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Clinical phenotype associations with various types of anti-dsDNA antibodies in patients with recent onset of rheumatic symptoms. Results from a multicentre observational study

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ABSTRACT

Despite anti-dsDNA antibodies constitute a wide range of specificities, they are considered as the hallmark for systemic lupus erythematosus (SLE).

Objective: To identify clinical phenotypes associated with anti-dsDNA antibodies, independently of any clinical diagnoses.

Methods: Patients with recent onset of any rheumatic symptoms were screened for antinuclear antibodies (ANA). All ANA-positive and matching ANA-negative patients were examined, and their clinical phenotypes were registered, using a systematic chart formulated after consensus between the participating centres. All patients were tested for different anti-dsDNA antibody specificities with assays habitually used in each participating laboratory. Crithidia Luciliae Immuno Fluorescence Test (CLIFT) was performed three times (with two different commercial kits); solid and solution phase ELISA were performed four times. Associations between clinical phenotypes and results of anti-dsDNA assays were evaluated by linear regression analysis (LRA) and principal component analysis (PCA).

Results: Totally, 292 ANA-positive and 292 matching ANA-negative patients were included in the study. A full dataset for statistical analysis was obtained in 547 patients. Anti-dsDNA antibodies were most frequently detected by ELISA. LRA showed that overall positivity of anti-dsDNA antibodies was associated with proteinuria and pleuritis. Alopecia was significantly associated only with CLIFT-positivity. Besides confirming the same findings, PCA showed that combined positivity of CLIFT and ELISA was also associated with lymphopenia.

Conclusions: Our results show that different anti-dsDNA antibody specificities are associated with nephropathy, pleuritis, alopecia and lymphopenia, regardless of the diagnosis. It may challenge the importance of anti-dsDNA antibodies as a diagnostic hallmark for SLE.

KEY MESSAGES

- In patients with recent onset of rheumatic symptoms, the assessment of anti-dsDNA antibodies with different techniques results in a considerable discrepancy of outcomes and of correlations to various clinical and biochemical manifestations.
- Anti-dsDNA antibodies are associated with presence of proteinuria, regardless of clinical diagnosis, outcome of ANA screening and laboratory method used for the assessment of anti-dsDNA antibodies.
- In distinct subgroups of patients, anti-dsDNA antibodies are also variously associated with presence of other clinical manifestations, such as haematuria, leukopenia, pleuritis and alopecia.

Systemic lupus erythematosus (SLE) is a systemic autoimmune disease with unknown aetiology. Whether SLE represents one disease entity or is a continuous overlap of aetiologically unrelated organ manifestations is not established.

This is particularly challenging when attempting to determine biomarkers for SLE. Anti-double-stranded DNA (dsDNA) antibodies are regarded as fairly specific for SLE.^{1–3} B-cell-mediated and T-cell-mediated autoimmunity to the individual components of nucleosomes are considered important in establishing a diagnosis,^{4–7} but the pathogenic and diagnostic roles played by anti-dsDNA and other antibodies are still debated.^{7,8}

Antibodies to dsDNA may have a direct pathogenic effect in lupus nephritis,⁹ lupus dermatitis^{10–11} and possibly also in certain

aspects of cerebral lupus.¹² How anti-dsDNA antibodies relate to the rest of the clinical components of current classification criteria^{5,8} remains to be determined.

When emphasising anti-dsDNA antibodies as a central biomarker in SLE, it is important to perceive that these antibodies basically are not representing a homogenous antibody population.^{13–16} Growing insight into the different possible mechanisms of production of antibodies specifically binding to dsDNA^{17–24} challenges the notion of a specific relationship between all anti-dsDNA antibodies per se with SLE. Which and how anti-dsDNA antibodies are pathogenic has also been questioned.^{9,25–38}

We aim to explore in the present investigation whether the positivity of anti-dsDNA antibodies is a biomarker indicating presence of defined clinical phenotypes, such as, for example, arthropathy or nephropathy or serositis, rather than a defined diagnosis, such as SLE.

The current literature is mainly composed of investigations of patients with established SLE and other defined and classified diagnoses. Studies concerning the association of anti-dsDNA antibodies with clinical manifestations in unselected patients with early onset of rheumatic symptoms are scarce.

The main purpose of this study is to correlate the presence in serum of anti-dsDNA antibodies with individual clinical manifestations and laboratory variables. We intend to use an unbiased approach that mirrors an ordinary clinical setting, where the physician is challenged to make the right diagnosis in patients with newly developed rheumatic manifestations, based upon clinical signs and symptoms, with the support of various diagnostic procedures, including laboratory tests. This approach also allows us to perform a comparison between serum levels of anti-dsDNA antibodies obtained in different laboratories with different methods.

METHODS AND PATIENTS

Patients

Consecutive patients with recent onset of suspected rheumatic disorder, referred for the first time to the participating rheumatologic units (Rigshospitalet in Copenhagen, Denmark; University Hospital in Tromsø, Norway; University Hospital in Lund, Sweden) were recruited to the study between February 2003 and December 2007.

Exclusion criteria were: established autoimmune disease, treatment with any biological drug, corticosteroids (equivalent Prednisolon >20 mg/day), immune-modulating, immunosuppressive or cytostatic drugs. Patients who previously had been examined by a rheumatologist, and patients unable to fully collaborate in the study (not confident with the language, actual cognitive, speech, hearing or memory impairment) were also excluded.

The patients were examined by a rheumatologist who made an initial working clinical diagnosis, based on

anamnesis, symptoms, physical examination and laboratory test results.

All the patients were screened for antinuclear antibodies (ANA) by local testing. All the ANA-positive patients and the same amount (1:1 ratio) of randomly selected sex-matched and age-matched ANA-negative patients built together a nested cohort, which underwent further clinical and laboratory assessments. A systematic chart including clinical data and routine laboratory variables (see online supplementary table S1) was completed, ensuring a common trunk of data on which the clinical phenotypes of the patients could be characterised. Aliquots of serum samples from all patients were collected and sent to each centre for simultaneous parallel analysis of anti-dsDNA antibodies by several different assays, as described below. All clinical data were collected without knowledge of the results of the anti-dsDNA testing.

Ethics

All patients entered the study after giving informed written consent. The participating centres performed the study according to the approval from the local ethics committees.

