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## Peptidyl arginine deiminase from *Porphyromonas gingivalis* abolishes C5a activity.

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\* Running title: *P. gingivalis* PPAD inactivates C5a

**Keywords:** C5a, *Porphyromonas gingivalis*, peptidylarginine deiminase

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**Background:** Pathogenic bacteria avoid killing by phagocytes through inhibition of C5a chemotactic activity.

**Results:** Periopathogen *Porphyromonas gingivalis* expresses unique peptidylarginine deiminase, which inactivates C5a by converting C-terminal arginine to citrulline.

**Conclusion:** Citrullination of C5a constitutes a novel virulence strategy that may contribute to immune evasion by *P. gingivalis*.

**Significance:** *P. gingivalis* peptidylarginine deiminase is a potential target for drug development.

## ABSTRACT

Evasion of killing by the complement system, a crucial part of innate immunity, is a key evolutionary strategy of many human pathogens. A major etiological agent of chronic periodontitis, the Gram-negative bacterium *Porphyromonas gingivalis* produces a vast arsenal of virulence factors that compromise human defense mechanisms. One of these is peptidylarginine deiminase (PPAD), an enzyme unique to *P. gingivalis* among bacteria, which converts Arg residues in polypeptide chains into citrulline. Here, we report that PPAD citrullination of a critical C-terminal arginine of the anaphylatoxin C5a disabled the protein function. Treatment of C5a with PPAD *in vitro* resulted in decreased chemotaxis of human neutrophils and diminished calcium signaling in monocytic cell line U937 transfected with the

C5a receptor (C5aR) and loaded with a fluorescent intracellular calcium probe: Fura 2-AM. Moreover, a low degree of citrullination of internal arginine residues by PPAD was also detected using mass spectrometry. Further, after treatment of C5 with outer membrane vesicles (OMVs) naturally shed by *P. gingivalis* we observed generation of C5a totally citrullinated at the C-terminal Arg74 residue (Arg74Cit). In stark contrast only native C5a was detected after treatment with PPAD-null OMVs. Our study suggests reduced antibacterial and proinflammatory capacity of citrullinated C5a, achieved via lower level of chemotactic potential of the modified molecule, and weaker cell activation. In the context of previous studies, which showed crosstalk between C5aR and toll-like receptors, as well as enhanced arthritis development in mice infected with PPAD expressing *P. gingivalis*, our findings support a crucial role of PPAD in the virulence of *P. gingivalis*.

*Porphyromonas gingivalis* is a major causative agent of periodontitis, a chronic inflammatory disease of tooth-supporting structures which affects up to 30% of the world's population (1). This Gram-negative, anaerobic bacterium uses a large arsenal of virulence factors such as hemagglutinins/adhesins, fimbriae, and proteolytic enzymes to facilitate colonization of the gingival sulcus, to generate nutrients and growth factors and to provide protection from the host immune system

(2). The latter is achieved by sophisticated manipulation of the host inflammatory response through activation of coagulation factors and contact activation system, corrupting complement functions, shedding receptors, modifying cytokines and intracellular signaling (3). In this way, *P. gingivalis* maintains the local chronic inflammatory reaction and thrives in this environment to access host components essential for bacterial growth.

Recently, *P. gingivalis* peptidylarginine deiminase (PPAD<sup>2</sup>), an enzyme absent in other prokaryotes, which converts Arg residues in polypeptide chain into citrulline (Cit) has been hypothesized to be a potential virulence factor (4). Posttranslational modification mediated by PPAD's deiminase activity may change protein function in a similar manner to that described for citrullination of chemokines (5) and antibacterial peptide LL-37 (6) by endogenous PADs. However, in stark contrast to mammalian enzymes, PPAD has a strong preference for C-terminal Arg residues, probably, to neutralize the positive charge at the C-terminus of (poly)peptide fragments generated by degradation of proteins by *P. gingivalis* Arg-specific gingipains (7). Furthermore, PPAD can also abrogate essential biological activities of host proteins and peptides, which are dependent on the C-terminal Arg residues. This hypothesis is supported by finding that C-terminal citrullination of epidermal growth factor (EGF) by PPAD impaired biological activity of EGF (8).

