Apolipoprotein E Genotype and the Diagnostic Accuracy of Cerebrospinal Fluid Biomarkers for Alzheimer Disease.

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APOE genotype and the diagnostic accuracy of CSF biomarkers for Alzheimer disease

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Conflicts of interest: ML declares that, over the past three years, he has received compensation for lectures from AstraZeneca, Bayer, Biophausia, Bristol Myers-Squibb, Lundbeck pharmaceuticals, Eli Lilly Sweden, Wyeth, Servier Sweden, and served at advisory board for AstraZeneca and Lundbeck pharmaceuticals. No other equity ownership, profit-sharing agreements, royalties, or patent. RO is an employee at GE Healthcare. HH declares no competing financial interests related to the present article. During the last two years (2011-2013) he has received lecture honoraria and/or research grants and/or travel funding and/or participated in scientific advisory boards and/or as a consultant to diagnostic, biotechnology and pharmaceutical companies involved in the manufacture and marketing of biomarkers and/or diagnostics and/or drugs or medicinal products for cognitive impairment and Alzheimer's disease including Boehringer-Ingelheim, Bristol-Myers Squibb, Elan Corporation, Wyeth, Novartis, Eisai Inc., Pfizer, Schwabe, Sanofi-Aventis, Roche Pharmaceuticals and Diagnostics, GE Healthcare, Astra-Zeneca, Avid, Eli Lilly and Company, Janssen-Cilag, Merz Pharmaceuticals, GlaxoSmithKline-Biologicals, Jung-Diagnostics, Thermo Fisher Scientific Clinical Diagnostics, Cytox. He is co-inventor in pending patent submissions relating to biological markers and/or diagnostics and has not received any royalties. HZ declares no conflicts of interest. KB has served at Advisory Boards for Pfizer, Roche, Lilly and Innogenetics.

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Abstract

Background

Several studies suggest that the APOE ε4 allele modulates cerebrospinal fluid (CSF) levels of β-amyloid_{1-42} (Aβ42). However, it is unknown whether this effect is secondary to the association of the APOE ε4 allele with cortical Aβ deposition or whether APOE ε4 directly influences CSF Aβ42 levels in an Aβ pathology-independent manner.

Objective

We evaluated whether the APOE genotype affects the diagnostic accuracy of CSF biomarkers for AD, CSF Aβ42 in particular, and whether the association of APOE ε4 with CSF biomarkers depends on cortical Aβ status.

Design

Multicenter study.

Setting

Data from four different centers in Sweden, Finland and Germany as well as from the North American multicenter study ADNI.

Participants

Cohort A: 1345 individuals (23-99 y) with baseline CSF samples, including 309 with AD, 287 with prodromal AD, 251 controls, 399 with stable mild cognitive impairment (sMCI) and 99 with dementias other than AD. Cohort B: 105 non-demented younger individuals (20-34 y) with CSF taps. Cohort C: 118 patients (60-80 y) with mild cognitive symptoms and \[^{18}F]\text{fluorimetamol} PET amyloid imaging and CSF taps.
Main outcome measures

CSF Aβ42, total tau (T-tau) and phosphorylated tau (P-tau) in relation to the APOE ε2/ε3/ε4 polymorphism in different diagnostic groups and in cases with or without [18F]flutemetamol cortical uptake.

Results

CSF Aβ42, but not T-tau and P-tau, was lower in APOE ε4 carriers as compared to non-carriers irrespective of diagnostic group (cohort A). Despite this, CSF Aβ42 differed between subjects with AD when compared to controls and sMCI, even when stratifying for APOE genotype. Multiple binary logistic regression revealed that CSF Aβ42 and APOE ε4 genotype were independent predictors of AD diagnosis. In cohort B (individuals <35 years), APOE ε4 carrier status did not influence CSF Aβ42 levels. Moreover, when stratifying for [18F]flutemetamol cortical uptake in cohort C, APOE ε4 genotype did not influence CSF Aβ42 levels. This result was replicated in ADNI using 11C-Pittsburgh compound B (11C-PiB).