Clinical phenotype description

Consensus as to the content of the clinical and laboratory dataset in the systematic chart (see online supplementary table S1) was obtained through a Delphi-like process, spanning over four meetings. A uniform definition of the final clinical dataset was assured by agreeing on the definitions of the various manifestations prior to the study (see online supplementary table S2). Manifestations were recorded as being absent ever, ongoing/active, previous/inactive or unknown and the date any manifestations first appeared was noted. In the present study, calculations were based on the presence ever of a manifestation.

Detection of ANA

In the various centres, respective current routine methodology was used for detection of ANA. In Copenhagen and Lund, screening for ANA was performed by indirect immunofluorescence (IIF) technique. In Copenhagen, HEp-2 cells (ImmunoConcepts, Sacramento, California, USA) and patient sera in dilution 1/160 were used together with an FITC-labelled antihuman IgG conjugate (DAKO Denmark A/S, Glostrup, Denmark). The diagnostic ANA titre was 1:160 as established by determination of the 95th percentile for negativity in healthy blood donors. In Lund, Hep-2 or Hep-20-10 cells (Euroimmun, Lübeck, Germany) and patient sera in dilution 1/400 were used which corresponded to detection of ANA with homogeneous pattern at 14 IU/mL (WHO reference serum 66/233). The diagnostic ANA titre was 1:400 as established by determination of the 96.5th percentile for negativity in healthy blood donors. In Tromsø, the detection of ANA was performed with the ELISA Vareliisa ReCombi ANA Screen (Pharmacia

Diagnostics, Freiburg, Germany). This is an indirect non-competitive enzyme immunoassay for the qualitative and semiquantitative determination of eight preselected ANA (dsDNA, Sm (B,B',D), ribo-nucleo protein (68 kDa, A, C), SS-A/Ro(52 and 60 kDa), SS-B/La, Scl-70, centromere and Jo-1) in serum or plasma. The assay was performed as recommended by the manufacturer, while validation and determination of cut-off values were performed in accordance with an internal and external quality assessment programme, with validation against both controls and other disease categories.

Detection of anti-dsDNA antibodies

Without knowledge of any clinical information, all the patients included in the nested cohort had IgG anti-dsDNA antibodies determined by different indirect immunofluorescence (IIF) tests and immunoassays available at the participating centres (table 1).

Immunofluorescent tests

Presence of anti-dsDNA antibodies was assessed in all the three participating laboratories by Crithidia Luciliae Immunofluorescence Test (CLIFT) with two different commercial kits, according to the manufacturers' instructions. By using a fluorescence microscope, the results were based on the fluorescence intensity and categorised as negative or positive.

Immunoassays

The determination of anti-dsDNA antibodies by the EliA test and the Varelisa were performed according to the manufacturer's instructions.

The solution phase anti-dsDNA ELISA (SPADE) is previously described in detail.¹³ It measures antibody binding to dsDNA in solution using biotinylated dsDNA.³⁹ Biotinylation of the pUC18 DNA (1 µg/µL H₂O) was carried out as recommended by the manufacturer (Pierce Chemical Company, Rockford, Illinois, USA). SPADE was performed by mixing 0.5 µg of pUC18 DNA (1 µg/µL H₂O) with serially diluted serum samples (solution phase step). After incubation for 30 min, the

mixtures were added to microtitre plates (Nunc MaxiSorp, Nunc, Denmark) coated overnight at 37°C with 50 µL streptavidin (5 µg/mL PBS). After incubation for 30 min, the plates were washed and incubated with horseradish peroxidase conjugated anti-human Fcγ antibodies (Sigma-Aldrich, St Louis, Missouri, USA) in phosphate buffered saline with tween 20. The reaction was developed by adding o-phenylenediamine dihydrochloride, and stopped by 1 M HCl. The reaction was read at 490 nm.

Statistical analysis

After finalising data retrieval, all data were registered in a common database accessible for all centres. Statistical analyses using SPSS statistics V.20.0 software (IBM) included contingency table analysis and binary logistic regression analysis. OR and corresponding 95% CIs were initially calculated as crude values using dichotomised anti-dsDNA results as the dependent variable and each dichotomised clinical manifestation registered in our database as explanatory variables. OR and corresponding 95% CI were then adjusted, using all the crude significant associations as covariates.

Principal component analysis (PCA) using R (<http://www.R-project.org>) included clinical phenotype data as well as positivity in the ANA test, any of the CLIFTs and any of the ELISAs. The PCA results were presented as biplots with variable vectors (arrows) indicating by direction which variables had the highest degree of covariation and influence, positive or negative, in discriminating the patients.

RESULTS

Altogether, 1073 patients were recruited from February 2003 to December 2007. Among these patients, 292 were found to be ANA positive by local testing. From the remaining ANA-negative patients, each centre in a 1:1 ratio randomly selected sex-matched and age-matched patients, reaching a total of 292 ANA-negative control patients. The resulting nested cohort of 584 patients (138 from

Table 1 Overview of anti-dsDNA tests used in the participating laboratories for analysis of serum aliquots from all patients

Name	Methodology	Antigen	Reference interval	Manufacturer	Centre
CLIFT 1	IIF	Crithidia luciliae kinetoplast	<titre 10	ImmunoConcept	CPH
CLIFT 2	IIF	Crithidia luciliae kinetoplast	<titre 10	ImmunoConcept	Tromsø
CLIFT 3	IIF	Crithidia luciliae kinetoplast	<titre 10	Euroimmune	Lund
EliA	Solid phase ELISA	Recombinant plasmid dsDNA	<10 IU/mL	Phadia	Tromsø
SPADE	Solution phase ELISA	Biotinylated plasmid dsDNA and biotinylated, S1 nucleated human dsDNA	<1 AU/mL	Inhouse	Tromsø
Varelisa 1	Solid phase ELISA	Recombinant plasmid dsDNA	<55 IU/mL	Phadia	Tromsø
Varelisa 2	Solid phase ELISA	Recombinant plasmid dsDNA	<35 IU/mL	Phadia	CPH

AU, arbitrary units; CLIFT, Crithidia Luciliae Immunofluorescence Test; CPH, Copenhagen; IIF: indirect immunofluorescence technique; IU, international units; SPADE, solution phase anti-dsDNA ELISA.

Copenhagen, 144 from Tromsø and 302 from Lund) was investigated concerning clinical phenotype at inclusion and anti-dsDNA antibodies assessed by different assays.

