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Isolation and Characterization of Two European Strains of Ehrlichia phagocytophila of Equine Origin

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We report the isolation and partial genetic characterization of two equine strains of granulocytic Ehrlichia of the genogroup Ehrlichia phagocytophila. Frozen whole-blood samples from two Swedish horses with laboratory-verified granulocytic ehrlichiosis were inoculated into HL-60 cell cultures. Granulocytic Ehrlichia was isolated and propagated from both horses. DNA extracts from the respective strains were amplified by PCR using primers directed towards the 16S rRNA gene, the groESL heat shock operon gene, and the ank gene. The amplified gene fragments were sequenced and compared to known sequences in the GenBank database. With respect to the 16S rRNA gene, the groESL gene, and the ank gene, the DNA sequences of the two equine Ehrlichia isolates were identical to sequences found in isolates from clinical cases of granulocytic ehrlichiosis in humans and domestic animals in Sweden. However, compared to amplified DNA from an American Ehrlichia strain of the E. phagocytophila genogroup, differences were found in the groESL gene and ank gene sequences.

Granulocytic Ehrlichia causes febrile diseases in many different animals and in humans. The first human cases were described for the United States in 1994, but clinical cases are now accumulating from many countries, mainly in temperate regions (3, 24, 29; A. van Dobbenburgh, A. P. van Dam, and E. Fikrig, Letter, N. Engl. J. Med. 340:1214–1216, 1999). In Scandinavia, clinical granulocytic ehrlichiosis (GE) has been reported for humans, cattle, sheep, horses, dogs, and cats (5, 7, 14, 26). The infectious agents are strictly intracellular rickettsia-like bacilli, with the capacity for intracellular life in the wildlife reservoir and the infected host, as well as prolonged survival in the principal vector, hard-bodied ticks of the genus Ixodes. Until the successful isolation of granulocytic ehrlichias in HL-60 cells, the possibilities of studying these bacteria were limited to indirect and molecular biological methods (16). Today, reports of successful isolation of granulocytic Ehrlichia from humans and animals are available from the United States but not from other parts of the world. The aim of this study was to isolate, maintain in culture, and genetically characterize European strains of granulocyticotropic Ehrlichia phagocytophila. Moreover, earlier reports on the isolation of granulocytic Ehrlichia spp. were based only on isolation from fresh blood. In this paper, we report the isolation of European E. phagocytophila of equine origin from stored frozen whole blood. Blood smears showed cytoplasmic inclusions in approximately 25 to 30% of the neutrophils in both cases, supporting the clinical diagnosis of GE. Both horses were treated with intravenous oxytetracycline (7 mg/kg of body weight daily for 7 days) and recovered clinically within 24 h.

Culture of ehrlichias in HL-60 cells. Promyelocytic HL-60 leukemia cells (ATCC CCL240) were maintained in antibiotic-free RPMI 1640 medium supplemented with 2 mM l-glutamine and 20% fetal bovine serum. The HL-60 cells were incubated at 37°C in an atmosphere of 5% CO2 (16). An aliquot of 0.5 ml of the sedimentsed leukocyte-rich fraction of equine EDTA-blood (kept at −20°C for 7 months) was inoculated into 25-cm2 flasks with HL-60 cells at a density of 2 × 106 cells/ml. The infected cells were then monitored daily by microscopy and examination of Giemsa-stained cytopsin-prepared cell spreads. The cultures were kept at a density of 2 × 105 to 4 × 106 cells/ml by feeding them with medium two to three times a week. Infection of the cells was quantified by the presence of morulae and by indirect immunofluorescence assay using a bovine anti-Ehrlichia immunoglobulin G-positive serum and a fluorescein isothiocyanate-conjugated goat anti-bovine immunoglobulin G antibody (product no. 209-0088; Jackson ImmunoResearch Laboratories), diluted 1:100 in phosphate-buffered saline, as secondary antibody. Noninfected HL-60 cells were used as negative controls.

PCR amplification and sequence analysis. DNA was extracted from infected and noninfected cells with the QiAamp Tissue Kit protocol (Qiagen GmbH, Hilden, Germany). Nested PCR protocols targeting the 16S rRNA gene, the ank gene, and the groESL gene were followed as described previously, but with minor changes (21, 22, 27). The primers used to amplify the 16S rRNA gene were 16SF1 (5′-AGATTGTGATCCGTGCGT-3′) and GE10 (5′-TTCCTTAAAGAGTCTAATACTCC-3′) for the primary reaction, and GEHS1 (5′-CTTACGCGTTAGCTCACACAG-3′) and GEHS6 (5′-CAATAATCATGGCAGCAGCAG-3′) for the nested reaction. In the ank gene assay, AQF3 (5′-GAAGAAATTTACAACTCTTGAG-3′) and AQ2R2 (5′-TGAGGCGAAGGCGAAACG-3′) were used for the primary reaction, and AQF2 (5′-TTGATGCCTGAAAGCTAAC-3′) and AQ2R1 (5′-ACCATTGTGCTTGTGAGGAG-3′) for the nested reaction. Each of the primers was subsequently used for direct sequencing of the appropriate purified PCR product. Two additional primers were needed to sequence the 781-bp region amplified by the 16S rRNA PCR, EB4 (5′-GTATTACCAGCGCGCTGCTGGCAC-3′) and EHR521 (5′-CTGGCAC-3′), and EHR251 (5′-TGGAGGCGGTCGTTGAAAGG-3′). Sequencing reaction products were separated and analyzed using an automated sequencer (ABI 377; Applied Biosystems) and fluorescent-labeled dideoxynucleotide technology. Sequences were edited and analyzed with the Staden software.
Two strains showed similar propagation patterns. Seven days after equine blood and HL-60 cells were mixed, the first signs of morulae could be noted in the infected cells when they were analyzed with Giemsa-stained cytospin preparations (Fig. 1). Noninfected cells, grown and analyzed in parallel, showed no corresponding cell changes. During the following days, the extent of infection increased: 10% infected cells at day 7 and 60% at day 16. After day 16, the infection declined so that at day 26 only 20% of the cells were infected.

