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Gadolinium Contrast Media are More Nephrotoxic Than a Low Osmolar Iodine Medium Employing Doses With Equal X-Ray Attenuation in Renal Arteriography: An Experimental Study in Pigs

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Rationale and Objectives. To investigate in a unilaterally nephrectomized porcine model whether gadolinium contrast media (Gd-CM) are less nephrotoxic than iodine media (I-CM) in x-ray arteriography of a kidney made temporarily ischemic by arterial balloon occlusion.

Materials and Methods. In a noncrossover design, 3 mL of each test solution were injected in eight pigs (mean weight 19 kg) at a rate of 20 mL/min into the right renal artery at the start of a 10-minute period of ischemia. In group 1 (40 pigs) we injected 0.5 M gadopentetate, 0.5 M gadodiamide, 0.5 M iohexol (190 mg I/mL), 0.18 M iohexol (70 mg I/mL; with an x-ray attenuation equal to that of 0.5 M Gd-CM at 80 kV), and saline. In group 2 (24 pigs), we tested 0.18 M iohexol with ischemia and saline with and without ischemia. Gd- and iodine contrast media functioned as markers of glomerular filtration rate (GFR). When saline was tested, a low dose of iohexol (3 mL per pig; 300 mg I/mL) was injected as GFR marker intravenously in group 1 and into the renal artery in group 2. The plasma half-life elimination times of the CM 1–3 hours after injection were used to compare the effects of the different test solutions on GFR. Longer half-life means lower GFR.

Results. Group 1: median plasma half-life elimination time of the GFR marker was 3 340 minutes after injection of 0.5 M gadopentetate, 256 after 0.5 M gadodiamide, 179 after 0.5 M iohexol, 143 after 0.18 M iohexol, and 133 minutes after saline. All differences except that between 0.18 M iohexol and saline were statistically significant (P < .01). Group 2: median plasma half-life was 174 minutes after 0.18 M iohexol with ischemia, 196 minutes after saline with ischemia, and 195 minutes after saline without ischemia. There were no significant differences between the test solutions in group 2 (P > .05).

Conclusion. In pigs, 0.5 M Gd-CM were more nephrotoxic than both equal-attenuating (70 mg I/mL) and equimolar (190 mg I/mL) concentrations of the I-CM iohexol. These results do not support the “off-label” use of Gd-CM for renal x-ray arteriography in man instead of commercially available concentrations of iodine contrast media at 140, 150 and 180 mg I/mL or diluted to 70 mg I/mL.

Key Words. Angiography; contrast media; toxicity; kidney; failure; gadolinium.
and vascular interventions, which all require high doses of I-CM. Many of these procedures are performed in elderly patients with decreased renal function and diabetes. Thus it is necessary to adopt strategies to decrease the risk of CMN.

One strategy to decrease this risk has been to exchange I-CM to other CM such as carbon dioxide (4,5) or gadolinium CM (Gd-CM) (6–11). The assumed lower nephrotoxic effect of Gd-CM is partly based on one study (8) in which intraarterial and intravenous doses of I-CM for x-ray arteriography (XRA) and computed tomography, respectively, proved more nephrotoxic than Gd-CM for magnetic resonance angiography in the same patients. However, the two CM were compared at an extremely different x-ray attenuation capacity. Thus the “off-label” recommendation to use Gd-CM as a safer alternative than I-CM in azotemic patients in a variety of XRA examinations—including aortofemoral (12,13), renal (14–17), carotid (18), and coronary arteriography (19,20)—is, in our opinion, based on an incorrect comparison. The only reason for using I-CM or Gd-CM in XRA is to exploit their ability to attenuate x-rays. A recent review dealing with the toxicity of these agents and their ability to attenuate x-rays indicated that I-CM would probably show a lower renal toxicity than Gd-CM if compared in equal volumes in equal-attenuating concentrations (21). However, no such studies have been performed.

The aim of the present investigation was therefore to answer the following question: “Do Gd-CM really have a lower renal toxicity than a commonly used low-osmolar I-CM when compared in equal volumes of equal-attenuating concentrations during renal arteriography?” To answer the question the nephrotoxic effect of a nonionic low-osmolality I-CM (iohexol) was compared with two 0.5 M Gd-CM with different osmolalities (gadopentetate and gadodiamide) during arteriography of ischemic kidneys in pigs.