Clinical and biochemical manifestations in ANA positive and ANA-negative patients

A full serological dataset was obtained for 547 patients, 288 ANA positive (78.5% females, median age 51.8 years, range 15.4–83.7 years) and 259 ANA negative (83.4% females, median age 51.1 years, range 15.9–84.2 years). The remaining 37 patients, 4 ANA positive (75% females, median age 64 years, range 49–77 years) and 33 ANA negative (76% females, median age 59 years, range 22–81 years) withdrew from the study because of technical and logistic problems that hampered the collection of a full serological dataset.

The initial clinical diagnoses of the patients based on the conclusion of the examining rheumatologist are listed in [table 2](#).

The most prevalent American College of Rheumatology (ACR) classification criteria for SLE,^{5 40} and the most relevant remaining clinical and biochemical manifestations are listed in [table 3](#). Arthritis, photosensitivity, oral/nasal ulcers, haematuria, and proteinuria were the five most prevalent SLE specific manifestations. Malar rash was the only clinical variable significantly associated with the presence of ANA.

Arthralgia, morning joint stiffness, Raynaud's phenomenon, headache, and subjective xerostomia were the five most prevalent manifestations among those not included in ACR classification criteria. Raynaud's phenomenon, headache, puffy fingers and affective disorders were significantly more prevalent in the presence of ANA, whereas psoriasis was significantly more prevalent in ANA-negative patients.

ANA and anti-dsDNA antibody assays

The number of patients positive in the different anti-dsDNA assays performed ranged from 33 (6.1%) to 61 (11.1%). Among all 547 sera, a substantial difference between the centres was seen, being the number of CLIFT-positive results 33 (6%), 36 (6.6%) and 45 (8.2%) for Copenhagen, Tromsø and Lund, respectively. A total of 59 patients were positive by at least one CLIFT and 99 by any ELISA assessment, with low agreement of results ($k=0.34$).

Of the 288 ANA-positive sera, 39 (13.5%) were positive in any CLIFT, while 50 sera (17.4%) were positive in any of the immunoassays (Vareliba, SPADE or EliA). Of the 259 ANA-negative sera, 20 (7.7%) were positive by any CLIFT, and 49 (18.9%) were positive by any immunoassay. One hundred and twenty-four patients (68 ANA positive and 56 ANA negative) were positive at least by one assay, whereof 34 patients (21 ANA positive and 13 ANA negative) were positive by at least one CLIFT and one ELISA assessment.

In general, the CLIFT tests were least often positive. None of the single anti-dsDNA tests was associated with the presence of ANA, with the only exception of the CLIFT performed in Lund. Nonetheless, the positivity for any CLIFT was strongly associated with ANA positivity ([table 4](#)). According to our results, having a negative ANA did not rule out having a positive anti-dsDNA test.

Anti-dsDNA antibodies and clinical manifestations

The prevalence of clinical and laboratory manifestations in anti-dsDNA-positive patients are reported in the online supplementary table S3. Malar rash, cutaneous vasculitis, alopecia, leukopenia, lymphopenia, non-haemolytic anaemia, pleuritis, proteinuria and haematuria are more prevalent in patients with combined CLIFT and ELISA positivity (CLIFT+ ELISA+). Sixty-five patients resulted negative by CLIFT and positive by

Table 2 Clinical diagnoses formulated by the examining rheumatologists in the participating centre

Diagnoses	Nested cohort—547 patients							37 dropout patients	
	ANA pos (N=288)		ANA neg (N=259)		Rate ratio*	Total		Total	
	N	Per cent	n	Per cent		n	Per cent	n	Per cent
Systemic lupus erythematosus	57	19.8	6	2.3	8.61	63	11.5	2	5.4
Inflammatory connective tissue disease	44	15.3	12	4.6	3.33	56	10.2	0	0
Inflammatory joint disease	49	17	95	36.7	0.46	144	26.3	13	35.1
Systemic inflammatory disease	10	3.5	17	6.6	0.53	27	4.9	2	5.4
Arthralgia	45	15.6	32	12.35	1.26	77	14.1	6	16.2
Osteoarthritis	25	8.7	29	11.2	0.78	54	10	5	13.5
Soft-tissue rheumatism	8	2.8	25	9.65	0.29	33	6.0	3	8.1
Non-rheumatic disease	34	11.8	26	10	1.18	60	11	3	8.1
Dermatological disorder	10	3.5	7	2.7	1.30	16	2.9	2	5.4
Unspecified	7	2.4	10	3.9	0.61	17	3.1	1	2.7

*Ratio between prevalence of manifestation in ANA positive and ANA negative patients.
ANA, antinuclear antibodies.

Table 3 Clinical and biochemical manifestations recorded during the initial work-up of the included 547 patients. Listed are manifestations included in systemic lupus erythematosus (SLE) classification criteria and other most prevalent clinical manifestations

