Global investigation and meta-analysis of the C9orf72 (G4C2)n repeat in Parkinson disease


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**ABSTRACT**

**Objectives:** The objective of this study is to clarify the role of (G\(_{4}C_{2}\))\(_{n}\) expansions in the etiology of Parkinson disease (PD) in the worldwide multicenter Genetic Epidemiology of Parkinson’s Disease (GEO-PD) cohort.

**Methods:** C9orf72 (G\(_{4}C_{2}\))\(_{n}\) repeats were assessed in a GEO-PD cohort of 7,494 patients diagnosed with PD and 5,886 neurologically healthy control individuals ascertained in Europe, Asia, North America, and Australia.

**Results:** A pathogenic (G\(_{4}C_{2}\))\(_{60}\) expansion was detected in only 4 patients with PD (4/7,232; 0.055%), all with a positive family history of neurodegenerative dementia, amyotrophic lateral sclerosis, or atypical parkinsonism, while no carriers were detected with typical sporadic or familial PD. Meta-analysis revealed a small increase in risk of PD with an increasing number of (G\(_{4}C_{2}\))\(_{n}\) repeats; however, we could not detect a robust association between the C9orf72 (G\(_{4}C_{2}\))\(_{n}\) repeat and PD, and the population attributable risk was low.

**Conclusions:** Together, these findings indicate that expansions in C9orf72 do not have a major role in the pathogenesis of PD. Testing for C9orf72 repeat expansions should only be considered in patients with PD who have overt symptoms of frontotemporal lobar degeneration/amyotrophic lateral sclerosis or apparent family history of neurodegenerative dementia or motor neuron disease.

**GLOSSARY**

ALS = amyotrophic lateral sclerosis; FTLD = frontotemporal lobar degeneration; GEO-PD = Genetic Epidemiology of Parkinson’s Disease; PD = Parkinson disease; RP = repeat-primed; STR = short tandem repeat.

A substantial number of patients with frontotemporal lobar degeneration (FTLD)/amyotrophic lateral sclerosis (ALS) (14%–35%) carrying C9orf72 (G\(_{4}C_{2}\))\(_{>60}\) expansions\(^1\)–\(^3\) present with atypical parkinsonism in early disease stages and increased incidence of parkinsonism with or without features of the FTLD/ALS complex in their relatives.\(^4\)–\(^9\) Ten research groups have reported on C9orf72 repeat expansions in Parkinson disease (PD) or atypical parkinsonism patients\(^10\)–\(^19\) but none of these investigated the C9orf72 repeat in large-scale cohorts, and European and Australian populations were underrepresented in the published data. Apart from the pathogenicity of (G\(_{4}C_{2}\))\(_{>60}\) expansions, we provided in vitro evidence that the (G\(_{4}C_{2}\))\(_{2}\) repeat size negatively correlated with the transcriptional activity of the C9orf72 promoter.\(^20\) Hence, it is conceivable that an increasing number of C9orf72 repeats may affect transcription gradually and increase susceptibility to disease.\(^20\) Three studies indicated a role of C9orf72 repeats in PD susceptibility but associations were found using different dichotomizations of repeat length, muddling biological interpretation. In one study, a marginal increased risk of PD was observed for carriers of (G\(_{4}C_{2}\))\(_{10}\) repeats.\(^12\) In the second, a significant increased frequency of (G\(_{4}C_{2}\))\(_{>20}\) repeats was observed in patients clinically diagnosed with PD.\(^13\) In the third study, the authors

*These authors contributed equally to this work.

Authors’ affiliations are listed at the end of the article.

GEO-PD Consortium coinvestigators are listed on the Neurology® Web site at Neurology.org.

Go to Neurology.org for full disclosures. Funding information and disclosures deemed relevant by the authors, if any, are provided at the end of the article. The Article Processing Charge was paid by research funding obtained by the principal investigator.

