.6\%$  of the genetic variance in men and women, respectively, with the lead SNP explaining most of the genetic variation at  $\sim 7.8\%$  for men and  $\sim 3.3\%$  for women. Thus additional signals at the *SHBG* locus identified through conditional analyses approximately doubled the variance of the trait explained. While we provide evidence for multiple variants associated with SHBG concentrations, further studies are needed to pinpoint the causal loci and functional variants. For the 11 regions outside the *SHBG* locus, most have biologically plausible related genes within 300 kb.

#### Biology of Plausible Genes near Identified SNPs

Several genes near the identified SNPs regulate sex steroid production and function. The NR2F2 locus (15q26.2) encodes a nuclear receptor important in testicular Leydig cell function, the primary source of gonadal testosterone production [15], and has been linked to male infertility [16]. NR2F2 has also been associated with estrogen receptor alpha (ER $\alpha$ ) signaling and may influence hormone responsivity in breast cancer [17]. PRMT6 (1p13.3) also encodes a nuclear receptor regulatory protein that mediates estrogen signaling as a co-activator of the estrogen receptor [18]. LHCGR (2p16.3) encodes the luteinizing hormone receptor which was associated with polycystic ovary syndrome (PCOS) in a recent GWAS [19,20]. PCOS is both a reproductive and metabolic disorder characterized by higher testosterone serum concentrations as well as an increased prevalence of obesity, insulin resistance, and T2D in women. Inappropriate secretion of luteinizing hormone leads to increased ovarian production of testosterone. Coincident lower SHBG concentrations contribute to increased bioavailable testosterone concentrations and the expression of both reproductive and metabolic phenotypes in PCOS [21,22,23].

The *SLCO1B1* locus encodes a liver-specific transporter of thyroid hormone as well as estrogens which impact liver production of SHBG [24].  $\mathcal{JMJD1C}$  (10q21.3), also known as *TRIP 8* (thyroid hormone receptor interactor protein 8 [25]), may impact SHBG concentrations via thyroid hormone effects on liver protein production. Thyroid hormone may alter SHBG production through effects on  $HNF4\alpha$  which is known to regulate *SHBG* transcription [26,27].

Many of the genes identified are involved in carbohydrate and lipid metabolism and liver function. The GCKR locus (2p23.3) encodes a protein that regulates glucokinase activity and has been associated with T2D in several ethnic populations [28,29,30,31]. GCKR has been associated with metabolic and inflammatory traits including triglyceride concentrations and other lipid fractions [30,32], fasting plasma glucose [33,34], insulin concentrations, uric acid, c-reactive protein (CRP), and non-alcoholic fatty liver disease which are all characteristic of the metabolic syndrome and T2D [28,35,36,37,38,39,40,41,42]. The SLCO1B1 locus (12p12.1) codes for a protein, hepatocyte protein anion-transporting polypeptide 1B1, involved in liver metabolism of both endogenous and exogenous compounds [43]. Consistent with SLCO1B1's role in liver metabolism, the same SNP (rs4149056) has been associated with circulating bilirubin concentrations in previous GWAS [44]. BAIAP2L1 (7q21.3) encodes a protein important in cytoskeleton organization [45] that has been associated with the inflammatory marker CRP in patients with arthritis [46]. BAIAP2L1 is also known as IRTKS (insulin receptor tyrosine kinase substrate) which

is involved in insulin receptor signaling [47] and may relate to insulin resistant states including obesity and T2D [48,49, 50,51,52,53,54]. We conducted a targeted analysis of *PPAR* $\gamma$ , a gene that influences *SHBG* gene expression in the liver [1,55] and is associated with T2D [56,57]. Our analysis identified one significant SNP (rs2920502, p = 9.9×10<sup>-5</sup>) and a second nominally significant SNP (rs13081389, p = 0.01) at *PPAR* $\gamma$ . Some of the identified genes involved in hepatic metabolism of lipids and carbohydrates may be affect SHBG concentrations indirectly through effects on the SHBG transcription regulator *HNF4* $\alpha$ although *HNF4* $\alpha$  itself was not identified in this meta-analyses [27,58,59,60].

