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Endogenous collagen peptide activation of CD1d-restricted NKT cells ameliorates tissue-specific inflammation in mice

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NKT cells in the mouse recognize antigen in the context of the MHC class I–like molecule CD1d and play an important role in peripheral tolerance and protection against autoimmune and other diseases. NKT cells are usually activated by CD1d-presented lipid antigens. However, peptide recognition in the context of CD1 has also been documented, although no self-peptide ligands have been reported to date. Here, we have identified an endogenous peptide that is presented by CD1d to activate mouse NKT cells. This peptide, the immunodominant epitope from mouse collagen type II (mCII707–721), was not associated with either MHC class I or II. Activation of CD1d-restricted mCII707–721–specific NKT cells was induced via TCR signaling and classical costimulation. In addition, mCII707–721–specific NKT cells induced T cell death through Fas/FasL, in an IL-17A–independent fashion. Moreover, mCII707–721–specific NKT cells suppressed a range of in vivo inflammatory conditions, including delayed-type hypersensitivity, antigen-induced airway inflammation, collagen-induced arthritis, and EAE, which were all ameliorated by mCII707–721 vaccination. The findings presented here offer new insight into the intrinsic roles of NKT cells in health and disease. Given the results, endogenous collagen peptide activators of NKT cells may offer promise as novel therapeutics in tissue-specific autoimmune and inflammatory diseases.

Introduction

The immune system responds to invading pathogens by recognizing their antigenic structures, while remaining unresponsive to self antigens. This simplified model, however, is challenged by the understanding that autoreactivity is a common feature of healthy organisms and a part of peripheral tolerance mechanisms. To generate animal models for autoimmune diseases, susceptible species are often immunized with self antigens. However, not all immunogenic self antigens have the capacity to provoke a pathogenic autoimmune response, and some of them confer protection (1). A widely accepted animal model for human rheumatoid arthritis is collagen-induced arthritis (CIA) in mice. CIA is induced by immunizing susceptible mouse strains with cartilage-derived triple helical type II collagen (CII). In mice expressing the MHC class II molecule H-2Aq, the disease-mediating epitope is an immunodominant glycopeptide derived from position 260–270 of rat CII (2). In comparison with heterologous CII, mouse CII, although capable of inducing arthritis, provokes a much weaker immune response to the immunodominant epitope and induces a lower incidence of arthritis (3). However, mouse CII immunization induces a recall proliferative response toward multiple epitopes in addition to the MHC class II–restricted CH260–270 peptide (4). The major epitopes share a common motif, with the strongest located at position 707–721 (mCII707–721). By investigating the immune response to this epitope, surprisingly, we found that this peptide was not MHC I/II restricted, but was binding and presented by CD1d, a cluster of differentiation glycoprotein. This led to the identification of a population of NKT cells that responded to mCII707–721 and were CD1d-restricted, TCRαβ ‘CD4’ ‘NK1.1’ cells.

CD1d is an alternative, MHC class I–like glycoprotein that presents antigens to NKT cells. The most extensively described population of CD1d-restricted NKT cells are reactive to the exogenous glycolipid α-galactosylceramide (αGalCer) (5). An endogenous ligand, isoglobotrihexosylceramide, has been shown to be involved in NKT cell autoreactivity (6). In addition to the association of CD1d with the binding and/or presentation of self and exogenous lipid or glycolipids (6), early reports have also indicated peptide-presenting capacity of CD1d (7–9).

Although the regulatory function of NKT cells is debated (5, 10), reports over the past decade have shown that both foreign and self-lipid–reactive CD1d-restricted NKT cells can regulate autoimmune diseases, moderate graft rejection, assist in host defense against infection, and promote tumor rejection, in spite of their relatively small population size (10–13). The immune regulatory function of CD1d-restricted NKT cells is ascribed to their capacity for rapidly releasing large quantities of cytokines, including IL-4 and IFN-γ (10). In the current report, we show that the mCII707–721–specific NKT cells are competent to produce IL-4, IFN-γ, IL-17A, TNF-α, and TGF-β upon peptide stimulation. However, the regulatory function of mCII707–721–specific NKT cells is likely through the Fas-FasL–mediated killing of activated T cells, not by cytokine production. Prevaccination with the mCII707–721 peptide resulted in downregulation of both Th1- and Th2-mediated immune responses and tissue inflammation in vivo. To our knowledge, this is the first report showing the existence of a CD1d-presented endogenous peptide

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The results have implications for anti-inflammatory vaccines for human autoimmune disorders.

**Results**

**Collagen peptide mCII<sub>707–721</sub> activates CD4<sup>+</sup> TCRαβ<sup>+</sup> NKT cells in an MHC I/II–independent manner.** We immunized B10.Q mice with mCII<sub>707–721</sub> and observed a strong and specific immune response, as measured by rechallenge of LN cells (LNCs) in vitro with mCII<sub>707–721</sub> (Figure 1A). No response was induced in the absence of antigen (media) or by using a nonrelated peptide control, the mouse collagen type I α1 chain peptide mCI<sub>707–721</sub>, which shares high homology with mCII<sub>707–721</sub>. mCI<sub>707–721</sub> was chosen as the negative control because this 15-aa peptide is similar to mCII<sub>707–721</sub> in size and sequence, differing in only 4 aa positions. It also does not elicit any T cell proliferation. Purified protein derivative (PPD) has been used as a positive control (recall antigen) for accuracy of immunization and proliferation, given that it was used for all the in vivo assays. PPD reactivity showed a proliferation index (mean ± SD) of 12.8 ± 2.1. We also immunized C57BL/6 mice per group with mCII<sub>707–721</sub> and observed a similar immune response as with B10.Q LNCs. Blocking with anti–MHC II and anti–MHC I antibodies produced similar levels of PI compared with mCII<sub>707–721</sub> with control antibody. PI was calculated by normalizing all cpm values to media control as 1. Data are mean ± SD, n = 6. **P ≤ 0.01; ***P ≤ 0.001.
We characterized the T cell response toward the mCII$_{707-721}$ peptide, using KO mice of genotypes MHC II$^{-/-}$, TAP1$^{-/-}$, CD1d$^{-/-}$, Tcrab$^{-/-}$, Tcrgd$^{-/-}$, CD4$^{-/-}$, and CD8$^{-/-}$, all backcrossed to the same genetic background, B10.Q. These were immunized and LNCs were rechallenged with or without antigens in vitro. Proliferative response to mCII$_{707-721}$ required TCR$^{\alpha\beta}$ expression, CD4$^+$ T cells, and CD1d, but was not dependent on TCR$^{\gamma\delta}$ expression or CD8$^+$ T cells (Figure 1B). No difference in proliferation was observed among the various KO mice and WT B10.Q mice in response to control peptides used in the assay (data not shown).