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reported association of \((G_4C_2)_\geq7\) repeats with PD in the Chinese Han population.\(^{16}\) All of these studies, however, were executed in ethnically distinct and medium scaled cohorts. We set out to clarify the role of the \(C9orf72\) \((G_4C_2)_n\) repeat in PD etiology in the first global multicenter study cohort of more than 7,000 patients with PD of 12 nationalities and 4 continents. First, we assess the global prevalence of pathogenic \((G_4C_2)_\geq60\) expansions. Second, the size of the combined study populations enables a detailed investigation of the specific \(C9orf72\) repeat allele or size threshold associated with increased risk of PD.

**METHODS** Standard protocol approvals, registrations, and patient consents. Genetic studies applied in this research were approved by the ethics committees of the ZNA (Hospital Network Antwerp), the Antwerp University Hospital, and University of Antwerp. Clinical protocols were approved by the ethics committees of the ZNA, the Antwerp University Hospital, and local ethical review boards of the participating research centers. All human biological samples were collected, fulfilling ethical approvals, and used in accordance with the terms of subjects’ written informed consent.

Participants. The Genetic Epidemiology of Parkinson’s Disease (GEO-PD) Consortium includes investigators from 60 sites from 30 countries and 6 continents (http://www.GEO-PD.org/about/). All sites were invited to participate in this study. A total of 18 sites representing 12 countries and 4 continents contributed either DNA or genotypic data, and clinical data of in total 15,123 individuals (tables 1 and 2). After thorough quality control as described in the procedures section below, 13,669 samples were included in this study. We excluded all duplicate samples, sex mismatches, and samples that failed in the DNA fingerprint analysis because of low quantity or quality of DNA or because of contamination of the sample. Demographics and diagnostic criteria of each series included in this study and the sample size breakdown from each site are provided in table 2. Controls were collected at the local sites as demographically matched neurologically healthy individuals.

**Procedures. Sample quality control.** Concentration and purity were checked spectrophotometrically using the Trinean DropSense96 UVVIS droplet reader (Trinean, Gentbrugge, Belgium) for all consortium genome DNA samples. Sex and DNA fingerprint were determined for all samples using an in-house-developed multiplex PCR panel composed of 13 short tandem repeat (STR) markers distributed over multiple autosomal locations: D20S480, D22S1174, D3S1287, D3S1744, D5S1764, D7S872, D7S2426, D8S1764, D14S1005, D20S866, D10S1237, D20S912, and D6S965. This panel also includes a marker specific for the X chromosome (DXS1187) and one for the SRY gene on the Y chromosome, and enables fast and accurate sample identification and sex determination in a single assay. After selective amplification of 20 ng genomic DNA, amplification products were size separated on an ABI 3730 automatic sequencer (Applied Biosystems, Foster City, CA) using GeneScan-600 LIZ (Applied Biosystems) as internal size standard and genotypes were assigned using in-house–developed TracI genotyping software (http://www.vibgeneticserivcefacility.be).

**Genetic analyses.** To screen the GEO-PD cohorts for the pathogenic \((G_4C_2)_\geq60\) \(C9orf72\) repeat expansion, we designed a 2-step procedure: an STR fragment length assay with flanking PCR primers optimized for alleles with high GC content (STR-PCR) followed by 2 repeat-primed PCR assays (forward and reverse RP-PCR) as described earlier.\(^{1,20}\)

Four participating research groups performed the genotyping in their local facilities according to previously published procedures.\(^{2,17}\) For consistent allele scoring of repeat lengths between GEO-PD groups and accurate interpretation of the repeat length, we designed a reference DNA set of 14 samples covering a range of normal repeat sizes that was genotyped by each of these facilities. Furthermore, for 2 of the cohorts, a random set of samples homozygous for the STR-PCR assay were included in the RP-PCR analysis at the Antwerp site for independent validation of the absence of a pathogenic repeat expansion.