Conclusion

CSF Aβ42 is strongly associated with both AD diagnosis and cortical Aβ accumulation independent of APOE genotype. The clinical cut off for CSF Aβ42 should be the same for all APOE genotypes.
Introduction

The apolipoprotein E (APOE) genotype is the most prominent susceptibility gene for late-onset Alzheimer disease (AD).1 Two polymorphisms (rs7412 and rs429358) make up three different alleles, ε2, ε3 and ε4, of the APOE gene. These polymorphisms lead to amino acid substitutions at positions 112 and 158 in the ApoE protein. The ε4 allele is known to increase the risk and lower the age at onset of AD in a gene dose-dependent manner. As compared to subjects lacking the ε4 allele, individuals homozygous for the ε4 allele have an approximately 12-fold increased risk of AD and an age at onset around 65 years, while heterozygous carriers have about three-fold increased risk and an age at onset around 75 years.2 The exact pathophysiological mechanisms underlying this strong genetic association are yet to be revealed, but some data point towards an impaired clearance of Aβ from the brains of APOE ε4-positive individuals as a possible key factor.3,4

With the emergence of biomarker-supported dementia diagnostics,5-7 there is an increasing interest in cerebrospinal fluid (CSF) biomarkers associated with AD, especially β-amyloid1-42 (Aβ42) and tau proteins.8 Low CSF levels of Aβ42 indicate ongoing AD but several studies have also shown decreased levels of Aβ42 in CSF in APOE ε4-positive individuals without clinical AD.9-12 It is unknown whether the effect of APOE ε4 on CSF Aβ levels is secondary to the association of the APOE ε4 allele with cortical Aβ deposition or whether APOE ε4 directly influences CSF Aβ42 levels in an Aβ pathology-independent manner. Further, for optimal clinical usage of genetic and CSF biomarkers, studies are needed to clarify to what extent
APOE genotype and CSF biomarkers correlate and provide overlapping versus complementing information for diagnosis and prognosis of AD and whether different clinical cut offs for CSF Aβ42 should be used depending on APOE genotype. Several studies have emphasized that the APOE ε4 allele could affect the diagnostic power of CSF Aβ42 and that APOE genotype should be taken into account when using CSF Aβ42 as a biomarker for AD. Here, we approached these issues by evaluating the effects of the APOE ε2/ε3/ε4 polymorphism on the diagnostic accuracy of CSF Aβ42, total tau (T-tau) and phosphorylated tau (P-tau) for AD in a cohort comprising 1345 individuals. We also assessed the association of CSF biomarker levels with APOE genotype and/or cortical amyloid deposition i) in a cohort with younger individuals, ii) in patients with mild cognitive symptoms with and without abnormal cortical Aβ42 uptake of [18F]flutemetamol and iii) in the Alzheimer Disease Neuroimaging Initiative (ADNI) cohort in subjects who had undergone both CSF biomarker analyses and 11C-Pittsburgh compound B PET.

Material and methods

Cohorts

Cohort A: Four memory clinics in Sweden, Finland and Germany took part in the study. The total cohort comprised 251 controls, 399 patients with stable mild cognitive impairment (sMCI), 287 patients with prodromal AD (MCI-AD), 309 demented patients with AD, and 99 patients with other dementias than AD. Patients in the sMCI group were followed for at least 2 years (median 3 years, range 2-11 years). All participants were assessed by physicians
specializing in cognitive disorders who were blinded to all CSF results. Parts of this cohort, including 186 patients from the ongoing prospective clinical longitudinal Gothenburg MCI study\textsuperscript{16}, have been included in earlier publications from our groups\textsuperscript{17-20}

Cohort B: The study also included a separate cohort comprising 105 individuals younger than 35 years (mean age 27.7 $\pm$ 3.8 years) without neurodegenerative conditions (67 patients with bipolar disorder and 38 healthy controls). This cohort was only used to assess the association of APOE $\varepsilon$4 with CSF biomarker levels but was not included in the studies of the diagnostic accuracy of the biomarkers due to their low age.

Cohort C: These subjects were included from the larger BioFINDER study (Biomarkers For Identifying Neurodegenerative Disorders Early and Reliably), which enrolls consecutive non-demented patients with mild cognitive symptoms from three memory clinics in Sweden. More information regarding the BioFINDER study will be available at www.biofinder.se. From this study, we selected the first 118 patients who had undergone both [\textsuperscript{18}F]flutemetamol PET imaging and CSF taps. Fifty-three percent of these were classified as having subjective MCI and 47% as objective MCI based on an extensive neuropsychological battery and the clinical assessment of a neuropsychologist. Among those with MCI, 76% had amnestic MCI (46% single domain and 30% multi domain) and 24% had non-amnestic MCI.