Anaphylatoxin C5a is a polypeptide of 74 residues released from C5 by C5 convertase during complement activation. Widespread expression of two C5a receptors (C5aR and C5L2) throughout the body assures a variety of biological responses, including chemotaxis of inflammatory cells, phagocytosis, respiratory burst, vascular permeability, and releases of pro-inflammatory cytokines and chemokines. The C-terminal Arg residue is crucial for C5a function and *in vivo*, the molecule is rapidly converted by carboxypeptidases to the far less potent C5a-desArg of significantly lower affinity for C5aR (9). Since C5a is an essential component of the inflammatory response to bacterial infection, it was of interest to determine if PPAD can abolish its biological activity. Here, we showed that PPAD efficiently deiminated C-terminal Arg in C5a *in vitro* and that citrullinated Arg74 of C5a can be

found in C5 samples treated with outer membrane vesicles isolated from *P. gingivalis* culture. Moreover, this modification decreased the ability of C5a to induce calcium influx in monocytic cell line expressing C5aR and strongly reduced its chemotactic potential for neutrophils.

## EXPERIMENTAL PROCEDURES

**Ethics statement** - The regional ethical board in Lund has approved collection of blood from healthy volunteers after informed consent.

**Proteins** - As a member of the C-terminal domain (CTD)-protein family of *P. gingivalis*, native PPAD was engineered to be secreted from a mutant of *P. gingivalis* W83 as a soluble form with a hexahistidine affinity tag using the same molecular strategy as was reported previously for the RgpB protease in the mutant 662i6H (10). Subsequently, PPAD was purified from the culture medium via ion-exchange and gel filtration chromatography. Briefly, the bacteria were cultured in enriched tryptic soy broth medium for 72 h, cells were removed by centrifugation and proteins in cell-free culture medium were precipitated with acetone, resuspended in phosphate buffer (pH 6.5), dialyzed and passed through a DE-52 column (Whatman) to remove excess hemin. The flow-through was dialyzed against 50 mM Tris-HCl, 0.02% NaN<sub>3</sub>, pH 8.0 and loaded on a Mono Q column (GE Healthcare). Adsorbed proteins were eluted with NaCl gradient and fractions containing PPAD activity were pooled. The final PPAD purification was achieved by gel filtration chromatography using Superdex 75 column (GE Healthcare). The purity of PPAD was evaluated by SDS-PAGE followed by silver staining. Activity of PPAD was tested using a colorimetric assay as previously reported (11). Before any assay, PPAD was pre-activated by incubation for 20 min 37°C in 10 mM Tris pH 8.0 buffer in presence of 1 mM L-cysteine. Since a very minor contamination (less than 0.5%) with arginine-specific gingipain (Rgp) was observed in some batches of purified PPAD, assay buffers were supplemented with 1 µM KYT-1, a specific Rgp inhibitor (Peptide International). At concentrations used, KYT-1 and L-cysteine did not influence the assays (data not shown). C5a and C5a-desArg were purchased from Complement Technology.

**Outer membrane vesicles (OMV) isolation from *P. gingivalis* strains.** *P. gingivalis* strains W83,

and mutant *Δppad* (in W83 background) (12) were cultivated in enriched Schaedler broth (supplemented with 5 mg/ml hemin, 0.5 mg/ml menadione, 5 mM L-cystein and for mutant strain with 5 µg/ml erythromycin) overnight in anaerobic conditions to reach OD=1. An aliquot (100 ml) of each culture was gently sonicated in sonicator bath to release OMV to cell supernatant. Bacteria were removed by centrifugation (20 min, 10,000g) and the remaining supernatants from each strain were subjected to ultracentrifugation (1h, 150,000g, 4°C). Pellets containing OMV fraction were resuspended in 20 mM Bis-Tris pH 6.8, 150 mM NaCl, 5 mM CaCl<sub>2</sub>) and analyzed for PPAD and Arg-specific gingipain (Rgp) activity. Rgp activity was tested using spectrophotometric assay with BAPNA as a substrate (13).and Rgp concentration was calculated based on the activity of purified enzyme.

*HPLC analysis of full-length form of C5a* - Samples (100 µl) containing 10 µg of C5a or C5a were treated with PPAD (in buffer containing 100 mM Tris, 5 mM L-cysteine, 5 µM KYT-1, pH 7.6) for 3 h at 37°C. Samples were subsequently acidified with trifluoroacetic acid (TFA; Sigma) and separated through a Phenomenex Aeris C18 Widespore 4.6x150 mm (Phenomenex) on AKTA micro (GE Healthcare). Peptides were then eluted in H<sub>2</sub>O/0.1% TFA (A) and 80% acetonitrile / 0.08% TFA (B) gradient and monitored at 215 nm.