**Statistical analyses.** To investigate the association between repeat units and PD susceptibility, explorative approaches were followed, based on (1) allele counts of the distinct repeat sizes, to determine whether one or more specific repeat sizes were associated with PD, (2) the total number of repeat units (sum of both alleles) per individual, and (3) the size of the longest repeat per individual (maximum allele). Summary statistics were computed in a random-effects meta-analysis (DerSimonian-Laird)\(^{21}\) for each approach in the rmeta package implemented in the R environment version 2.15.3. Based on the results obtained in the above-mentioned analyses, we performed hypothesis-driven dichotomized genotypic meta-analyses. Details are provided in the e-Methods on the Neurology® Web site at Neurology.org. To take into account the number of tests performed (\(n = 22\)), a Bonferroni-corrected 2-sided \(p\) value of \(\leq0.002\) was considered statistically significant. Population attributable risk of \((G_4C_2)_10\), \((G_4C_2)_20\), and \((G_4C_2)_\geq77\) was estimated using the epis package in R. For the meta-analyses, only the cohorts including both the patients with PD and the controls that were consistent with \(p\) \(<0.002\) were included for each analysis.
### Table 2 Characteristics of the GEO-PD cohorts included in the study

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<th>Ethnicity</th>
<th>Controls</th>
<th>Patients</th>
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<tr>
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<tr>
<td>% Familial</td>
<td>24</td>
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<tr>
<td>AAO ± SD</td>
<td>63.8 ± 13</td>
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</table>

Diagostic criteria for PD

- Gelb
- UKPDBB
- Ward and Gibb
- Hoehn and Yahr
- UK Parkinson’s Disease Brain Bank.

**Abbreviations:** AAO = age at onset; GEO-PD = Genetic Epidemiology of Parkinson’s Disease; NA = not available; PD = Parkinson disease; PI = principal investigator; QC = quality control; STR = short tandem repeat; UKPDBB = UK Parkinson’s Disease Brain Bank.

At the initial quality control step (QC), we excluded all duplicate samples, sex mismatches, and samples that failed in the DNA fingerprint analysis because of low quantity or quality of DNA or because of contamination of the sample. Additional samples did not pass the 2-step genetic analysis because of DNA shortage or limited concentration of the DNA sample.
size-corrected based on the reference panel were included in the study.

RESULTS Definite pathogenic C9orf72 repeat expansions in PD. A total of 12,710 samples, including 7,232 patients with PD and 5,478 control individuals, were successfully genotyped with the 2-step \((G_4C_2)_n\) repeat genotyping assay. RP-PCR analysis revealed the typical sawtooth tail pattern indicative of a pathogenic repeat expansion \((G_4C_2)_{>60}\) in one German (MS_RK cohort) (1/1,304; 0.08%) and 3 French (SL cohort) (3/1,182; 0.25%) patients but none in the other GEO-PD patient cohorts (table 1). Based on these results, we calculated a prevalence of pathogenic C9orf72 repeat expansions in this global consortium cohort of 0.06%.

The German patient was diagnosed with idiopathic PD at the age of 57 years. One year after onset, clinical examination revealed hypomimia, hypokinesia, resting tremor of the right arm, minor postural instability, mild bilateral rigidity, and slowed shuffling gait but also short-term memory disturbances, social withdrawal, and minor apathy. The patient had a positive family history of neurodegenerative dementia. All 3 French patients were diagnosed with PD without cognitive dysfunction at disease onset, and a detailed clinical description has been reported previously. Briefly, the first patient developed left hemiparkinsonism at age 29 years and symptoms worsened progressively while dopamine agonists were only partially effective. In the second patient, parkinsonism began at age 48 years and a cognitive decline was noted at age 56 years. The third patient developed parkinsonism at age 64 years and developed a mild cognitive deficit at age 69. Although these 3 patients were clinically diagnosed with PD, they all had family histories of atypical parkinsonism, degenerative dementias, or ALS. No expansions were detected in patients with sporadic PD or patients with a familial history of PD. Moreover, mutations in known PD genes had previously been excluded in these 4 pathogenic expansion carriers.

We identified one Asian control of Chinese origin with an age at inclusion of 52 years carrying a pathogenic \((G_4C_2)_{>60}\) expansion (table 1). Currently, there is no record of any symptoms related to PD, FTLD, or ALS in this individual. This brings the estimated prevalence of pathogenic repeat expansions in controls to 0.02% (1/5,478). Apart from the expansion mutations, the distribution of repeat lengths ranged from 0 to 32 in the Caucasian and from 7 to 14 in the Asian control persons.