We investigated potential antigen-presenting molecules for mCII$_{707-721}$ using MHC II$^{-/-}$ mice and TAP-deficient mice (TAP1$^{-/-}$, Terab$^{-/-}$, Tergd$^{-/-}$, CD4$^{+/-}$, and CD8$^{-/-}$, all backcrossed to the same genetic background, B10.Q). These were immunized and LNCs were rechallenged with or without antigens in vitro. Proliferative response to mCII$_{707-721}$ required TCR$^{\alpha\beta}$ expression, CD4$^+$ T cells, and CD1d, but was not dependent on TCR$^{\gamma\delta}$ expression or CD8$^+$ T cells (Figure 1B). No difference in proliferation was observed among the various KO mice and WT B10.Q mice in response to control peptides used in the assay (data not shown).

We investigated potential antigen-presenting molecules for mCII$_{707-721}$ using MHC II$^{-/-}$ mice and TAP-deficient mice (TAP1$^{-/-}$), which lack transporter associated with antigen processing, important for presentation of MHC I-restricted peptides. By utilizing these mices (Figure 1B), as well as monoclonal antibodies to block MHC class I and II (Figure 1C), sustained proliferation to mCII$_{707-721}$ was observed. These data indicate that the mCII$_{707-721}$ response was neither MHC II- nor MHC I-restricted. An MHC II-binding assay also verified that the mCII$_{707-721}$ peptide did not bind to H-2$^k$ (data not shown). These results demonstrate that the T cell response to mCII$_{707-721}$ was dependent on CD4 and TCR$^{\alpha\beta}$, but unexpectedly, did not involve MHC I or II presentation.

mCII$_{707-721}$ binds to CD1d and activates NKT cells. Mice were immunized with mCII$_{707-721}$, and recall antigen reactivity of LNCs or splenocytes was measured 10–13 days after immunization. As shown in Figure 1B and Figure 1D, the proliferative response to mCII$_{707-721}$ was essentially completely diminished when mice genetically lacking CD1d-dependent NKT cells (CD1d$^{-/-}$ mice) were investigated or monoclonal antibodies to CD1d or NK1.1 were added to cultures prior to restimulation with mCII$_{707-721}$.
No effect on mCII\textsubscript{707–721} or PPD was observed (data not shown). Similarly, depletion of NK1.1\(^+\) cells, or antibody blocking of CD1d in vivo prior to subsequent immunization with mCII\textsubscript{707–721}, resulted in reduced proliferative response to mCII\textsubscript{707–721} (data not shown). Thus, the response to mCII\textsubscript{707–721} was confirmed to be CD1d restricted.

The binding capacity of CD1d for mCII\textsubscript{707–721} was determined using recombinant soluble dimeric mouse CD1d immunoglobulin and biotinylated mCII\textsubscript{707–721}. Significant binding of biotinylated mCII\textsubscript{707–721} to CD1d was observed, while no binding of the negative control, biotinylated mCII\textsubscript{707–721}, was detected. Excess nonbiotinylated mCII\textsubscript{707–721} peptide effectively competed for binding of biotinylated mCII\textsubscript{707–721} to CD1d (Figure 2A). Figure 2B shows that the interaction was dose dependent and saturable. Using GalCer with biotinylated mCII\textsubscript{707–721} revealed that at certain concentrations it competes for the binding to CD1d, indicating partial competition for the binding site (Figure 2C). PBS (non-CD1d-Ig dimer) is used for negative control.

\textit{mCII\textsubscript{707–721}} activates a heterogeneous NKT cell population. To investigate the frequency of NKT cell expansion in response to mCII\textsubscript{707–721}, WT B10.Q mice were treated in vivo with mCII\textsubscript{707–721} and CFA and 13 days later, splenocytes were analyzed. mCII\textsubscript{707–721} is capable of significantly inducing NKT cell expansion (gated on CD4\(^{–}\)NK1.1\(^+\) T cells) compared with all other control groups, namely vehicle-treated (CFA), mCII\textsubscript{707–721}, and naive mice (Figure 3, A and B).

To further characterize the mCII\textsubscript{707–721}-responding T cells, a peptide-specific cell line was established by subjecting LNCs from mice immunized with mCII\textsubscript{707–721} to different cycles of restimulation and rest. Expression of the NK1.1 marker was confirmed after each round of antigenic stimulation. Figure 3C shows that 57% of the CD4\(^+\)mCII\textsubscript{707–721}-specific T cells in the primary cell line were NK1.1\(^+\) T cells. This percentage increased to 65% after the second stimulation and to 88% after the third, indicating enrichment of NKT cells upon antigenic stimulation.

NKT cells can utilize a diverse combination of V\textalpha\textsubscript{v} and V\textbeta\textsubscript{v} chains, with evidence suggesting that any of the V\textbeta\textsubscript{v} chains can pair with the V\textalpha\textsubscript{v}14 invariant chain (14). The TCR V\textalpha\textsubscript{v} usage of the mCII\textsubscript{707–721}-specific NKT line was characterized by real-time quantitative PCR (qPCR) and FACS. A conventional T cell line specific to the CNS self-antigenic peptide myelin oligodendrocyte glycoprotein 79–96 (MOG\textsubscript{79–96}) was included for comparison. The mCII\textsubscript{707–721}-specific NKT cell line showed higher expression of V\textalpha\textsubscript{v}14-J\textalpha\textsubscript{8} than the conventional T cell line by real-time quantitative PCR (qPCR) (Figure 3D). Using a panel of V\textbeta\textsubscript{v} TCR antibodies to stain mCII\textsubscript{707–721}-specific NKT cells showed that these cells utilize polyvalent and diverse V\textbeta\textsubscript{v} chains (Figure 3E). Although some skewing toward usage of V\textbeta\textsubscript{8.2} is seen, mCII\textsubscript{707–721} reactivity is heterogeneous, not clonal.