ADNI cohort: 53 subjects (9 with AD, 33 with MCI and 11 healthy controls) with data on both CSF analysis and \textsuperscript{11}C-PiB scans were obtained from the Alzheimer’s disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu).

For a more detailed description of the cohorts see eMethods 1 in the supplement.
**Lumbar puncture**

CSF samples were obtained by lumbar puncture in the L3/4 or L4/5 interspace without any reported serious side effects, collected in polypropylene tubes, centrifuged and stored frozen at -80°C until analysis according to standard operating procedures. Most biomarker measurements were performed at the Clinical Neurochemistry Laboratory at the Sahlgrenska University Hospital, Mölndal, Sweden, but samples from Kuopio, Finland and Munich, Germany were analyzed locally.

**CSF analyses**

CSF T-tau levels were measured using a sandwich enzyme-linked immunosorbent assay (ELISA, INNOTEST hTAU-Ag, Innogenetics, Ghent, Belgium), which detects all tau isoforms irrespective of phosphorylation status, as previously described. CSF P-tau (Tau phosphorylated at threonine 181) levels were determined using a sandwich-ELISA assay (INNOTEST Phospho-Tau[181P]), as previously described. The concentration of CSF Aβ42 was measured using a sandwich-ELISA (INNOTEST β-amyloid[1-42]), designed to detect both the 1st and the 42nd amino acid in the Aβ protein, as previously described. A subset of the samples were analyzed for T-tau, P-tau and Aβ42 using the xMAP Luminex AlzBio3 assay (Innogenetics, Ghent, Belgium), normalized to INNOTEST concentrations as previously described. All analyses were carried out by experienced laboratory technicians who were blinded to the study participants’ diagnosis and other clinical information.
To adjust for variation in biomarker levels between the different laboratories, data were normalized by defining one center cohort as reference group and then calculating factors between the APOE ε4-negative controls from each participating center and the APOE ε4-negative controls in the reference group. These factors were then applied to all data, hence relating biomarker levels in all the different cohorts to those in the reference group. Cross-fertilization of standard samples in each assay was not used, which is a limitation of the study.

**APOE**

APOE (gene map locus 19q13.2) genotyping was performed using TaqMan® Allelic Discrimination technology (Applied Biosystems, Foster City, CA) or equivalent techniques. Genotypes were obtained for the two SNPs that are used to unambiguously define the ε2, ε3, and ε4 alleles (rs7412 and rs429358).

\[^{[18F]}\text{Flutemetamol PET acquisition and analysis}\]

Flutemetamol \(^{[18F]}\) Injection was manufactured by GE Healthcare\(^{25}\) and PET/CT scanning of the whole brain was conducted at two sites (Malmö and Lund in Sweden) as described previously.\(^{26}\) For a detailed description of PET acquisition and analysis see eMethods 2 in the supplement.

**Statistical analysis**

Pair-wise comparisons of biomarker levels between and within the diagnostic groups were performed using a Mann-Whitney-U test for independent samples. Comparisons between
more than two groups were performed using a Kruskal-Wallis-H test for several independent samples. The area under the receiver operating characteristics (ROC) curve was calculated for all biomarkers and separately for each APOE ε4 carrier group in patients with AD versus controls as well as sMCI versus prodromal AD (MCI-AD). Multiple backward stepwise binary logistic regression was performed to simultaneously study the associations between clinical diagnosis versus biomarker levels as well as age as continuous variables, and gender and APOE genotype (carriers of zero, one or two APOE ε4 alleles) as nominal variables. General linear model (ANCOVA) was used to examine the association between CSF Aβ42 (independent variable) and APOE ε4 (carriers of zero, or 1-2 APOE ε4 alleles) when adjusting for [18F]flutemetamol (dichotomized). Statistical significance was determined at P<0.05. All statistical calculations above were performed using SPSS version 19 (SPSS Inc., Chicago, IL, USA). All figures were created using GraphPad Prism version 5 (GraphPad Software, Inc., La Jolla, CA, USA).

