A subsequent study revealed that astaxanthin-rich algal meals and vitamin C are capable of inhibiting Helicobacter pylori infection in BALB/cA mice. This finding underscores the potential therapeutic benefits of such compounds in the management of H. pylori infections, which are a significant public health concern.
Astaxanthin-Rich Algal Meal and Vitamin C Inhibit Helicobacter pylori Infection in BALB/cA Mice

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Helicobacter pylori infection in humans is associated with chronic type B gastritis, peptic ulcer disease, and gastric carcinoma. A high intake of carotenoids and vitamin C has been proposed to prevent development of gastric malignancies. The aim of this study was to explore if the microalga Haematococcus pluvialis rich in the carotenoid astaxanthin and vitamin C can inhibit experimental H. pylori infection in a BALB/cA mouse model. Six-week-old BALB/cA mice were infected with the mouse-passaged H. pylori strain 119/95. At 2 weeks post-inoculation mice were treated orally once daily for 10 days (i) with different doses of algal meal rich in astaxanthin (0.4, 2, and 4 g/kg of body weight, with the astaxanthin content at 10, 50, and 100 mg/kg, respectively), (ii) with a control meal (algal meal without astaxanthin, 4 g/kg), or (iii) with vitamin C (400 mg/kg). Five mice from each group were sacrificed 1 day after the cessation of treatment, and the other five animals were sacrificed 10 days after the cessation of treatment. Culture of H. pylori and determination of the inflammation score of the gastric mucosae were used to determine the outcome of the treatment. Mice treated with astaxanthin-rich algal meal or vitamin C showed significantly lower colonization levels and lower inflammation scores than those of untreated or control-meal-treated animals at 1 day and 10 days after the cessation of treatment. Lipid peroxidation was significantly decreased in mice treated with the astaxanthin-rich algal meal and vitamin C compared with that of animals not treated or treated with the control meal. Both astaxanthin-rich algal meal and vitamin C showed an inhibitory effect on H. pylori growth in vitro. In conclusion, antioxidants may be a new strategy for treating H. pylori infection in humans.

MATERIALS AND METHODS

Chemicals. Homogenized and dried cells of the unicellular green alga Haematococcus pluvialis rich in astaxanthin (2 to 3% [wt/wt]) and algal meal without astaxanthin as the control meal were obtained from AstaCarotene AB (Gustavberg, Sweden). Vitamin C (L-ascorbic acid) was purchased from ICN Biomedicals Inc. (Lund, Sweden). Algal meal and control meal were suspended in distilled water and vitamin C was dissolved in distilled water just before use.

Bacterial strains. The H. pylori mouse-passaged strain 119/95 was grown on GAB-Camp agar (Gc Agar Base; Becton Dickinson, Lund, Sweden) supplemented with 10% horse serum and incubated for 48 h at 37°C under microaerophilic conditions (35). The cells were harvested in phosphate-buffered saline (PBS), centrifuged at 2,800 × g for 10 min, and resuspended in PBS to a final concentration of 10⁹ CFU/ml.

Animals. Six- to eight-week-old conventional BALB/cA mice were used in this study (B&K Universal Company, Stockholm, Sweden). Mice were housed on a 12-h light–12-h dark schedule and fed with rat and mouse standard diet no. 2 (B&K Universal) (34) and water ad libitum.

Experimental design. Sixty mice were inoculated orally through a feeding tube three times at 2-day intervals with 0.1 ml of an H. pylori suspension containing 10⁹ CFU/ml, and 10 mice were inoculated with PBS as a negative control group. The H. pylori-inoculated mice were divided into six groups. Five groups were orally treated with three doses of algal meal rich in astaxanthin (0.4, 2, and 4 g/kg of body weight, in which the astaxanthin content was 10, 50, and 100 mg/kg, respectively), a control algal meal (4 g/kg), or vitamin C (400 mg/kg) once daily for 10 days at 2 weeks postinoculation (p.i.). The infected and normal uninfected control mice were given distilled water through a feeding tube. Half of the animals from each group were sacrificed 1 day after the cessation of treatment, and the other half were sacrificed 10 days after the cessation of treatment.