### MATERIALS AND METHODS

#### Animals

Sixty-four Swedish landrace male pigs (Swedish University of Agricultural Sciences), with a mean weight of 19 kg (range 16–23 kg) were used. The study was approved by the local ethical committee. The pigs were acclimatized at the Experimental Department, Malmö University Hospital, for 4–6 days before surgery. They had free access to food and water until 15 hours before the experiment and thereafter only free access to water. After the experiment, all animals were sacrificed by an overdose of pentobarbital natrium and 96% ethanolum (Pentobarbital 100 mg/mL, Apoteket Produktion och Laboratorier, Gothenburg, Sweden).

#### Anesthesia and Surgery

The pigs were premedicated with 8–10 mL ketamine (Ketalar 50 mg/mL, Warner Lambert Nordic AB, Parke-Davis, Solna, Sweden) intramuscularly 15 minutes before the anesthesia, which was induced with an intravenous (IV) injection of 0.5 mL/kg bispotential sodium (25 mg/mL; Pentothal natrium, Abbott, Abbott Park, Ill). After tracheal intubation, the pigs were connected to a volume controlled respirator (Servo-Ventilator 900, Elema-Schönander, Sweden) delivering a gas volume mixture of 70% nitrogen and 30% oxygen. The rectal temperature was continuously registered and kept close to 38°C by use of a heating pad. Surgical anesthesia was initiated and maintained by continuous IV infusion of 0.4–0.6 mL/min of a mixture of 10 mL midazolam (Dormicum 5 mg/mL, F. Hoffmann-La Roche AG, Basel, Switzerland), 40 mL ketamine (Ketalar 50 mg/mL, Park Davies, Barcelona, Spain), and 20 mL vecuronium bromide (Norcuron 2 mg/mL, Organon Teknika, Netherlands). During the entire experiment Ringers solution (2.5 mL/min; Ringer acetate, Fresenius Kabi, Uppsala, Sweden) was given.

The right femoral artery was dissected free and an 8 Fr introducer (Medikit Co., Ltd, Tokyo, Japan) was inserted and connected to a pressure transducer for continuous recording of the arterial pressure (Model MP 100, Biopac Systems Inc., Santa Barbara, Ca). Intraarterial pH, oxygen pressure, and carbon dioxide pressure (AVL Critical Care Analyzers, AVL Scientific Corp., Roswell, Ga) were also monitored during the experiment. A 14 Fr introducer was inserted into the right internal jugular vein for injection of heparin (5000 IU/mL, Leo, Denmark) and blood sampling. A bolus dose of heparin (30 IU/kg) was given and the introducers were flushed with heparinized saline (5000 IU in 1000 mL of saline) to prevent clotting.

A left-sided nephrectomy was performed through a subcostal incision. After surgery a 60-minute calibration period was used to monitor the pigs for hemodynamic stability. A 4 Fr Cobra catheter (Cordis, Oostende, Netherlands) was inserted via the right femoral artery introducer into the right renal artery. The cathe-
The test solution, their doses, concentrations, and osmolalities are given in Table 1. The viscosity of 0.5 M gadopentetate (Magnevist, Schering AG, Berlin, Germany) is 2.9 mPas, gadodiamide (Omniscan, Amersham Health AS, Oslo, Norway) 1.9 mPas, and iohexol Omnipaque (Amersham Health AS) 140 mg I/mL 1.5 mPas at 37°C, according to the manufacturers. The viscosity of the noncommercially available concentrations of iohexol at 70 and 190 mg I/mL have not been measured.

**Group 1**

The nephrotoxic effect of 2 Gd-CM, 0.5 M gadopentetate, and 0.5 M gadodiamide were compared with that of (1) an equimolar concentration of 0.5 M iohexol (190 mg I/mL), (2) 0.18M iohexol (70 mg I/mL), roughly equal-attenuating with 0.5M Gd-CM at about 80 kV (21), and (3) saline. Each pig was subjected to only one of the test solutions, which was injected in eight animals at a dose of 3 mL/kg and a rate of 20 mL/min. The test solutions were randomly assigned to a total number of 40 pigs. All injections were made at the beginning of the 10-minute period of renal artery occlusion. In the animals
subjected to saline, a smaller total dose of 3 mL iohexol (300 mg I/mL) was injected intravenously as a glomerular filtration rate (GFR) marker.

**Group 2**

The nephrotoxic effect of 0.18 M iohexol (70 mg I/mL) with ischemia was compared with that of saline with ischemia and saline without ischemia. The same technique was used as in group 1 except that each dose of saline was enriched with 3 mL iohexol (300 mg I/mL) as the GFR marker. Each of the three procedures was performed in 8 animals and was randomly assigned to a total number of 24 pigs.