	All patients		ANA pos (N=288)		ANA neg (N=259)		Rate ratio*	p Value
	n	Per cent	N	Per cent	n	Per cent		
Manifestations among SLE classification criteria								
Peripheral arthritis	157	28.7	82	28.5	75	29	1.0	0.90
Photosensitivity	57	10.7	35	12.2	22	8.5	1.4	0.16
Oral/nasal ulcers	32	5.9	21	7.3	11	4.2	1.7	0.13
Haematuria	20	3.7	11	3.8	9	3.5	1.1	0.82
Proteinuria	17	3.1	9	3.1	8	3.1	1	0.98
Malar rash	15	2.7	12	4.2	3	1.2	3.5	0.03
Leukopenia	14	2.6	10	3.5	4	1.5	2.3	0.15
Alopecia	12	2.2	6	2.1	6	2.3	0.9	0.85
Lymphopenia	11	2	8	2.8	3	1.2	2.3	0.17
Discoid LE	8	1.5	2	0.7	6	2.3	0.3	0.13
Thrombocytopenia	8	1.5	6	2.1	2	0.8	2.6	0.19
Pleuritis	5	0.9	4	1.5	1	0.3	5	0.16
Other manifestations								
Arthralgia	308	56.3	168	58.3	140	54.1	1.1	0.29
Morning joint stiffness	128	23.4	69	24.0	59	22.8	1.1	0.74
Raynaud's phenomenon	73	13.3	47	16.3	26	10.0	1.6	0.03
Headache	72	13.2	49	17.0	23	8.9	1.9	0.004
Xerostomia	70	12.8	43	14.9	27	10.4	1.4	0.11
Arterial hypertension	67	12.2	35	12.2	32	12.4	1	0.94
Tendinitis	49	9	27	9.4	22	8.5	1.1	0.72
Psoriasis	40	7.4	14	4.9	26	10.2	0.5	0.02
Affective disorder	40	7.3	27	9.4	13	5.0	1.9	0.05
Keratoconjunctivitis sicca	39	7.2	25	8.7	14	5.5	1.6	0.14
Asthma bronchiale	38	7.0	25	8.7	13	5.0	1.7	0.09
Puffy fingers	36	6.6	25	8.7	11	4.2	2.1	0.35
Thyroiditis	33	6.1	18	6.3	15	5.8	1.1	0.81
Peripheral neuropathy	32	5.9	14	4.9	18	6.9	0.7	0.30
Weight loss	28	5.2	15	5.2	13	5.0	1	0.92
Axial arthritis	25	4.6	15	5.2	10	3.9	1.3	0.45
Fibromyalgia	21	3.9	12	4.2	9	3.5	1.2	0.67
Anaemia, non-haemolytic	20	3.7	13	4.5	7	2.7	1.7	0.26
Non-infectious fever	19	3.5	9	3.1	10	3.9	1.3	0.65
Arrhythmia	16	2.9	9	3.1	7	2.7	0.8	0.77
Cutaneous vasculitis	14	2.6	7	2.5	7	2.7	0.9	0.85
Miscarriage/abortion	14	2.6	9	3.1	5	1.9	1.6	0.32
Chronic urticaria	12	2.2	7	2.4	5	1.9	1.3	0.69
Livedo reticularis	12	2.2	9	3.1	3	1.2	2.6	0.11
Palindromic arthritis	12	2.2	6	2.1	6	2.3	0.9	0.85

*Ratio between prevalence of manifestation in ANA positive and ANA-negative patients.
ANA, antinuclear antibodies.

ELISA (CLIFT– ELISA+), with higher prevalence of oral or nasal ulcers, tendinitis, xerophthalmia, xerostomia and fibromyalgia. The combination of CLIFT positivity with negative outcome of ELISA (CLIFT+ ELISA–) was more frequent in patients with arthralgia, peripheral arthritis, morning joint stiffness, photosensitivity, Raynaud's phenomenon, puffy fingers, thrombocytopenia, livedo reticularis, chronic urticaria, discoid LE, asthma, peripheral neuropathy and headache.

Associations between clinical and biochemical manifestations and positivity of anti-dsDNA antibodies are listed in [table 5](#). ORs with 95% CIs are given as crude

values (univariate) and as adjusted values (multivariate) for all the individual manifestations significantly associated with presence of anti-dsDNA antibodies, assessed by any CLIFT and any ELISA, namely proteinuria, haematuria, alopecia, leukopenia, thrombocytopenia, cutaneous vasculitis and pleuritis.

Lymphopenia and non-haemolytic anaemia were significantly associated with positive CLIFT, as well as morning stiffness and arthralgia were inversely associated only with a positive ELISA result.

The clinical manifestations most strongly associated with positive result of CLIFT had higher OR in

Table 4 Prevalence of positivity in the anti-dsDNA tests performed

	All patients		ANA pos (N=288)		ANA neg (N=259)		Rate ratio*	p Value
	n	Per cent	n	Per cent	n	Per cent		
CLIFT 1	33	6.1	18	6.3	15	5.8	1.1	0.82
CLIFT 2	36	6.6	22	7.6	14	5.4	1.4	0.29
CLIFT 3	45	8.2	30	10.4	15	5.8	1.8	0.05
Any CLIFT	59	10.8	39	13.5	20	7.7	1.75	0.03
EliA	61	11.1	27	9.4	34	13.1	0.7	0.16
SPADE	57	10.4	28	9.7	29	11.2	0.9	0.57
Vareliisa 1	50	9.1	26	9.0	24	9.3	1	0.92
Vareliisa 2	59	10.8	30	10.4	29	11.2	0.9	0.77
Any ELISA	99	18.1	50	17.4	49	18.9	0.9	0.64
Any test	124	22.7	68	23.6	56	21.6	1.1	0.58
CLIFT+ELISA	34	6.2	21	7.3	13	5.0	1.6	0.27

Test details are presented in [table 1](#).

*Ratio between prevalence of positive anti-dsDNA test in ANA positive and ANA-negative patients.

ANA, antinuclear antibodies; CLIFT, Crithidia Lucillia Immunofluorescence Test; SPADE, solution phase anti-dsDNA ELISA.

ANA-positive patients, except for pleuritis. In patients with any positive ELISA, only OR for cutaneous vasculitis, thrombocytopenia and leukopenia were higher in

ANA-positive patients. Having a positive anti-dsDNA test was associated with pleuritis and proteinuria even in ANA-negative patients.

Table 5 Association between most relevant clinical manifestations and positive outcome of anti-dsDNA tests (any CLIFT and any ELISA)