\textbf{Activation of mCII\textsubscript{707–721}-reactive NKT cells requires TCR and costimulatory signaling.} To examine TCR signaling in the mCII\textsubscript{707–721}-specific response, we investigated ZAP-70 phosphorylation in the mCII\textsubscript{707–721}-specific NKT cell line. NKT cells stimulated with mCII\textsubscript{707–721} expressed phosphorylated ZAP-70 on the cell membrane, indicating TCR engagement and signaling (Figure 4A). Furthermore, blocking CD1d by adding a neutralizing antibody to the culture medium completely inhibited the appearance of phosphorylated ZAP-70 on the cell surface. Thus, interaction among CD1d, mCII\textsubscript{707–721}, and TCR is operative in the activation of mCII\textsubscript{707–721}-specific NKT cells.

The requirement for costimulatory signals in the activation of mCII\textsubscript{707–721}-specific NKT cells was studied using antibodies or fusion proteins to block the B7- and CD40-dependent pathways in peptide-stimulated LNC cultures from mCII\textsubscript{707–721}-immunized mice. Blocking either the CD40/CD40L or the B7/CD28 pathways led to significant inhibition of the proliferative response (Figure 4B) and showed that B7.1, but not B7.2, was the crucial ligand for CD28 on the NKT cells. The data support the interpretation that B7.1-CD28 and CD40-CD40L, in addition to CD1d-TCR ligation, are the necessary signals for mCII\textsubscript{707–721}-induced NKT cell activation.

mCII\textsubscript{707–721}-specific NKT cells produce cytokines. Cytokine production by the mCII\textsubscript{707–721}-specific NKT cells was measured by ELISA for IFN-\gamma, IL-2, IL-4, IL-10, TGF-\beta1, and TNF-\alpha in peptide-stimulated LNC cultures from mCII\textsubscript{707–721}-immunized mice. TGF-\beta1, IFN-\gamma, and IL-4 production was significantly increased compared with cells stimulated with the negative control media and mCII\textsubscript{707–721} (Figure 5A). IL-2 and IL-10 production was undetectable (data not shown). Interestingly, TNF-\alpha production was significantly reduced upon mCII\textsubscript{707–721}–specific NKT cell activation (Figure 5B).

The kinetics of cytokine production resembled those of conventional T cells, i.e., peaking at 72 hours after restimulation in vitro and showing no early burst of cytokines (data not shown).

To investigate whether NKT cells are the direct source of cytokine production, we treated WT B10.Q mice with mCII\textsubscript{707–721} plus CFA in vivo. As control groups, mice were treated with mCII\textsubscript{707–721} plus CFA; an additional group received only vehicle (CFA), and finally a group of mice did not receive any treatment (naive). Splenocytes were dissected and single-cell suspensions treated in vitro with mCII\textsubscript{707–721} for 48 hours. Cells were then stained for intracellular cytokines and analyzed using FACS. As shown in Figure 6, mCII\textsubscript{707–721}-specific NKT cells are capable of intracellular production of TGF-\beta1, IFN-\gamma, IL-4, and TNF-\alpha. However, the levels of TNF-\alpha intracellularly (Figure 5B) exhibited a trend similar to the levels seen as assessed by ELISA (Figure 7D), i.e., reduction in the in vivo mCII\textsubscript{707–721}-treated group compared with controls.

\textbf{Cytokine production of mCII\textsubscript{707–721}-specific NKT cells is CD1d dependent.} Splenocytes from similarly treated B10.Q WT and CD1d\(^{–}\) mice were analyzed for the production of cytokines in response to mCII\textsubscript{707–721}. As depicted in Figure 7, A–C, only WT mice that...
were treated in vivo with mCII<sub>707-721</sub> showed significant capacity to produce TGF-β1, IFN-γ, and IL-4, as measured by ELISA. Moreover, mCII<sub>707-721</sub>-specific TGF-β1, IFN-γ, and IL-4 production was significantly reduced in CD1d<sup>−/−</sup> compared with WT mice, though interestingly, a significantly lower TNF-α production was observed (Figure 7D).

mCII<sub>707-721</sub>-specific NKT suppression of T cells is Fas-FasL dependent. In vitro suppression of T cells was investigated by activating syngeneic splenocytes with plate-bound anti-CD3 (here referred to as responder cells), which were then cocultured with the mCII<sub>707-721</sub>-specific NKT cell line. Responder cells proliferated vigorously as indicated by a high proliferation index (Figure 8). However, mCII<sub>707-721</sub>-specific cells significantly inhibited proliferation of anti-CD3-stimulated responder T cells, compared with the control conventional MOG<sub>79-96</sub> T cell line. Since the mCII<sub>707-721</sub>-specific NKT cells produced TGF-β1, IFN-γ, and IL-4 upon activation, we assessed the role of these cytokines in T cell suppression. To block these cytokines, neutralizing antibodies were added to mCII<sub>707-721</sub>-specific NKT cell activation engages TCR and requires costimulation. (A) Activation of mCII<sub>707-721</sub>-specific NKT cells operated through TCR signaling. Percentage of phosphorylated ZAP70-positive cells in NK1.1<sup>+</sup> T cells by FACS shows a significant increase over non-stimulated cells upon mCII<sub>707-721</sub> stimulation in vitro, with abrogation by blocking anti-CD1 (1B1, 10 μg/ml). (B) Costimulatory signals for proliferation after in vitro stimulation of LNCs from mCII<sub>707-721</sub>-immunized mice. Blocking with antibodies to costimulatory proteins significantly reduced proliferation. Data are mean ± SD, n = 4. **<i>P</i> ≤ 0.01; ***<i>P</i> ≤ 0.001.

