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Reappraisal of the Regulation of Lactococcal L-Lactate Dehydrogenase

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Lactococcal lactate dehydrogenases (LDHs) are coregulated at the substrate level by at least two mechanisms: the fructose-1,6-biphosphate/phosphate ratio and the NADH/NAD ratio. Among the Lactococcus lactis species, there are strains that are predominantly regulated by the first mechanism (e.g., strain 65.1) or by the second mechanism (e.g., strain NCDO 2118). A more complete model of the kinetics of the regulation of lactococcal LDH is discussed.

Lactococci are known for their homolactic metabolism, whereby more than 90% of the sugars present are converted into lactic acid. However, under certain conditions, the metabolism may shift to the production of mixed acids (acetate, ethanol, and formate). From the 1960s onward, the view that the control of this shift was modulated mainly by the intracellular concentration of fructose-1,6-biphosphate (FBP) activating both L-lactate dehydrogenase (l-LDH; EC 1.1.1.27) and pyruvate kinase (EC 2.7.1.40) was held (3, 9, 18, 19). Inorganic phosphate (Pi) was recognized as a severe inhibitor of both enzymes. Apparently, both FBP and Pi were seen to compete for the same allosteric site of LDH (9).

Recently, this metabolic model was questioned by Garrigues et al. (5), who showed that the sugar metabolism of strain NCDO 2118 was instead regulated by the NADH/NAD ratio. A high ratio inhibited glyceraldehyde 3-phosphate dehydrogenase (GAPDH; EC 1.2.1.12) and increased LDH activity. Many, but not all, researchers took up this view without reflecting, creating a confusing situation. Currently, metabolic flux models based on enzyme kinetics are applied as predictive tools in metabolic engineering (see, e.g., reference 8), illustrating the importance of expressing kinetic characteristics adequately. Therefore, we undertook an investigation regarding the nature of the regulation of LDH activity among several Lactococcus lactis strains, including those frequently used in metabolic flux studies. We also examined the effects of ATP, ADP, AMP, and phosphoenolpyruvate (PEP), since Jonas et al. (9) observed strong competitive inhibition by ATP of LDH activity in L. lactis strain US3 (=NCIMB701197).

L. lactis strains (listed in Table 1) were each grown anaerobically at 30°C in pH-controlled batch cultures on glucose (10 g/liter) with SD3 medium (17). The external pH was kept at 6.5, which corresponds to an internal pH of 7.2 (12). The cultures were harvested in the late exponential phase, centrifuged at 4°C for 10 min at 5,000 × g, washed, and resuspended in triethanolamine buffer (50 mM; pH 7.2), 5 mM MgCl2 · 6H2O. Cell extracts (CE) were prepared by using glass beads (17). LDH activity was measured spectrophotometrically by monitoring the oxidation of NADH (340 nm; ε = 6,220 M⁻¹·cm⁻¹) at 30°C. One milliliter of the standard assay mixture consisted of triethanolamine buffer (50 mM; pH 7.2), NADH (0.3 mM), FBP (10 mM), and CE (approximately 160 mg of protein/liter). The reaction was initiated by the addition of pyruvate (initial concentration, 10 mM). One unit of LDH was defined as the amount of enzyme that oxidized 1 μmol of NADH min⁻¹. To ensure consistent reproducible values, the level of LDH activity was determined by testing the activities of four different concentrations of CE. The enzyme activity was always proportional to the concentration of the enzyme. The LDH of each strain was characterized in vitro by the addition of pyruvate, Pₐ, pyruvate, PEP, ATP, ADP, AMP, NADH, and NAD at various concentrations. Protein concentration was assayed as described by Bradford (2). Parameter estimation was carried out by application of Lineweaver-Burk plots or the least-squares method.

