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Grb10 is a dual regulator of receptor tyrosine kinase signaling

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Abstract

The adaptor protein Grb10 is a close homolog of Grb7 and Grb14. These proteins are characterized by an N-terminal proline-rich region, a Ras-GTPase binding domain, a PH domain, an SH2 domain and a BPS domain in between the PH and SH2 domain. Human Grb10 gene encodes three Grb10 splice variants. These variants show differences in functionality. Grb10 associates with multiple proteins including tyrosine kinases in a tyrosine phosphorylation dependent or independent manner. Association with multiple proteins allows Grb10 to regulate different signaling pathways resulting in different biological consequences.

Keywords: InsR, IGF-1R, FLT3, KIT, PDGFR, VEGFR, RTK, Adaptor.

1. Introduction

In all the aspects of cellular processes protein-protein interactions play a role. Interactions are mediated through many different ways. Functional domains are one of the basic components for protein-protein interactions. Among the many different types of protein-protein interactions, association of the receptor and signaling proteins is one of the important event in signal transduction from extracellular stimuli. In many cases tyrosine phosphorylation of the receptor is involved in this process that in turn recruits SH2 domain containing proteins. The human genome encodes approximately 112 SH2 domain containing proteins which are divided into 38 families [1]. The Grb7 family proteins include three members referred to as Grb7, Grb10 and Grb14 [2]. Besides the presence of SH2 domain, these proteins possess a multi-domain structure including RA, PH and BPS domains. The presence of multiple functional domains facilitates recruitment of multiple signaling proteins and thus Grb7 family proteins act as adaptors to link signaling proteins to the receptor. This review will address our current understanding of Grb10 biology and its roles in cell signaling.

2. Grb10 structure and splice variants

Human Grb10 was first cloned in 1995 after cloning of mouse Grb10 in the same year [3,4]. While mouse Grb10 encodes a 621 amino acids proteins, the first cloned human Grb10 was a 548 amino acids protein and was named as Grb-IR, as it was found to be associated with insulin receptor, InsR [3]. The human Grb10 gene is located on the chromosome 7p11.2-p12 and expresses mainly three splice variants, Grb10 α , Grb10 β and Grb10 γ (Figure 1). Grb10 α is expressed as a 548 amino acids protein which is also known as Grb-IR or Grb10 β . Human Grb10 β was also named as Grb-IR β or Grb10-IR-SV1 and it is expressed as a 536 amino acids protein. The longest human Grb10 splice variant is the Grb10 γ expressing as 594 amino acids protein, which has also been described as Grb10 ζ . Grb10 possesses four functional domains and an N-terminal proline-rich region. All three splice variants contain an intact RA domain, a BPS domain and an SH2 domain. Grb10 α lacks a part of the PH domain, while Grb10 β lacks a part of the N-terminal proline-rich region. The SH2 domain facilitates interaction with phosphotyrosine residue of receptors or signaling proteins [5,6]. The PH domain mainly recruits Grb10 to the cell membrane through interaction with phospholipids. The RA domain is known to bind with the Ras superfamily proteins and the BPS (between PH and SH2 domains) domain interacts with the insulin receptors and acts as a negative regulator of insulin signaling [7].

3. Grb10 expression

Grb10 expression was initially identified in the mouse fibroblast [4]. Although a low level Grb10 mRNA expression was detected in the mouse liver, an abundant mRNA expression was detected in the skeletal muscle, heart, liver, brain, cartilage and adipose tissue [6,8]. In human, higher Grb10 expression was detected in pancreas and skeletal muscle, intermediate expression was detected in brain and cardiac muscle, and lower expression was reported in liver, lung, kidney, placenta, spleen, ovary, prostate, colon, testis and small intestine [3,9-11]. The mRNA expression of Grb10 has been found to be up-regulated in the primary cervical squamous cell cancers and depletion of Grb10 mRNA by siRNA resulted in marked cell growth inhibition of the cervical squamous cell suggesting that Grb10 act as a survival factor in this disease [12].