Figure 4
mCII<sub>707-721</sub>-specific NKT cell activation engages TCR and requires costimulation. (A) Activation of mCII<sub>707-721</sub>-specific NKT cells operated through TCR signaling. Percentage of phosphorylated ZAP70-positive cells in NK1.1<sup>+</sup> T cells by FACS shows a significant increase over non-stimulated cells upon mCII<sub>707-721</sub> stimulation in vitro, with abrogation by blocking anti-CD1 (1B1, 10 μg/ml). (B) Costimulatory signals for proliferation after in vitro stimulation of LNCs from mCII<sub>707-721</sub>-immunized mice. Blocking with antibodies to costimulatory proteins significantly reduced proliferation. Data are mean ± SD, n = 4. **<i>P</i> ≤ 0.01; ***<i>P</i> ≤ 0.001.

Figure 5
mCII<sub>707-721</sub>-specific NKT cells produce cytokines. (A) mCII<sub>707-721</sub>-specific cytokine production in primary LNCs from mCII<sub>707-721</sub>-immunized WT mice measured by ELISA. Significantly elevated levels of TGF-β1, IFN-γ, and IL-4 (<i>n</i> = 3) in response to mCII<sub>707-721</sub> are seen, but (B) TNF-α levels were lowered. Data are mean ± SD, n = 6. *<i>P</i> ≤ 0.05; **<i>P</i> ≤ 0.01.
specific NKT cell cultures prior to coculture with anti-CD3–activated splenocytes. The neutralizing antibodies did not affect the significant decrease in responder T cell proliferation (Figure 8A), indicating that the suppressive effect of mCII\textsubscript{707–721}–specific NKT cells was independent of their cytokine production.

Since these soluble cytokines seem to exert no major effects in the context of mCII\textsubscript{707–721}–specific NKT cell suppression, we investigated to determine whether cell-cell contact was required. As depicted in Figure 8B, the suppressive function of the mCII\textsubscript{707–721}–specific NKT cells was dependent on cell-cell contact, since separating direct cell-cell interaction using a Transwell system abrogated their suppressive effect.

We investigated whether mCII\textsubscript{707–721}–specific NKT cells could induce apoptosis, as measured by annexin V staining of anti-CD3–activated responder cells. Upon coculture, mCII\textsubscript{707–721}–specific NKT cells significantly induced apoptosis of anti-CD3–activated responder cells (Figure 8C). As Fas-FasL is one of most common mechanisms for induction of apoptosis (15), we used FasL-blocking antibody to determine whether Fas-FasL is involved in apoptosis induction. Cell death induced by mCII\textsubscript{707–721}–specific NKT cells was Fas-FasL dependent, since adding FasL-blocking antibody to the cultures inhibited apoptosis (Figure 8C). To confirm this result, we purified mCII\textsubscript{707–721}–specific NKT cells from mCII\textsubscript{707–721}–immunized mice, then cocultured with mCII\textsubscript{707–721}–specific NKT cells from lpr or WT mice, and cell death quantified as 7-aminoactinomycin D–positive (7AAD-positive) cells by FACS. Figure 8D shows that the cytotoxic function of mCII\textsubscript{707–721}–specific NKT cells was Fas-FasL dependent.

IL-17A is not required for mCII\textsubscript{707–721}–specific NKT function. NKT cells are reported to produce IL-17 (16), so we stained splenocytes from mCII\textsubscript{707–721}–immunized mice, then stained for IL-17A. Upon immunization with mCII\textsubscript{707–721}, 37.3% of NKT cells produced IL-17A (Figure 9A). However, IL-17A did not appear to play a role in the suppressive
function of these cells. CFSE-labeled CD4+ T cells were cocultured with mCII\textsubscript{707–721}–specific NKT cells in an anti-CD3–coated plate, with or without neutralizing IL-17A antibody. After 48 hours, cell death was analyzed with 7AAD staining on gated CD4+ T cells. The percentage of 7AAD+ T cells was similar with or without IL-17A blocking, suggesting that IL-17A was not involved in mCII\textsubscript{707–721}–specific NKT cells' suppressive function (Figure 9B).

mCII\textsubscript{707–721}–specific NKT cells prevent Th1- and Th2-mediated responses. The in vivo suppressive capacity of mCII\textsubscript{707–721}–specific NKT cells was investigated using B10.Q mice vaccinated with mCII\textsubscript{707–721} or the negative control peptide mCI\textsubscript{707–721}, prior to induction of a delayed-type hypersensitivity (DTH) reaction. Mice vaccinated with mCI\textsubscript{707–721} developed significantly less inflammation than the control group (Figure 10, A and B). Moreover, the antiinflammatory effect of mCI\textsubscript{707–721} vaccination was dependent upon activation of CD1d-restricted mCI\textsubscript{707–721}–specific NKT cells, since lack of this population in CD1d\textsuperscript{−/−} mice resulted in significant abrogation of the antiinflammatory effect (Figure 10C). These results support the in vitro findings that CD1d-restricted mCII\textsubscript{707–721}–specific NKT cells suppress T cell activation and hence inhibit Th1-polarized DTH inflammation.