The UGT2B15 locus (4q13.2) was significantly associated with SHBG concentrations in men but not women in this metaanalysis. UGT2B15 belongs to a family of genes (the UGT2B gene family) that code for enzymes involved in the metabolism of sex hormones through glucuronidation which allows for excretion of sex steroids through the kidney and the gut via bile excretion [61,62], primary clearance mechanisms for sex steroids [63]. UGT2B15 is involved in the conjugation and inactivation of testosterone [64]. An association between rs293428 in the UGT2B15 locus and circulating SHBG concentrations in men is supported by a previous study demonstrating that a nonsynonymous SNP in UGT2B15 (rs1902023; D85Y) is associated with serum SHBG concentrations in younger adult men [65]. UGT2B15 is thought to play a significant role in local tissue inactivation of androgens in androgen dependent prostate cancer [66,67]. The mechanism behind the influence of genetic variants in UGT2B15 on SHBG concentrations is unknown, but one may speculate that UGT2B15 affects the local androgenic environment in selected tissues, which in turn results in regulation of SHBG concentrations.

In addition to UGT2B15, three other genes near the identified SNPs are associated with carcinogenesis, particularly in the prostate and breast. ZBTB10 (8q21.13), has been linked to breast cancer [68]. In breast cancer cell lines ZBTB10 is suppressed by ROS-microRNA27a thereby enhancing ER $\alpha$  alpha expression and mediating estrogen effects [17]. The ZNF652 (17q21.32) locus codes for a DNA binding protein thought to act as a tumor suppressor gene in breast cancer [69,70,71] that is also co-expressed with the androgen receptor in prostate cancer [72]. TDGF3, teratocarcinoma derived growth factor 3, is the only significant region identified on the X chromosome ((Xq22.3). TDGF3 is a *pseudogene* of TDGF1 located on chromosome 3p23-p21 that has been associated with testicular germ cell tumors [73].

#### Strengths and Limitations

This GWAS meta-analysis incorporated data from approximately 22,000 men and women from 16 epidemiologic cohorts. The overall size of the study yields power but the meta-analysis of data from different epidemiologic studies requires the inclusion of different laboratory methods. The different studies used a variety of assay methodologies to measure serum SHBG concentrations although the vast majority were immunoassays (Tables S1 and S2, Text S1) with similar methodologies. Variation introduced by the use of different SHBG assays would result in loss of statistical power and likely bias toward the null. Additionally, the majority of women were post-menopausal as ascertained by self-report in all studies (Table S1). SHBG concentrations, like testosterone, decline only slightly across the menopause [74] so adjustment for menopause status is not necessary. SHBG may also increase with ovulation and be slightly higher in the luteal versus the follicular phase of the menstrual cycle in premenopausal women, but most studies did not collect data on menstrual phase at the time of SHBG measurement so adjustment for menstrual phase was not possible [75]. Finally, individuals were not excluded based on health status, therefore some individuals with chronic conditions that may affect hepatic production of or clearance of proteins including SHBG such as liver disease, renal disease, or severe malnutrition, may have been included in this analysis.

#### Conclusion

SHBG synthesis in the liver is known to be affected directly or indirectly by estrogens, androgens and thyroid hormones and has been observed to be inversely associated with the higher insulin concentrations characteristic of insulin resistant states such as T2D [1,6]. In summary, the results of this GWAS reflect these influences. Three regions map to proteins related to hepatic function (12p12.1-SLCO1B1 [76], 2p23.3-GCKR [77] and 10q21.3-7M7D1C [77]). In addition, 2p23.3-GCKR and 7q21.3-BAIAP2L1 [alias insulin receptor tyrosine kinase substrate (IRTKS)] are involved in susceptibility to T2D [48] and insulin signaling [47], respectively. Two signals also mapped to loci involved in thyroid hormone regulation (10q21.3-7M7D1C and 12p12.1-SLCO1B1). One signal mapped to the receptor for luteinizing hormone 2p16.3-LHCGR [20], the hormone that stimulates testosterone production. Five regions mapped to genes previously implicated in androgen and estrogen signaling (1p13.3-PRMT6 [18], 8g21.13-ZBTB10 [17], 12p12.1-SLCO1B1 [76], 15q26.2-NR2F2 [78], 4q13.2-UGT2B15 [63]).

