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Tumor necrosis factor restricts hematopoietic stem cell activity in mice: involvement of two distinct receptors

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Whereas maintenance of hematopoietic stem cells (HSCs) is a requisite for life, uncontrolled expansion of HSCs might enhance the propensity for leukemic transformation. Accordingly, HSC numbers are tightly regulated. The identification of physical cellular HSC niches has underscored the importance of extrinsic regulators of HSC homeostasis. However, whereas extrinsic positive regulators of HSCs have been identified, opposing extrinsic repressors of HSC expansion in vivo have yet to be described. Like many other acute and chronic inflammatory diseases, bone marrow (BM) failure syndromes are associated with tumor necrosis factor–α (TNF) overexpression. However, the in vivo relevance of TNF in the regulation of HSCs has remained unclear. Of considerable relevance for normal hematopoiesis and in particular BM failure syndromes, we herein demonstrate that TNF is a cell-extrinsic and potent endogenous suppressor of normal HSC activity in vivo in mice. These effects of TNF involve two distinct TNF receptors.

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(Tartaglia et al., 1993), implicate that some functions of TNF might be facilitated by or even strictly depend on the expression of both receptors, although evidence for this is limited.

The physiological relevance of the ability of TNF to suppress mouse and human HSC maintenance in vitro (Zhang et al., 1995; Bryder et al., 2001; Dybedal et al., 2001) can rightfully be questioned, among other reasons because the investigated concentrations of TNF might be physiologically irrelevant, because TNF exists in two isoforms (TNF and TNF-β; Aggarwal, 2003), and because the in vitro experiments could not mimic the physiological balance and potentially distinct roles of soluble and membrane-bound TNF in vivo (Grell et al., 1995). Furthermore, TNF is likely to have pleiotropic direct and indirect effects on HSCs in vivo. In fact, previous in vivo TNF loss of function experiments did not only fail to support a suppressive role of TNF in HSC regulation, they rather implicated a stimulatory role of TNF in HSC maintenance in vivo (Rebel et al., 1999), in apparent discrepancy with studies demonstrating a potent suppressive effect of TNF on normal HSCs in vitro (Bryder et al., 2001; Dybedal et al., 2001), as well as HSCs in Fanconi anemia models for BM failure syndromes being hyperresponsive to TNF suppression in vitro (Milsom et al., 2009). Herein, we sought to reconcile these previous findings and to establish the in vivo role of TNF in the regulation of HSC maintenance and expansion in mice lacking expression of either or both TNF receptors.

RESULTS AND DISCUSSION

TNF restricts HSC activity in vivo

We first investigated HSC numbers and function in mice deficient for both Tnfrsf1a (Tnfrsf1a−/−) and Tnfrsf1b (Tnfrsf1b−/−) receptors, hereafter referred to as Tnfrsf1 double KO (Tnfrsf1−/−dKO; Peschon et al., 1998). Unlike previous studies of Tnfrsf1a−/− mice (Zhang et al., 1995; Rebel et al., 1999), we did not observe any differences in phenotypically enriched HSCs (LSKFLT3−; Fig. 1, b and c; Adolphson et al., 2001) nor in frequencies of long-term reconstituted mice when unfractionated BM cells were transplanted at limiting numbers from steady-state BM of WT and Tnfrsf1−/−dKO (Fig. 1 d), suggesting that HSCs are not affected by TNF in steady-state. Steady-state cell cycle analyses of cells within the LSKFLT3− HSC-enriched compartment revealed only a minor increase of cells in active cell cycle (G1 and S/G2/M) in Tnfrsf1−/−dKO mice as compared with WT mice, and in both cases LSKFLT3− cells were predominantly found to be highly quiescent (Fig. 1, e and f).

In contrast to the steady-state analysis, Tnfrsf1−/−dKO BM cells revealed a considerable competitive advantage over WT BM cells in their ability to long-term reconstitute myeloid and lymphoid cell lineages in myeloabluted WT recipients (Fig. 2, a–f). This competitive advantage of Tnfrsf1−/−dKO BM cells was further enhanced after three rounds of serial transplantation and maintained for a total of at least five rounds of transplantation (Fig. 2 b). An enhanced Tnfrsf1−/−dKO donor contribution to the phenotypically defined LSK compartment (enriched for multipotent stem and progenitor cells; Fig. 2 f) mimicked the increased multilineage peripheral blood (PB) reconstitution levels (Fig. 2, b and d), supporting the idea that the suppressive effect of TNF is targeted to multipotent HSCs. Although we did not specifically address the homing of HSCs, altered HSC homing as a consequence of TNF receptor deficiency is unlikely to explain the increased long-term reconstitution, as short-term (3 wk) reconstitution levels were unaffected in recipients of Tnfrsf1−/−dKO BM cells (Fig. 2 b).

