Serum antibodies to Helicobacter hepaticus and Helicobacter pylori in patients with chronic liver disease

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Serum antibodies to *Helicobacter hepaticus* and *Helicobacter pylori* in patients with chronic liver disease

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Abstract

**Background**—Bile tolerant helicobacter species such as *H hepaticus* and *H bilis* have frequently been reported to cause hepatitis in mice and other rodents.

**Aims**—To investigate the possible pathogenic role of these and other helicobacter species in chronic liver disease in humans.

**Methods**—Serum samples from 144 patients with various chronic liver diseases, 30 patients with primary sclerosing cholangitis (PSC), and 48 healthy blood donors were analysed for antibodies against *H hepaticus* murine strain CCUG 33637 and *H pylori* strain CCUG 17874. Cell surface proteins of *H hepaticus* were extracted by acid glycine buffer and used in an enzyme immunoassay (EIA) and immunoblot (IB).

**Results**—56 of 144 (39%) patients with chronic liver diseases and six of 30 (20%) with PSC showed increased antibody concentrations in the *H hepaticus* EIA; in the *H pylori* EIA the numbers were 58% and 13% respectively. Compared with the healthy blood donors the antibody reactivity against the two helicobacter species was not increased (46% and 48% respectively). Patient serum samples retested by the *H hepaticus* EIA after absorption with sonicated *H pylori* cells remained positive in 12 of 37 (33%) serum samples. Distinct antibody reactivity to 55–65 kDa proteins was observed by *H hepaticus* IB, after the absorption step, and was considered specific for *H hepaticus*. These 12 serum samples were from patients with chronic alcoholic liver disease.

**Conclusions**—Antibodies to *H hepaticus*, often cross reacting with *H pylori*, occur frequently in patients with chronic liver diseases, with no clear cut relation to specific diagnostic groups. The pathogenic significance of these findings is not known.

Abbreviations used in this paper: CLD, chronic liver disease; EIA, enzyme immunoassay; ERC, endoscopic retrograde cholangiography; IB, immunoblot; PCR, polymerase chain reaction; PSC, primary sclerosing cholangitis; RAA, relative antibody activity.
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RAA value) greater than 35 and less than 25 g gammaglobulin, Pharmacia & Upjohn, Stockholm, Sweden) was added. The EIA test results are expressed as corrected mean absorbance values (A 405 nm) in percent of a reference standard (human serum). For absorption of potential cross-reacting antibodies, sonicated whole cells of *H* pylori (CCUG 17874) were used. To 1 ml of sonicated cells (A 540 of OD 1.0) 10 µl serum was added and incubated for one hour at 37°C during constant shaking. Cells were removed by centrifugation and supernatants collected for *Helicobacter* serology.

BACTERIAL STRAINS AND CULTURE CONDITIONS

*H* hepaticus, a murine liver isolate (strain CCUG 33637) was cultured on Fastidious agar (Amersham Pharmacia Biotech, Rainersh, UK), and grown for 7–10 days in a microaerophilic atmosphere (3% H₂, 10% CO₂, 5% O₂, 82% N₂) at 37°C. Agar plates were examined for colonies with a green metallic transparent surface, and tested positive for urease, catalase, and oxidase enzyme activities.

PREPARATION OF CELL SURFACE ANTIGENS FOR ENZYME IMMUNOASSAY AND IMMUNOBLOT

Ten agar plates (Ø5 cm) with confluent bacterial growth were harvested and washed once in 10 mM phosphate buffered saline (PBS) pH 7.2, with a total cell yield of 0.6 g wet weight. Cell surface antigens were extracted using a standard method with 0.2 M acid glycine buffer pH 2.2 as described previously. The protein concentration was 290 µg per ml as determined by the BioRad protein assay (BioRad, Richmond, UK). The protein profile of *H* hepaticus was compared with acid glycine extracted cell surface proteins of *H* pylori (CCUG 17874) by sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) followed by silver staining.