**Plasma Half-Life of the Contrast Media**

The pharmacokinetic properties of Gd-CM and I-CM are very similar, both demonstrating rapid extracellular distribution. Their body clearance from plasma is dominated by glomerular filtration with no significant tubular excretion or reabsorption (22–26). Hence, both types of CM function as markers of GFR and may as such be used to evaluate their own nephrotoxic effects (ie, their possible depression of GFR). The larger a CM-induced depression of GFR is, the lower the body clearance of the GFR marker (the contrast medium) from plasma will be, with subsequent prolongation of its plasma half-life elimination time. The plasma half-life of the contrast media was calculated by using a one-compartment model and linear regression analysis of logarithmic plasma concentrations (22) in the period 1–3 hours after injection.

**Determination of CM Concentrations in Plasma**

Ten blood samples of 4 mL each (LH tubes, Sarstedt Aktiengesellschaft & Co, Germany) were drawn 15, 30, 45, 60, 75, 90, 105, 120, 150, and 180 minutes after the start of test solution administration. The samples were centrifuged at 4000 r/min. The plasma was used to determine the concentration of the contrast media using x-ray fluorescence technique (92X-II Spectrum Master Gamma-Ray Spectrometer; EG & G Ortec, Tenn) (27–29). The spectrometer consists of four principal ingredients: the radiation source, the sample, the detector, and the multichannel analyzer.

The radiation source is an americium-241 isotope, which is used to irradiate the plasma samples with 59.5 keV photons. These photons excite the K-shell electrons of iodine or gadolinium atoms, so that the atoms enter an excited state. The ejected K-shell electrons are immediately replaced by L-shell electrons. Thus the excited atoms return to their original energy level and emit x-rays with energies characteristic for iodine or Gd. The number of characteristic photons registered by the detector is proportional to the amount of iodine or Gd atoms in the sample. The plasma samples, 500 µL each, were measured during 300 seconds; the analysis was repeated three times and the resulting mean value calculated.

**Radiograms**

Three radiographic images using a C-arm equipment (BV 300 PLUS, Philips, Holland) were obtained 3 (end of injection), 10 (end of ischemic period), and 30 minutes after the start of injection. The x-ray kilovoltage were set to 70 kV. In each animal the radiograms were evaluated by two of the authors (UN, TA), who were blinded to the CM used. The radiograms were compared in a semiquantitative fashion (ie, if the nephrographic radiodensity appeared similar or significantly different and if excretion to the collecting system was present or not).

**Statistics**

To evaluate any statistically significant nephrotoxic differences among the various test solutions, as indicated by any differences in plasma half-life elimination of the CM GFR-markers, the nonparametric Mann-Whitney U-test was used. P values of .05 or less were considered statistically significant.

**RESULTS**

**Plasma Half-Life Elimination Time**

The results are summarized in Fig 1 and 2.

**Group 1**

Pigs injected into the renal artery with saline got a median plasma half-life elimination time of 133 minutes of the GFR marker (range 92–167). The median plasma half-life elimination time after injection of 0.18 M iohexol (70 mg I/mL) was 143 minutes (range 121–160); the small difference compared with saline was not statistically significant. All other test solutions caused a statistically significant (P < .01) longer plasma half-life elimination time of the GFR marker than that caused by saline or 0.18 M iohexol. Compared with saline, 0.5 M iohexol (190 mg I/mL) increased median plasma half-life of the GFR marker by 35% (median time 179 minutes, range 158–211), 0.5 M gadodiamide by
92% (median time 256 minutes, range 179–393), and 0.5 M gadopentetate 25 times (median time 3,340 minutes, range 1,008–H11009). The difference in plasma half-life between 0.5 M gadodiamide and 0.5 M iohexol (190 mg I/mL) was also statistically significant (P < .01), as was the difference in plasma half-life between gadopentetate and gadodiamide (P < .001).

**Group 2**

Again, there was no statistically significant difference between 0.18 M iohexol (70 mg I/mL; median time 196 minutes, range 162–316) and saline with ischemia (median time 196 minutes, range 167–316) regarding plasma half-life of the GFR markers. Nor was there any statistically significant difference between 0.18 M iohexol (70 mg I/mL) with ischemia and saline without ischemia (median time 195 minutes, range 127–252).

**Radiograms**

Three and ten minutes after the start of CM injection, the use of 0.18 M iohexol (70 mg I/mL), gadodiamide, and gadopentetate resulted in a similar nephrographic attenuation, whereas 0.5 M iohexol (190 mg I/mL) cause a markedly higher radiodensity. Thirty minutes after the
start of CM injection, excretion of the CM was visualized and there was no persisting nephrogram except for the animals that had received gadopentetate, which showed a persistent nephrogram (Fig 3).