*Deglycosylation and proteolytic treatment* - Samples of C5a and C5a-Cit prepared in the same way as for HPLC analysis where lyophilized and dissolved in 20 mM Tris, 100 mM NaCl, pH 8.0 containing 5 mM DTT and denatured at 95 °C for 1 h. To remove N-linked glycosylation, samples were treated with 0.25 Units PNGase F (Sigma) per µg of C5a and incubated at 37°C for 3 h. To generate two different variants of the C-terminal peptide, deglycosylated sample was treated with either trypsin (1:25 ratio) or clostripain (4 units per µg C5a) (both from Sigma) in the presence of 1 mM calcium acetate and incubated at 37°C for 16 h. To uniformly modify all cysteine residues, samples were reduced by 5 mM DTT for 30 min at 25°C, followed by 30 min with 15 mM iodoacetamide. The samples were micropurified on StageTips (Thermo Scientific) according to the manufacturer's instructions.

*Modification of C5 by enzymes present in OMVs from different strains of P. gingivalis* - Purified C5 (CompTech) was mixed at 25:1 molar ratio to Rgp with OMVs isolated from *P. gingivalis* strains W83 and *Δppad* (in PBS buffer pH 7.4 supplemented with 5 mM L-cystein). Samples were incubated for 30 min 37°C and the reaction was stopped by addition of TFA. Citrullination of the C-terminal Arg residue in C5a was assessed by mass spectrometry.

*Mass spectrometry* - nLC-MS/MS analyses were performed on an EASY-nLC II system (ThermoScientific) connected to a TripleTOF 5600+ mass spectrometer (AB Sciex) equipped with a NanoSpray III source (AB Sciex) operated under Analyst TF 1.5.1 control. The lyophilized samples were suspended in 0.1% formic acid, injected, trapped and desalted isocratically on a Biosphere C18 column (5 µm, 2 cm x 100 µm I.D; Nano Separations). Peptides were eluted using 250 nl/min and a 20 min (for C5 samples treated with OMVs) or 50 min (for C5a samples treated with purified PPAD) gradient from 5% to 35% phase B (0.1% formic acid and 90% acetonitrile). Eluted peptide samples were separated on a 15-cm analytical column (75 µm i.d.) with RP ReproSil-Pur C18-AQ 3 µm resin (Dr. Marisch GmbH). The collected MS files were converted to Mascot generic format (MGF) using the AB SCIEX MS Data Converter beta 1.1 (AB SCIEX) and the "protein pilot MGF" parameters. The generated peak lists were searched against an in-house database containing the C5a sequence using the Mascot search engine (Matrix Science). Search parameters were either trypsin or Arg-C as protease allowing 3 missed cleavage sites. Carbamidomethyl was set as fixed modification and citrullination (R), deamidation (NE) and oxidation (M) as variable modification. Peptide tolerance and MS/MS tolerance were set to 10 ppm and 0.2 Da, respectively.

*Cell culture conditions* - U937 cells expressing C5a receptor (U937-C5aR) were maintained in RPMI medium containing 10% FCS and G418 (400 µg/ml) at 37°C in a humidified 5% CO<sub>2</sub> atmosphere (14).

*Calcium mobilization assay* - U937-C5aR cells were harvested by centrifugation, washed twice with PBS, resuspended in HBSS (0.137 M NaCl, 5.4 mM KCl, 0.25 mM Na<sub>2</sub>HPO<sub>4</sub>, 0.44 mM KH<sub>2</sub>PO<sub>4</sub>, 1.0 mM MgSO<sub>4</sub>, 4.2 mM NaHCO<sub>3</sub>, 1g/L