4. The role of Grb10 in insulin receptors signaling

Grb10 was found to be associated with InsR in response to the insulin stimulation and this association resulted in negative regulation of insulin signaling [3,5]. Interaction was dependent on InsR-pY1322 and Grb10-SH2 domain [5]. Similar to InsR, Insulin-like growth factor 1 receptor (IGF-1R) was associated with Grb10 and the association was mediated through the carboxy-terminal of activated receptor [13,5,14]. However, unlike other InsR interacting proteins, Grb10 did not associate with the insulin receptor substrate-1 (IRS-1) [5] suggesting that Grb10 plays unique role in the insulin signaling independent of IRS-1. Although Grb10 has been shown to negatively regulate the insulin signaling, contradictory results have also been reported. Microinjection of the SH2 domain of Grb10 in fibroblasts blocked insulin and IGF-1 induced mitogenesis but had no effect on the EGF-induced mitogenesis [9] suggesting that Grb10 cooperates with insulin in downstream signaling. Furthermore, in response to PDGF, IGF-1 and insulin, Grb10 potentiates cell proliferation [15]. This biological effect is mediated through association of Grb10 with Gab1. Overexpression of the mouse Grb10 α in p6 or other mouse embryo fibroblast cell lines partially blocked insulin induced transformation but not cell proliferation [16]. Thus, Grb10 acts differentially in InsR signaling.

Activation of InsR induces tyrosine phosphorylation of Grb10 on Y67 residue. Although Grb10 directly associates with InsR, Grb10 is not a direct substrate of this receptor. Insulin induced Grb10 tyrosine phosphorylation is mediated through the Src family non-receptor tyrosine kinase [17]. Grb10 displays specificity in association to the insulin receptors, as compared to the IGF-1R [6]. Besides interaction through the SH2-domain, it appears that Grb10 BPS domain interacts with the catalytically active InsR and IGF-1R [7].

While InsR displays equal affinity to the SH2 and BPS domains, IGF-1R displays higher affinity to the BPS domain and EGFR does not bind to the BPS domain but to the SH2 domain [7]. Recombinant BPS domain inhibits IGF-1-induced substrate phosphorylation [18].

Grb10 interacts with the regulatory domain of PI3-K, p85 in response to the insulin stimulation suggesting that Grb10 can recruit PI3-K to the InsR [19]. Grb10 creates bridges between IGF-1R receptor and GYF domain containing proteins GIGYF1 and GIGYF2 [20]. These associations are mediated through the proline-rich region of Grb10 and GYF domain of GIGYF1 and GIGYF2 suggesting that Grb10 is capable of interacting multiple proteins at the same time using different functional regions. Additionally Grb10 can create dimers through its SH2 domain which further creates additional sites for complex formation [21]. Mouse Grb10 interacts constitutively with the E3 ubiquitin ligase, NEDD4 and this interaction is mediated through the BPS and/or SH2 domain of Grb10 [22]. Association with NEDD4 did not induce Grb10 ubiquitination. However, it effected the stability of the IGF-1R in response to the ligand stimulation, suggesting that Grb10 links ubiquitin ligase to the receptor and induces receptor ubiquitination, internalization and degradation [23]. Thus, by limiting stability Grb10 negatively regulates of IGF-1 signaling (Figure 2).

Grb10 negatively regulates receptor signaling not only by destabilizing the receptor but also competing with substrate proteins for association. For example, human Grb10 γ blocks insulin induced IRS1 and IRS2 phosphorylation without affecting the kinase activity of InsR [24]. Grb10 disrupts association of IRS with InsR which further blocks insulin induced Akt phosphorylation. Therefore Grb10 γ acts as a negative regulator of the insulin signaling. Expression of Grb10 in the Chinese hamster ovary cells expressing InsR reduced insulin-induced MAPK phosphorylation [25]. Grb10 reduced insulin induced MAPK activation by blocking InsR-mediated Shc phosphorylation and an intact SH2 domain of Grb10 was required for this inhibition suggesting that negative regulation occurred by direct inhibition of the receptor. Knockdown of Grb10 enhanced IGF-1-induced phosphorylation of IRS, Akt and Erk1/2 [26]. Grb10 γ underwent serine phosphorylation in response to the insulin stimulation and this phosphorylation is mediated through activation of Erk1/2 by InsR. Three serine residues 150, 418 and 476 are directly phosphorylated by Erk1/2 and Ser 150 and Ser 476 sites are required for Grb10-mediated inhibition of the insulin signaling [27]. Another mechanism is probably involved in the serine phosphorylation of Grb10. The Grb10 homolog, Grb14 has been shown to be phosphorylated by PKC ζ in response to insulin stimulation [28]. PKC ζ is a member of PKC serine/threonine family protein kinases which are involved in various cellular processes and can be regulated by receptor tyrosine kinases [29-36]. Thus it is likely that PKC family proteins might capable of Grb10 phosphorylation on serine residues. Overexpression of

Grb10 inhibited insulin induced receptor auto-phosphorylation and glucose uptake, while only BPS-SH2 domain fragment had no effect on these processes. However, both full length and BPS-SH2 fragment inhibited insulin-induced IRS phosphorylation as well as activation of downstream signaling and could efficiently associate with InsR [37].