**H** hepaticus ENZYME IMMUNOASSAY (IgG-EIA)

The *H* hepaticus enzyme immunoassay (EIA) was performed as described for *H* pylori EIA. In brief, Maxisorp immunoplates (NUNC, Roskilde, Denmark) were coated with 100 µl per well with a protein concentration of 5 µg per ml; serum diluted 1/800 was added and the plates were incubated for 90 minutes at 37°C. An alkaline phosphatase goat antihuman antibody (Sigma, St Louis, Missouri, USA) was used as the second antibody. After repeated washings, 100 µl per well of a substrate solution containing *p*-nitrophenyl phosphate (Sigma), diethanolamine, and MgCl₂ was added. The EIA test results are expressed as corrected mean absorbance values (A 405 nm) in percent of a reference standard (human gammaglobulin, Pharmacia & Upjohn, Stockholm, Sweden). A relative antibody activity (RAA value) greater than 35 and less than 25 units was defined as positive and negative, respectively; values in between were regarded as low positive values (borderlines). The cut off values are based on EIA and immunoblot analyses of serum samples from patients with positive and negative gastric culture for *H* pylori, healthy blood donors, and children. Similar IgG EIA analyses were performed with acid glycine extracted cell surface proteins of *H* pylori.

**IMMUNOBLOT (IgG)**

In the immunoblot assay, acid glycine extracted cell surface proteins of *H* hepaticus were separated by SDS-PAGE with an 8–18% gradient (ExelGel, Pharmacia Biotech AB, Uppsala, Sweden) according to the Pharmacia manual. Proteins were electroblotted onto a PVDF membrane (Millipore, Intertech, Bedford, Massachusetts, USA), and subsequent blocking and probing with patient and control serum samples were performed as described previously. Similar immunoblot analyses were performed with acid glycine extracted cell surface proteins of *H* pylori.

**IgA IMMUNOBLOT**

As control for non-specific bacterial binding between proteins of *H* hepaticus/*H* pylori and IgA, an immunoblot was performed. Strips, containing separated cell surface proteins of the two helicobacter species were overlaid with human IgA antibodies (myeloma produced) and then incubated with an enzyme conjugated anti-IgA antibody followed by substrate staining. Another two strips were probed with serum samples from *H* hepaticus/*H* pylori EIA positive patients. Enzyme conjugated antihuman IgA antibodies (P216, Dako A/S, Glostrup, Denmark) were added and after incubation the strips were developed.

**IMMUNOSTAINING OF HEAT SHOCK PROTEINS (Hsp60)**

Immunoblot strips containing separated *H* hepaticus and *H* pylori cell surface proteins respectively were probed with a polyclonal antiserum to the *H* pylori Hsp60 protein. The Hsp60 protein was purified from *H* pylori strain CCUG 17874 by gel filtration (Superose 12, Pharmacia Biotech, Uppsala, Sweden) and ion exchange chromatography (MonoQ, Pharmacia Biotech), and then used for immunisation. Corresponding strips were incubated with an antihuman Hsp60 serum (Santa Cruz, Biotechnology Inc., USA). Immunoblot analyses were performed as described above.

**ABSORPTION EXPERIMENTS**

For absorption of potential cross reacting antibodies, sonicated whole cells of *H* pylori (CCUG 17874) were used. To 1 ml of sonicated cells (A 540 of OD 1.0) 10 µl serum was added and incubated for one hour at 37°C and for 16 hours at 6°C during constant shaking. Cells were removed by centrifugation and supernatants collected for *Helicobacter* serology.
Table 1 Outcome of the H hepaticus EIA and H pylori EIA in serum samples from patients with chronic liver disease (CLD) and primary sclerosing cholangitis (PSC)

<table>
<thead>
<tr>
<th></th>
<th>H hepaticus EIA positive* (%)</th>
<th>H pylori EIA positive* (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CLD (n=144)</td>
<td>56 (39)</td>
<td>79 (55)</td>
</tr>
<tr>
<td>PSC (n=30)</td>
<td>6 (20)</td>
<td>4 (13)</td>
</tr>
<tr>
<td>Controls† (n=48)</td>
<td>22 (46)</td>
<td>24 (48)</td>
</tr>
</tbody>
</table>

*Borderline values are included.
†Healthy blood donors.

n, number of serum samples.