glucose) with 1.3 mM  $\text{Ca}^{2+}$  at  $2 \times 10^6$  cells/ml and incubated in the presence of a cell permeable fluorescent intracellular calcium probe (5  $\mu\text{M}$  Fura 2-AM, Invitrogen), for 30 min  $37^\circ\text{C}$  with gentle agitation. After double washing with HBSS+/ $\text{Ca}^{2+}$ , cells were resuspended in the same buffer at  $1.25 \times 10^6$  cells/ml and kept on ice until use. Cell response to a test ligand was monitored with continuous fluorescence measurements at  $\lambda_{\text{excitation}} = 340$  nm and 380 nm, and  $\lambda_{\text{emission}} = 510$  nm, using the Infinite 200 PRO microplate reader (Tecan) and i-Control software. After few seconds of the basal signal measurement, 50  $\mu\text{l}$  of C5a (final conc. 10 nM) or C5a pre-treated for 1 h at  $37^\circ\text{C}$  with PPAD were mixed with 200  $\mu\text{l}$  of cell suspension and measurement continued for 5 min. Values of fluorescence intensity (FI) for each time point were recalculated as ratio  $\text{FI } \lambda_{\text{emission}} = 510 \text{ nm at } \lambda_{\text{excitation}} = 340 \text{ nm} / \lambda_{\text{emission}} = 510 \text{ nm at } \lambda_{\text{excitation}} = 380 \text{ nm}$ . As controls, C5a-desArg and C5a-desArg pre-incubated with PPAD were used.

**Neutrophil chemotaxis** - Human neutrophils were isolated from healthy donors with Histopaque-1119/Percoll (15), washed twice, resuspended in PBS supplemented with 0.5% human albumin (HSA, Sigma-Aldrich) at  $10^6$  cells/ml and labeled with 1  $\mu\text{M}$  carboxyfluorescein succinimidyl ester (CFSE, Fluka) for 15 min at room temperature with gentle agitation. After staining, cells were washed twice and resuspended in PBS with 0.5% HSA and 4% heat-inactivated (30 min,  $56^\circ\text{C}$ ) hirudin-treated human plasma at concentration  $5 \times 10^6$  cells/ml prior to use. Purity of neutrophils population (defined as  $\text{CD14}^{\text{low}}/\text{CD15}^+/\text{CD16}^+$ ) was determined by flow cytometry CyFlow Space (Partec) using  $\alpha\text{-CD14-PE}$ , IgG2a (BD);  $\alpha\text{-CD15-FITC}$ , IgM and  $\alpha\text{-CD16-APC}$ , IgG1 (Immunotools) antibodies and confirmed as 90-95%.

Migration of neutrophils was assessed using a disposable 96-well cell migration system with 3  $\mu\text{m}$  polycarbonate membranes (ChemoTx; Neuro-Probe). C5a was preincubated with serial dilutions of PPAD for 1 h at  $37^\circ\text{C}$ . The samples were then supplemented with heat-inactivated human plasma at the same concentration as for neutrophils medium and applied to the wells of ChemoTx plate. C5a at final concentration of 12.5 nM was used as positive control, while 125 nM C5a-desArg, treated and untreated with PPAD, served as a negative control. Neutrophil suspension (50

$\mu\text{l}$ ) was then applied to each well of the upper filter and the plate was incubated for 60 min  $37^\circ\text{C}$  in humidified air with 5%  $\text{CO}_2$ . The membrane filter was then removed and the cell suspensions from the bottom wells were transferred to flat-bottom, 96-well black plate (Nunc). The bottom wells of ChemoTx plate were washed twice with 30  $\mu\text{l}$  of PBS and the washes pooled with the corresponding cell suspensions from the bottom wells. Fluorescence intensity signal from cells were measured for 0.1 s using WallacVictor 2 1420 Multilabel Counter (Perkin-Elmer) using  $\lambda_{\text{excitation}} = 485\text{nm}$  and  $\lambda_{\text{emission}} = 535\text{nm}$ .

**Statistical analysis** - One-way ANOVA with Dunnett's post-test was used to estimate whether the observed differences between groups were statistically significant. Data were analyzed using GraphPadPrism 5.0.

## RESULTS

**PPAD citrullinates C5a.** Purity of PPAD was determined with SDS-PAGE followed by silver staining (Fig. 1A). To confirm citrullination of C5a by PPAD, native and PPAD treated C5a were analyzed using HPLC. C5a in the native form eluted in two peaks from the C-18 column while PPAD-treated C5a was eluted in four peaks, at retention times different than those of the native peptide. This indicated citrullination of arginine residue(s) in C5a by PPAD (Fig. 1B). To determine which Arg residue(s) were deiminated, the native and PPAD-treated C5a were digested with trypsin or clostripain, and subsequently analyzed by mass spectrometry on-line with RP-HPLC. Among tryptic peptides derived from PPAD-incubated C5a, one abundant peptide showed a clear shift in retention time compared to the control C5a-derived peptides. Similarly, single peptide with shifted retention time was also found in clostripain-digested, PPAD-treated C5a versus native C5a (data not shown). Subsequently, the peptides were identified by MS/MS as derived from the C-terminus of C5a (DMQLGR and ANISHKDMQLGR). The citrullination of terminal Arg residue in these peptides was indicated by a 1 Da mass shift, peptide score (sequence information from MS), as well as the retention time shifts (Fig. 1C). In PPAD-treated C5a, only the citrullinated version of the C-terminus was detected, indicating complete modification at the C-terminus. Native C5a did not exhibit significant amounts of this