**DISCUSSION**

The present investigation was focused on comparing the nephrotoxicities of equal-attenuating concentrations of Gd-CM and I-CM, which in equal volumes were injected into the renal artery. The concentration at which I-CM are equal-attenuating with 0.5 M Gd-CM may vary with the x-ray tube kilovoltage (kV) used (30), which, in turn, may depend on patient size. We therefore used a concentration of a commonly used nonionic monomeric I-CM causing roughly the same attenuation of x-rays as an 0.5 M Gd-CM at the commonly used 70–90 kV for XRA (ie, 0.18 M solution of iohexol [70 mg I/mL]) (21). We also used an 0.5 M iohexol solution (190 mg I/mL), which contains the same number of CM molecules as 0.5 M Gd-
CM, but three times the number of attenuating atoms. Thus at 80 kV the attenuation of 0.5 M iohexol is almost three times that of 0.5 M Gd-CM. Iohexol (0.5 M) should also have a higher attenuation than 0.5 M Gd-CM at 96 kV, considered the optimal kV for Gd-CM (31), and at any higher used kV (110–120 kV) for angiography according to experimental results comparing the physical performances of different x-ray contrast agents (30,32).

The results of the present study support our previous assumption (21) that a plasma-isotonic solution (0.29 osmol/kg water) of a “low osmolality” 0.18 M I-CM is significantly less nephrotoxic than roughly equal-attenuating (at 80 kV) concentrations and doses of plasma-hypertonic solutions of two Gd-CM, 0.5 M gadodiamide (1.96 osmol/kg water), and gadopentetate (1.96 osmol kg/water) when injected into the renal artery. Iohexol 70 mg I/mL appeared to have no deleterious effect on renal function compared with saline. Although gadodiamide (2.7 times the osmolality human plasma) almost doubled the plasma half-life of the GFR marker compared with iohexol 70 mg, gadopentetate (7 times the osmolality of human plasma) appeared to cause an almost complete cessation of renal function.

According to some advocates, 0.5 M Gd-CM (31) is equal-attenuating with I-CM in the 75–150 mg I/mL range when an x-ray tube voltage of 96 kV is applied. Applying the low-osmolality I-CM iohexol at an even higher iodine concentration (190 mg I/mL), which contains the same concentration of potentially nephrotoxic CM molecules as the 0.5 M Gd-CM, still resulted in a significantly shorter elimination half-life of the GFR marker than gadodiamide and gadopentetate in the present investigation. The osmolality of 0.5 M iohexol is only half that of 0.5 M gadodiamide (ie, 0.4 osmol/kg water). Thus, even in equimolar solutions, I-CM seem to cause less effect on renal function and at the same time improve diagnostic potentials through better radiographic contrast at all practically applied kilovoltages for angiography.

Determination of GFR as body clearance could not be calculated in the present experiment because the contrast media serving as GFR markers were injected directly into the renal arteries in most animals. Hence, an unknown fraction of the GFR markers never appeared in the general circulation but was filtered into the urine during their first pass through the kidneys. The amount of CM that escaped from the circulation through the glomerula into the renal tubules during the time of ischemia was not possible to account for and may have varied between different substances because of different degrees of renal injuries. These factors should not affect the plasma CM half-life elimination time, because calculations of CM half-life are only dependent on relative changes in plasma concentration of CM over time. Therefore, calculated CM plasma half-lives in the present study have been used as an indicator of any difference in renal toxicity (ie, effect on GFR) of the various test solutions when calculated as body clearance. The volume of distribution (Vd) in the body of the various CM must be known or be reasonably estimated for calculation of body clearance (GFR) according to the formula in the appendix (33).

Vd could not be calculated in the present experiment for the two Gd-CM because they were only injected selectively into the renal artery with an unknown fraction escaping the general circulation. In spite of that, we think it is possible to make a reasonable comparison between the three CM with regard to their effect on total body clearance (representing GFR) according to the formula.

We therefore conclude that, in our experimental model, Gd-CM caused more renal damage (reduction of body clearance) than the I-CM. This conclusion supports the position of the producers of Gd-CM that applying Gd-CM for XRA implies an “off-label” use. Our results regarding the effect on renal function seems to be in accordance with another series of experiments (34,35). In an ischemic rat model it was found that intra-aortic injections of gadopentetate and diatrizoate, both with an osmolality of about 2.0 osmol/kg water, caused a similar decrease in GFR measured as creatinine clearance. However, the dose of iodine atoms appeared to have been 10 times higher than that needed to achieve the same attenuation as the Gd-CM during an XRA study at 80 kV (21). Thus if equal-attenuating doses had been used it seems reasonable to assume that the nephrotoxic effect of diatrizoate had been much less.
Radiographs of gadopentetate showed, unlike the other CM, a persistent nephrogram at 30 minutes after injection. It has been pointed out that a persistent nephrogram after administration of intravascular contrast media not only may be the result of a preexisting acute renal failure, but also may indicate renal impairment developed during the x-ray examination and caused by the applied CM, as in our pigs subjected to gadopentetate (36,37).

