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CD146 expression on primary nonhematopoietic bone marrow stem cells is correlated with in situ localization

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Nonhematopoietic bone marrow mesenchymal stem cells (BM-MSCs) are of central importance for bone marrow stroma and the hematopoietic environment. However, the exact phenotype and anatomical distribution of specified MSC populations in the marrow are unknown. We characterized the phenotype of primary human BM-MSCs and found that all assayable colony-forming units-fibroblast (CFU-Fs) were highly and exclusively enriched not only in the lin−/CD271+/CD45−/CD146+ stem-cell fraction, but also in lin−/CD271+/CD45−/CD146−/low cells. Both populations, regardless of CD146 expression, shared a similar phenotype and genotype, gave rise to typical cultured stromal cells, and formed bone and hematopoietic stroma in vivo. Interestingly, CD146 was up-regulated in normoxia and down-regulated in hypoxia. This was correlated with in situ localization differences, with CD146 coexpressing reticular cells located in perivascular regions, whereas bone-lining MSCs expressed CD271 alone. In both regions, CD34+ hematopoietic stem/progenitor cells were located in close proximity to MSCs. These novel findings show that the expression of CD146 differentiates between perivascular versus endosteal localization of nonhematopoietic BM-MSC populations, which may be useful for the study of the hematopoietic environment. (Blood. 2011;117(19):5067-5077)

Introduction

Human bone marrow contains a rare population of nonhematopoietic mesenchymal stem cells (BM-MSCs), which are multipotent and can differentiate in vivo toward skeletal lineages such as osteoblasts, adipocytes, and chondrocytes, as well as toward fibroblastic stromal cells.1-3 In vitro, clonogenic cells—denoted as colony-forming units, fibroblast (CFU-Fs)—can be assayed from the bone marrow as plastic adherent cells giving rise to fibroblastic colonies. These CFU-Fs are considered to reflect primary BM-MSCs, and on further proliferation in culture, their descendants make up the well-known and extensively studied cultured stromal colonies.4

Bone marrow CFU-Fs express surface markers such as STRO-1,4 CD271 (nerve growth factor receptor [NGFR]),5,7 stage-specific embryonic antigen-4 (SSEA-4),9 GD2 (disialoganglioside 2),9 CD49a (integrin α1),10 and CD146 (melanoma cell adhesion molecule [MCAM]).3,11 To date, these different CFU-F markers have not been used in combination, and it is therefore not known whether they identify the same cells or whether different subtypes of early nonhematopoietic stem and progenitor cells coexist in the bone marrow.

Culture-expanded CD146+ cells have been demonstrated to reestablish the hematopoietic microenvironment (HME) in a xenotransplantation model, and the transplanted cells colocalized with suggested HSC niches in the bone marrow.3 Therefore, BM-MSCs are likely to be relevant for human HME and stem cell niche anatomy and function. However, a precise phenotypic definition of the human stem cell niche cellular components has thus far been elusive, in contrast to the murine system, in which different niche cell types have been recently described.12-15

We report herein that nonhematopoietic human BM-CFU-Fs are highly and exclusively enriched in lin−/CD271+/CD45−/CD146+ cells and in lin−/CD271+/CD45−/CD146−/low cells. Whereas CD271 expression identifies all assayable BM-CFU-Fs, different expression patterns of CD146 are correlated with in situ localization differences: subendothelial sinusoidal CFU-Fs display the primary CD271+/CD146+ phenotype, whereas bone-lining CD271− CFU-Fs are predominantly CD146−/low. In both locations, CD34+ hematopoietic stem/progenitor cells are located in close proximity, which might enable for the first time the prospective investigation and dissection of differently localized putative HSC niche cells in human bone marrow.

Methods

BM-MNCs

Sixty milliliters of bone marrow was aspirated from the iliac crest bone of consenting healthy donors. This procedure was approved by the University of Lund ethics committee. Bone marrow mononuclear cells (BM-MNCs)
were isolated by density gradient centrifugation using LSM 1077 Lymphocyte Separation Medium (PAA Laboratories) either with or without prior incubation with RosetteSep Human Mesenchymal Stem Cell Enrichment Cocktail (StemCell Technologies) for lineage depletion (CD3, CD14, CD19, CD38, CD66b, and glycophorin A).