We have combined a conventional GWAS approach with detailed additional analyses, including sex stratification, conditional analysis and imputation from 1000 Genomes. Our results demonstrate that these approaches can lead to an appreciable gain in heritable variance explained. It does however highlight the complexity of elucidating individual variant causality through statistical approaches. In addition to the extensive allelic heterogeneity at the *SHBG* locus, our data identify loci with a role in sex steroid hormone metabolism, which may help elucidate the role of sex steroid hormones in disease, particularly T2D and hormonesensitive cancers.

#### Methods

We performed a genome wide association study (GWAS) metaanalysis of 21,791 individuals (Table S1: 9,390 women, 12,401 men) from ten observational studies. Data from an additional six studies totaling 7,046 individuals (Table S2: 4,509 women; 2,537 men) were used for validation. The proportion of variance explained was estimated in an independent study (InCHIANTI, n = 1,129). The individual study protocols were approved by their respective institution's ethics committee/institutional review board and all participants provided informed consent prior to participation. Individuals known to be taking hormonal contraceptives or hormone replacement therapy at time of SHBG measurement were excluded from analysis. Age, sex and body mass index (BMI) were included as covariates. After applying standard quality control measures, imputed genotypes were available for approximately 2.5 M SNPs. See Figure 2 for an overview of the analytic plan and the Text S1 for further information for individual studies included in this meta-analysis.

#### GWAS Conditional Meta-Analysis Steps

**Conditional analysis** #1. The initial starting point for the conditional analysis was the four *SHBG* locus SNPs that all showed low Hapmap LD ( $r^2 < 0.05$ ) with each other: rs12150660 (lead SNP Table 1), rs6258 p= $2.7 \times 10^{-46}$ , rs1625895 p= $1.2 \times 10^{-14}$  and rs3853894 p= $2.5 \times 10^{-11}$ . Each cohort fitted a single

regression model, fitting SHBG concentrations against these four genome-wide significant SHBG locus SNPs (rs12150660, rs6258, rs1625895 and rs3853894), in addition to age, sex and BMI. After meta-analyzing the results from all cohorts, three of the SNPs retained genome wide significance when regressed against each other, with the fourth SNP narrowly missing that threshold (rs3853894,  $p = 4.1 \times 10^{-6}$ ).

**Conditional GWAS #1 (Table 1, conditional analysis).** We next performed a conditional GWAS meta-analysis, where each study included, as additional covariates to the original analysis plan, the ten genome-wide significant autosomal SNPs (the eight 'Main' signals from Table 1 and the two unique *SHBG* locus signals described above in addition to the lead SNP rs12150660: rs6258 and rs1625895). Three additional signals (independence based on HapMap  $r^2 < 0.05$ ) at the *SHBG* locus reached genome-wide significance (rs1641537 p =  $7.8 \times 10^{-32}$ , rs6259 p =  $1.5 \times 10^{-12}$  and rs10432029 p =  $3 \times 10^{-8}$ ), giving a total of six independent signals in this gene region. In addition, two novel signals reached genome-wide significance in the conditional analysis, at 7q21.3 (rs3779195 p =  $1 \times 10^{-8}$ ) and 2p16.3 (rs10454142 p =  $3 \times 10^{-8}$ ). After replication, only rs3779195 at the *BAIAP2L1* locus retained genome-wide significance.

**Conditional analysis** #2 (Table 2, full model). Given the six signals observed at the *SHBG* locus (three through conditional analysis #1 rs12150660, rs6258, rs1625895, three through LD estimates from conditional GWAS #1: rs1641537, rs6259, rs10432029), we sought to confirm which of these six were truly independent by a second round of conditional analysis. All discovery and replication cohorts fitted a single regression model of the six SNPs (SNPs # 1–6, Table 2) against SHBG concentrations, using the same parameters and covariates as conditional analysis #1. Four of the six SNPs (#1–4: rs12150660, rs6258, rs1641537, and rs1625895) retained genome-wide significance when conditioned against each other, with two showing nominal evidence of association (SNP #5 rs6259, p = 0.0001; SNP #6 rs10432029, p = 0.01).

**Conditional GWAS #2 (Table 2, conditional model).** Finally, we performed a second conditional GWAS analysis, adjusting for the six *SHBG* locus SNPs which had evidence of association from conditional analysis #2. All the discovery cohorts were used in this analysis, in addition to three replication cohorts (total sample size 24,354). This analysis revealed evidence for a further three independent signals at the *SHBG* locus (based on HapMap  $r^2 < 0.01$ ), SNP #7 rs9901675 p =  $1.5 \times 10^{-7}$ , SNP #8 rs8077824 p =  $4.6 \times 10^{-5}$ , and SNP #9 rs9393218 p =  $9.9 \times 10^{-6}$ .