Grb10 expression led to reduction in InsR level where depletion increased InsR level but in both cases mRNA levels of InsR remained unchanged [38]. Furthermore, the proteasome inhibitor MG132 but not lysosome inhibitor chloroquine reversed Ins-induced InsR reduction in Grb10 expressing cells and Grb10 expression increased InsR ubiquitination suggesting that Grb10 not only reduces InsR kinase activity but also reduces InsR turnover. Grb10 binds to the E3 ubiquitin ligase Nedd4 and promotes IGF-I-stimulated ubiquitination, internalization, and degradation of the IGF-IR through multi-ubiquitination, and clathrin-dependent and-independent internalization [39]. In addition, Grb10 associates with mTORC1 and mTORC1 phosphorylates and stabilizes Grb10 that led to feedback inhibition of the PI3K signaling [40,41]. Grb10 is also a negative regulator of the insulin signaling in pancreatic beta cells [42].

5. Regulation of type III receptor tyrosine kinases by Grb10

The type III receptor tyrosine kinases include PDGFR α , PDGFR β , CSF-1R, KIT and FLT3. These receptor tyrosine kinases are associated with cancer and many of family members found to be mutated in several cancers [43,44]. Downstream signaling of this family receptor tyrosine kinases have been shown to be regulated by interacting with various proteins [45-50]. The N-terminal truncated isoform Grb10 β was found to be associated with multiple proteins including PDGFR, EGFR and c-Abl [10]. Grb10 was associated with PDGFR β in response to the PDGF and this association was mediated through pY771 of PDGFR β and the SH2 domain of Grb10 [51]. Overexpression of full-length Grb10 potentiated mitogenic signaling from PDGFR as well as PDGF-induced cell proliferation but the SH2 domain alone suppressed these events suggesting that Grb10 positively regulates PDGF signaling. Grb10 was associated with KIT in response to the stem cell factor (SCF)-induction through the SH2 domain and then it interacted with AKT resulting in aberrant activation of mitogenic signaling [52]. Probably in this way KIT can bypass PI3-K activity in AKT activation. KIT mutant that lacks PI3K binding site still can activate AKT in Grb10 expressing cells suggesting that Grb10 can transduce signal from the receptor to AKT directly. Grb10 directly associates with ligand stimulated wild-type FLT3 as well as oncogenic FLT3 and recruits p85 leading to activation of AKT and STAT5 in hematopoietic cells [53]. Thus it

is suggested that although Grb10 is mainly negatively regulator of insulin signaling, it cooperates with type III receptor tyrosine kinase signaling by recruiting multiple signaling proteins (Figure 3).

6. Grb10 in VEGFR regulation

The vascular endothelial growth factor receptor (VEGFR) is involved in both vasculogenesis and angiogenesis. In response to the VEGF-stimulation Grb10 was tyrosine phosphorylated and this phosphorylation was mediated partly through the activation of Src. In endothelial (HUVEC) cells VEGF-induction increased transcriptional activation of Grb10. Elevated Grb10 expression further potentiated expression and tyrosine phosphorylation of the VEGFR3 (KDR) following accelerated mitogenic signaling [54]. This effect was mediated by direct association of Grb10 to the activated receptor through the Grb10 SH2 domain. Grb10 stabilized VEGFR2 by inhibiting NEDD4-mediated VEGFR2 degradation [55]. The inhibition was mediated through direct interaction of Grb10 with NEDD4 and then over-expression of Grb10 in VEGFR2 expressing cells accelerated VEGF signaling. Thus, like type III receptor tyrosine kinases Grb10 potentiates VEGFR signaling.