Results

Protein profiles of acid glycine extracted cell surface proteins from H hepaticus were compared with a corresponding antigen preparation of H pylori after silver staining of the SDS-PAGE gel. Differences of the band pattern were observed with proteins migrating in the region of 43–94 kDa in molecular mass (M₀), whereas proteins with lower M₀ showed a similar pattern (fig 1).

IgG-EIA

Antibody reactivity to extracted H hepaticus cell surface proteins in serum samples from patients with chronic liver disease was initially analysed by EIA. Of 144 serum samples, 37 (26%) showed increased antibody concentrations (RAA greater than 35 units), 19 (13%) showed moderate values (RAA 25–35 units), and 88 (61%) were negative (table 1). The distribution of antibody reactivity in the diagnostic subgroups among H hepaticus EIA positive patients, including primary biliary cirrhosis and other autoimmune liver diseases and cryptogenic liver disease, did not differ from the whole patient population. In the H pylori EIA, 64 of 144 (44%) serum samples were positive and 15 serum samples gave borderline values (table 1). Of the 30 serum samples from PSC patients, six (20%) showed increased antibody concentrations, and 24 (77%) were negative in the H hepaticus EIA. In the H pylori EIA four serum samples were positive (table 1).

Serum samples from 48 healthy blood donors were assayed in the two EIAs and the

Table 2 Cross reactivity to cell surface antigens between the two helicobacter species

<table>
<thead>
<tr>
<th>Patients</th>
<th>Positive in H hepaticus EIA before absorption</th>
<th>Positive in H hepaticus EIA after absorption</th>
</tr>
</thead>
<tbody>
<tr>
<td>Controls*</td>
<td>24</td>
<td>0 (3†)</td>
</tr>
</tbody>
</table>

Cross reactivity to cell surface antigens between the two helicobacter species was analysed in 37 serum samples from the patients with chronic liver disease and in 24 from the controls. The serum was absorbed with sonicated whole cell material from H pylori and then retested.

*Healthy blood donors.
†Values just above the lower cut off.

seroprevalence was found to be 46% for H hepaticus and 48% for H pylori (table 1).

ABSORPTION EXPERIMENTS

Cross reactivity between cell surface proteins of the two helicobacter species was analysed with 37 serum samples from patients with chronic liver diseases showing high antibody concentrations in the H hepaticus EIA (14 of these were H pylori EIA negative) and with 24 blood donor serum samples found positive by H hepaticus EIA (three of these were H pylori EIA negative). In 25 patients the H hepaticus EIA became negative but 12 serum samples remained positive. These serum samples were from patients who suffered from alcoholic related liver disease. All blood donor serum samples became negative after the absorption step except three that showed low values close to the lower cut off level (table 2).

IMMUNOBLOT (IgG)

Patient serum samples absorbed with H pylori whole cells and with a remaining positive value in the H hepaticus EIA were further analysed by immunoblot, and the antibody reactivity was compared with results with the same serum samples that were not absorbed. In the H hepaticus immunoblot the unabsorbed serum samples intensively stained proteins of 55–65 kDa in M₀ (fig 2A). This was not observed in the immunoblot based on H pylori antigens (fig 2B). When these serum samples, absorbed with sonicated H pylori cells, were analysed in the H hepaticus immunoblot, reactivity to the 55–65 kDa proteins remained (fig 2C). However, when the serum samples were probed on strips containing separated H pylori proteins no reactivity was observed (figure 2D).

IMMUNOBLOT (IgA)

Patients with chronic alcoholic liver disease often show high serum IgA levels compared with healthy individuals. The immunoblots performed for control of a non-specific binding of bacterial cell material from H hepaticus and H pylori showed no visible IgA reactivity (data not shown).