Results

Demographics, genetic and biochemical data of cohort A

As expected, most AD and MCI-AD patients carried one or two copies of the APOE ε4 allele, with less than 30% being APOE ε4-negative (Table 1). Non-AD groups showed opposite results. Frequencies of different genotypes were similar between AD dementia and MCI-AD patients. AD and MCI-AD groups showed the lowest mean levels of CSF Aβ42 and the highest
mean levels of CSF tau proteins (Table 1). Biomarker levels in the sMCI group were similar to those in the control group.

*CSF Aβ42 in relation to APOE genotype*

CSF levels of Aβ42 were lower in APOE ε4 carriers than in non-carriers in a gene dose-dependent manner irrespective of diagnostic group (P < 0.001 in all groups) (Figure 1A). However, the levels of Aβ42 differed significantly between subjects with AD compared with controls, as well as between MCI-AD subjects compared with sMCI cases, even when analyzing subgroups according to APOE ε4 carrier status separately (p<0.001 to p=0.006) (Figure 1A).

ROC analysis showed that Aβ42 had high diagnostic accuracy for AD versus controls in individuals with either none or one APOE ε4 allele (Figure 1B). The diagnostic accuracy of Aβ42 in individuals with two alleles was lower than in the other APOE groups, but the uncertainty was large due to the relatively small number of APOE ε4 homozygous controls. A similar pattern was seen for MCI-AD versus sMCI patients (Figure 1C). The 95% CI of the different AUCs were clearly overlapping (Figure 1B-C), indicating that there was no real difference between them.

To determine to what extent CSF Aβ42 levels and APOE genotype contributed to distinguishing between AD and controls, as well as between MCI-AD and sMCI cases, we performed multiple binary logistic regression models which revealed that CSF Aβ42
concentration and APOE genotype were independent statistical predictors of AD diagnosis. Table 2 shows logistic regression using a backward stepwise conditional method. APOE genotype, CSF Aβ42, age and gender were entered in the first step. Gender was non-significant and was removed from the model. Analysis was done using AD dementia patients versus controls and revealed that CSF Aβ42, APOE genotype and age were independent statistical predictors of AD diagnosis. Results were similar in the MCI cohort, but with a somewhat smaller contribution from APOE genotype (data not shown).

CSF tau proteins in relation to APOE genotype

CSF T-tau levels were similar in all APOE genotype subgroups across the diagnostic spectrum and did not show the same dose-dependent differences as CSF Aβ42 within the diagnostic groups (Figure 2A). Statistical differences were only observed within the sMCI and MCI-AD groups (P = 0.013 and P = 0.009, respectively), which could be attributed to differences between the APOE ε4 -/- and APOE ε4 +/- subgroups. However, as expected CSF T-tau levels differed significantly between AD and controls (P < 0.001 to P = 0.010) as well as between MCI-AD and sMCI cases (P < 0.001) irrespective of APOE genotype group (Figure 2A).

As far as the diagnostic performance is concerned, ROC analyses showed that APOE genotype did not affect the diagnostic accuracy of CSF T-tau (Figures 2B-C). As for Aβ42, the diagnostic accuracy for T-tau among homozygous APOE ε4 carriers was somewhat lower than in the other APOE genotype subgroups when comparing AD versus controls (Figure 2B). When comparing MCI-AD versus sMCI, the diagnostic performance of CSF T-tau showed
high accuracy across all APOE ε4 subgroups (Figure 2C). Relating the levels of CSF P-tau to the different APOE genotypes revealed the same pattern as for CSF T-tau (data not shown).

No effect of APOE ε4 genotype on CSF Aβ42 levels in individuals younger than 35 years

To dissect if the association of APOE genotype with CSF Aβ42 levels was due to a direct effect of apoE isoforms on CSF Aβ42 concentration, or if it was an indirect association confounded by more amyloid pathology in the brains of APOE ε4 carriers, we analyzed young individuals (<35 years of age; cohort B) who most likely would have no amyloid accumulation in the brain. This cohort consisted of patients with bipolar disorder (n=67) and healthy, age-matched controls (n=38). No differences in APOE ε4 genotype frequencies or CSF Aβ42 concentrations were seen between the two groups (data not shown). Pooled data revealed no association of APOE genotype with CSF Aβ42 levels (Figure 3). However, the low number of APOE ε4 homozygous individuals (n=3) in this group was a limitation.