**FACS**

Lineage-depleted BM-MNCs were incubated in blocking buffer (Dulbecco PBS [DPBS] without Ca²⁺, Mg²⁺, and 3.3 mg/mL of human normal immunoglobulin [Gammanorm; Octapharm] and 1% FBS [Invitrogen]) to prevent unspecific binding, followed by staining with monoclonal antibodies against CD45, CD146, and CD271 (see supplemental Methods, available on the Blood Web site; see the Supplemental Materials link at the top of the online article). Sorting gates were set according to the corresponding fluorescence-minus-one (FMO) controls. Cells were sorted on a FACSAria I or a FACSDiva flow cytometer (both BD Biosciences). Dead cells were excluded by 7-amino-actinomycin (7-AAD; Sigma) staining, and doublets were excluded by gating on forward scatter-height versus forward scatter-width and side scatter-height versus side scatter-width.

**Generation of cultured mesenchymal stromal cells**

Sorted BM-MNCs were cultured in standard MSC culture medium (NH Expansion Medium [Miltenyi Biotec] plus 1% antibiotic-antimycotic solution [Sigma-Aldrich]). Trypsinized cells were replated at 500-1000 cells/cm². Mesenchymal stromal cells for intra-bone marrow transplantation or subcutaneous transplantation with hydroxyapatite/tricalcium phosphate (HA/TCP) carriers were initiated from sorted CD271+/CD45−/CD146−/low and CD271+/CD45+/CD146+ BM-MNCs, respectively. Sorted cells were initially plated at 20-50 cells/cm², and adherent cells were culture expanded as described in the beginning of this section. Green fluorescent protein (GFP)–labeled cells for intra-bone marrow transplantation were produced by transducing 0. passage cells (before the first passing) with a lentiviral vesicular stomatitis virus glycoprotein GFP vector, followed by sorting of GFP+ cells 2 weeks after transduction and an additional 3 weeks of culture until transplantation.

**CFU-F assay**

CFU-F frequencies of BM-MNC populations were determined as described previously. Briefly, FACS–sorted cells were cultured at plating densities of 1-50 cells/cm² when assaying CD271+/CD45+/CD146−/low and CD271+/CD45−/CD146+ BM-MNCs, respectively. Sorted cells were initially plated at 20-50 cells/cm², and adherent cells were culture expanded as described in the beginning of this section. Green fluorescent protein (GFP)–labeled cells for intra-bone marrow transplantation were produced by transducing 0. passage cells (before the first passing) with a lentiviral vesicular stomatitis virus glycoprotein GFP vector, followed by sorting of GFP+ cells 2 weeks after transduction and an additional 3 weeks of culture until transplantation.

**Flow cytometry**

Cultured cells were harvested (0.05% trypsin/EDTA) and washed, and unspecific binding was blocked with human normal immunoglobulin. Cells were stained for 45 minutes at 4°C with combinations of antibodies. Samples were analyzed on a FACSCalibur flow cytometer (BD Biosciences).

**In vitro differentiation assays**

Cultured BM mesenchymal stromal cells were differentiated toward the adipogenic, osteoblastic, and chondrogenic lineage, as described previously. Briefly, cells were cultured for 14 days in AdipoDiff medium (Miltenyi Biotec), and cells were cultured with Oil Red O (Sigma-Aldrich). For osteogenic differentiation, cells were cultured in osteogenesis induction medium (standard MSC medium supplemented with 0.05mM ascorbic acid [Wako Chemicals] and 1.0μM dexamethasone and 10mM β-glycerophosphate [both from Sigma-Aldrich]) for 21 days, and calcium depositions in the cultures were detected with Alizarin Red (Sigma-Aldrich). Chondrogenic differentiation was accomplished by culturing cell pellets (2.5 × 10⁶ cells/pellet) for 28 days in chondrogenesis induction medium (DMEM high-glucose medium supplemented with 0.1μM dexamethasone, 1mM sodium pyruvate, and 0.35mM L-proline [all from Sigma-Aldrich]), 0.17mM ascorbic acid [Wako Chemicals], 1% ITS plus (insulin transferrin selenium) culture supplements [BD Biosciences], and 0.04 μg/mL of TGF-β3 [R&D Systems]). Pellets were parafomaldehyde fixed and frozen in O.C.T. Compound (Sakura). Cryosections were stained against aggrecan (see supplemental Methods) and toluidine blue and Alcian blue (both Sigma-Aldrich). For aggrecan staining, nuclei were stained with 4’,6-diamidino-2-phenylindole (Invitrogen). Sections were analyzed with a fluorescence microscope (BX51; Olympus) and a digital camera (DP70; Olympus) using DP manager software version 1.1.171 (Olympus).