	Any CLIFT positive (39 ANA positive+20 ANA negative)		Any ELISA positive (50 ANA positive+49 ANA negative)	
	CRUDE OR (95% CI)	ADJUSTED OR (95% CI)	CRUDE OR (95% CI)	ADJUSTED OR (95% CI)
Peripheral arthritis	1.1 (0.6–2)		0.6 (0.4–1.04)	
Photosensitivity	0.6 (0.2–1.7)		0.5 (0.2–1.2)	
Oral ulcers	0.8 (0.25–2.9)		1.8 (0.8–4.1)	
Haematuria	3.8 (1.4–10.4)	0.6 (0.1–3.4)	3.2 (1.3–8)	0.5 (0.1–2.8)
Proteinuria	14 (5.1–38.4)	13 (2.9–57.7)	16.7 (5.3–52.6)	18.8 (3.7–95.2)
Malar rash	3.1 (0.97–10.2)		1.7 (0.5–5.4)	
Anaemia	3.8 (1.4–10.4)		1.1 (0.4–3.5)	
Leukopenia	6.8 (2.3–20.3)	2.3 (0.5–10.7)	6.5 (2.2–19.1)	3.5 (0.8–14.2)
Alopecia	4.4 (1.3–15)	4.3 (1.1–16)	3.35 (1–10.8)	3.1 (0.9–10.8)
Lymphopenia	10.9 (3.2–37)		2.7 (0.8–9.2)	
Discoid LE	1.2 (0.1–9.8)		0.6 (0.8–5.3)	
Thrombocytopenia	8.8 (2.1–36.2)	3.1 (0.5–20.2)	4.7 (1.1–19)	1.1 (0.2–7.6)
Pleuritis	13 (2.1–79.6)	11.1 (1.5–83.8)	18.8 (2.1–170.3)	14.5 (1.4–148.2)
Arthralgia	0.6 (0.3–1.02)		0.5 (0.3–0.8)	
Morning joint stiffness	0.5 (0.2–1.04)		0.4 (0.2–0.7)	
Raynaud's phenomenon	1.6 (0.8–3.2)		1.2 (0.6–2.2)	
Headache	0.7 (0.3–1.75)		0.8 (0.4–1.6)	
Xerostomia	0.75 (0.3–1.8)		1 (0.5–2)	
Arterial hypertension	0.96 (0.4–2.2)		1 (0.5–1.9)	
Tendinitis	0.2 (0.02–1.2)		1.3 (0.7–2.7)	
Psoriasis	0.6 (0.2–2.2)		0.5 (0.2–1.4)	
Affective disorder	0.7 (0.2–2.2)		0.8 (0.3–1.9)	
Keratoconjunctivitis sicca	0.4 (0.1–1.8)		0.8 (0.3–2)	
Cutaneous vasculitis	6.7 (2.25–20.2)	2.1 (0.5–9.4)	6.4 (2.2–18.9)	2.4 (0.6–9.5)
Asthma or COL	2.4 (1.04–5.5)		1.4 (0.7–3.15)	
Lymphadenopathy	6.5 (1.4–29.7)		1.8 (0.3–9.6)	

Crude OR with corresponding 95% CIs in brackets is reported for all the variables. Adjusted OR is reported only for variables significantly associated with positivity of any CLIFT and any ELISA.

ANA, antinuclear antibodies; CLIFT, Crithidia Lucillia Immunofluorescence Test.

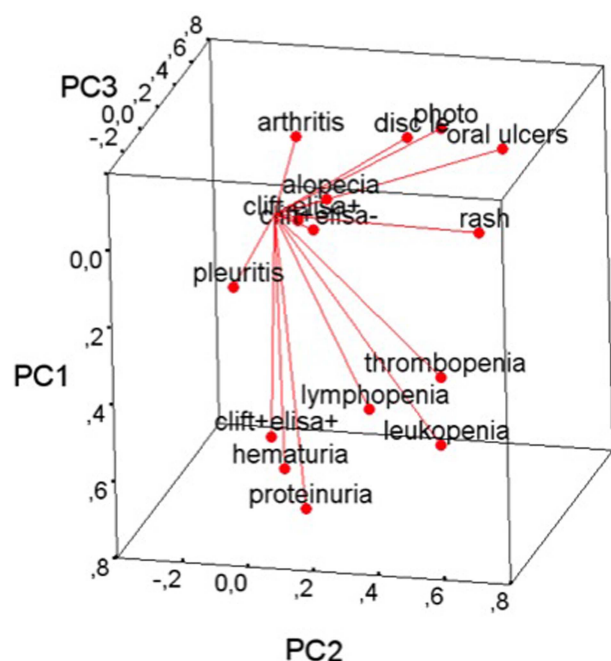


Figure 1 Principal component (PC) analysis of typical manifestations of systemic lupus erythematosus, positivity of any Chritidia Luciliae Immunofluorescence Test (CLIFT) and positivity of any ELISA included in this study. PC 1, 2 and 3 explained 16.1%, 10.7% and 8.6%, respectively. This PC plot aims to optimally display variances and not correlations; however, the angles between the various plot vectors serve as good indicators of the correlations among the variables, and the length of the vectors provide good indications as to which variables have had the largest effect on the variation in the dataset. The plot shows how having a positive CLIFT and ELISA (clift+elisa+) is associated with signs of nephropathy, haematological abnormalities and pleuritis; having a single positive test (CLIFT or ELISA) did not cluster with any of the manifestations included in the PC analysis.

The overall association of anti-dsDNA with typical manifestations of SLE

A PCA of the ACR classification criteria^{5 40} items included in [table 4](#), alopecia (included in the Systemic Lupus International Collaborating Clinics (SLICC) criteria⁸) and outcomes of anti-dsDNA tests by ELISA (any test positive) and CLIFT (any test positive) is shown as a three-dimensional plot for the three major principal components explaining 16.1%, 10.7% and 8.6%, respectively, of the variance in the dataset ([figure 1](#)). From this plot it is seen that having positive CLIFT and ELISA tests for anti-dsDNA (n=34) was closely associated with nephropathy (proteinuria and haematuria), haematological abnormalities and pleuritis; patients with a positive CLIFT and negative ELISA (n=25) or negative CLIFT and positive ELISA (n=65) did not cluster with any particular subset of the clinical manifestations.

DISCUSSION

The uniqueness of the present study is that the association between clinical phenotypes and presence in

serum of anti-dsDNA antibodies has been investigated in consecutive patients with recent onset of rheumatic symptoms, regardless of the diagnosis. By this approach, we aimed to reproduce the usual clinical setting, where the physician, at an early stage, is challenged to formulate a diagnosis and predict the outcome, based on clinical manifestations and suitable diagnostic tools available locally.

This study demonstrates that different techniques for the detection of anti-dsDNA antibodies result in a considerable variation in anti-dsDNA antibody status and relation hereof to various clinical and biochemical manifestations. Shared findings for the regression analyses and the PCA are the relationships between nephropathy and pleuritis with a positive CLIFT. The results obtained for immunoassays were, instead, highly variable and less selective with respect to several typical SLE manifestations. Furthermore, anti-dsDNA antibodies in general were only associated with a few manifestations typically used to classify SLE, and notably with lupus nephritis, where anti-dsDNA antibodies are demonstrated to be involved in the pathogenesis.