No effect of APOE ε4 genotype on CSF Aβ42 levels when subjects with mild cognitive symptoms are stratified according to cortical [18F]flutemetamol uptake

Next we analyzed a cohort of 118 individuals with CSF taps and [18F]flutemetamol PET imaging (cohort C). Subjects with positive cortical [18F]flutemetamol uptake (> 1.42 SUVR) had lower levels of CSF Aβ42 (Figure 4A). When the patients with positive or negative [18F]flutemetamol PET scans were analyzed separately, there was no difference in CSF Aβ42 levels between those with no APOE ε4 alleles or 1-2 APOE ε4 alleles (Figure 4A). Moreover, when adjusting for cortical [18F]flutemetamol uptake status, there was no association between
CSF Aβ42 and APOE ε4 carrier status (P=0.72). Similar results were obtained for CSF T-tau and P-tau (data not shown). We next aimed to replicate the results in the ADNI cohort. Since [18F]flutemetamol scans were not performed, we instead examined data from scans with the similar PET tracer 11C-Pittsburgh compound B (11C-PiB). Fifty-three subjects with both CSF analysis and 11C-PiB scans were located in the ADNI database, 9 with AD, 33 with MCI and 11 healthy controls. The cut off to identify an abnormal mean 11C-PiB SUVR was established with mixture modeling (> 1.63 SUVR). The results were very similar to our study (Figure 4B), i.e. no differences were found in Aβ42 levels between no APOE ε4 alleles and 1-2 alleles, when the patients with positive or negative 11C-PiB PET scans were analyzed separately. Further, there was no association between APOE ε4 and Aβ42 (P=0.36), when adjusting for 11C-PiB amyloid status. Even when using a previously defined 11C-PiB cutoff by Jagust et al. (>1.46 SUVR) the results were similar (data not shown).

Discussion

Distribution of APOE genotypes across the diagnostic spectrum

In cohort A, we conducted a large study with data from four specialized memory clinics to assess the effect of the APOE ε2/ε3/ε4 polymorphism on the diagnostic accuracy of CSF biomarkers for AD (Aβ42, T-tau and P-tau). The memory clinics were not prospectively harmonized against each other regarding the details of the diagnostic algorithms but all used the same clinical criteria. Likewise, the laboratory procedures for the measurement of CSF biomarkers were not harmonized, which necessitated a normalization approach (described in
detail in the methods section). Finally, the median follow-up time of stable MCI patients was 3 years, which may be considered somewhat short to rule out prodromal AD in the light of recent studies. These are three major limitations of our study, all considered unlikely to influence the interpretability of the data.

As expected, the APOE ε4 allele was more prevalent in AD and prodromal AD cases than in controls and sMCI cases. However, also sMCI cases had higher APOE ε4 prevalence compared with controls, especially in cases with low CSF Aβ42 levels. One possible explanation for this somewhat skewed distribution might be that some of these individuals, in spite of being non-demented at the time of sampling, actually had prodromal AD. To fully verify that an MCI case is non-progressive, a follow-up time of 5-10 years is probably needed. The short clinical follow-up time of MCI patients and the lack of autopsy data are the major limitations of our study.

*The diagnostic accuracy of CSF biomarker levels does not depend on APOE genotype*

We could clearly verify that APOE ε4 genotype is associated with lower CSF levels of Aβ42, but not the levels of T-tau and P-tau, in a gene dose-dependent manner, which is in agreement with earlier studies. However, all three biomarkers showed significant differences between AD patients and controls as well as between MCI-AD and sMCI patients, irrespective of APOE genotype. Even the high diagnostic accuracy of CSF Aβ42 as well as that of T-tau and P-tau was shown to be independent of APOE genotype (with the exception of somewhat lower diagnostic performance in APOE ε4 homozygous subjects, which is due to the low number of
observations in this subgroup), which further underlines the biomarkers’ strength in discriminating between the diagnostic groups. Finally, multiple logistic regression analysis confirmed that both CSF Aβ42 and APOE genotype are in fact independently associated with AD diagnosis. This is in line with earlier findings, including the North American multicenter study ADNI.9,13