**Single-cell multiplex PCR**

For single-cell PCR, CD271+/CD45−/CD146−/low and CD271+/CD45+/CD146+ BM-MNCs were sorted into 96-well plates (Thermo Scientific) containing 4μL/well of lysis buffer (2.5mM dNTP [Takara Bio] plus 10% NPN, 0.1M DTT, and 40 μM of RNase Out [all Invitrogen]). Plates were spun, frozen, and stored at –80°C until analysis. Reverse transcription first-strand synthesis was performed at 37°C for 60 minutes after adding Moloney murine leukemia virus–reverse transcriptase (200 U/μL) and 5X first-strand buffer (both Invitrogen) and outer reverse primers to each well. The first round of PCR was performed with 10 μL of reverse transcriptase reaction, outer forward primers, and Taq polymerase (Takara Bio) at 94°C for 1 minute, 60°C for 1 minute, and 72°C for 2 minutes for 34 cycles and then 72°C for 7 minutes. The second round of PCR was performed for each gene with 1 μL of first-round PCR product, the inner primer pair, and Taq polymerase at 94°C for 30 seconds, 60°C for 1 minute, and 72°C for 1 minute for 34 cycles and then 72°C for 7 minutes. PCR products were loaded on 2% agarose E-gels with E-Gel Low-Range DNA Ladder (both Invitrogen) and analyzed. Primer sequences are provided in supplemental Methods.

**In vivo transplantation**

For orthotopic transplantations, 1 × 10⁶ GFP+ mesenchymal stromal cells generated from sorted CD271+/CD45−/CD146−/low and CD271+/CD45+/CD146+ BM-MNCs were injected intrafemorally into irradiated (2 Gy) 6- to 8-week-old NOD-Cg-Prkdcscid Il2rgtm1Wjl/SzJ mice. After 8 weeks, harvested by trypsinization and assayed for hematopoietic colony formation, as described in the beginning of this section.
mice were killed by cervical dislocation and femurs were removed and fixed in parafomaldehyde. After decalcification, permeabilization, and dehydration, specimens were embedded in paraffin for analysis.

For analysis of in vivo bone and stroma formation, cultured MSCs were derived from sorted CD271+/CD45-/CD146−/low and CD271+/CD45+/CD146+ cells. Cells were loaded overnight on HA/TCP ceramic powder, and 4 x 10^5 cells were implanted subcutaneously into 8-week-old female NOD/SCID mice (4 implants per culture). Implants were removed after 8 weeks, fixed, decalcified, and paraffin embedded. Sections were stained with hematoxylin/eosin and analyzed as described previously.18 All animal procedures were approved by the local ethical committees on animal experiments.

**Immunofluorescence staining of bone sections**

Paraffin sections from human bone marrow and mouse femurs were deparaffinized and rehydrated following standard protocols. Heat-induced epitope retrieval was applied using citrate buffer, pH 6 (Target Retrieval Solution; Dako) for 30 minutes at 98°C. Sections were blocked/permeabilized with Dulbecco PBS and 0.3% Triton X-100 (Sigma-Aldrich), 10% normal goat serum, 0.1% sodium azide, and 0.1% cold fish-skin gelatin (Sigma-Aldrich), and stained for 1 hour at room temperature or overnight at 4°C with primary antibodies. After washing, secondary staining was performed for 1 hour at room temperature (for antibodies, see supplemental Methods). TO-PRO-3 (Invitrogen) was used as a nuclear stain. Photographs were taken on a confocal microscope (DMRE; Leica) equipped with green helium/neon, standard helium/neon, and argon lasers using Confocal Software v2.61 (Leica).

**Hypoxia experiments**

Stromal cultures generated from unfractionated bone marrow were cultured with the addition of 100μM deferoxamine mesylate in a standard incubator with daily medium changes. After 1 week, cells were analyzed by FACS for the expression of CD146 and other surface markers.