We next studied the effect of CD1d-restricted mCII\textsubscript{707–721} NKT cells on the Th2-mediated immune response. Mice were vaccinated as above and then to provoke a Th2 response were injected with OVA emulsified in alum. Interestingly, a downregulated Th2 response was observed in the mCII\textsubscript{707–721}–vaccinated group compared with control. A significant decrease in IL-4, IL-5, and IL-13 production was detected in bronchoalveolar lavage fluid (BALF) after OVA rechallenge in WT mice. Moreover, we observed that vaccination with mCII\textsubscript{707–721} repressed IgE production in BALF of WT mice. Furthermore, mCII\textsubscript{707–721}–specific NKT cell activation and the antiinflammatory effects of such activation were determined to be CD1d restricted, as CD1d\textsuperscript{−/−} mice lacked such capacity (Figure 11). Taken together, the observations show that the mCII\textsubscript{707–721}–specific CD1d-restricted NKT cells are involved in immune regulation of both Th1-mediated cellular responses and Th2-mediated humoral immune responses.
Activation of mCII<sub>707–721</sub>–specific NKT cells ameliorates CIA. The mCII<sub>707–721</sub> peptide is a major epitope in mouse collagen type II, so mCII<sub>707–721</sub>–specific cells might be crucial for regulating CIA. To test our hypothesis, B10.Q mice were vaccinated with either mCII<sub>707–721</sub> or control peptide 10 days prior to CIA induction. We found that mCII<sub>707–721</sub> vaccination ameliorated the severity of arthritis compared with the control (Figure 12A). The incidence in the mCII<sub>707–721</sub>–treated group was 74% compared with 93% in the control group. Both clinical signs of disease and histopathological studies revealed that mCII<sub>707–721</sub>–vaccinated mice had less joint inflammation than control peptide–vaccinated mice (Figure 12B). Prevention of inflammation in these mice was associated with significantly reduced numbers of CD4<sup>+</sup> T cells in synovial infiltration (mean ± SD; 12 ± 3 vs. 37 ± 7, n = 10, P ≤ 0.05) and reduced production of IFN-γ, IL-4, and TNF-α (Figure 12, C and D), as determined by joint tissue immunohistochemistry staining. These results support the possibility of controlling arthritis by activation of mCII<sub>707–721</sub>–specific CD1d-restricted NKT cells.

Vaccination with mCII<sub>707–721</sub> ameliorates EAE. The DTH and OVA sensitization experiments indicated that the suppressive quality of the mCII<sub>707–721</sub>–specific NKT cells was not antigen or tissue specific. Therefore, we evaluated the effect of vaccination with mCII<sub>707–721</sub> on EAE, a well-described Th1-mediated autoimmune disease affecting the CNS and a widely used experimental model for MS. EAE was
induced in B10.Q WT mice vaccinated with mCII\textsubscript{707–721} by immunization with MOG\textsubscript{79–96}. EAE progression was significantly suppressed in mCII\textsubscript{707–721}−vaccinated mice, resulting in lower mean clinical scores compared with the control group vaccinated with mCII\textsubscript{707–721} (Figure 13A). Although no differences were seen between the 2 groups in disease incidence (both groups had 89% disease), affected mice in the mCII\textsubscript{707–721}−vaccinated group recovered entirely, while control group mice exhibited lingering symptoms up to 1 month after disease induction. In support, the CNS of mCII\textsubscript{707–721}−vaccinated mice showed significantly less demyelination than control mice (Figure 13, B and C), which correlated well with the clinical findings and the general inflammation observed in the CNS (data not shown).

These data strongly support the capacity of mCII\textsubscript{707–721}−specific NKT cells to suppress a variety of inflammatory conditions, not limited to the Th1/Th2 paradigm nor restricted to collagen-tissue specificity.

**Discussion**

CD1d-restricted NKT cells exert profound and diverse regulatory effects in disease. They have deservedly earned the distinction of being the innate and adaptive immune systems, showing capacity for antiinflammation activity.

However, despite almost 2 decades of study, research on natural ligands of NKT cells has not resulted in the identification of any self-peptide ligands that could activate NKT cells. Many experimental systems and clinical models have focused on exogenous glycolipids (such as αGalCer) presented by CD1d and highly potent in activation of NKT cells. Although this approach has been important to the characterization of NKT cells’ properties and functions, the biology of endogenous antigen–presentation by CD1d and activation of CD1d-dependent NKT cells remains largely unknown. The discovery of endogenous ligands, including self-peptide ligands that bind to CD1d, can shed light on the innate physiological functions of CD1d-dependent NKT cells. Our results support the idea that self-peptide reactivity could be considered part of the intrinsic immune function of NKT cells to maintain tolerance to self in cases of tissue inflammation.

We show here for what we believe is the first time a population of CD4\textsuperscript{+}TCR\textalphaβ\textsuperscript{+} NKT cells with specificity for a self-tissue peptide that binds to CD1d. These CD1d-restricted mCII\textsubscript{707–721}–specific NKT cells constitute an important component of the peripheral tolerance maintenance mechanism in that they protect against tissue-specific autoimmunity. Although mCII\textsubscript{707–721}–specific NKT cells are part of the peripheral pool of immune cells, they require activation via CD1d–self-antigen–TCR signaling to become competent immune regulators. Significantly, they suppress a range of inflammatory and autoimmune conditions.

Although CD1d has a large hydrophobic groove more suitable for the binding of glycolipids, studies have reinforced the notion that peptides can bind (9). How the mCII\textsubscript{707–721} structurally fits into this groove is not clear. Other studies have shown that a CD1d-restricted peptide can elicit a cellular immune response in vivo (17), leading the researchers to opine that intracellular murine CD1d might normally be associated with peptides and therefore peptide presentation may be a true physiologic function of murine CD1d (8).

Here, we determined that mCII\textsubscript{707–721} does bind to CD1d and has the capacity to be presented to NKT cells. In support of this observation, CD1d presentation of mCII\textsubscript{707–721} to NKT cells upregulated phosphorylated ZAP-70, a tyrosine kinase that is crucial for productive stimulation of T cells through TCR signaling after activation of TCR/CD3 (18). We found that mCII\textsubscript{707–721}–specific NKT cells required costimulatory signals via B7.1/CD28 and CD40/CD40L pathways, supporting earlier studies on NKT cell activation requirements (19). The limited usage of V\textalpha and the V\beta chain diversity in the TCR of NKT cells, combined with a nonpolymorphic CD1d, suggest that the CD1d-restricted NKT cells may recognize a single or conserved set of antigens. Although some skewing toward V\textalpha4-\textalpha518, V\beta8.2 TCR usage was observed in mCII\textsubscript{707–721}–reactive NKT cells, this response is far from being a clonal response. Instead, the mCII\textsubscript{707–721} peptide activates a heterogeneous NKT cell population, as identified with a panel of V\beta TCR that showed it to be polyclonal and diverse. Nonetheless, variations in CD1d-restricted antigen pre-
sentation could arise from differences in expression, trafficking, processing, or loading of antigens (20).