These observations raise the discussion about the performance of the various anti-dsDNA detection techniques, the general pathogenic role of the various anti-dsDNA antibodies identified, their clinical associations, and how the syndrome of SLE is currently delineated. Anti-dsDNA antibodies represent a heterogeneous population of antibodies with respect to origin, intrinsic affinity, fine structural DNA specificity and potential to cross-react with non-DNA structures. From this consideration, diverse anti-dsDNA antibody assays may have been designed without implementing this knowledge, but rather with a general view that anti-dsDNA is a homogeneous antibody population. Therefore, in order to analyse this, diverse assay principles are included in this study ([table 1](#)). These assays are partly claimed to detect antibodies of low versus high avidity, and those recognised in the CLIFT assays are reported to recognise highly bent structures on DNA, that are disclosed from the general B helical DNA used in some of the ELISA assays. Consistent with this, the kinetoplast DNA has one of the greatest known degrees of stable curvature.^{41 42} Thus, the assays may disclose antibody binding to DNA structures that are only formed by strong deformations from the more common linear B helical DNA structure. Antibodies recognising the kinetoplast DNA of the haemoflagellate *Crithidia luciliae* may specifically bind unique structures shared by nucleosomes.^{41 42} This stringent antibody specificity may well reflect structures on eukaryotic nucleosomal DNA that is believed to induce such immune responses in vivo.

We analysed anti-dsDNA antibodies by a variety of anti-dsDNA tests commonly used in clinical practice. CLIFT was performed on all sera at all three laboratories. Each laboratory also performed additional anti-dsDNA antibody assays on all patient sera, depending on their interests and expertise. This approach uncovered a high degree of variation in the reporting of

presence of anti-dsDNA between the individual assays. Even when using a common antigen source (CLIFT) in all laboratories, a high degree of variation in the assay results was demonstrated. It cannot be regarded as acceptable, and it supports the need for further international efforts in assay standardisation, and a broad discussion of what we want to analyse. Do we want to detect as many antibodies as possible, or only those with stringent characteristics, like high avidity and strong correlation with organ manifestations?

The present study shows a high degree of variation in the associations of anti-dsDNA antibodies with clinical and biochemical manifestations, which is in line with previous observations.^{1 43 44} We do confirm also that immunoassays had higher rates of positivity than CLIFT, as previously reported.^{13 45}

Anti-dsDNA antibodies do not represent a well-defined autoantibody entity. Given the multiple and diverse mechanisms for their production, it is less obvious that anti-dsDNA antibodies per se are associated with all aspects of the broad syndrome constituting SLE.

As others and we have observed, individuals may produce anti-dsDNA antibodies without having organ manifestations, like, for example, nephritis or dermatitis. It has been known for decades that not all anti-dsDNA antibodies are pathogenic.

A renal target for potentially nephritogenic anti-dsDNA antibodies has been demonstrated to be extracellular, poorly degraded chromatin fragments in both murine^{30 46 47} and human⁴⁸ lupus nephritis. Antibodies that bind exclusively in ELISA recognise B helical DNA, but not the highly bent structure in the kinetoplast or in the chromatin fragments.^{41 42} Thus, what makes anti-dsDNA antibodies pathogenic is their potential to bind chromatin structures, as reflected in, for example, the CLIFT assay, and not just because they bind any DNA structure. This basic information should encourage us to consider this as a problem, and to develop a new consensus with respect to how we should test for anti-dsDNA antibodies.

In an effort to clinically identify the pathogenic potential of the various anti-dsDNA antibodies measured in this study, we have correlated these findings to clinical and biochemical features of patients referred to our clinics for further evaluation of rheumatic disease. The data support the notion that various anti-dsDNA antibodies impact differently on the classification of SLE.

We focused on whether any of the anti-dsDNA tests were discriminatory towards manifestations included in the current ACR classification criteria.⁵ Anti-dsDNA antibodies detected by CLIFT are associated with a clinical picture characterised by proteinuria, haematuria, pleuritis and leukopenia. When the presence of anti-dsDNA antibodies is confirmed by any immunoassay, then the prevalence of malar rash, cutaneous vasculitis, alopecia, lymphopenia and non-haemolytic anaemia seems to increase.

Arthralgia, cutaneous vasculitis, morning stiffness and alopecia are variously associated with the anti-dsDNA tests. Except for alopecia, however, these associations are not strong, and these findings may be spurious. Our results may indicate the existence of a cluster of patients with anti-dsDNA antibodies and diffuse non-scarring alopecia. It supports the recent reinstatement of non-scarring alopecia as a clinical criterion in the SLICC classification criteria of SLE.⁸

With regard to the immunological ACR classification criterion of SLE 'antibody to native DNA in an abnormal titre',⁵ this study has demonstrated an up to twofold difference in prevalence of positivity, and considerable variation in the clinical association profile between the anti-dsDNA tests performed. This may relate to varying test properties with regard to avidity and structural antibody specificity.¹³ Immunoassays are often used to make a preliminary screening of anti-dsDNA antibodies. Positive results are then verified by other more specific assays, such as CLIFT. In our investigation, the positivity of ELISA combined with CLIFT negativity was found in 65 patients, whereof 13 were diagnosed with SLE. A moderately increased prevalence of tendinitis, fibromyalgia, xerophthalmia, xerostomia, oral and nasal ulcers was observed, a clinical picture often found in patients affected by Sjögren's Syndrome. However, these and other parallel findings^{49–54} indicate that currently the anti-dsDNA antibodies cannot be considered precise enough as a classification criterion. Low specificity of ELISA testing for anti-dsDNA antibodies has in newly proposed classification criteria been offset by raising the cut-off to twice above lower laboratory reference range.⁸ Whether this is sufficient to generate a valid, clinically relevant anti-dsDNA antibody test situation has not yet been validated. To this end, it is noteworthy that even the CLIFT test, which according to the present results is the anti-dsDNA assay best correlated with a typical nephritic SLE phenotype, had only low diagnostic value in the diagnosis of SLE in this population of patients with recent onset of rheumatic symptoms.⁵⁵

When analysing the ANA-positive subset of the patients, the CLIFT association with clinical phenotypes consisting of proteinuria, haematuria, leukopenia, thrombocytopenia, cutaneous vasculitis and alopecia persisted. On the other hand, pleuritis and proteinuria defined the clinical phenotypes associated with any of the anti-dsDNA tests in ANA-negative patients. These observations suggest that the presence of anti-dsDNA antibodies in ANA-negative patients may be of less clinical significance, supporting the view that testing for antibodies to dsDNA is not indicated in ANA-negative sera.⁵⁶ Detection in ANA-negative patients of anti-dsDNA antibodies with low avidity may, in a worst case scenario, lead to incorrect diagnosis and classification.