C9orf72 repeat and PD susceptibility. We investigated the role of \((G_4C_2)_n\) repeats in risk of PD. First, we assessed the distribution of the alleles in patients with PD vs controls in the GEO-PD cohort (figure 1). The frequencies of the \((G_4C_2)_{10}\) allele and of \((G_4C_2)_{>17}\) were nominally increased in PD vs the controls but the differences were not statistically significant after Bonferroni correction (figure 1, table 3, figure e-1, A and B). Genotypic frequencies for \((G_4C_2)_{10}\) (table 3, figure e-1, C) and \((G_4C_2)_{>17}\) (table 3, figure e-1, D) were not significantly different between patients and controls after correction for multiple testing. The estimated attributable fractions in the population were very low (table 3). When considering the sum of the alleles and the size of the maximum allele as a quantitative variable, we observed a small but significant increase of disease risk with a rising number of repeat units (sum of alleles \(p = 0.0012\), summary effect \([B] = 0.0128\; [0.00504–0.0205]\), figure e-2, A; maximum allele \(p = 0.0010\), summary effect \([B] = 0.0181\; [0.00731–0.029]\), figure e-2, B). Together, these results suggested that the risk effect may not only be linked to the \((G_4C_2)_{10}\) repeat but may be increasing with length while the effect in the larger alleles is probably masked by the small number of carriers. Therefore, we decided to analyze the risk effect of C9orf72 repeat expansions as a binary categorical value with a cutoff between 9 and 10. However, neither allelic nor genotypic meta-analysis of the GEO-PD cohorts revealed significant association with PD for \((G_4C_2)_{>10}\) repeat alleles after Bonferroni correction (table 3, figure e-3, A and B). Furthermore, the estimated population attributable risk is low (table 3).

DISCUSSION Molecular reclassification of complex brain diseases based on genetic etiology is of utmost importance to improve differential diagnosis and to rationalize drug development. Assessment of the contribution of novel disease genes to clinically and pathologically overlapping diseases is instrumental in this reclassification. In this global study, we assessed the prevalence of \((G_4C_2)_n\) repeat alleles and expansions in an extended PD cohort ascertained within the GEO-PD Consortium and excluded a major role for pathogenic \((G_4C_2)_{>60}\) repeat expansions in the causation of PD. The low frequency of these expansions (0.06%) in the GEO-PD cohort is in agreement with earlier findings in distinct patient groups and falls in the range of frequencies observed in controls by us (0.02%) and others (0–0.6%). Furthermore, 75% of the pathogenic expansion carriers in this global study showed a decline in cognitive functions within 1 to 8 years after onset. In the absence of autopsy diagnoses, we therefore cannot exclude that some if not all of these expansion carriers are primarily FTLD/ALS patients with pronounced early parkinsonian symptoms or comorbidity of PD and FTLD/ALS. This hypothesis is supported by the identification of only one pathogenic mutation carrier
in 826 (0.1%) autopsy-confirmed PD cases. Of note, this carrier presented, in addition to Lewy body pathology, with frontotemporal degeneration and C9orf72-ALS/FTLD pathology with numerous p62-positive inclusions. Furthermore, although substantia nigra involvement is common in C9orf72-positive ALS, it can be clearly distinguished from PD-related mechanisms by the presence of p62-positive inclusion and absence of Lewy body pathology.

Altogether, it is not advisable to include C9orf72 (G4C2)n repeat expansion testing in a medical genetic diagnostic setting for typical PD patients. Exceptions can be made for patients with PD who have cognitive and/or behavioral deficits early in the disease process or in patients with a personal or familial history of FTLD/ALS. Given differences in the existing literature on C9orf72 (G4C2)n repeat length as risk factor for PD, we used the size of this global cohort to estimate a PD-related threshold of C9orf72 repeats. Calculation of the risk for each of the observed C9orf72 (G4C2)n alleles in the GEO-PD cohorts suggested a role for the 10-units repeat and for the pooled alleles of 17 units or more in PD susceptibility. Genotypic meta-analysis supported a possible link between (G4C2)10 and increased risk of PD but the association did not reach significance after correction for multiple testing. In addition, the number of carriers of these intermediate alleles is small and one should be cautious with the interpretation of these results. Furthermore, it is difficult to envisage the biological relevance of risk associated with a single