TNF-mediated suppression of HSC activity is dependent on expression of both TNF receptors

As previous studies of the specific hematopoietic actions of TNF mediated through Tnfrsf1a and Tnfrsf1b have been conflicting (Zhang et al., 1995; Rebel et al., 1999), we next evaluated the consequence of loss of either of the two TNF receptors by studying mice deficient for either Tnfrsf1a (Tnfrsf1a−/−) or Tnfrsf1b (Tnfrsf1b−/−) receptors. Notably, we observed a competitive multilineage PB and BM LSK reconstitution advantage of BM cells from Tnfrsf1a−/− (Fig. 2, g and i) as well as Tnfrsf1b−/− (Fig. 2, h and j) mice. This advantage was sustained after secondary transplantations (Fig. 2, g–j), although the competitive advantage was more pronounced with BM cells from Tnfrsf1−/−dKO mice (Fig. 2, a–f), implicating the importance of both TNF receptors in restricting HSC expansion.

We next undertook a series of in vitro experiments to try to corroborate the requirement of both TNF receptors for the observed TNF-induced restriction of HSC activity. As expected, the clonal growth of WT BM cells was strongly inhibited in response to TNF (Fig. 3, a–c). Notably, TNF had no discernable effect on the clonal growth of different myeloerythroid progenitors from Tnfrsf1a−/− or Tnfrsf1b−/− mice (Fig. 3, a–c), and the same observation was made with regard to the effect of TNF on the growth of HSC-enriched LSK cells (Fig. 3 d), which is compatible with both Tnfrsf1a and Tnfrsf1b being required for the suppressive effects of TNF on HSCs and hematopoiesis. Importantly, TNF-mediated growth inhibition could not be rescued in cells overexpressing the antiapoptotic protein BCL2 (Fig. S1, a–c; Domen et al., 1998), suggesting that induction of apoptosis plays little or no role in TNF-induced growth inhibition and HSC expansion. However, as these experiments were performed in vitro and as there are alternative, BCL2-resistant pathways for apoptosis, we cannot rule out the possibility that the suppressive effects of TNF on HSCs might involve apoptosis.

In vivo administration of TNF targets HSCs

Although no other ligands than TNF and TNF-β are known for the two TNF receptors, we could not rule out the possibility that the HSC phenotype in Tnfrsf1a−/− and Tnfrsf1b−/− mice is not mediated by TNF. Thus, we next investigated the effects of TNF administered in vivo on HSCs. Furthermore, as the suppressive impact of TNF on HSC activity after transplantation but not in steady-state hematopoiesis could in part be related to HSCs being recruited from quiescence
induced a reduction in total BM cellularity (Fig. 4 b) and a reduction in HSC activity as determined in a competitive transplantation assay (Fig. 4, c–f). This suppressive effect of TNF on HSC activity was enhanced in 5-FU–treated mice (Fig. 4, b–f). These findings suggest that actively cycling rather than quiescent HSCs are the primary targets of TNF suppression, which is of relevance for the enhanced reconstitution after transplantation.