Mice were killed with carbon dioxide, and their stomachs were collected. The stomachs were opened through the longer curvature using sterile surgical instruments. One-third of stomach biopsies were placed in 0.5 ml of PBS in a 1.5-ml Eppendorf tube and homogenized with a Pellet Pestle Mixer from KEBO Laboratories (Lund, Sweden). The homogenized sample was serially diluted 10-fold. Each 0.1 ml of homogenate was plated on GAB-Camp agar and incubated at 37°C for 5 to 10 days under microaerophilic conditions (36). The H. pylori colonies were counted and calculated as log₁₀ CFU per milliliter of homogenate. The presence of H. pylori on the culture plates was confirmed by

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TABLE 1. Concentrations of total carotenoids and astaxanthin in mouse stomachs

<table>
<thead>
<tr>
<th>Treatment group (concentration of astaxanthin [mg/kg])</th>
<th>Mean concn ± SD (μg/kg)</th>
<th>1 day after cessation of treatment</th>
<th>10 days after cessation of treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Total carotenoid</td>
<td>Astaxanthin</td>
<td>Total carotenoid</td>
</tr>
<tr>
<td>Normal</td>
<td>58 ± 8</td>
<td>0</td>
<td>51 ± 3</td>
</tr>
<tr>
<td>H. pylori</td>
<td>98 ± 28</td>
<td>0</td>
<td>58 ± 5</td>
</tr>
<tr>
<td><strong>H. pylori + algal meal (100)</strong></td>
<td>5,806 ± 1,162</td>
<td>516 ± 192</td>
<td>164 ± 34</td>
</tr>
<tr>
<td><strong>H. pylori + algal meal (50)</strong></td>
<td>2,255 ± 1,344</td>
<td>344 ± 101</td>
<td>107 ± 13</td>
</tr>
<tr>
<td><strong>H. pylori + algal meal (10)</strong></td>
<td>2,926 ± 668</td>
<td>120 ± 29*</td>
<td>116 ± 34</td>
</tr>
</tbody>
</table>

* P < 0.05 versus results from the groups treated with 100 and 50 mg of astaxanthin per kg.

RESULTS

Total carotenoid and astaxanthin analysis. The mouse stomachs showed correspondingly high total carotenoid and astaxanthin contents when they were treated with various concentrations of astaxanthin (Table 1). Significant differences were noted between the treated and untreated group, especially for animals just posttreatment. Mice treated with the highest dose of astaxanthin demonstrated a higher astaxanthin content in their stomachs than those of the animals treated with lower doses.

Culture. All noninoculated mice were H. pylori negative in culture. Both astaxanthin-rich algal meal (dose from 10 to 100 mg/kg) and vitamin C significantly reduced the number of H. pylori organisms in gastric tissue 1 day after the cessation of treatment (3.5 weeks p.i.) compared with the numbers recoverable from the untreated mice and the control mice treated with meal lacking astaxanthin (P < 0.05) (Fig. 1). At 10 days after the cessation of treatment (5 weeks p.i.), the numbers of H. pylori organisms in the groups treated with astaxanthin-rich algal meal and vitamin C were again significantly lower than the numbers in the groups not treated or treated with the control meal (P < 0.05) (Fig. 1). However, the astaxanthin-rich algal meal (100 mg/kg) and vitamin C (400 mg/kg) treatment groups had more numbers of H. pylori-negative animals (40%) was scored in a blind manner on a scale of 0 to 3 for body, antrum, and duodenum (36).

Lipid peroxidation assay. Mice stomach tissues were homogenized in 20 mM Tris-HCl, pH 7.4, to a concentration of 10% (wt/vol). Homogenate supernatants (200 μl) were tested for malondialdehyde (MDA) and 4-hydroxy-2(E)-nonenal with a lipid peroxidation assay kit from Calbiochem (Lund, Sweden). The colorimeters were measured at an absorbency at 586 nm, and tissue lipid peroxidation was calculated as micromoles per gram of tissue.

In vitro inhibition test. MICs were determined as described elsewhere (22) such that each preculture containing 10^5 cells was plated onto GAB-Camp agar with or without various concentrations of algal meal (0.3125 to 20 mg/ml), with algal meal without astaxanthin (5 mg/ml), or with vitamin C (0.5 to 4 mg/ml). The surviving cells were counted on the agar as colonies, and the MIC was defined as the concentration leaving no survivors after 5 to 10 days of incubation under microaerophilic conditions. Ten strains of H. pylori were tested, and MICs were shown as a range.

Statistical analysis. The Mann-Whitney U test was used for analysis of colonization and inflammation distribution. The level of significance was chosen to be a P of <0.05.
than the astaxanthin-rich algal meal (10 and 50 mg/kg) treatment groups (20%). There were no significant differences among the groups treated with three doses of astaxanthin-rich algal meal and vitamin C.