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**Figure 1** Overall distribution of C9orf72 repeat alleles in the GEO-PD cohorts

![Graph A](image1)

**A** Allele frequencies. The p values for individual alleles were calculated using a Dersimonian-Laird random-effect meta-analysis. **B** Allele counts. *Nominally significant p values. Con = controls; GEO-PD = Genetic Epidemiology of Parkinson's Disease; PD = Parkinson disease.
allele. Of note, we observed a small but significant increase in risk with an accumulative number of repeats suggesting the idea of a threshold size rather than a single allele as the culprit of increased risk. We therefore decided to study the combined effect of (G\textsubscript{4}C\textsubscript{2})\textsubscript{10} alleles of 10 units and larger in the global GEO-PD cohort. Although meta-analyses implicated a potential role for these intermediate-sized repeats in PD, none of the associations survived Bonferroni correction suggesting that if C9orf72 repeats of 10 units or larger have a role in PD susceptibility, the effect is small. This is supported by the fact that none of the published genome-wide association studies revealed the C9orf72 locus as a risk factor for PD.\textsuperscript{25} A limitation of this study is that we did not yet include all published association studies of C9orf72 in PD; however, we chose to include only those studies that were corrected for allele scoring bias based on a reference panel.

Altogether, these data support the current hypothesis that pathogenic (G\textsubscript{4}C\textsubscript{2})\textsubscript{n} repeat expansions in C9orf72 appear to be specific for the FTLD/ALS spectrum with little or no contribution to the wider spectrum of movement disorders. It will be of interest to study the role of intermediate repeats ≥10 units in other neurodegenerative disorders, however, to obtain a more profound knowledge on their role in neurodegenerative diseases and a better understanding of the underlying mechanism.

**AUTHOR AFFILIATIONS**

From the Neurodegenerative Brain Diseases Group (J.T., A.V., K.S., E.W., I.G., S.S., D.C., E.C., E.E., M.C., C.V.B.), Department of Molecular Genetics, VIB, Antwerp, Institute Born-Bunge (J.T., A.V., K.S., E.W., I.G., S.S., D.C., E.C., E.E., S.E., P.F.D.D., P.C., M.C., C.V.B.), University of Antwerp, Department of Neurology (D.C., P.C.), Antwerp University Hospital, Edegem; Department of Neurology and Memory Clinic (S.E., P.F.D.D.), Hospital Network Antwerp, Middelheim and Hoge Beuken, Antwerp, Belgium; Department of Neurology and Alzheimer Research Center (P.F.D.D.), University of Groningen and University Medical Center Groningen, the Netherlands; Department of Neurodegenerative Diseases (M.S., R.K.), Hertie-Institute for Clinical Brain Research and DZNE-German Center for Neurodegenerative Dis-eases, Tuebingen; Institute for Clinical Epidemiology and Applied Biometry (M.S.), University of Tuebingen, Germany; INSERM, UMR_S975 (S.L., A.B.), Université Pierre et Marie Curie-Paris, CNRS, UMR 7225, AP-HP, Pitié-Salpêtrière Hospital; CNRS (S.L., A.B.), UMR 7225, Paris, AP-HP (A.B.), Pitié-Salpêtrière Hospital, Department of Genetics and Cyto- genetics, Paris, France; Department of Neurology (S.J.C., M.-J.K., Y.J.K.), Asian Medical Center, University of Ulsan Col- lege of Medicine, Seoul, Korea; Departments of Neuroscience (O.A.R.) and Neurology (Z.K.W.), Mayo Clinic, Jacksonville, FL; Tanz Centre for Research in Neurodegenerative Diseases (E.R., Z.X.), Department of Medicine, University of Toronto; Toronto Western Hospital Research Institute (A.E.L.), University Health Network, Toronto, Canada; Institute of Neurogenetics (C.K., A.W.), University of Luebeck, Germany; Eskitis Institute for Drug Discovery (G.D.M.), Griffith University, Queensland, University of Queensland (P.A.S.), Centre for Clinical Research, Queens- land, Australia; Department of Neurology (G.M.H., E.D.); Neuroscience Unit, Faculty of Medicine, School of Health Sciences, University of Thessaly, Larissa, Greece; Department of Neurology (N.H., K.O.), Juntendo University School of Medicine, Tokyo, Japan; Department of Neurology (E.-K.T., Y.Z.), Singapore General Hospital, National Neuroscience Institute, Singapore; Duke NUS Graduate Medical School (E.-K.T., Y.Z.), Singapore; Department of Neurology (J.A.), St. Olaf’s Hospital, Tondheim; Department of Neuroscience (J.A.), Norwegian University of Science and Technology (NTNU), Trondheim, Norway; IRCCS Casa Sollievo della Sofferenza Hospital (E.M.V., S.P.), Mendel Laboratory, San Giovanni Rotondo; Institute of Molecular Bioimaging and Physiology (G.A., A.Q.), National Research Council, Section of Gemmaneto (CZ); Institute of Neurology (A.Q.), Department of Medical Sciences, University Magna Graecia, Catanzaro, Department of Neuroscience (C.F., L.B.), Section of Neurology, University of Milano-Bicocca, San Gerardo Hospital, Monza, Italy; Department of Neurology (A.D.), Max Plank Institute of Psychiatry, Munich, Germany, Department of Clinical Sciences (A.P.), Section of Neurology, Lund University; Department of Neurology (A.F.), Lund, Skåne University Hospital, Department of Clinical Sciences (C.N.), Clinical Memory Research Unit, Lund University, Sweden; Human Genetic Centre (G.G.), University Hospital of Liége, Belgium; Department of Neurology (M.S.L., R.F.P.), University of Tennessee Health Science Cen- ter, Memphis; Department of Neurology (M.B.-J., G.O.), Medical University of Silesia, Katowice, Poland; and Department of Neurology (D.M.M.), North- shore University HealthSystem, Evanston, IL.