7. The role of Grb10 in EGFR signaling

The epidermal growth factor receptor (EGFR) is a family of four receptor tyrosine kinases including EGFR, ERBB2, ERBB3 and ERBB4 [56,57]. This family proteins are over-expressed in many cancers. Grb10 was first identified as an EGFR interacting protein in a screen of bacterial expression libraries in 1995 [4]. Grb10 was found to be poorly associated with EGFR and EGF induction did not induce a tyrosine phosphorylation of Grb10. However, EGF induced serine phosphorylation of Grb10 suggesting that Grb10 is not a direct substrate of EGFR and its role in EGFR signaling has to be defined.

8. Regulation of other tyrosine kinases

The receptor tyrosine kinase Ret plays a role in the development of the enteric nervous, endocrine, and renal systems. Grb10 was found to be associated with RET in a yeast two-hybrid screening of a mouse embryonic library using the cytoplasmic domain of RET [58]. Using an EGFR extracellular domain with RET cytoplasmic chimera, it has been demonstrated that Grb10 associates with RET in an activation dependent manner [58]. The activated hepatocyte growth factor receptor (MET) has also been found to be associated Grb10 [51]. However the role of Grb10 in RET and MET downstream signaling remains still unknown.

Grb10 associates with receptor tyrosine kinase ELK in vascular endothelial cells through ELK-pY929 and Grb10-SH2 domain [59]. Besides interaction with activated receptor tyrosine kinases, Grb10 interacts with a number of non-receptor tyrosine kinases. Although the interaction between receptor and Grb10 is mainly dependent on Grb10 SH2 domain, in many cases Grb10 associates with non-receptor tyrosine kinases through an SH2-domain independent manner. For example, interaction with c-Abl is mediated through SH3 domain of c-Abl [10].

AKT directly binds to the Grb10 and phosphorylates on serine 428 residue that further facilitate binding to 14-3-3 [60]. Furthermore, Grb10 creates complex with Raf1 and 13-3-3 and inhibits Bad mediated apoptosis [61]. Despite of interaction with phospho-tyrosine residue, Grb10 SH2 domain interacts with Raf1 and MEK1 in a phosphotyrosine independent manner [62]. MEK1 associates with Grb10 through Thr-386 and this association is needed for MEK1 induced survival of the HTC-IR and COS-7 cells [62]. The oncogenic fusion protein BCR-ABL also associates with Grb10 in through phosphotyrosine residues in BCR site and kinase activity of BCR-ABL is indispensable for this interaction [63]. This interaction is important for BCR-ABL induced cell transformation. Another non-receptor tyrosine kinase Tec binds with Grb10 and phosphorylates Grb10 on tyrosine residues, and this association results in a negative regulation of Tec induced c-Fos-induction in Ba/F3 cells [64]. In addition, Grb10 has been shown to be associated with mitochondria and IGF-1-induction induces translocation of Grb10 to the membrane. Mitochondrial localization probably mediated through Raf-1, as it was reported to be associated with activated Raf-1 in mitochondria following ultraviolet radiation [65].

10. Growth Hormone (GH) receptor and Grb10

Loss of Grb10 function in mice results in fetal and placental overgrowth suggesting that Grb10 play a role in negative regulation of growth factors signaling. Although Grb10 transgenic mice displayed growth retardation [66], disruption of Grb10 in mice peripheral tissues had no significant effect on fasting glucose or insulin levels but peripheral-tissue-specific knockout led to significant over growth [67]. Dual Grb10 and Grb14 knockout mice results in deregulation of whole-body glucose homeostasis [68]. Adult mice deficient in Grb10 have elevated body mass and muscle mass throughout the adulthood [69]. Thus it is likely that Grb10 maintains normal growth of mice. Grb10 associated with growth hormone receptor (GHR) in response to growth hormone (GH) in the Huh-7 hematoma cells [70]. Grb10 associated with Jak2 as well, and overexpression of Grb10 blocked GH-induced induction of serum response element (SRE) but not STAT5 tyrosine phosphorylation [70] suggesting that Grb10 negatively regulates selective downstream pathways of GHR.

11. Conclusion

Grb10 has been implicated mainly in insulin signaling. Despite of its regulation of insulin signaling Grb10 play important roles in other receptors regulation (Table 1). The presence of multiple domains, proline-rich region and ability of self-dimerization facilitate multiple protein complex formation by Grb10. In this way Grb10 can recruit multiple signaling proteins to the receptor. In case of insulin receptor regulation Grb10 associates with phosphotyrosine residue and BPS domain of receptor and then recruits either ubiquitin ligase or signaling proteins or block access of substrate proteins. In this manner Grb10 can negatively regulate insulin signaling or can transmit signal from InsR depending of interacting partners. In a similar fashion Grb10 regulates other receptors. Grb10 associates with FLT3 and KIT and recruits p85 and AKT resulting in accelerated activation of downstream signaling. In some cases Grb10 associates with ubiquitin ligase and blocks receptor ubiquitination. Since Grb10 gene is expressed as three different splice variants and the variants are structurally different, it is possible that depending on tissue specific abundance of different splice variants of Grb10 regulates receptor downstream signaling differentially.