We show that antigen-specific production of TGF-β1, IFN-γ, and IL-4 by CD1d-restricted mCII707–721–specific NKT cells resembles conventional T cells in the adaptive immune response. IL-17 production is associated with CD4+ ‘NK1.1’ invariant NKT cells (21) and is suggested to be pathogenic in several autoimmune diseases (22, 23). However, the pathogenic role of IL-17–producing Th17 cells has also received challenges (24), and the regulatory role for regulatory T cells has been shown to be independent of their influence on Th17 (25). Still, a potential influence for IL-17–producing NKT cells on the regulation of autoimmune diseases could be plausible. In the present study, we found CD1d-restricted mCII707–721–specific NKT cells produced IL-17A, but it was not involved in their suppressive function. In support, CD1d-dependent NKT cells’ regulation of arthritis is IL-17 independent (26).

Of note, the mCII707–721–specific NKT cells suppressed anti-CD3–activated T cells in a cell-cell contact–dependent manner. Cytokines including IL-4, IL-10, IL-13, and TGF-β1 have been suggested as important mediators in NKT cell suppressive functions (10, 27), so suppression mediated by a final release of cytokines upon cell interaction could be feasible. However, we detected no role for cytokine production by the mCII707–721–specific NKT cells in their in vitro suppressive effects on T cells since neutralizing these cytokines did not prevent the suppressive effects. In our previous work, the regulatory role of CD1d-restricted NKT cells in EAE was associated with CD4+ ‘NK1.1’ TGF-β1–producing regulatory T cells (21). In this study, we found that mCII707–721–specific NKT cells produced Th2 cytokines, and lower IgE production. Thus, CD1d-deficient mice lack this regulatory role for TGF-β1 expression, which may prevent Th2 cytokine response, and lower IgE production. Therefore, CD1d-restricted mCII707–721–specific NKT cells regulate both the Th1-mediated DTH reaction and the Th2 cytokine response, and lower IgE production. Thus, CD1d-restricted mCII707–721–specific NKT cells regulate both the Th1-driven DTH and a Th2-driven antigen-induced airway inflammation, we found, unexpectedly, that mCII707–721–specific NKT cell activation reduced both the Th1-mediated DTH reaction and the Th2 cytokine response, and lower IgE production. Thus, CD1d-restricted mCII707–721–specific NKT cells regulate both the Th1-driven and Th2-driven states and diminish humoral Th2 responses. Vaccination of mice with mCII707–721 prior to CIA induction significantly ameliorated arthritis, as evidenced by lower clinical and histopathological scores. Significantly lower joint inflammation was accompanied by reduction of all tested cytokines but no alteration of the Th1/Th2 profile of cells infiltrating the joints. In accordance, we reported that general activation of CD1d-dependent NKT cells in WT mice prevents arthritis in different models of joint inflammation without induction of a cytokine shift, while CD1d-deficient mice lacking this population are defective in suppressing joint inflammation (26).

Given that NKT cells can either suppress or activate immunity, their therapeutic use necessitates a thorough understanding of their intrinsic biology. Limitations in this regard have been revealed by αGalCer or its synthetic analogs, which despite some success in early clinical trials have on the whole been less promising (33–35).
This could perhaps be attributed to the fact that these substances are not found in self tissues and hence fail to replicate the natural course of NKT cell activation in disease. mCII\textsubscript{707–721}–specific NKT cells may be able to overcome these obstacles and achieve fuller potential as treatments for autoimmune diseases and cancer.

A natural question emerging from the currently reported results regards the tissue specificity of the autoimmune suppressive function of activated mCII\textsubscript{707–721}–specific NKT cells. Therefore, we investigated its effect on inflammation of the CNS via an EAE model. Mice vaccinated with mCII\textsubscript{707–721} prior to induction of EAE showed significantly reduced clinical symptoms and demyelination compared with the control group. We surmise that even if the population of NKT cells is self-collagen specific, their suppressive function is not necessarily restricted to arthritis and cartilage tissue but can operate in other tissue-specific autoimmune and inflammatory diseases.

The question of how the mCII\textsubscript{707–721}–specific NKT cells regulate autoimmunity in multiple models, specifically with regard to their effects on autoreactive T cells, is of interest. This ability is consistent with properties of conventional regulatory T cells (36) as well as antigen-specific regulatory T cells generated in inflamed tissues such as those of the CNS (37). Antigen-nonspecific responses of NKT cells, or bystander suppression, could involve different mechanisms. One of the widely discussed ideas is the ability of NKT cells, even in a non-self-antigen–specific manner (e.g., αGalCer or its analogs), to regulate multiple autoimmune diseases via their capacity to produce an early burst of cytokines and deviation of the Th1/Th2 axis (reviewed in ref. 38). However, the suppressive function of mCII\textsubscript{707–721}–specific NKT cells differs since we detected a cell-cell–mediated mechanism. This is consistent with an earlier report (39) in which it was shown that NKT cells inhibited the differentiation of naive autoreactive T cells into effector cells. Here we have shown that the suppressive function of mCII\textsubscript{707–721}–specific NKT cells is to a great extent via the engagement of the Fas/FasL pathway. This supports a model whereby activation of autoreactive NKT cells could inhibit further effector T cell activation via a direct cytolytic effect.

Alternatively, the suppressive function of the NKT cells may also be mediated by differential engagement of or effects on tissue-specific APCs such as macrophages (40), mature/immature DCs (41), and/or B-1 B cells (42). The ability of NKT cells to modulate the function of APCs, as discussed in a recent review (6), may therefore play an important role in shaping the outcome of the immune response. Hence, differential engagement of APCs in different tissues, as the consequence of mCII\textsubscript{707–721}–specific NKT cell activation, may also serve a broad immune regulatory function.