This study provides the first in vivo evidence of a non-redundant, cell-extrinsic factor involved in negative regulation of the HSC compartment, specifically in a setting relevant for BM transplantation. Importantly, our experiments of mice
Figure 2. Enhanced activity of TNF receptor–deficient HSCs after transplantation. (a–f) BM cells were pooled from three Tnfrsf1-dKO (CD45.2+) or three WT (CD45.1+) mice (8–12 wk old), and for each of the genotypes 10⁶ unfractionated BM cells were competitively cotransplanted into lethally irradiated congenic WT recipients (CD45.1+/2+). Recipient mice were evaluated for PB multilineage reconstitution levels at the indicated time points. (a) Representative FACS analysis illustrating PB myeloid chimerism evaluation. (b) PB myeloid reconstitution derived from transplanted Tnfrsf1-dKO and WT cells was analyzed at the indicated time points. At the time points indicated by the vertical dashed lines, mice were sacrificed, and 0.5 femur equivalent of BM cells were serially transplanted into newly irradiated (CD45.1+/2+) recipients. Displayed are PB myeloid chimerism levels. Results are mean (SEM) values from six separate experiments with five to seven recipients in each experiment. (c–f) 16 wk after the primary transplantation, PB analysis of T (CD4/CD8), myeloid (MAC1), and B (B220) cell reconstitution was performed (d), BM was analyzed for LSK chimerism (f), and mean percentages for all recipients are presented. (c and e) Representative FACS analysis for reconstitution levels within the PB and BM are shown. (d and f) Results are from six separate experiments with five to seven recipients in each experiment. Mean (SEM) values are shown. (g–j) BM cells from 8–12 wk old Tnfrsf1a−/− (g and i) and Tnfrsf1b−/− (h and j) mice (CD45.2+; backcrossed for 10 generations with C57BL/6 mice) were transplanted in competition with WT BM cells (CD45.1+) into lethally irradiated WT (CD45.1+/2+) recipients. PB myeloid chimerism levels (g and h) and BM LSK chimerism levels (i and j) at the indicated time points after primary and secondary (indicated by dashed line) transplantations are shown. All results are mean (SEM) values from two separate experiments with five to seven recipients in each experiment. PB and BM reconstitution analysis were performed as illustrated in panels a and e, respectively.
HSCs, either through signaling through both receptors or potentially through the suggested role of the Tnfrsf1b in ligand passing to the Tnfrsf1a receptor (Aggarwal, 2003).

The current findings are of considerable relevance for several BM failure syndromes, in which enhanced expression of TNF has been implicated in the pathogenesis of hematopoietic failure deficient in Tnfrsf1a or Tnfrsf1b expression, corroborated by in vitro TNF response experiments, indicate that the ability of TNF to suppress HSCs is dependent on expression of both TNF receptors. This clearly implicates the importance of both receptors in eliciting the signaling required to suppress HSCs, either through signaling through both receptors or potentially through the suggested role of the Tnfrsf1b in ligand passing to the Tnfrsf1a receptor (Aggarwal, 2003).

The current findings are of considerable relevance for several BM failure syndromes, in which enhanced expression of TNF has been implicated in the pathogenesis of hematopoietic failure deficiency.
(Bagby and Meyers, 2007; Aalto et al., 2011). For instance, in the case of Fanconi anemia, TNF has not only been demonstrated to be overexpressed in the BM (Dufour et al., 2003), but HSCs have also been found to be hyperresponsive to TNF suppression in vitro (Milsom et al., 2009). The physiologic relevance of these in vitro findings, as well as previous findings demonstrating that TNF has potent suppressive effects on WT mouse and human HSCs in vitro (Bryder et al., 2001; Dybedal et al., 2001), are supported by our demonstration of a suppressive role of Tnfrsf1a and Tnfrsf1b on HSCs in vivo, as well as an HSC-suppressive effect of TNF administration in vivo. The reason for the opposite conclusions reached in the experiments by Rebel et al. (1999) remains unclear. In that study, Tnfrsf1a was rather suggested to play a critical positive role in maintaining HSCs. This role of TNF appeared to be restricted to older mice (>6 mo old), although phenotypic analysis of the HSC compartment in older mice showed no significant difference from WT mice (Rebel et al., 1999).

Thus, it was only in long-term transplantation experiments that reduced multilineage reconstitution was observed from Tnfrsf1a−/− BM cells as an indication of reduced HSC activity (Rebel et al., 1999), although phenotypic analysis of the HSC compartment itself was not performed in transplanted mice. It is not obvious how the age of the TNF receptor–deficient HSCs could explain the different conclusions reached between this previous study and our study, as we in our study performed serial transplantation experiments for up to 76 wk with HSCs with an intrinsic TNF receptor deficiency. Moreover, in the study, the enhanced long-term multilineage reconstitution levels of TNF receptor–deficient BM cells were corroborated by correspondingly enhanced chimerism also in the phenotypic HSC compartment. Although the mice used herein carry the same mutated alleles, the genetic background of the TNF receptor–deficient strains and the nature of the WT control mice were not described in detail in the study of Rebel et al. (1999), and therefore the apparent discrepancy could potentially be explained by this. In that regard, our study included experiments in which experimental mice had been backcrossed to a C57BL/6 genetic background and in which WT littermates were used as controls.