Several experimental animal models have been used to study CM-induced nephrotoxicity (1,34,35,38–40). We designed a porcine model with a high probability of CM nephrotoxicity by injecting high doses of CM selectively into the renal artery during a period of ischemia. The model was partly chosen because the use of Gd-CM has been advocated to guide treatment of renal artery stenoses with balloon angioplasty or stent placement (14–17), and such procedures will imply periods of renal ischemia. We also choose the pig because we were familiar with catheterization of porcine renal arteries from interventional training courses and because their kidneys are more like human kidneys with regard to anatomy and physiology than the kidneys of most other species (40–43). In an initial pilot study, we did not notice any effect on renal function when the CM were injected into patent renal arteries as had been found in an earlier canine study from our laboratory (44). Therefore, an ischemic period was added to the experiment, a well-established procedure to potentiate the nephrotoxic effect of CM (35,40,45,46). It is well known from both clinical and experimental studies that prior preconditioning is often necessary to make living species susceptible to CM-induced renal failure. Transient ischemia is only one of many models of experimental preconditioning (47,48).

The pathophysiologic mechanisms behind the noted effects on glomerular function in the present model may be multifactorial and dependent on chemotoxic and osmotic effects of the CM in combination with ischemia. It is noteworthy that the iso-osmotic solution of iohexol 70 mg I/mL with 10 minutes of renal ischemia had no effect on plasma half-life elimination time of GFR markers—different from that of saline without ischemia. This may indicate that the 10-minute ischemic period per se did not immediately affect GFR.

The effect of balloon occlusion may instead of ischemia rather be due to prolongation of contact time between the more concentrated and hyperosmolar CM and red blood cells, vascular endothelial, and tubular cells. This may result in marked shrinkage and rigidification of red blood cells obstructing microcirculation (49–51) and vascular endothelial injuries resulting in thrombotic occlusions (52,53). A nonosmolality-dependent release of vasoconstrictors has also been postulated (1). A direct toxic effect on tubular cells might also be possible (54).

Osmotic factors have been considered to play a significant role in the development of CMN (1) and may explain some of the differences noted between the various solutions in the present study. All hypertonic solutions significantly increased elimination half-life, increasing with the osmolality of the agents. Apart from the direct exposure of red blood cells, endothelial cells, and tubular cells to the hypertonic solutions injected into the renal artery, the CM also represent an osmotic load to the kidneys with secondary diuresis and natriuresis (1,55,56). Osmotic load may activate the tubuloglomerular feedback mechanism mediating vasoconstrictive agents, increase medullary oxygen demand for active reabsorption of the increased tubular sodium load resulting in hypoxic injuries, or increase intratubular pressure with elevation of interstitial pressure and secondary vascular compression beneath the rigid renal capsule.

There are also clinical studies (2,3,57) indicating that the use of CM causing a lower osmotic load on the kidneys will reduce the risk of the CMN in azotemic patients. When low-osmolality CM (eg, iohexol) was compared with high-osmolality CM (eg, diatrizoate) the CM representing the lower osmotic load reduced the risk of CMN (2,3). In a recent study comparing iohexol with the iso-osmolar ioxitalam, a nonionic dimer representing an osmotic load still lower than that of iohexol, iodixanol caused less deterioration of renal function (57). Gadopentetate has an osmolality similar to that of diatrizoate 370 mg I/mL (76%) and gadodiamide the same as iohexol 350 mg I/mL. Thus, when equal volumes of the CM in the present study are compared, the Gd-CM represented a significantly higher osmotic load to the porcine kidneys than the iohexol solutions used (ie, 190 and 70 mg I/mL).

**CONCLUSION**

To the best of our knowledge the present investigation represents the first comparison, including both experimental and human studies, between Gd- and I-CM regarding their nephrotoxic effects during XRA using equal-attenuating and equimolar concentration and doses. The results do not support the “off-label” use of Gd-CM for XRA in man instead of commercially available concentrations of
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APPENDIX

Formula regarding the relation between total body clearance, volumes of distribution, and plasma half-life: Total body clearance (mL/min) = ln2 × Vd/plasma half-life.

In, natural logarithm; Vd, volume of distribution in mL; T, half-life elimination time (minutes) of GFR marker.