Cultures generated from unfractionated bone marrow were cultured in a hypoxic chamber set to 1% O2 in standard MSC medium with weekly medium changes. After 2 weeks, cells were analyzed by FACS for the expression of CD146 and other surface markers (see supplemental Methods).

**Results**

**CFU-Fs are highly and exclusively enriched in CD271+/CD45− BM-MNCs expressing either CD146−/low or CD146+**

Human BM-MNCs were stained with antibodies against CD146, CD271, and CD45 (Figure 1A-B). Without CD45 exclusion, CD271+/CD146+ populations contained large numbers of hematopoietic cells, (43%, 97%, and 88% of CD271+/CD45+, CD271+/CD45−, and CD271+/CD146− cells, respectively; n = 9). As shown in Figure 1C, we detected 0.02% ± 0.005% CD271+/CD45−/CD146+ 0.01% ± 0.003% CD271+/CD45+/CD146−, and 0.06% ± 0.02% CD271+/CD45+/CD146+ cells within viable human BM-MNCs (data are mean ± SEM; n = 9).

We next assessed the CFU-F frequency of the different CD271+/CD146− cell populations in lineage-depleted bone marrow (Figure 1D-G). CFU-Fs were highly and exclusively enriched in the CD271+/CD45−/CD146−/low and CD271+/CD45+/CD146− cells. In contrast, colony growth was not observed in either CD45− or the CD271− cell fractions regardless of CD146 expression (Figure 1F). No CFU-Fs were lost by lineage depletion (supplemental Figure 1).

Mean CFU-F frequencies were 4.2 ± 1.6 CFU-Fs per 100 plated CD271+/CD45−/CD146−/low cells and 2.0 ± 0.7 CFU-Fs per 100 CD271+/CD45+/CD146− cells; n = 10. CFU-Fs per 100 viable cells in lineage-depleted BM-MNCs (Figure 1F).
Primary CD271+/CD146−/low and CD271+/CD146+ CFU-Fs share a similar phenotype

We compared the uncultured CD271+/CD45−/CD146−/low and CD271+/CD45−/CD146+ CFU-F-containing populations based on morphology, FACS profile, and gene expression. As shown in Figure 3, freshly sorted cells before attachment were round and displayed reticular extensions (Figure 3A-B left). After attachment, cells displayed a large, flat, branched-out fibroblastic morphology (Figure 3A-B right). In addition, cytospin preparations were prepared to study nuclear morphology and cytoplasm properties of freshly isolated cells in more detail (supplemental Figure 4A-C). CD271+/CD45−/CD146−/low and CD271+/CD45−/CD146+ cells were characterized by cytoplasmic vacuoles and large, immature nuclei with an open chromatin pattern, and CD271+/CD146−/low cells were generally smaller than the double-positive cells (supplemental Figure 4A-B). In contrast, CD146 single-positive cells, which did not contain any CFU-Fs, also had immature nuclei but exhibited a more basophilic cytoplasm and perinuclear halos, which clearly distinguished them from CD271-positive CFU-Fs (supplemental Figure 4C). We also investigated sorted primary CD271+/CD45+ BM-MNCs, which displayed clear morphologic differences compared with the CFU-F-containing CD271+/CD45− cell population (supplemental Figure 2B).

Multicolor FACS analysis of lineage-depleted BM-MNCs (n = 4) was performed to assess the coexpression of signature mesenchymal and stromal cell-surface marker profiles on CD271+/CD45−/CD146−/low and CD271+/CD45−/CD146+ cells. As illustrated in Figure 3C, primary cells from both CFU-F populations showed coexpression of CD105, CD90, and STRO-1, as well as PDGFR-beta. The majority of the cells, however, failed to express SSEA-4 and CD2. However, CD271+/CD45− cells coexpressed integrin α 1 (CD49a) (supplemental Figure 4D) and nestin (supplemental Figure 4E). Endothelial markers such as...
CD146 expression (Figure 1B and 1E). CD45 expression was generally not detected.

CD31 and CD34 were only expressed on CD146 single-positive (ie, non-CFU–F–containing) cells (Figure 3C).