The FBP concentrations at which LDH was 50% activated (KFBP0.5V) differed among the strains (Table 1). Strain 65.1 was found to have an LDH that was activated by substantial concentrations of FBP and was strongly inhibited by Pi (Table 1 and Fig. 1A) but was only slightly affected by the NADH/NAD ratio (Fig. 1B). Both the inhibition constant and the critical concentration of NAD were relatively high. ATP and ADP had threshold concentrations (at which they start being inhibitory) of 4 and 2 mM, respectively, for strain 65.1. These findings are in strong contrast to those for the LDHs of strains NCDO 2118 and ATCC 19435; for these strains, high activities (approximately 67% of the maximum activity) were found in the presence of FBP at the micromole level and Pi, was hardly inhibitory (Table 1 and Fig. 1A). However, their LDHs were significantly regulated by the NADH/NAD ratio (Fig. 1B). For full activation, strain NCDO 2118 still required intracellular FBP concentrations of about 4 to 5 mM (Table 1), although FBP concentrations were found to be far in excess even under substrate-limiting conditions (5). Therefore, in this strain, FBP will not have a significant influence on the performance of LDH, and the results obtained in the present study support those of Garrigues et al. (5). The LDHs of strains 65.1 and NCDO 2118 were at two extremes with regard to their regulation at the substrate level. All other strains tested possessed mixtures of both regulation mechanisms (Table 1 and Fig. 1). Therefore, we conclude that the activity of lactococcal LDH is regulated by a combination of two mechanisms, the FBP/P ratio (the P [phosphorus] type) and the NADH/NAD ratio.
(the R [redox] type). The former mechanism is based on allosteric regulation (3), while the latter displayed competitive inhibition by NAD (Fig. 2), possibly at the active site. In addition, ADP and ATP, showing mixed inhibition (15), contribute to the inhibitory effect of NAD. There was inhibition by AMP, but since this compound is present in the cell at low concentrations, it was considered negligible. PEP at concentrations of up to 10 mM did not inhibit LDH in any of the strains studied.

As described previously (3, 9), FBP was seen to affect the $K_{m}$ for NADH and pyruvate. Increasing the FBP concentration in the assay lowered the affinity constants of all strains tested in our study (Table 2), revealing that the allosteric mechanism is also present to some extent in the LDH of strain NCDO 2118. From the kinetics study by Crow and Pritchard (3), it can be concluded that pyruvate did not affect the $K_{m}$ for NADH and vice versa. The kinetics of LDH can be described by the Monod-Wyman-Changeux rate equation (10) adapted to the use of two substrates (7) and incorporating inhibition by NAD, ATP, and ADP (Table 3). Estimation of this inhibition by the kinetics according to the work of Han and Levenspiel (6) was superior to estimation by mixed-inhibition kinetics (13) (results not shown). The inhibitory effects of ATP and ADP appeared to be one of a kind and could therefore be grouped within one inhibitory term (Table 3). This conclusion was supported by the outcome of assays in which mixtures of the three inhibitors were used (data not shown). The equation could be simplified because $V_{T,\text{max}}$ (the maximum rate of the enzyme in the T state) was zero for most strains studied. The backward reaction of the catalytic step of LDH was not significant here, with $\gamma/K_{eq}$ (where $\gamma$ is the mass-to-action ratio [lactate]/[pyruvate] and $K_{eq}$ is the equilibrium constant of the reaction catalyzed by LDH) being negligible when the data provided by Hoefnagel et al. (8) were used.

To obtain some indication of how the P and R types of LDH (of strains 65.1 and NCDO 2118, respectively) might behave under in vivo conditions, a small in silico model study was carried out. For this study, the kinetics of both types of LDH were subjected to the data for the glycolytic dynamics of resting cells of *L. lactis* MG1363 measured with $^{13}$C and $^{31}$P nuclear