IMMUNOSTAINING OF Hsp60

By immunoblot the polyclonal anti-H pylori Hsp60 serum stained two major bands with an approximate M₀ of 62–65 kDa with the two helicobacter species. The antihuman Hsp60 serum weakly stained a 65 kDa protein of H pylori, and with H hepaticus no bands were observed (data not shown).
Antibodies against *H. hepaticus* and *H. pylori* were detected in serum samples from patients with various liver diseases. The frequency of antibody reactivity was, however, not increased in any specific diagnostic groups and serum positivity was not increased in patients with chronic liver diseases compared with the healthy individuals. Some of these antibodies may represent cross reactivity between the two species, while a specific antibody response to cell surface proteins of *H. hepaticus* was detected in serum samples from patients with alcoholic related liver disease.

Several new species of the genus helicobacter have recently been identified in various mammals, with some species inducing gastrointestinal infection while others seem to colonise the gastrointestinal tract without causing infection and tissue inflammation. Recent reports on multifocal hepatitis in dogs induced by bile tolerant helicobacter, suggested that helicobacter organisms with liver tropism may also cause hepatitis in humans and other primates. However, up to now no human liver bile duct isolates of helicobacter have been reported although bile samples were shown to contain *H. pylori* DNA. In susceptible strains of mice, *H. hepaticus* causes an acute focal hepatitis, which progresses to chronic hepatitis with the presence of relatively few infectious organisms. Franklin and colleagues reported on a novel helicobacter species, *H. cholecystus*, isolated from the bile tract of Syrian hamsters with chronic gall bladder inflammation and pancreatitis. Interestingly, Siringo and colleagues reported a high prevalence of *H. pylori* infection in cirrhotic patients, which was not confirmed by Tsai. Thus, it is possible that not yet isolated urease positive as well as urease negative helicobacter species may be able to colonise the human intestinal tract, as recently described in dogs and cats. Fox and colleagues concluded that helicobacter might induce a chronic gall bladder inflammation in humans with late sequelae such as bile tract and gall bladder malignancies, and a potential role of *H. hepaticus*, *H. bilis*, and other bile tolerant helicobacter species in chronic cholecystitis and liver disease. The importance of further studies on the possible role of these new helicobacter species in human liver disease was emphasised as was the need to develop non-invasive serological assays to determine the seroprevalence of hepatic helicobacter infections.

It is assumed that bile tolerant helicobacter species are antigenically and genetically similar. Our findings in this study, showing 12 patient serum samples with high antibody reactivity to cell surface proteins of *H. hepaticus* even after absorption with *H. pylori*, indicate that some helicobacter-like organism may be involved in human liver diseases. By immunoblotting, antibodies mainly reacted to the 55–65 kDa proteins and at least two of these
proteins were recognised by an anti-\textit{H pylori} Hsp60 antibody, suggesting that patient serum samples intensively staining the 60–65 kDa proteins of \textit{H hepaticus} contained increased concentrations of antibodies to helicobacter Hsp60 proteins (fig 2A). Hsps are highly conserved, immunogenic molecules induced by heat, inflammatory mediators, and other forms of physiological stress\textsuperscript{5}; ethanol might be such a factor. The pathogenic mechanism of this finding is unknown, but it is a fact that ethanol abuse induces a local immune defect\textsuperscript{11}. Ward and colleagues\textsuperscript{26} found that antibodies to H. pylori in patients with chronic active hepatitis C were associated with a higher prevalence of cirrhosis. This study was supported by grants from the Swedish Medical Research Council and the Forssman Foundation. Part of this study was reported at EASL, Lisbon, April 1998 (abstract no. C11/015) and at the IXth International Workshop on Gastrointestinal Pathology and Helicobacter pylori, Budapest, 2–5 September 1998 (Gut 1998;suppl 2):45, abstract no. 14/402).