The results of the present study support the contention that self-antigenic peptide-reactive NKT cells could be an important part of the peripheral tolerance maintenance mechanisms. Their physiological role may be to maintain tissue homeostasis through induction of cell-cell–mediated cytotoxic action on activated lymphocytes and to dampen inflammation regardless of the Th profile of its source. This is in contrast to the rapid release of cytokines traditionally assigned to the mode of action of NKT cells. Given that the joint is prone to wear and tear, the presence of this peptide-specific NKT cell population in the periphery could be part of the native physiology that might partly explain the relative resistance to mouse type II collagen–induced arthritis that is observed. As a general phenomenon
across a range of disease states, such cells could act rapidly toward any danger signal evoked by pathogens or inflammatory conditions. The effect would be to prevent chronicity of the inflammation and hence sustain immune balance and tissue homeostasis. Given the high degree of conservation of NKT cells among species, therapeutic immunization with an NKT-activating peptide may hold promise for clinical benefit in human inflammatory diseases.

Methods

Mice. Animals were kept and bred at the animal facilities of the University of Lund or the University of Copenhagen. All animal experiments were reviewed and approved by both of these universities’ institutional review boards, located in Lund and Copenhagen, respectively. All studies with mCII707–721 were performed on WT male C57BL/10.H-2q (B10.Q) mice (originated from Jan Klein, University of Tubingen, Tubingen, Germany). KO mice were backcrossed to this background to produce well-controlled genetic strains to avoid strain-specific effects that might influence the results.

Tcrab−/− or Tcrgδ−/− mice were the results of targeted germ-line mutations on 129 and backcrossed to C57BL/6J (The Jackson Laboratory). These were backcrossed to the B10.Q background for 6 generations. Subsequent intercrossing of mice heterozygous or homozygous for mutated TCR yielded homozygous (αβ or γδ T cell deficient) and heterozygous (normal T cell phenotype) littermates. T cell phenotypes were determined by flow cytometry analysis of blood cells using mAbs against either TCRαβ (H57-597-FITC; Pharmingen) or TCRγδ (GL3-PE; Pharmingen).

CD4- and CD8-deficient founders backcrossed to C57BL/6J mice (43) were backcrossed to B10.Q for 10 generations, and positive offspring were intercrossed twice to yield littermates homozygous for the WT gene, homozygous for the disrupted gene, or heterozygous, as determined by flow cytometry with anti-CD4 (Gk1.5-PE; BD) and anti-CD8 (53-6.7-biotin: BD Biosciences — Pharmingen) mAbs.

MHC II–deficient and TAP-deficient mice (originally received from Diana Mathis, Institute of Genetics and Molecular and Cellular Biology, University of Strasbourg, Strasbourg, France) were backcrossed to C57BL/6J for 20 generations and then backcrossed to B10.Q for 3 generations. The offspring deficient in targeted genes were intercrossed once to yield homozygous KO or WT, and heterozygous littermates. lpr mice of H-2q haplotype were received from Saleh Ibrahim (Department of Immunology, Rostock University).
CD1d<sup>Δ</sup> mice (44) on a C57BL/6 background (over 20-generation backcross) were backcrossed to B10.Q for more than 5 generations. Animals were sex and age matched (8–20 weeks) for all experiments.

Media and reagents. All cells were grown in cDMEM, DMEM with Gluta-max-1 (GIBCO BRL, Life Technologies) supplemented with 10 mM HEPES buffer, 10% heat-inactivated FCS, 0.16 mM/ml penicillin, 0.03 mM/ml streptomycin, and 50 μM 2-mercaptoethanol. Growth was in a humidified incubator at 37°C with 7.5% CO<sub>2</sub>. All reagents were from Sigma-Aldrich, unless otherwise noted.

All peptides were from Schäfer. Mouse collagen type II peptide mCII<sub>707–721</sub> (PPGANGNPAGPPG) and negative control mouse collagen type I peptide mCII<sub>707–721</sub> (PPGPGNAGPGGPG) differ at positions 710 (proline to alanine), 711 (serine to asparagine), 714 (alanine to proline), and 717 (proline to alanine). Biotinylated peptides were used in binding assays.

Proliferation assays. mCII<sub>707–721</sub>-specific and myelin-specific T cell lines. B10.Q male mice, aged 8–14 weeks, were immunized in the hind paws and tail base with 200 μl of a 1:1 emulsion of 100 μg of either mCII<sub>707–721</sub> or mCII<sub>707–721</sub> in PBS and CFA containing Mycobacterium tuberculosis H37Ra (Difco). For some experiments, B10.Q and a panel of KO mice on a B10.Q background were used. Draining LNs or spleens were collected on days 10–13 after immunization, and a single-cell suspension was prepared in PBS by passing through a sieve. Cells were washed and resuspended in cDMEM.

To measure T cell proliferation in response to mCII<sub>707–721</sub>, cells were cultured in quadruplicate in round-bottom 96-well plates (Nunc) at 5 × 10<sup>4</sup> cells/well and stimulated with 100 μg/ml mCII<sub>707–721</sub> or mCII<sub>707–721</sub> and/or media only or PPD for 4 days in the presence of 1 μCi <sup>3</sup>H-thymidine per well in the final 12 hours of culture. <sup>3</sup>H-thymidine incorporation was measured in a beta scintillation counter (Matrix 96 Direct Beta Counter; Packard). mCII<sub>707–721</sub>-specific and myelin-specific T cell lines were established as previously published (37).

Blocking antibodies were anti-CD40L (20 μg/ml); anti-CTLA-4 (5 μg/ml); anti-B7.1 Fab-fragment (20 μg/ml); anti-B7.2 Fab-fragment (20 μg/ml); CTLA-4-lg fusion protein (10 μg/ml); and isotype controls (10 μg/ml).

CD1d binding assay. Maxisorp plates (Nunc) were coated with recombinant soluble dimeric mouse CD1d-lg dimer (BD Biosciences – Pharmingen) at 4 μg/well. Biotin-labeled mCII<sub>707–721</sub> (0.6 μg/ml) was added and incubated at 37°C for 2 hours. After washing, plates were incubated with europium-avidin followed by enhancement buffer according to the manufacturer’s instructions and fluorescence intensity measured on a fluorometer (Wallac Oy EG & G). Biotin-labeled anti-CD1d was a positive control, and biotin-labeled mCII<sub>707-721</sub> was the negative control. Nonlabeled mCII<sub>707-721</sub> in excess or varying amounts of αGalCer was used to assess competitive binding.