Competing interests:

Authors declares no conflicts of interest.

Authors' contributions:

NNK and JUK selected articles and wrote the manuscript.

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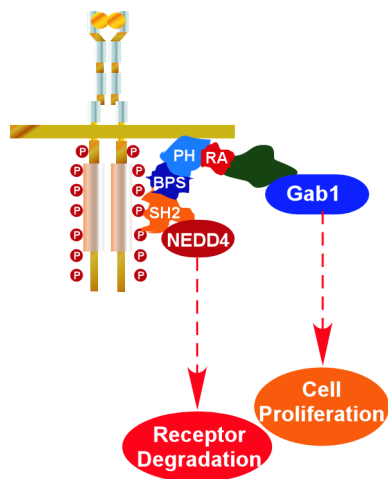
Figure legends:

Fig 1. Grb10 splice variants: Grb10 gene expresses three splice variants. The hGrb10 α expresses a 548 amino acids protein that lacks a part of PH domain. The hGrb10 β expresses a 536 amino acids proteins that lacks N-terminal proline rich regions. The hGrb10 γ expresses the longest protein with all functional domains presence in mouse Grb10.

Fig 2. Grb10 in insulin receptor signaling: Grb10 associates with activated insulin receptor through SH2 domain when it is attached to the cell membrane through PH domain. Then it recruits ubiquitin ligase NEDD4 that transfers ubiquitin moieties to the receptor leading to receptor degradation. On the other hand Grb10 can recruit signaling proteins to the receptor and then it can activate downstream signaling.

Fig 3. Grb10 in type III receptors signaling: Grb10 associates with phosphotyrosine residue of activated receptor through SH2 domain and then it recruits p85 or AKT and activates downstream signaling.





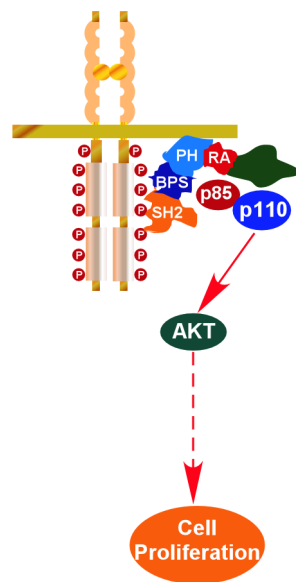


Table 1: Grb10 interacting proteins:

Regulation	Variant	Interacting proteins	Cell line	Reference
Negative	hGrb10 α	InsR	CHO	[3]
	hGrb10 β	InsR, IGF-1R, mTORC1		[18,40,41]
	hGrb10 γ	InsR, Erk1/2, DOK, Tec	CHO, Ba/F3	[24,25,38,27,71,64]
	mGrb10 α	IGF-1R, InsR, JAK2, NEDD4, GHR	3T3, P6, 3T3-L1	[16,23,26,39,37,70]
	mGrb10 ϵ	InsR, LRP6		[66,72]
Positive	hGrb10 β	FLT3, IGF-1R, InsR, KDR, NEDD4, p85	Hematopoietic, fibroblasts, HUVEC	[9,53-55]
	hGrb10 γ	MEK1, Raf1, BimL	HEK293, HTC-IR and COS-7	[62,65,61,73]
	mGrb10 α	IGF-1R, InsR, PDGFR β , p85, Gab1, c-Kit, Akt	NIH3T3, Hematopoietic	[15,19,51,52]
Unknown	hGrb10 β	c-Abl, EGFR, Grb10, PDGFR, pp135		[7,10,21]
	hGrb10 γ	FYN, Src,	CHO, HEK293	[17]
	mGrb10 α	GIGYF1, GIGYF2, IRS1, IRS2, MET, RET, ELK, EGFR, Bcr-Abl, 14-3-3	3T3-L1, L6, 293T, vascular endothelial cells	[4,19,20,58,51,59,63,60]