### Sensitivity Analysis—Allelic Heterogeneity at the SHBG Locus

We performed a sensitivity analysis using samples from the 1966 Northern Finland Birth Cohort (NFBC1966) study to further investigate allelic heterogeneity at the SHBG locus (Text S1). The conditional meta-analysis showed evidence for up to nine signals at the SHBG locus, but it is possible that these signals could be explaining a much smaller number of causal variants in the region. Since 1000 Genomes imputation allows us to assess the genetic variation associated with a phenotype across a much denser set of markers, it increases our power to detect allelic heterogeneity within a region. Therefore, 1000 Genomes imputation was carried out on all the samples in the NFBC1966 study and forward selection was used to identify the set of SNPs that best explain the variation in the SHBG phenotype. 1000 Genomes imputation was carried out using IMPUTE2. The mean genotype probabilities for each SNP were calculated and used in the model selection step. Only SNPs 250 kb upstream and 250 kb downstream from the SHBG locus (7283453–7786700 bp) were used in the analysis. All SNPs with MAF <0.1% or an imputation quality score less than 0.4 were excluded from the analysis. In total, 1978 SHBG region SNPs measured or imputed in 4467 samples from the NFBC1966 study were used in the sensitivity analysis. Forward selection was implemented in R (version 2.13.0) using the stepAIC package to estimate the Akaikie Information Criterion (AIC), an inclusion parameter. Given the high degree of correlation between the SNPs in this region, we increased the penalty (k) on the number of terms included in the model to 12 (where it is usually two), to minimize possible over fitting. The final model included seven SNPs, adjusted for sex and BMI.

#### Pathway Analysis

We examined potential interactions among the proteins encoded by the SHBG locus and the proteins encoded by the 11 genes (ZBT10, TDGF1, ZNF652, PRMT6, JMJD1C, GCKR, BAIAP2L1, LHCGR, SLCO1B1, UGT2B15, NR2F2) closest to the 11 identified SNPs using pathway analysis with Search Tool for the Retrieval of Interacting Genes/Proteins (STRING) Pathways Analysis (www.string-db.org). The interactions explored by STRING include direct (physical) and indirect (functional) associations. We then expanded the analysis to examine protein interactions among the SHBG gene and the proteins encoded by 67 genes within 500 kb of the 11 identified SNPs.

#### Targeted Candidate Gene Analysis

We conducted targeted analysis of two strong candidate genes, hepatocyte nuclear factor- $4\alpha$  (*HNF4* $\alpha$ ) and peroxisome-proliferating receptor  $\gamma$  (*PPAR* $\gamma$ ). Statistical significance thresholds were set correcting for the number of SNPs tested in each gene region (±100 kb).

#### **Supporting Information**

 Table S1
 Characteristics of 21,791 individuals from 10 discovery cohorts included in the meta-analysis.

 (DOC)
 (DOC)

(DOC)

**Table S2** Characteristics of 8,175 individuals from the six cohorts included in the validation analysis (WHI, CARDIA, Prospect-EPIC, MrOs, NHS, YFS) and the independent cohort used to estimate the proportion of genetic variance explained by the indentified SNPs (InChianti).

**Table S3** Hapmap (release 22) linkage disequilibrium estimates for the nine *SHBG* gene locus single nucleotide polymorphisms. (DOC)

**Text S1** Supplementary Methods with Specific Cohort Information.

(DOC)

#### Acknowledgments

We are grateful to all study participants and staff in our participating studies. Full study acknowledgements are available in Text S1.

#### **Author Contributions**

Conceived and designed the experiments: FH de Jong, O Raitakari, A Teumer, C Ohlsson, JM Murabito, JRB Perry. Analyzed the data: JRB Perry, AD Coviello, R Haring, M Wellons, D Vaidya, T Lehtimäki, S Keildson, KL Lunetta, C He, M-R Järvelin. Wrote the paper: AD Coviello, R Haring, M Wellons, D Vaidya, T Lehtimäki, S Keildson, KL Lunetta, C He, TM Frayling, A Murray, S Franks, M-R Järvelin, FH de

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