The purpose of this study was to reach more insight into the linkage between anti-dsDNA antibody detection and clinical and laboratory manifestations without the

restriction posed by existing SLE classification criteria. Our findings provide clinical support for a diagnostic role of CLIFT-determined anti-dsDNA antibodies, however, only in a limited number of key SLE manifestations, proteinuria in particular. These findings challenge the broad role of ELISA-based anti-dsDNA antibody testing in diagnosis and classification of SLE as presently defined.

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REFERENCES

- Isenberg DA, Ehrenstein MR, Longhurst C, *et al.* The origin, sequence, structure, and consequences of developing anti-DNA antibodies. A human perspective. *Arthritis Rheum* 1994;37:169–80.
- Khalil M, Spatz L, Diamond B. Anti-DNA antibodies. In: Lahita R. ed. *Systemic lupus erythematosus*. 3rd edn. Academic Press, 1999:197–217.
- van Bruggen MC, Walgreen B, Rijke TP, *et al.* Antigen specificity of anti-nuclear antibodies complexed to nucleosomes determines glomerular basement membrane binding in vivo. *Eur J Immunol* 1997;27:1564–9.
- van Bavel CC, Fenton KA, Rekvig OP, *et al.* Glomerular targets of nephritogenic autoantibodies in systemic lupus erythematosus. *Arthritis Rheum* 2008;58:1892–9.
- Tan EM, Cohen AS, Fries JF, *et al.* The 1982 revised criteria for the classification of systemic lupus erythematosus. *Arthritis Rheum* 1982;25:1271–7.
- Hahn BH. Antibodies to DNA. *N Engl J Med* 1998;338:1359–68.
- Foster MH. T cells and B cells in lupus nephritis. *Semin Nephrol* 2007;27:47–58.
- Petri M, Orbai AM, Alarcon GS, *et al.* Derivation and validation of the Systemic Lupus International Collaborating Clinics classification criteria for systemic lupus erythematosus. *Arthritis Rheum* 2012;64:2677–86.
- Fenton K, Fismen S, Hedberg A, *et al.* Anti-dsDNA antibodies promote initiation, and acquired loss of renal Dnase1 promotes progression of lupus nephritis in autoimmune (NZBxNZW)F1 mice. *PLoS One* 2009;4:e8474.
- Fismen S, Hedberg A, Fenton KA, *et al.* Circulating chromatin-anti-chromatin antibody complexes bind with high affinity to dermo-epidermal structures in murine and human lupus nephritis. *Lupus* 2009;18:597–607.
- Grootscholten C, van Bruggen MC, van der Pijl JW, *et al.* Deposition of nucleosomal antigens (histones and DNA) in the epidermal basement membrane in human lupus nephritis. *Arthritis Rheum* 2003;48:1355–62.
- Huerta PT, Kowal C, DeGiorgio LA, *et al.* Immunity and behavior: antibodies alter emotion. *Proc Natl Acad Sci USA* 2006;103:678–83.
- Haugbro K, Nossent JC, Winkler T, *et al.* Anti-dsDNA antibodies and disease classification in antinuclear antibody positive patients: the role of analytical diversity. *Ann Rheum Dis* 2004;63:386–94.
- Jang YJ, Stollar BD. Anti-DNA antibodies: aspects of structure and pathogenicity. *Cell Mol Life Sci* 2003;60:309–20.
- Kalsi JK, Martin AC, Hirabayashi Y, *et al.* Functional and modelling studies of the binding of human monoclonal anti-DNA antibodies to DNA. *Mol Immunol* 1996;33:471–83.
- Stollar BD. Immunochimistry of DNA. *Int Rev Immunol* 1989;5:1–22.
- Berden JH. Lupus nephritis: consequence of disturbed removal of apoptotic cells? *Neth J Med* 2003;61:233–8.
- Fismen S, Mortensen ES, Rekvig OP. Nuclease deficiencies promote end-stage lupus nephritis but not nephritogenic autoimmunity in (NZB x NZW) F1 mice. *Immunol Cell Biol* 2011;89:90–9.
- Kruse K, Janko C, Urbonaviciute V, *et al.* Inefficient clearance of dying cells in patients with SLE: anti-dsDNA autoantibodies, MFG-E8, HMGB-1 and other players. *Apoptosis* 2010;15:1098–113.
- Munoz LE, Janko C, Schulze C, *et al.* Autoimmunity and chronic inflammation—two clearance-related steps in the etiopathogenesis of SLE. *Autoimmun Rev* 2010;10:38–42.
- Napirei M, Karsunky H, Zevnik B, *et al.* Features of systemic lupus erythematosus in Dnase1-deficient mice. *Nat Genet* 2000;25:177–81.
- Urbonaviciute V, Fumrohr BG, Meister S, *et al.* Induction of inflammatory and immune responses by HMGB1-nucleosome complexes: implications for the pathogenesis of SLE. *J Exp Med* 2008;205:3007–18.
- Van Ghelue M, Moens U, Bendixsen S, *et al.* Autoimmunity to nucleosomes related to viral infection: a focus on hapten-carrier complex formation. *J Autoimmun* 2003;20:171–82.
- Rekvig OP. Anti-dsDNA antibodies as a classification criterion and a diagnostic marker for SLE: critical remarks. *Clin Exp Immunol* 2014. doi:10.1111/cei.12296
- Mostoslavsky G, Fischel R, Yachimovich N, *et al.* Lupus anti-DNA autoantibodies cross-react with a glomerular structural protein: a case for tissue injury by molecular mimicry. *Eur J Immunol* 2001;31:1221–7.
- Amital H, Heilweil M, Ulmansky R, *et al.* Treatment with a laminin-derived peptide suppresses lupus nephritis. *J Immunol* 2005;175:5516–23.
- Deocharan B, Qing X, Lichauro J, *et al.* Alpha-actinin is a cross-reactive renal target for pathogenic anti-DNA antibodies. *J Immunol* 2002;168:3072–8.
- Mageed RA, Zack DJ. Cross-reactivity and pathogenicity of anti-DNA autoantibodies in systemic lupus erythematosus. *Lupus* 2002;11:783–6.
- Sabbaga J, Line SR, Potocnjak P, *et al.* A murine nephritogenic monoclonal anti-DNA autoantibody binds directly to mouse laminin, the major non-collagenous protein component of the glomerular basement membrane. *Eur J Immunol* 1989;19:137–43.
- Kalaaji M, Mortensen E, Jorgensen L, *et al.* Nephritogenic lupus antibodies recognize glomerular basement membrane-associated chromatin fragments released from apoptotic intraglomerular cells. *Am J Pathol* 2006;168:1779–92.