modification. Interestingly, we confirmed endoarginine deiminase activity of PPAD by detecting modifications of position 40 (AAR\*ISLGPR) and 46 (ISLGPR\*CIK) in the tryptic digest of PPAD treated C5a. These, however, were much less abundant than the C-terminal modification.

**OMVs released from *P. gingivalis* generate citrullinated form of C5a from C5.** Based on previous reports showing release of active C5a from C5 by Arg-specific gingipains (16) and presence of both Rgp and PPAD in OMV from *P. gingivalis* (17,18) we incubated intact C5 with OMVs from the wild-type *P. gingivalis* and the isogenic mutant strain lacking PPAD. Mass spectrometry analysis revealed full citrullination of the residue Arg74 of C5a in C5 samples treated with wild type strain OMVs. No such modification was present in the C5 sample treated with OMVs derived from  $\Delta ppad$ . In this sample only the native C5a C-terminus was detected (Fig. 1D). Significantly, the gingipain activity in wild-type and PPAD-null vesicles was identical and both degraded C5 to the same extent releasing C5a. Nevertheless citrullination of generated C5a was observed only after treatment with OMVs from wild-type *P. gingivalis*. This clearly indicates that PPAD very efficiently modifies C5a released by gingipains.

**Citrullinated C5a has decreased chemotactic activity.** C-terminal arginine of C5a anaphylatoxin is crucial for biological activity of this peptide. We hypothesised that deimination of this residue by PPAD, resulting in generation of neutral citrulline, should suppress proinflammatory activity of C5a, in a manner similar to physiological removal of C-terminal arginine by carboxypeptidases. Indeed, preincubation of C5a with PPAD strongly reduced its chemotactic activity for neutrophils, in a concentration dependent manner, while it had no effect on the activity of C5a-desArg (Fig. 2A). This result suggests that C5a-Cit, similar to C5a-desArg, has significantly lower affinity for C5aR on neutrophils. In keeping with this finding, treatment of C5a with PPAD also impaired its ability to induce calcium influx in myeloid-derived cell line transfected with C5aR (Fig. 2B and C). Interestingly, at higher concentrations and/or prolonged incubation with PPAD, the enzyme totally abrogated the capacity of C5a to activate neutrophils and U937 C5aR cells. In contrast, C5a-

desArg treated with PPAD, did not show altered potential to stimulate neutrophil chemotaxis and calcium release in U937-C5aR cells. As expected, native C5a-desArg had much lower activity than C5a in these assays.

## DISCUSSION

The complement system constitutes an essential part of innate immunity. Its activation, in a cascade-like manner, unleashes a spectrum of molecules aimed to destroy invading microbes. The membrane attack complex (MAC) perforates cell membranes, anaphylatoxins C3a and C5a attract and activate neutrophils, opsonization with fragments of C3b facilitates phagocytosis and intracellular killing and, finally, activated components of complement enhance adaptive immune response. Therefore, it is not surprising that successful human pathogens developed a variety of mechanisms to inhibit complement activation and/or to neutralize activated components. Several of the latter strategies target C5a or C5aR. Proteolytic inactivation of C5a by *Serratia marcescens* 56K protease, protease ScpA of *Streptococcus pyogenes* and ScpA-like enzymes of group B streptococci results in inhibition of C5a-mediated pro-inflammatory and chemotactic signalling that slows the influx of inflammatory cells and hinders removal of bacteria from the initial site of invasion (19). The importance of ScpA as a specialized virulence factor is underscored by the narrow specificity of this protease. The enzyme has no activity against intact C5 or other proteins but cleaves exclusively C5a at the C-terminus, removing part of the region that interacts with C5aR on neutrophils (20).