Cells infected with the respective strain were positive in PCR assays targeting the 16S rRNA gene, the ank gene, and the groESL gene of granulocytic *Ehrlichia*. The amplified 781-bp fragments of the 16S rRNA gene sequences were identical to corresponding sequences of amplified DNA obtained in Swedish human cases of GE (19). Similarly, the groESL gene and the ank gene sequences of the equine *Ehrlichia* isolates were identical for the two isolates and identical to the sequences of the corresponding gene fragments from Swedish human strains (21).

**DISCUSSION**

GE in horses was described as a clinical entity in Scandinavia in 1990 (6). The two infected horses in the present study resided in an area in southern Sweden where GE is commonly diagnosed in horses and dogs. They showed typical clinical signs of ehrlichiosis and responded well to treatment. Thus, the two *Ehrlichia* isolates described represent pathogenic granulocytic *Ehrlichia* strains from an area of southern Sweden where GE is endemic. To our knowledge, these two strains of equine origin are the first two European granulocytic *Ehrlichia* strains to be isolated and propagated in cell culture.

As gene sequence data for granulocytic ehrlichias accumulate, proposals have been made to merge *E. equi*, *E. phagocytophila*, and the human GE agent into a single species, *E. phagocytophila* (10, 13, 27). In accordance with these proposals and with data obtained in this study, *E. phagocytophila* has been used to designate the clinical isolates in this study.

The establishment of an infection in a cell line may vary in time and rate. In this study 10% of the HL-60 cells were infected 1 week after infection with the two equine isolates. Other studies of in vitro cultures of *E. phagocytophila* in HL-60 cells have shown that some culture systems result in an infection rate of ≥50 to 60% while other systems never reach more than a 2 to 5% infection rate. The infections are also established with different rapidities (16). These differences probably relate to the bacterial load of the inoculum, to *Ehrlichia* strain variations, and to variations in the HL-60 cell populations.

The 16s rRNA gene has become the “gold standard” for classification of bacteria. In this study, the nucleotide sequences of the 781-bp fragments of the 16S rRNA gene of the equine *E. phagocytophila* isolates were completely identical to the most common sequence variant of 16S ribosomal DNA obtained in clinical cases of GE in humans, cattle, horses, dogs, and cats in Scandinavia and other parts of Europe, as well as in human and canine cases in the United States (4, 7, 15, 17–19, 24).

In order to investigate the genetic and antigenic relationship between closely related bacterial species, the comparison of more-variable genes, e.g., genes of structural proteins, may be of value, since the 16S rRNA gene is too conserved to be able to resolve strain differences at this level. One possible gene to study is the ank gene, coding for a 160-kDa cytoplasmic protein antigen (8, 25). Analyses of this gene from several granulocytic *Ehrlichia* strains from geographically different areas resulted in the division of *E. phagocytophila* into three distinct clades: northeastern United States, upper midwestern United States, and Europe (21). The ank gene sequences of our two equine isolates were identical to previously described ank gene sequences in Swedish and Slovenian *E. phagocytophila* isolates from humans and animals but showed only 94 to 96% identity with ank gene sequences of North American *E. phagocytophila* isolates. The groESL sequences obtained from our equine *E. phagocytophila* isolates were identical to each other and to a previously characterized Swedish *E. phagocytophila* isolate from an infected human but differed from all other *Ehrlichia* groESL sequences present in GenBank by at least two nucleotides (27).

The principal wildlife reservoir of *E. phagocytophila* is believed to be small mammals, mainly rodents, and deer (1). The fact that both the reservoir and vector species differ between North America and Europe and that coevolution of the bacteria, the vector, and the host must to a large extent be independent processes on the two continents means that significant differences in strain characteristics can be expected, both genetically and in terms of phenotypic traits, such as antigenic profile, host preferences, and virulence. Antigenic pleomorphism has been reported earlier for various isolates of *E. phagocytophila* (2, 20, 30). Our results show differences be-
between the North American and European variants of \textit{E. phagocytophila} and suggest that the \textit{ank} gene provides useful information complementary to that from the 16S rRNA gene that can be used to divide \textit{E. phagocytophila} into clades corresponding to geographic distribution (9). This is interesting, as some of the differences found may lead to better geographically adapted diagnostic tools and to understanding of differences in bacterial virulence and host preferences (2, 20, 23, 28, 30).

Thus, these results warrant further comparisons of \textit{E. phagocytophila} strains of different geographic origins in terms of genetic relationships, expression of antigens, ecology, and epidemiology.

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