31. Lefkowitz JB, Gilkeson GS. Nephritogenic autoantibodies in lupus: current concepts and continuing controversies. *Arthritis Rheum* 1996;39:894–903.
32. Mjelle JE, Rekvig OP, Fenton KA. Nucleosomes possess a high affinity for glomerular laminin and collagen IV and bind nephritogenic antibodies in murine lupus-like nephritis. *Ann Rheum Dis* 2007;66:1661–8.
33. Mortensen ES, Fenton KA, Rekvig OP. Lupus nephritis: the central role of nucleosomes revealed. *Am J Pathol* 2008;172:275–83.
34. Mortensen ES, Rekvig OP. Nephritogenic potential of anti-DNA antibodies against necrotic nucleosomes. *J Am Soc Nephrol* 2009;20:696–704.
35. Seredkina N, Rekvig OP. Acquired loss of renal nuclease activity is restricted to DNase1 and is an organ-selective feature in murine lupus nephritis. *Am J Pathol* 2011;179:1120–8.
36. Seredkina N, Zykova SN, Rekvig OP. Progression of murine lupus nephritis is linked to acquired renal Dnase1 deficiency and not to up-regulated apoptosis. *Am J Pathol* 2009;175:97–106.
37. Van Bruggen MC, Kramers C, Berden JH. Autoimmunity against nucleosomes and lupus nephritis. *Ann Med Interne (Paris)* 1996;147:485–9.
38. van Bruggen MC, Kramers C, Hylkema MN, *et al.* Pathophysiology of lupus nephritis: the role of nucleosomes. *Neth J Med* 1994;45:273–9.
39. Radic MZ, Seal SN. Selection of recurrent V genes and somatic mutations in autoantibodies to DNA. *Methods* 1997;11: 20–6.
40. Hochberg MC. Updating the American College of Rheumatology revised criteria for the classification of systemic lupus erythematosus. *Arthritis Rheum* 1997;40:1725.
41. Griffith J, Bleyman M, Rauch CA, *et al.* Visualization of the bent helix in kinetoplast DNA by electron microscopy. *Cell* 1986;46:717–24.
42. Hirota Y, Ohshima T. Adjacent upstream superhelical writhe influences an Escherichia coli promoter as measured by in vivo strength and in vitro open complex formation. *J Mol Biol* 1995;254:566–78.
43. Isenberg DA, Dudeney C, Williams W, *et al.* Measurement of anti-DNA antibodies: a reappraisal using five different methods. *Ann Rheum Dis* 1987;46:448–56.
44. Tan EM, Smolen JS, McDougal JS, *et al.* A critical evaluation of enzyme immunoassays for detection of antinuclear autoantibodies of defined specificities. I. Precision, sensitivity, and specificity. *Arthritis Rheum* 1999;42:455–64.
45. Werle E, Blazek M, Fiehn W. The clinical significance of measuring different anti-dsDNA antibodies by using the Farr assay, an enzyme immunoassay and a Crithidia luciliae immunofluorescence test. *Lupus* 1992;1:369–77.
46. Kalaaji M, Sturfelt G, Mjelle JE, *et al.* Critical comparative analyses of anti-alpha-actinin and glomerulus-bound antibodies in human and murine lupus nephritis. *Arthritis Rheum* 2006;54:914–26.
47. Zykova SN, Seredkina N, Benjaminsen J, *et al.* Reduced fragmentation of apoptotic chromatin is associated with nephritis in lupus-prone (NZB x NZW)F(1) mice. *Arthritis Rheum* 2008;58:813–25.
48. Kalaaji M, Fenton KA, Mortensen ES, *et al.* Glomerular apoptotic nucleosomes are central target structures for nephritogenic antibodies in human SLE nephritis. *Kidney Int* 2007;71:664–72.
49. Antico A, Platzgummer S, Bassetti D, *et al.* Diagnosing systemic lupus erythematosus: new-generation immunoassays for measurement of anti-dsDNA antibodies are an effective alternative to the Farr technique and the Crithidia luciliae immunofluorescence test. *Lupus* 2010;19:906–12.
50. Conrad K, Ittenson A, Reinhold D, *et al.* High sensitive detection of double-stranded DNA autoantibodies by a modified Crithidia luciliae immunofluorescence test. *Ann N Y Acad Sci* 2009;1173: 180–5.
51. Kim KH, Han JY, Kim JM, *et al.* Clinical significance of ELISA positive and immunofluorescence negative anti-dsDNA antibody. *Clin Chim Acta* 2007;380:182–5.
52. Neogi T, Gladman DD, Ibanez D, *et al.* Anti-dsDNA antibody testing by Farr and ELISA techniques is not equivalent. *J Rheumatol* 2006;33:1785–8.
53. Roggenbuck D, Conrad K, Reinhold D. High sensitive detection of double-stranded DNA antibodies by a modified Crithidia luciliae immunofluorescence test may improve diagnosis of systemic lupus erythematosus. *Clin Chim Acta* 2010;411:1837–8.
54. Tzioufas AG, Terzoglou C, Stavropoulos ED, *et al.* Determination of anti-ds-DNA antibodies by three different methods: comparison of sensitivity, specificity and correlation with lupus activity index (LAI). *Clin Rheumatol* 1990;9:186–92.
55. Compagno M, Jacobsen S, Rekvig OP, *et al.* Low diagnostic and predictive value of anti-dsDNA antibodies in unselected patients with recent onset of rheumatic symptoms: results from a long-term follow-up Scandinavian multicentre study. *Scand J Rheumatol* 2013;42:311–16.
56. Manoussakis MN, Garalea KL, Tzioufas AG, *et al.* Testing for antibodies to ENA and to dsDNA is not indicated in FANA-negative sera. *Clin Rheumatol* 1988;7:465–9.



Clinical phenotype associations with various types of anti-dsDNA antibodies in patients with recent onset of rheumatic symptoms. Results from a multicentre observational study

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