**AUTHOR CONTRIBUTIONS**

Jessie Theuns: drafting and revising the manuscript for content, study concept and design, analysis and interpretation of data, acquisition of data, statistical analysis, study supervision and coordination, obtaining funding. Aline Versintenen: revising the manuscript for content, study design, analysis and interpretation of data, acquisition of data, statistical analysis, study coordination. Kristel Sleegers: drafting and revising the manuscript for content, study concept, analysis and interpretation of data, statistical analysis, obtaining funding. Eline Wauters: revising the manuscript for content, analysis and interpretation of data, acquisition of data. Stefanie Smolders: revising the manuscript for content, study concept, analysis and interpretation of data, acquisition of data. 

**Table 3** Overview of DerSimonian-Laird meta-analyses

<table>
<thead>
<tr>
<th>Allelic (OR [95% CI])</th>
<th>Genotypic (OR [95% CI])</th>
<th>p</th>
<th>AF (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>(G\textsubscript{4}C\textsubscript{2})\textsubscript{10}</td>
<td>1.31 (1.06–1.62)</td>
<td>0.012</td>
<td>1.05 (0.26–1.84)</td>
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<tr>
<td>(G\textsubscript{4}C\textsubscript{2})\textsubscript{20}</td>
<td>1.44 (1.03–2.01)</td>
<td>0.03</td>
<td>0.35% (–0.30 to 0.72)</td>
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<tr>
<td>(G\textsubscript{4}C\textsubscript{2})\textsubscript{30}</td>
<td>1.18 (1.04–1.35)</td>
<td>0.02</td>
<td>1.47% (0.07–2.83)</td>
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</tbody>
</table>

Abbreviations: AF = attributable fraction; CI = confidence interval; OR = odds ratio.

For the meta-analyses, only the cohorts including both patients with Parkinson disease and geographically matched controls that were size-corrected using the reference panel were included. The p values were calculated using DerSimonian-Laird meta-analysis; p = 0.002 is considered statistically significant [22 tests].

*Allele count approach.

†Dichotomized approach.
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REFERENCES


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