This study establishes the in vivo relevance of the suppressive effects of TNF on HSCs in hematopoiesis and BM failure syndromes and supports the further exploration of anti-TNF treatment in such conditions (Dufour et al., 2009; Scott et al., 2010; Aalto et al., 2011). The in vivo condition-modeling experiments used in this study, 5-FU treatment and irradiation, have been shown to cause increases in blood levels of TNF (Xun et al., 1994; Okamoto et al., 2000), suggesting that anti-TNF treatment might also be beneficial to enhance postransplantation reconstitution (Ferrara et al., 2009).

MATERIALS AND METHODS

Mice. Tnfrsf1a−/− mice (TNF receptor double KO; Peschon et al., 1998) were obtained from the Jackson Laboratory (stock no. 003243). The Tnfrsf1a−/− mice were originally generated by interbreeding mice deficient for Tnfrsf1a (or Tnf−p85, generated on a C57BL/6 background) with mice deficient for Tnfrsf1b (or Tnf−p75, generated on a 129 background), followed by subsequent backcrossing for five generations to C57BL/6 mice. Tnfrsf1a−/− mice were used for phenotyping and competitive transplantation experiments, with WT C57BL/6 mice (CD45.2+, CD45.1+, or F1 CD45.1+/CD45.2+) as controls. Tnfrsf1a−/− and Tnfrsf1b−/− single KO mice were generated by crossing Tnfrsf1a−/− to WT C57BL/6 mice to create single KO mice with identical allelic mutations as the original Tnfrsf1a−/− and Tnfrsf1b−/− mice, respectively. Heterozygous offspring were interbred to generate Tnfrsf1a−/− and Tnfrsf1b−/− mice for in vitro experimentation (Fig. 3, a–d), and WT littermates were used as controls. Single KO mice were also backcrossed with C57BL/6 mice for an additional four generations to generate 10th generation Tnfrsf1a−/− and Tnfrsf1b−/− single KO mice. These mice were used in the transplantation experiments. H2K−BCL-2 mice have been described previously (Domen et al., 1998). Mice were maintained at the Lund University animal facility. All mice procedures were performed with consent from the local ethics committee (Malmö/Lunds djurförsöksnämnd).

Phenotypic analysis and purification of HSCs. BM cells were collected from femurs, tibiae, and sometimes iliac crests. For both phenotypic analysis and FACs purification, unfraccionated BM cells were stained with either purified or fluorochrome-conjugated lineage antibodies against B220 (RA3-6B2), GR-1 (RB6-8C5), MAC1 (M1/70), CD8 (3–6.7), TER119, CD4 (H129.19), and sometimes CD5 (53–7), all from BD. FACs purifications were performed as previously described (Adolfsson et al., 2001). In brief, cells were next either lineage depleted using sheep anti–rat IgG(Fc)–conjugated immunomagnetic beads (Invitrogen) or ckit–conjugated magnetic beads (Milenyi Biotec). Purified lineage antibodies were visualized with CyChrome (Invitrogen)–or Qdot 605 (Invitrogen)–conjugated goat anti–rat antibodies; in the case of lineage depletion, cells were stained after the lineage enrichment, and in the case of ckit enrichment, cells were stained before magnetic selection. Subsequently, cells were stained with different combinations of fluorochrome-conjugated antibodies against SCA1 (D7 [BioLegend] or E13-161.7 [BD]), ckit (or CD117, 2B8; ebioscience or BD), FLT3 (or FLK2/CD135, 2F10.1; BD or ebioscience), and CD150 (or Slamf1, TC15-12F12.2; BioLegend). Propidium iodide (Invitrogen) or 7-amo-actinomycin D (Sigma–Aldrich) was used to exclude dead cells.

Gating was performed to exclude doublets from analysis.

For calculation of total numbers of LSK and LSKFLT3− cells, the frequency of LSKFLT3− cells in each individual mouse, as determined by flow cytometric evaluation of unfraccionated BM cells, was multiplied by total BM cell numbers. Total BM numbers were calculated based on total BM cells in two femurs and two tibiae, representing 25% of total BM cells preserved at the Lund University animal facility. All mice procedures were performed with consent from the local ethics committee (Malmö/Lunds djurförsöksnämnd).