APOE genotype does not modulate CSF levels of Aβ42 in younger individuals

The underlying mechanism of the association between APOE and CSF Aβ42 concentration is not fully understood, but might be partly linked to the hypothesis that the ε4-encoded ApoE isoform may be less effective at clearing Aβ from the brain, thus resulting in accelerated Aβ deposition and lower Aβ42 levels in the CSF in APOE ε4 carriers.3,4 Although this is an observational study that cannot address molecular mechanisms, we decided to explore the APOE-Aβ42 association in young individuals who were likely to be amyloid-free to test the hypothesis that there might be a primary effect (not amyloid-mediated) of different apoE isoforms on CSF Aβ42 levels. In this group of individuals, the gene-dose dependent effect on CSF levels of Aβ42 was absent. Thus, in the absence of Aβ pathology, there is no association of APOE ε4 with CSF Aβ42 levels. Earlier results showing a gene-dose dependent effect on CSF levels of Aβ42 in cognitively normal elderly individuals9-13 may thus be interpreted as driven by APOE ε4-associated preclinical Aβ pathology and not a direct effect of APOE ε4 on CSF Aβ42 levels.

CSF Aβ42 in relation to amyloid PET
It has been suggested that different cut off levels should be used for CSF Aβ42 based on APOE ε4 status. Our data show a strong association between CSF Aβ42 and cortical [18F]flutemetamol uptake, but no effect of the APOE ε4 genotype on CSF Aβ42 levels when stratifying patients into those with positive or negative [18F]flutemetamol PET scans (Figure 4A). This result was also replicated in the ADNI database using the almost identical PET tracer 11C-PiB (Figure 4B). These data strongly suggest that CSF Aβ42 levels reflect cortical Aβ deposition and not the APOE ε4 genotype per se. Consequently, the cut off for CSF Aβ42 should be the same for all APOE genotypes.

**Conclusion**

Taken together, we confirm that the APOE ε4 allele is associated with lower CSF levels of Aβ42, but not T-tau or P-tau, in age groups where amyloid pathology is prevalent, also in the absence of manifest AD. We extend these data by showing that CSF Aβ42 levels are not associated with the APOE ε4 genotype when stratifying for cortical uptake of [18F]flutemetamol, suggesting that CSF Aβ42 levels reflect cortical Aβ deposition in an APOE ε4-independent manner. Consequently, the clinical cut off for CSF Aβ42 should be the same for all APOE genotypes. Finally, CSF biomarkers are strongly associated with AD diagnosis and cortical Aβ deposition independent of APOE ε4 genotype.
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Study concept and design: Lautner, Mattsson, Wallin, Blennow, Zetterberg, Hansson.

Acquisition of data: Lautner, Palmqvist, Mattsson, Wallin, Pålsson, Jakobsson, Herukka, Owenius, Olsson, Hampel, Rujescu, Ewers, Landén, Minthon, Hansson.

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References


Figure legends

Figure 1. *APOE* genotype and the diagnostic accuracy of CSF Aβ42.

Panel A: CSF Aβ42 levels show gene dose-dependent differences within the diagnostic groups, with lower levels in *APOE* ε4-positive individuals (*P* < 0.001 in all groups). CSF Aβ42 levels differ significantly between AD and controls (Mann Whitney U test; *P* < 0.001 to *P* = 0.006) as well as between MCI-AD and sMCI (Mann Whitney U test; *P* < 0.001 to *P* = 0.001) irrespective of *APOE* genotype.

Panel B: When comparing AD vs. controls, the diagnostic performance of CSF Aβ42 is high, irrespective of *APOE* genotype. Among homozygous *APOE* ε4 individuals the diagnostic accuracy is lower with a large uncertainty due to the limited number of *APOE* ε4 +/+ controls (*n* = 7).

Panel C: When comparing MCI-AD vs. sMCI, the diagnostic performance of CSF Aβ42 is similar to that of AD vs. controls, with a somewhat lower diagnostic accuracy among *APOE* ε4 +/+ individuals.

Figure 2. *APOE* genotype and the diagnostic accuracy of CSF T-tau.