In contrast to other bacterial pathogens for which suppression of the inflammatory response is the immune evasion strategy, the key periodontal pathogen *P. gingivalis* seems to thrive in the inflammatory milieu. Through proteolytic degradation of C3 and C5 (21) and hijacking complement regulator C4BP (22), the bacterium efficiently blocks assembly of MAC and opsonization. However, the cleavage of C5 by gingipains is associated with the release of fully functional C5a (16). The release of this potent chemoattractant and activator of neutrophils has limited effects on *P. gingivalis*, which is fairly resistant to killing by granulocytes and macrophages. It has been reported that in murine

macrophages, C5a induces a cross talk between C5aR-dependent signaling and activated Toll like receptor-2 (TLR-2) resulting in *P. gingivalis* survival *in vivo* (23). Importance of C5a signaling for *P. gingivalis* survival was verified by the finding that specific blockade of C5aR enabled eradication of infection (24). This mechanism described for murine macrophages and murine model of periodontitis is at odds with two countermeasures *P. gingivalis* takes to temper C5a biological activity. First, the bacterium proteolytically inactivates C5aR on human phagocytes (25). Second, as we report here, *P. gingivalis* can use PPAD to attenuate proinflammatory functions of C5a by deimination of the C-terminal Arg residue. This reaction is likely facilitated by close proximity of Rgps and PPAD on both the *P. gingivalis* cell surface as well as in OMVs released by this bacterium, in the same manner as citrullination of C-terminal Arg residues of fibrinogen-derived peptides generated during incubation of *P. gingivalis* with fibrinogen (7).

In comparison to native C5a, the citrullinated anaphylatoxin shows significantly reduced chemotactic activity for neutrophils and ability to activate  $\text{Ca}^{2+}$ -influx in U937 cells expressing C5aR. This appears to be mediated mostly by complete C-terminal citrullination and not by modifications of intrapeptide arginines as C5a-desArg showed no further attenuation by PPAD treatment. It is likely that C5a-Cit and C5a-desArg, the physiological attenuated form of C5a generated by endogenous carboxypeptidase(s) (26), elicit similar biological effects by signaling through C5aR on immune cells and activating receptor-associated G-proteins. Taking into account the abundance of carboxypeptidases on immune cell surface (27) and the expression of PPAD by *P. gingivalis* (4), it is tempting to speculate that paralysis of bactericidal activity of murine macrophages is induced not by the native

anaphylatoxin crosstalk but by generated *in situ* C-terminally modified C5a. Such hypothetical scenario, which needs to be experimentally verified, reconciles PPAD as an important virulence factor contributing to *P. gingivalis* survival within the inflammatory site *in vivo*.

Modification of C5a by PPAD may generate a potential antigenic epitope due to the presence of C-terminal citrulline. Together with other C-terminally citrullinated peptides derived from bacterial and host proteins, including those derived from Rgps-degraded fibrinogen and enolase, C5a-Cit adds to the burden of post-translationally modified antigens. Occurring within a chronically inflamed periodontal tissue, these citrullinated epitopes may initiate breakdown of immune tolerance against host citrullinated proteins and the generation of autoantibodies in susceptible individuals, which eventually leads to symptoms associated with rheumatoid arthritis (28). This contention is supported by finding that collagen-induced arthritis was exacerbated in mice infected with wild-type *P. gingivalis* as manifested by earlier onset, accelerated progression and enhanced severity of the disease, including significantly increased bone and cartilage destruction (12). The ability of *P. gingivalis* to augment arthritis was dependent on the expression of PPAD and associated with increased levels of citrullinated epitopes.

Taken together, in this report we described a novel pathogenic strategy of bacterial pathogen to inactivate the antibacterial, proinflammatory activity of C5a by deimination of its C-terminal arginine. Such approach is, thus far, unique for *P. gingivalis*, being the sole bacterium expressing peptidyl arginine deiminase, with strong preference for C-terminal arginine. In this context, PPAD emerges as an important virulence factor, which warrants further investigation.