Ratio of binding for mCII<sub>707-721</sub> to CD1d was calculated by dividing the fluorometer OD value by the OD value for the positive control, for different mCII<sub>707-721</sub> concentrations.

FACS. Standard FACS procedures and analysis were followed (45). Antibodies were from BD Biosciences – Pharmingen unless stated otherwise and used at 1–5 μg/ml, and were PE-Cy5-anti-CD4 (GK1.5), FITC-anti-TCR-β (H57-597), mouse Vβ TCR screening panel (cat. no. 557004), PE-anti-IL-4 (11B11), PE-anti–IFN-γ (XM1g1.2), biotin–anti-TGF-β1 (A75-3.1), streptavidin-PE (cat. no. 554061), APC–anti-TNF-α (MP6-XT22), PE–anti–IL-17A (TC11-18H10), PE–anti–CD49b (DX5), FITC– and PE–anti-NK1.1 (Pk136).

Real-time PCR. To detect TCR V<sub>α</sub>14-J<sub>18</sub> expression, total RNA was extracted from the NKT cell line using a QIAGEN kit, reverse transcribed, and amplified and quantified by SYBR Green detection. Primers were V<sub>α</sub>14-J<sub>18</sub> primer, forward: 5′-TGGGAGATCTCAGCAGACTTGG-3′; reverse: 5′-CAGGTATGACAATCAGCTGAGTCC-3′; and constant region housekeeping forward: 5′-CCTCTGCTGTTTACCCGACTT-3′; reverse: 5′-TGGCGTTGTCTTGGAGAAGG-3′. PCR was 10 minutes at 95°C,
40 cycles of 95°C for 10 seconds, decreasing by 20°C/s to 60°C, and 60°C for 60 seconds. The melting curve was 20°C/s to 95°C, cooling at 20°C/s to 60°C, and heating at 0.2°C/s to 95°C with fluorescence collection at 0.2°C intervals.

ELISA. ELISAs were performed according to the manufacturers’ instructions. ELISA kits were used for IL-13 and IgE (BD Biosciences) and IL-4, IL-5, IFN-γ, TNF-α, and TGF-β1 (R&D Systems).

Purification of NKT cells. Single-cell suspensions were washed and resuspended in 2% FCS in PBS and analyzed or purified using FACS Aria. CD4+ T cells and CD4+ NK1.1+ NKT cells were purified with FACS Aria.

In vitro suppressive assays. Splenocytes from B10.Q mice or a conventional B10.Q MOG79–96–specific T cell line were activated by culturing on plate-bound anti-CD3 at 5 μg/ml and used as responder cells by coculturing at a 1:1 ratio with mCII707–721–specific NKT cells. Proliferation was measured by 3H-thymidine incorporation. When splenocytes were responder cells, the B10.Q MOG79–96–specific cells were used as controls. Blocking antibodies were added to the NKT cells for 30 minutes before coculture.

A Transwell insert of 0.4 μm (Falcon) was used for separation of mCII707–721–specific NKT and responder cells. Soluble molecules, but not cells, could pass through the insert.

For annexin V staining, cells were incubated in binding buffer with 5 μl of annexin V-PE at room temperature for 5 minutes in the dark and washed. Positive cells were monitored by FACS. Some experiments used 7AAD instead of annexin V.

Vaccination with mCII707–721. OVA challenge, induction of DTH, CIA, and EAE. Mice were immunized in the flank with 100 μg mCII707–721 emulsified in CFA (Difco). At 13 days after immunization, mice received an injection of 20 μg CII in 0.05 M acetic acid in the outer right ear and just acetic acid in the outer left ear as a negative control. Additional control groups vaccinated with mCII707–721 and/or CFA (vehicle).

DTH response. B10.Q mice and CD1d KO mice received an intradermal injection at the tail base with 100 μg of CII emulsified in CFA (Difco). At 48 hours, the DTH response was measured using a Kröplin caliper and calculated as the difference in swelling (thickness) of the right and left outer ears. Histopathological evaluation of inflammation in the outer ears was also performed.

OVA sensitization and BALF. B10.Q and CD1d KO mice received an intraperitoneal injection on days 0 and 4 with 50 μg OVA with 5 μg alun. At day 13, mice were challenged with 50 μg of OVA intranasally. Mice were sacrificed 24 hours later and BALF collected for ELISA.

Induction and evaluation of CIA. Male B10.Q mice, 8–12 weeks of age, were immunized intradermally at the tail base with 100 μg rat collagen emulsified in CFA (Difco) and boosted at day 35 with 50 μg rat CII emulsified in incomplete Freund adjuvant (Difco). Clinical scoring was performed as described (46). For histopathological and immunohistochemical analyses, ankle joints were removed on day 31 after collagen immunization (47). For immunohistochemistry, cryosections were stained with biotinylated antibodies to IFN-γ (Ani8), IL-4 (BVD6-24G2), TNF-α (MIP6-XT3), and CD4 (L3T4). ExtrAvidin-peroxidase and diaminobenzidine (50 mg/ml; Saveen Biotech) were used for detection, with hematoxylin staining. For cytokines, 0.1% saponin was used at all steps after fixation. Stained infiltrating cells were counted in the synovium by randomly selecting 6 areas per section and calculating the mean of positive cells per animal.

Induction and evaluation of EAE. B10.Q mice were immunized for induction of EAE, as described (37).

For demyelination, the lumbar part of spinal cords from 3–4 mice per group was collected at day 40 after MOG immunization and fixed in 4% formaldehyde and then as described (48). Results are the mean from 4 sections.

Statistics. Statistical evaluation was performed using StatView software. The Mann-Whitney test was used to analyze differences in clinical scores; otherwise a Student’s unpaired t-test was used.

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