For calculation of total numbers of LSK and LSKFLT3− cells, the frequency of LSKFLT3− cells in each individual mouse, as determined by flow cytometric evaluation of unfraccionated BM cells, was multiplied by total BM cell numbers. Total BM numbers were calculated based on total BM cells in two femurs and two tibiae, representing 25% of total BM cells present in a mouse. For cell cycle analyses (Thorén et al., 2008), ~30 × 105 CD45.1+ (WT) and 30 × 105 CD45.2+ (Tnfrsf1-dKO) BM cells were mixed and stained with antibodies against lineage markers, SCA1, ckit, CD150, and FLT3 as described in the previous paragraph, as well as with antibodies against CD45.1 (A20; BioLegend) and CD45.2 (104; BioLegend). Subsequently, cells were fixed in 2% paraformaldehyde (GTG) and permethylated in 0.1% saponin from quillaja bark (Sigma–Aldrich) and stained with anti-Ki67 (B56; BD) or isotype-conjugated antibodies. Before analysis, cells were incubated in 2.5 µg/ml DAPI (Invitrogen) to visualize DNA content.

Cells were analyzed or sorted on FACs Aria or DIVA cell sorters (BD). All flow cytometry and FACs data were analyzed with FlowJo software (Tree Star).

In vivo transplantation experiments. Competitive transplantation experiments using the congenic CD45.1+/CD45.2+ mouse model were performed as described previously (Szilvassy et al., 1990; Adolfsson et al., 2001). In brief, CD45.2+ BM cells from TNF receptor–deficient mice were injected intravenously along with competitor CD45.1+ BM cells into lethally irradiated (950 cGy) CD45.1+/2+ mice. PB was collected at the indicated times posttransplantation and analyzed for donor reconstitution by FACs. PB was obtained from retroorbital bleeding, red blood cells were sedimented with 2% dextran T-500 (Pharmacia), remaining red blood cells

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were lysed with ammonium chloride, and leukocytes were subsequently stained with antibodies against CD3, B220, and MAC1 or against CD4, CD8, B220, MAC1, and NK1.1 (PK136; BioLegend).

In limiting dilution experiments, positively multilineage reconstituted mice were defined as having a minimum of 0.1% total donor PB cells and stained with antibodies against CD3, B220, and MAC1 or against CD4, HSCs sorted from WT or

Fig. S1 shows clonogenic capacities of t


test was used to generate all p-values. Statistical analyses were performed using Excel software (Micro-

and 25 ng/ml rhG-SCF (Amgen), with or without 20 ng/ml TNF. After 11 d

and streptomycin/penicillin as above, with the addition of 25 ng/ml rrSCF

cells were single cell sorted in Terasaki plates into IMDM, supplemented

into serum-free medium (X-vivo15; BioWhittaker), supplemented with 1%

were transplanted in competition with 106 fresh CD45.1 + BM into each

previously described (Adolfsson et al., 2001), and cells were cultured in the presence or absence of 20 ng/ml TNF in addition to the indicated cytokines. For evaluation of proliferative capacities, HSC-enriched cells were sorted


Cheng, T., N. Rodrigues, H. Shen, Y. Yang, D. Dombkowski, M. Sykes, and D.T. Scadden. 2000. Hematopoietic stem cell quiescence maintained by 1% detoxified bovine serum albumin (STEMCELL Technologies), 0.1 mM β-mercaptoethanol, 2 mM t-glutamine (Intriviogen), streptomycin/penicillin (Sigma-Aldrich), and cytokines rSCF (25 ng/ml; Amgen or PeproTech), rhIL-6 (50 ng/ml; provided by Genetics Institute Cambridge, San Francisco, CA), and rmIL-3 (20 ng/ml; PeproTech), with or without 20 ng/ml TNF. After 7 d of culture, total cell numbers were counted, and expansion equivalents were calculated. For evaluation of clonogenic capacity, HSC-enriched cells were single cell sorted in Terasaki plates into IMDM, supplemented with 10% prescreened FCS (Intriviogen), β-mercaptoethanol, t-glutamine, and streptomycin/penicillin as above, with the addition of 25 ng/ml rSCF and 25 ng/ml rhG-SCF (Amgen), with or without 20 ng/ml TNF. After 11 d

in culture, wells were evaluated for colony numbers and size.

Statistics. Statistical analyses were performed using Excel software (Micro-

Student’s t test was used to generate all p-values.

Online supplemental material. Fig. S1 shows clonogenic capacities of HSCs sorted from WT or B62 transgenic mice cultured in the presence and absence of TNF. Online supplemental material is available at http://www

jem.org/cgi/content/full/jem.20110752/DC1.

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REFERENCES


Allopp, R.C., S. Cheshier, and L.L. Weissman. 2001. Telsonere short-

enabling increased increased cell cycle activity during serial trans-