Multiplex single-cell PCR of sorted, uncultured CD271+/CD45−/CD146−/low cells (Figure 3D) and CD271+/CD45+/CD146+ cells (Figure 3E) were performed. Tissue-nonspecific alkaline phosphatase was expressed by virtually all CD271+/CD45−/CD146−/low and CD271+/CD45+/CD146+ cells. Genes involved in adipogenic differentiation (eg, CEBPA and LPIN1) also exhibited a similar expression in both CFU-F populations. Further, both populations expressed genes such as Nanog, Oct-4, and Sox2 (for protein expression, see supplemental Figure 4F). Interestingly, CD146 expression could also be detected in approximately 40% of the CD271+/CD45−/CD146−/low cells; however, the band intensity was weaker compared with CD271+/CD45+/CD146+ cells, thus matching the flow cytometric profile of a gradually increasing CD146 expression (Figure 1B and 1E). CD45 expression was generally not detected.

**In vivo differentiation capacity of cultured CD271+/CD45−/CD146−/low and CD271+/CD146+ cells**

CD271+/CD45−/CD146−/low and CD271+/CD45+/CD146+ CFU-Fs were culture expanded in standard MSC medium to obtain sufficient cell numbers for heterotopic (subcutaneous) and orthotopic (intrafemoral) transplantation into immunodeficient mice. Eight weeks after subcutaneous injection (cultured cells with HA/TCP carrier particles), bone, adipocytes, fibroblastic tissue, and capillaries could be detected in both transplants (Figure 4A). In addition, we detected invading hematopoietic cells in the transplants (Figure 4B).

Orthotopic intrafemoral transplantations into irradiated NOD.Cg-Prkdcscid IleIl2rgtm1Wjl/SzJ mice were performed with GFP-labeled stromal cultures generated from either CD271+/CD45−/CD146−/low or CD271+/CD45−/CD146+ cells. After 8 weeks, GFP+ cells could be detected in the perivascular regions surrounding the endothelium of vessels, as cells lining the surface of cortical and trabecular bone or surrounding adipocytes, or as reticular cells in the marrow space (Figure 4C-E). Some of the bone-lining GFP+ cells were found to express N-cadherin (Figure 4C). The majority of transplanted GFP+ cells were localized in close proximity (ie, within 20 μm, corresponding to approximately 2 cells) of the bone surface and vasculature, respectively (supplemental Table 1). No differences in the distribution of transplanted CD271+/CD45−/CD146−/low- and CD271+/CD45−/CD146+ derived cells were observed compared with CD271+/CD45−/CD146−/low-derived cultures.
The principal capacity of transplanted cells to form secondary colonies was investigated qualitatively in first experiments by harvesting bone marrow cells from 2 mice 8 weeks after intrafemoral transplantation and plating them for CFU-Fs in standard MSC culture medium. GFP \(^{-}\)/fibroblastic colonies were detected in the bone marrow of mice transplanted with CD271\(^{-}\)/CD45\(^{-}\)/CD146\(^{-}\)/CD271\(^{+}\) or CD271\(^{-}\)/CD45\(^{-}\}/CD146\(^{-}\)/CD271\(^{-}\) cells, thus demonstrating the primarily colony-forming capacity of transplanted BM-MSC–derived cells. However, these experiments were not designed to allow for a quantitative analysis.

**In situ localization of CD271\(^{+}\)/CD146\(^{-}\)/CD271\(^{-}\)/CD146\(^{-}\) cells in human bone marrow**

To investigate the in situ localization of BM-CFU-Fs, paraffin sections of normal human bone marrow were stained with antibodies against CD146 and CD271. CD146/CD271 double-positive cells were observed as perivascular cells surrounding the capillary endothelium and larger vessels (Figure 5A-B and supplemental Figure 5B-E). CD271\(^{-}\) reticular cells spanned the bone marrow with long extensions, some of which encircled adipocytes. In contrast to these CD146/CD271 double-positive cells, we found that bone-lining cells proximal to the surface of trabecular bone primarily expressed CD271 alone (Figure 5C and supplemental Figure 5A). CD146 coexpression by bone-lining cells was rarely detected. Costaining with CD45 confirmed that no perivascular, reticular, or bone-lining cells were CD45\(^{-}\) (Figure 6A). In the marrow space, CD271\(^{+}\)-coexpressing hematopoietic CD45\(^{-}\) cells were visible as smaller, round cells (Figure 6A right), and these cells could easily be distinguished from CD271\(^{+}\)/CD146\(^{-}\) reticular cells based on their morphology. CD146 was not only expressed by perivascular cells, but also by \(\alpha\)-smooth muscle actin (\(\alpha\)-SMA)–positive cells within the vascular tunica media (supplemental Figure 5C), whereas CD271\(^{+}\) expression was exclusively confined to the extraluminal cells in the vascular tunica adventitia (Figure 6B and supplemental Figure 5B). CD271\(^{+}\) and CD146\(^{-}\) cells in the tunica adventitia exhibited weak expression of \(\alpha\)-SMA (Figure 6B and supplemental Figure 5B-C).