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## FOOTNOTES

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<sup>2</sup>The abbreviations used are: C5aR, C5a receptor; FI, fluorescence intensity; HSA, human serum albumin; MS, mass spectrometry; PPAD, peptidylarginine deiminase EGF, epidermal growth factor;

## FIGURE LEGENDS

**FIGURE 1 Citrullination of C5a by *P. gingivalis* PPAD.** (A) Laemmli SDS-PAGE followed by silver staining of PPAD purified from genetically modified *P. gingivalis* strain W83 culture medium. (B) C5a (10 µg) incubated in the reaction buffer alone or with 3.22 µM PPAD for 3 h at 37°C, were analyzed with HPLC on C18 column. (C) Samples same as for B, after trypsin/clostripain digestion were analysed with nLC-MS/MS. Results indicated significant citrullination of the C-terminal Arg in C5a treated with PPAD. (D) C5 (10 µg) incubated with OMVs from *P. gingivalis* W83 wild type strain and  $\Delta ppad$  mutant strain at 25:1 molar ratio to Rgp for 30 min at 37°C, were analysed with nLC-MS/MS after trypsin digestion. Results demonstrate significant citrullination of the Arg74 residue in C5a formed from C5 by Rgp and modified by PPAD present in OMVs from W83 strain. C5a was not citrullinated in the sample treated with OMVs from the  $\Delta ppad$  mutant strain.

**FIGURE 2 PPAD citrullinates C5a, decreasing its chemotactic ability against neutrophils and calcium influx into U937 C5aR cells.** (A) C5a (12.5 nM) was incubated with serial dilutions of PPAD for 1 h at 37°C. Neutrophil migration was measured after 1 h incubation as fluorescence intensity of CFSE-labelled cells in the lower chamber of the transmigration assay plate. Buffer with KYT-1 and L-cysteine, PPAD alone and C5a-desArg (125 nM) incubated with 0.1 µM PPAD or without were used as negative controls, human C5a (12.5 nM) was the positive control. Results are average of three independent experiments with SD. Statistical significance was calculated using a one-way ANOVA with Dunnett's post-test. \*\*,  $p < 0.01$ ; \*\*\*,  $p < 0.001$ . (B) C5a (10 nM) was incubated with increasing PPAD concentrations for 1 h at 37°C. Changes in  $[Ca^{2+}]$  were monitored in U937 C5aR cells loaded with Fura-2 AM in the presence of 1.3 mM calcium. (C) C5a-desArg (100 nM) was incubated with PPAD or alone in buffer supplemented with KYT-1 was used as negative controls. Depicted are the results of one representative experiment.