**Histopathology.** Mice treated with astaxanthin-rich algal meal or vitamin C showed significantly lower inflammation scores than control mice infected with *H. pylori* or treated with meal lacking astaxanthin 1 day and 10 days after the cessation of treatment (*P* < 0.05) (Fig. 2). The control-meal-treated mice developed gastritis as severe as that of the untreated control animals, and their inflammation scores were significantly higher than those of the non-*H. pylori*-inoculated mice (*P* < 0.01). The mice treated with the highest dose of astaxanthin (100 mg/kg) in algal meal showed significantly lower inflammation scores than the mice treated with 50 mg of astaxanthin per kg (*P* < 0.05).

Normal noninfected control mice showed normal fundic mucosa (Fig. 3A). Mice treated with astaxanthin-rich algal meal (100 mg/kg) or vitamin C (400 mg/kg) showed fewer inflammatory cells in their mucosae than control mice infected control mice (Fig. 3B to D).

**Lipid peroxidation.** The concentrations of MDA and 4-hydroxyalkenals in murine stomachs (in micromoles per gram of tissue) were significantly increased in *H. pylori*-infected untreated and control-meal (algal meal without astaxanthin)-treated mice compared with concentrations in normal control animals (*P* < 0.01). All astaxanthin-rich algal meal- or vitamin C-treated mice showed significant decreases in lipid peroxidation compared with levels in the untreated and control-meal-treated animals (*P* < 0.05) (Fig. 4).

**In vitro inhibition.** Astaxanthin-rich algal meal inhibited *H. pylori* growth at 0.3125 to 2.5 mg/ml (astaxanthin content, 6.25 to 50 μg/ml, pH 7.2), while algal meal without astaxanthin did not show this effect at 5 mg/ml. Vitamin C inhibited the growth of *H. pylori* at concentrations of 0.5 to 2 mg/ml (pH 7.2).

**DISCUSSION**

We have shown that antioxidants such as algal meal rich in astaxanthin as well as vitamin C inhibit *H. pylori* infection in BALB/cA mice. Among the three doses of astaxanthin tested, the highest dose (100 mg/kg) showed the best effect in reducing bacterial load and gastric inflammation. This finding is to our knowledge the first demonstration of an antimicrobial activity of astaxanthin-rich algal meal against *H. pylori* and associated gastric inflammation.

*H. pylori* infection has been associated with a decreased level of vitamin C and of major antioxidants (e.g., β-carotene) in human gastric tissue (5, 26). We found that vitamin C reduced bacterial colonization in the murine stomach and decreased the inflammation score. Interestingly, Jarosz et al. (11) reported that a high daily dose of vitamin C for 4 weeks (5 g per day) given to *H. pylori*-infected patients with chronic gastritis resulted in apparent *H. pylori* eradication in 30% of treated patients. In those patients the highly significant rise in total vitamin C concentration in the gastric juice persisted for at least 4 weeks posttreatment. Vitamin C not only seems to be an antioxidant and a free radical scavenger (17, 21, 26) but also shows antimicrobial activity against *H. pylori* both in vitro and in a Mongolian gerbil infection model (39).

Epidemiological evidence and clinical experiments suggest that vitamin C may exert a protective effect against the development of *H. pylori*-associated gastric carcinoma (4, 6, 37), but the mechanisms involved are not so clear.

The carotenoid astaxanthin has been established to be a powerful antioxidant in vitro (15, 24) and was previously shown to be able to prevent oral carcinogenesis in an experimental rat model (32). However, this carotenoid has not previously been shown to have an antimicrobial activity. We found that algal meal rich in astaxanthin has an inhibitory effect on *H. pylori* growth in vitro and also colonization in mouse stomach. BALB/cA mice treated with astaxanthin-rich algal meal showed decreased lipid peroxidation and granulocyte infiltration in their gastric mucosae.

*H. pylori*-infected individuals show high oxidative stress and high levels of ROMs in their gastric mucosae and an increased gastric antioxidative capacity after the eradication of *H. pylori* (14). A recent study of the formation of pro- and antioxidants to *H. pylori* infection in a Mongolian gerbil model showed an increase in the level of lipid peroxidation and activated glutathione turnover (31).

Astaxanthin acts as an antioxidant that protects against tis-
FIG. 3. (A) Normal fundic mucosa from an uninfected control mouse; (B) tissue from an *H. pylori*-infected mouse with a large amount of acute inflammatory cell infiltration within the mucosa and along the lamina muscular mucosa; (C and D) less inflammation (small amount of inflammatory cell infiltration) in mice treated with astaxanthin-rich algal meal and vitamin C, respectively.
In conclusion, our results suggest that the use of antioxidants to combat *H. pylori* infection in humans is an attractive new treatment strategy. Further studies with different feeding formulas and delivery systems as well as prophylaxis studies of animal models and clinical studies are now in progress.

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REFERENCES