Panel A: CSF T-tau levels do not show any clear gene dose-dependent differences within the diagnostic groups. Statistical significance is only reached within the sMCI and MCI-AD groups (Kruskal-Wallis-H test; *P* = 0.005 and *P* = 0.015 respectively), which is due to differences between the *APOE* ε4 -/- and ε4 +/- subgroups. However, CSF T-tau levels differ
significantly between AD and controls (Mann Whitney U test; P < 0.001 to P = 0.010) as well as between MCI-AD and sMCI (P < 0.001) irrespective of APOE genotype.

Panel B: When comparing AD vs. controls, the diagnostic performance of CSF T-tau is high irrespective of APOE genotype group. Among homozygous APOE ε4 individuals the diagnostic accuracy is lower with a large uncertainty due to the limited number of APOE ε4 +/- controls (n = 7).

Panel C: When comparing MCI-AD vs. sMCI, the diagnostic performance of CSF T-tau shows high accuracy across all APOE ε4 subgroups.

**Figure 3.** No association between CSF Aβ42 and APOE ε4 genotype in younger non-demented subjects.

In cohort B, including non-demented subjects under the age of 35, CSF Aβ42 levels do not show any APOE ε4 gene-dose dependent differences (Kruskal-Wallis-H test; P = 0.841).

**Figure 4.** No association between CSF Aβ42 and APOE ε4 genotype when adjusting for cortical Aβ deposition.

In cohort C, we found that in the subgroup with negative [18F]flutemetamol scans (<1.42 SUVR) there were no differences in the levels of CSF Aβ42 between cases with no APOE ε4 alleles (n=49) and cases with 1-2 APOE ε4 alleles (n=10) (Mann Whitney U test; P = 0.78). Similarly, in the subgroup with positive [18F]flutemetamol scans there were no differences in the levels of CSF Aβ42 between cases with no APOE ε4 alleles (n=17) and cases with 1-2
APOE ε4 alleles (n=42) (Mann Whitney U test; P = 0.23) (Panel A). This result was replicated in the ADNI cohort using \(^{11}\)C-PiB in a population of 53 subjects (9 with AD, 33 with MCI and 11 healthy controls) (Panel B). An abnormal \(^{11}\)C-PiB was defined as a mean SUVR of >1.6 based on mixture modeling analysis.
### Table 1. Demographics, genetic and biochemical data (cohort A)

<table>
<thead>
<tr>
<th>Clinical &amp; laboratory values</th>
<th>Controls (n=251)</th>
<th>sMCI (n=399)</th>
<th>Other dementias (n=99)</th>
<th>MCI-AD (n=287)</th>
<th>AD (n=309)</th>
<th>all cases (n=1345)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, mean (range), years</td>
<td>65 (23-99)</td>
<td>67 (29-89)</td>
<td>73 (54-86)</td>
<td>73 (49-87)</td>
<td>77 (56-89)</td>
<td>71 (23-99)</td>
</tr>
<tr>
<td>Gender, male/female</td>
<td>118/133</td>
<td>189/210</td>
<td>59/40</td>
<td>97/190</td>
<td>97/212</td>
<td>560/785</td>
</tr>
<tr>
<td>APOE ε4 -/-, No. (%)</td>
<td>177 (70.5)</td>
<td>235 (58.9)</td>
<td>57 (57.6)</td>
<td>76 (26.5)</td>
<td>87 (28.2)</td>
<td>632 (47.0)</td>
</tr>
<tr>
<td>APOE ε4 +/-, No. (%)</td>
<td>67 (26.7)</td>
<td>136 (34.1)</td>
<td>37 (37.4)</td>
<td>155 (54.0)</td>
<td>172 (55.7)</td>
<td>567 (42.2)</td>
</tr>
<tr>
<td>APOE ε4 +/+, No. (%)</td>
<td>7 (2.8)</td>
<td>38 (7.0)</td>
<td>5 (5.1)</td>
<td>56 (19.5)</td>
<td>50 (16.2)</td>
<td>146 (10.9)</td>
</tr>
<tr>
<td>CSF Aβ42, mean (SD), ng/L^a</td>
<td>670.5 (181.4)</td>
<td>632.7 (182.9)</td>
<td>554.4 (184.4)</td>
<td>386.2 (146.7)</td>
<td>382.8 (102.3)</td>
<td>524.1 (204.2)</td>
</tr>
<tr>
<td>CSF T-tau, mean (SD), ng/L^b</td>
<td>323.7 (166.9)</td>
<td>353.4 (184.6)</td>
<td>422.6 (350.4)</td>
<td>689.3 (348.8)</td>
<td>793.1 (481.5)</td>
<td>525.7 (377.9)</td>
</tr>
<tr>
<td>CSF P-tau, mean (SD), ng/L^c</td>
<td>61.4 (21.7)</td>
<td>64.3 (23.9)</td>
<td>61.8 (24.6)</td>
<td>98.6 (39.3)</td>
<td>105.7 (56.1)</td>
<td>79.7 (41.2)</td>
</tr>
</tbody>
</table>