Fig. 1

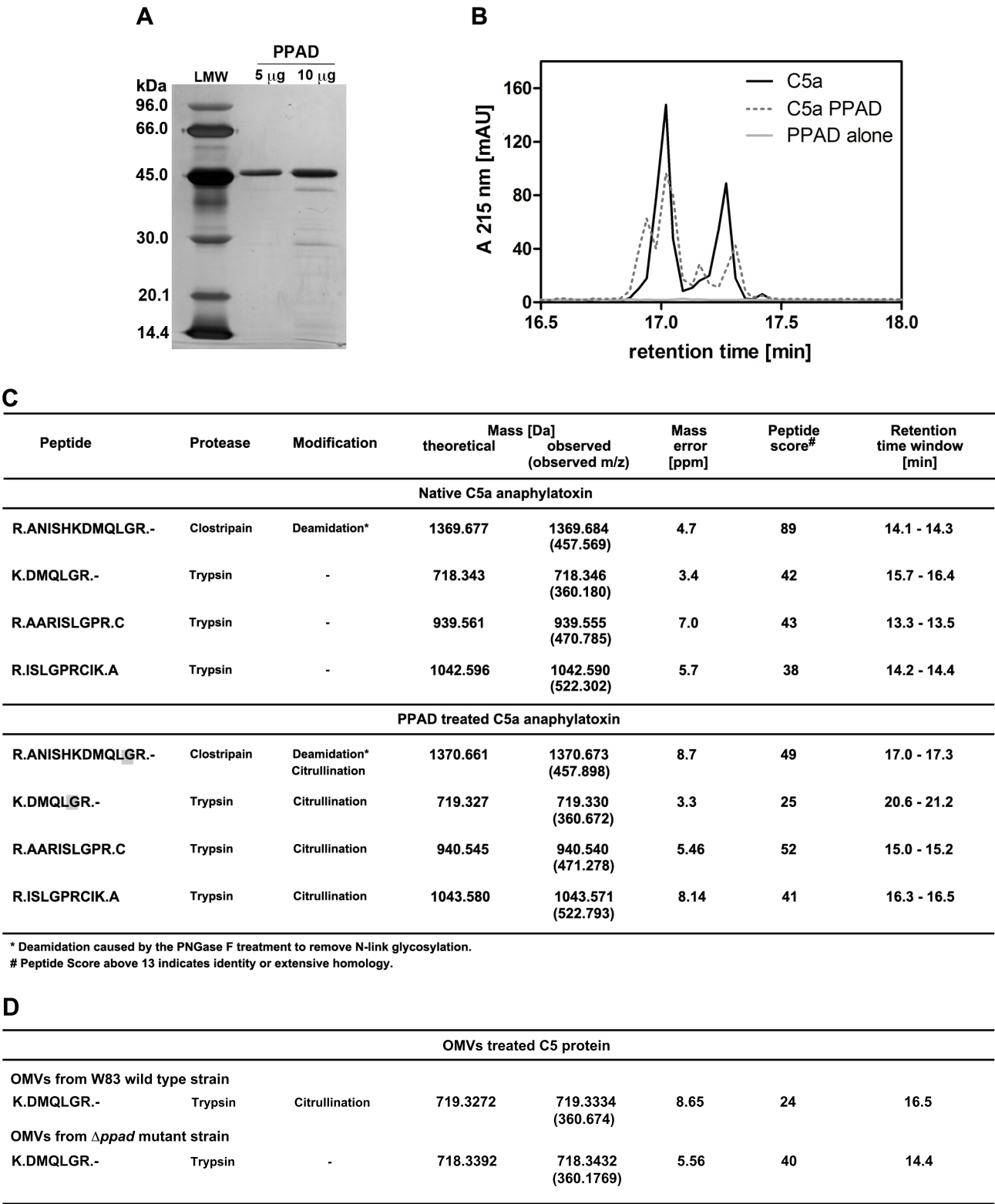
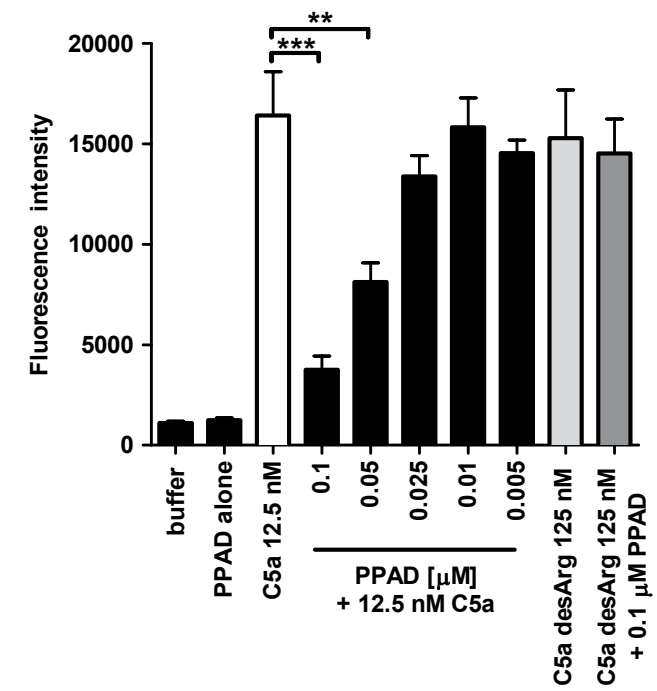
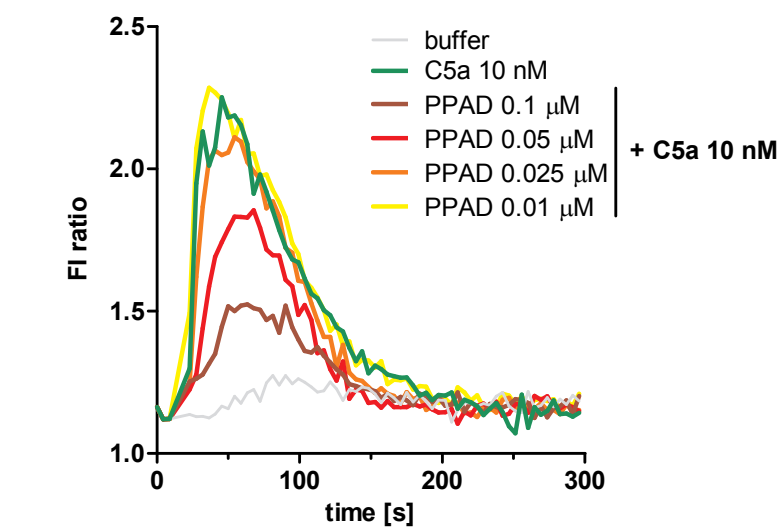


Fig. 2

A



B



C