<table>
<thead>
<tr>
<th>Strain</th>
<th>Activation by FBP (mM)</th>
<th>$K_{FPR,0.5V}$ (mM)</th>
<th>$K_{NADH}$ (mM)</th>
<th>NAD$_{crit}$ (mM)</th>
<th>ATP$_{crit}$ (mM)</th>
<th>ADP$_{crit}$ (mM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>65.1</td>
<td>0.550</td>
<td>10</td>
<td>9.2</td>
<td>25</td>
<td>35</td>
<td>25.4</td>
</tr>
<tr>
<td>MG 1363</td>
<td>0.006</td>
<td>5</td>
<td>10.3</td>
<td>1.8</td>
<td>26</td>
<td>21.6</td>
</tr>
<tr>
<td>NCIMB701197</td>
<td>0.010$^{a}$</td>
<td>1$^{a}$</td>
<td>17</td>
<td>0.54</td>
<td>13</td>
<td>14.4</td>
</tr>
<tr>
<td>NCIMB700509</td>
<td>0.002$^{b}$</td>
<td>5$^{b}$</td>
<td>23</td>
<td>0.38</td>
<td>9.9</td>
<td>16.7</td>
</tr>
<tr>
<td>NCIMB700763</td>
<td>0.002</td>
<td>8</td>
<td>31</td>
<td>0.44</td>
<td>10.3</td>
<td>13.8</td>
</tr>
<tr>
<td>IL 1403</td>
<td>0.003</td>
<td>5</td>
<td>38</td>
<td>0.5</td>
<td>13</td>
<td>15.8</td>
</tr>
<tr>
<td>ATCC 19435</td>
<td>&lt;0.001$^{c}$</td>
<td>2</td>
<td>2,400</td>
<td>2.3</td>
<td>15</td>
<td>31.4</td>
</tr>
<tr>
<td>NCDO 2118</td>
<td>&lt;0.001$^{d}$</td>
<td>4.5</td>
<td>592,500</td>
<td>2.1</td>
<td>15.3</td>
<td>19.6</td>
</tr>
</tbody>
</table>

$^{a}K_{FPR,0.5V}$, FBP concentration at which LDH was 50% activated in the absence of P$_i$; Total, FBP concentration at which LDH was totally activated in the absence of P$_i$.

$^{b}K_{FPR,0.5V}$, phosphate concentration at which LDH was 50% inhibited, as extrapolated to 0 mM FBP.

$^{c}K_{NADH}$, inhibition constant of NAD based on kinetics of competitive inhibition.

$^{d}$NAD$_{crit}$, ATP$_{crit}$, and ADP$_{crit}$, concentrations of NAD, ATP, and ADP, respectively, at which LDH is expected to be inhibited completely based on the equation of Han and Levenspiel (6).

$^{e}$Taken from literature (9).

$^{f}$Taken from literature (3).
activity of the R-type LDH was insensitive to L, that its value should be low (0 to 1) for the P-type LDH. This experimentally determined. However, the simulations revealed the R (relaxed) state of the enzyme without substrate, was not respectively, in the T state.

\[
V_{1,\text{LDH}} = V_{R,\text{max}} \cdot \left[ \frac{\alpha_p \delta \rho R^2 (R^2 + \Delta L^2)}{(R^2 + \Delta L^2)^2} \right] (1 - \delta) (1 - \epsilon - \eta), \text{ an equation for the expression of lactococcal LDH}^a
\]

**TABLE 3.** Variables used in \( V_{1,\text{LDH}} \) = \( V_{R,\text{max}} \) · \( \frac{\alpha_p \delta \rho R^2 (R^2 + \Delta L^2)}{(R^2 + \Delta L^2)^2} \) · (1 − δ) · (1 − ε − η), an equation for the expression of lactococcal LDH