We next investigated the colocalization of CD34\(^{+}\) hematopoietic stem/progenitor cells with CFU-Fs in bone marrow sections. As illustrated in Figure 6, human, round CD34\(^{+}\) cells could be found in proximity to perivascular cells stained with CD271 (Figure 6C). In addition, CD34\(^{+}\) cells could be detected in proximity to trabecular bone-lining CD271\(^{+}\) cells (Figure 6D), although, as expected, only at very low frequencies.
In vitro CD146 expression is up-regulated during culture and down-regulated under hypoxic conditions

Sorted CD271+/CD146−/low and CD271+/CD146+ cells were plated on chamber slides and cultured for 6 days at 21% O2. Cells that initially were CD146−/low continued to express CD146 at lower levels compared with CD146+ cells (Figure 7A-B). Conversely, CD271 expression was comparable (Figure 7A-B). The difference in CD146 expression could also be observed flow cytometrically when analyzing lower-passage cultures generated from either CD146−/low or CD146+ cells. However, the CD146 expression of CD271+/CD146−/low cells progressively increased over time and attained levels comparable to those observed in cultured CD271+/CD146+ cells or cultures generated from unfractionated bone marrow by the end of the second passage (Figure 7C).

Because the expression levels of CD146 increased in normoxic culture and was related to the in situ localization of primary CFU-Fs, we examined the influence of oxygen levels on CD146 expression. Established stromal cultures from unsorted bone marrow (third and fourth passage) were cultured with or without deferoxamine mesylate (DFO) (n = 2). FACS analysis on day 7 showed that approximately 25% of the DFO-treated cells had become negative for CD146, compared with approximately 3.5% in untreated controls (Figure 7D). No changes in CD90 and CD271 expression were observed (supplemental Figure 6A). This finding was confirmed in hypoxic chamber experiments (n = 2), which showed that CD146 expression decreased after 2 weeks of culture in 1% O2 compared with 21% O2 (Figure 7E). This hypoxia-induced decrease of CD146 expression was reversible when cells were incubated for an additional 2 weeks at 21% O2 (Figure 7E). No changes in expression levels could be detected for any of the remaining surface markers investigated (CD105, CD73, CD90, HLA class I, and CD271; supplemental Figure 6B).

Discussion

Nonhematopoietic BM-MSCs are capable of generating tissues such as bone, fat, and cartilage. Furthermore, BM-MSCs also give rise to the HME, which provides a niche for HSCs in vivo and plays a pivotal role in regulating, supporting, and maintaining hematopoesis.3,19,20

Considerable progress has been made regarding the phenotypic description of BM-MSCs, and several surface markers for an effective enrichment of CFU-Fs have been reported. Nonetheless, a comprehensive characterization of the nonhematopoietic stem cell system and its different subpopulations is just beginning to emerge. Furthermore, relatively little is known about BM-MSCs and their role in the HME.

Recently, Sacchetti et al1 identified a population of CD146+ subendothelial human bone marrow cells that contained all assayable CFU-Fs and generated bone and HME when transplanted subcutaneously into immunodeficient mice. CD271 is another broadly accepted marker for CFU-Fs,5,6 which stains a population of subendothelial and bone-lining stromal cells in human bone marrow.21 We show herein by costaining of CD146 and CD271 that initially were CD146+ stem cells, but also in the CD146−/low fraction, thus complementing the current knowledge.