^a based on 1342 cases (3 missing data); ^b based on 1338 cases (7 missing data); ^c based on 1256 cases (89 missing data)

### Table 2. AD vs. controls, logistic regression using a backward stepwise conditional method

<table>
<thead>
<tr>
<th>Variables</th>
<th>B (intercept)</th>
<th>Standard error</th>
<th>P Value</th>
<th>odds ratio (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>APOE ε4 -/-</td>
<td></td>
<td></td>
<td>.01</td>
<td>Reference category for APOE genotype</td>
</tr>
<tr>
<td>APOE ε4 +/-</td>
<td>0.786</td>
<td>0.309</td>
<td>.01</td>
<td>2.20 (1.20-4.03)</td>
</tr>
<tr>
<td>APOE ε4 +/+</td>
<td>1.224</td>
<td>0.551</td>
<td>.03</td>
<td>3.40 (1.16-10.01)</td>
</tr>
<tr>
<td>CSF Aβ42</td>
<td>-0.011</td>
<td>0.001</td>
<td>&lt;.001</td>
<td>0.99 (0.986-0.991)</td>
</tr>
<tr>
<td>Age</td>
<td>0.137</td>
<td>0.018</td>
<td>&lt;.001</td>
<td>1.15 (1.11-1.19)</td>
</tr>
</tbody>
</table>
Figure 1

**A. APOE vs Aβ42**

- **CSF Aβ42 (ng/L)**
- **controls**
- **sMCI**
- **other**
- **MCI-AD**
- **AD**

AUC (95% CI) for each APOE genotype:
- APOE ε4/-: 0.89 (0.85-0.93)
- APOE ε4+/−: 0.90 (0.85-0.95)
- APOE ε4+/+: 0.82 (0.63-1.00)
- All APOE genotypes: 0.91 (0.89-0.94)

**B. ROC curve CSF Aβ42**

AD vs control

AUC (95% CI) for each APOE genotype:
- APOE ε4/-: 0.84 (0.78-0.89)
- APOE ε4+/−: 0.84 (0.79-0.88)
- APOE ε4+/+: 0.73 (0.62-0.84)
- All APOE genotypes: 0.86 (0.83-0.89)

**C. ROC curve CSF Aβ42**

MCI-AD vs sMCI
A. APOE vs Tau

B. ROC curve CSF Tau

AD vs control

C. ROC curve CSF Tau

MCI-AD vs sMCI

APOE genotypes

AUC (95% CI)

APOE ε4/- 0.93 (0.89-0.96)
APOE ε4+/− 0.85 (0.80-0.91)
APOE ε4+/+ 0.80 (0.58-1.00)
All APOE genotypes 0.90 (0.88-0.93)

AUC (95% CI)

APOE ε4/- 0.79 (0.72-0.86)
APOE ε4+/− 0.81 (0.76-0.86)
APOE ε4+/+ 0.86 (0.77-0.95)
All APOE genotypes 0.83 (0.79-0.86)
Figure 3

APOE vs Aβ42

CSF Aβ42 (ng/L)

controls & bipolar disorder age < 35 years

APOE ε4-/-  APOE ε4+/-  APOE ε4+/+

0 200 400 600 800 1000 1200

APOE ε4-/−  APOE ε4+/−  APOE ε4+/+
Figure 4

A. The present study

Abnormal $^{18}$F-flutemetamol

B. Replication in ADNI

Abnormal $^{11}$C-PiB