<table>
<thead>
<tr>
<th>Variable</th>
<th>Equivalency</th>
</tr>
</thead>
<tbody>
<tr>
<td>( \alpha_p )</td>
<td>( \text{[pyruvate]/K}_{R,\text{Pyr}} )</td>
</tr>
<tr>
<td>( \epsilon )</td>
<td>( \frac{E_{\text{max}}}{K_{R,\text{Pyr}} + K_{T,\text{Pyr}}} )</td>
</tr>
<tr>
<td>( \delta )</td>
<td>( \frac{[\text{NAD}]/[\text{NAD}^\text{crit}]}{[\text{Pi}]/[\text{Pi}^\text{crit}]} )</td>
</tr>
<tr>
<td>( \eta )</td>
<td>( \frac{[\text{ADP}]/[\text{ADP}^\text{crit}]}{[\text{ATP}]/[\text{ATP}^\text{crit}]} )</td>
</tr>
<tr>
<td>( \beta )</td>
<td>( \frac{[\text{FbP}]/[\text{FbP}^\text{crit}]}{[\text{Pyr}]/[\text{Pyr}^\text{crit}]} )</td>
</tr>
<tr>
<td>( R )</td>
<td>( \frac{1 + \alpha_p + \delta}{1 + \alpha_p + \delta + \alpha_p \delta} )</td>
</tr>
<tr>
<td>( c_N )</td>
<td>( \frac{[\text{NAD}]/[\text{NAD}^\text{crit}]}{[\text{NAD}]/[\text{NAD}^\text{crit}]} )</td>
</tr>
<tr>
<td>( c_{NAD} )</td>
<td>( \frac{[\text{NAD}]/[\text{NAD}^\text{crit}]}{[\text{NAD}]/[\text{NAD}^\text{crit}]} )</td>
</tr>
<tr>
<td>( c_{pi} )</td>
<td>( \frac{[\text{Pi}]/[\text{Pi}^\text{crit}]}{[\text{Pi}]/[\text{Pi}^\text{crit}]} )</td>
</tr>
<tr>
<td>( L )</td>
<td>( \frac{L_a}{(1 + \gamma)} )</td>
</tr>
<tr>
<td>( \gamma )</td>
<td>( \frac{[\text{P}]/[\text{P}^\text{crit}]}{[\text{P}]/[\text{P}^\text{crit}]} )</td>
</tr>
<tr>
<td>( T )</td>
<td>( \frac{1 + \epsilon \rho \delta + c_N c_{NAD} + c_{p} \delta c_{NAD}}{1 + \epsilon \rho \delta + c_N c_{NAD} + c_{p} \delta c_{NAD}} )</td>
</tr>
</tbody>
</table>

\( V_{R,\text{max}} \) maximum enzyme rate in the R state; \( L_a \), the equilibrium between the \( T \) and \( R \) states; \( K_{R,\text{Pyr}} \) and \( K_{T,\text{Pyr}} \), affinity constants for pyruvate in the R and T states, respectively; \( K_{N\text{ADH}} \), \( K_{T,\text{Pyr}}, \) and \( K_{T,\text{Pyr}}^\text{crit} \), concentrations of NAD, ADP, and ATP, respectively; \( L \), NAD concentration at which LDH was 50% activated in the absence of FBP, \( K_{R,\text{NADH}} \) and \( K_{T,\text{NADH}} \), affinity constants for LDH in the R and T states, respectively; \( L_a \), equilibrium between the T state and the R state of the LDH enzyme without substrate; \( K_{P,\text{Pyr}}^\text{crit} \), phosphate concentration at which LDH was 50% inhibited, as extrapolated to 0 mM FBP; \( m \) and \( n \), number of modulator sites.

The substrate concentrations of LDH were present at near their respective \( K_m \) values, meaning that the enzyme operated at far less than maximum capacity (both LDH types operated at about 10% of their maximum capacities under the given experimental conditions). Under most conditions, the FBP concentration will be far beyond its respective \( K_{FP,0.5V} \) value unless the flux becomes very low. In contrast to the P type, for the R type, the internal phosphate concentration never came near its respective \( K_{FP,0.5V} \) value.

Although both views on the regulation of lactococcal LDH...
(5, 9) appear to be correct, each one addresses only one type of LDH. Since the critical concentrations of ATP and ADP are quite similar, if one considers their cellular moiety to be nearly constant during growth, their inhibitory effect on LDH will have no regulatory function (likewise for GAPDH [results not shown]). However, if the size of their moiety were to undergo changes, e.g., as a function of the growth rate, then this change might very well act as a regulation mechanism. Here, we show that there are at least two types of regulation at the substrate level. These two types of regulation operate to different extents within each strain, and the overall kinetics can be described by a general equation (Table 3). Interestingly, the R-type LDH could be found in the lactococcal strains that were isolated from plant material (strains ATCC 19435 and NCDO 2118), while the P type or a mixture of both types prevails in the typical dairy strains. We can only speculate about whether the distinction in regulation among the strains is a result of small differences in the homologies of lactococcal LDHs (15) or whether different LDH genes are expressed at different ratios (1).

The technical assistance of Therese Segerstein is greatly appreciated.

REFERENCES