Whereas Sacchetti et al used CD146+ and CD146− FACS sorting of MACS CD45-depleted BM-MNCs, we started with a
highly enriched population of lin<sup>-</sup>/CD45<sup>-</sup>/CD271<sup>+</sup> cells for sorting on CD146 expression, which is the likely reason that the very rare population of CD146<sup>-low</sup> CFU-Fs became assayable in our experiments.

In agreement with previous studies, the majority of CD271<sup>+</sup> cells coexpressed CD45<sup>+</sup>. Therefore, even though CD271—in contrast to CD146—identified all human BM-CFU-Fs, it is necessary to also exclude CD45<sup>+</sup> hematopoietic cells from the population to reach a high purity of BM-CFU-Fs.

In addition to differences in CD146 expression, both CD271<sup>+</sup> populations showed comparable BM-MSC properties (morphology, surface-marker expression, in vitro and in vivo differentiation capacities, stroma-supporting capacities, and secondary colony formation). However, differences in CD146 expression were
clearly correlated with in situ localization, and therefore enabled us to identify endosteally localized CD271+/CD146−/low cells and perivascular CD271+/CD146+ cells. These observations agree with the findings by Sacchetti et al. on the localization of bone marrow CD146+ adventitial reticular cells and with data by Cattoretti et al. reporting that CD271+ stromal cells can be found as either perivascular or bone-lining cells.

As a note of caution, it has not been ultimately proven at present whether all of the CD271+/CD45−/CD146−/low and CD271+/CD45+/CD146+ cells that we identified in situ represent CFU-Fs, because some of the cells could belong to the fraction of non-colony-forming cells that might still be present even in highly purified cell populations (here, an even more precise BM-MSCs phenotype definition would be necessary). From our coexpression analysis, CD105, CD90, CD49a, PDGFR-β, and Stro-1 appear to be potentially useful as additional CFU-F marker candidates. However, in contrast to what has been reported previously, we did not observe expression of SSEA-4 and GD2 on primary BM-MSCs, which might have been due to differences in staining and analysis protocols.

In bone marrow sections, both CD271+/CD146−/low and CD271+/CD146+ cells were associated with CD34+ hematopoietic cells, and to our knowledge this is the first report demonstrating the association of distinct BM-MSC subsets with different potential HSC niche cell types in the human system. These observations agree with recently published landmark findings showing that nestin-positive mesenchymal stem cells form a niche for HSCs in murine bone marrow. In that study, murine nestin+/CD45− cells were identified as a small population of quiescent, perivascular BM-MNCs, which could differentiate into multiple skeletal lineages, possessed self-renewal capacity, and were required for HSC homing and maintenance of HSC. Interestingly, the genome-wide expression profile of these cells was closest to that of human CD146+ BM-MSCs. In comparison, the human multipotent CD271+/CD45− BM-MSCs described herein also showed nestin expression and exhibited hematopoietic stroma characteristics in
clearly demonstrated that an oxygen gradient exists in the bone near the surface of trabecular bone. Nevertheless, it has been human bone marrow biopsies studied, with vessels also located will therefore be addressed in future experiments.

The concept that separate endosteal and perivascular HSC niche cell types exist, and that different hematopoietic subsets reside in distinct localizations of the murine bone marrow12-15,24-27, for example, murine long-term HSCs reside in close proximity to the endosteal surface of trabecular bone.26 Whether distinct human CD34+ hematopoietic stem- and progenitor-cell populations colocalize with different localizations and the different BM-MSC populations as identified herein is certainly of great interest and will therefore be addressed in future experiments.

In summary, we have defined the phenotype of human primary nonhematopoietic BM-MSCs based on the expression of CD271, CD45, and CD146. We demonstrate that CD146 expression in lineage-negative, NGFR-positive, common leukocyte antigen-negative stroma stem cells is correlated with in situ localization, and that different BM-MSC subpopulations colocalize with different putative HSC niche cell types. This is an important finding that is likely to be the first step toward a better characterization of the human HME, hopefully leading to a better understanding of niche anatomy and function in normal and diseased marrow.

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Authorship

Contribution: A.T. performed and designed research, analyzed and interpreted data, and wrote the manuscript; O.L., J.C.B., S.W., B.S., and N.D. performed research and analyzed and interpreted data; M.E. contributed vital samples and analyzed and interpreted data; M.K. analyzed and interpreted data; and S.S. designed research, analyzed and interpreted data, and wrote the manuscript.

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