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Research Article

Genetic Profiling Differentiates Second Primary Tumors from Metastases in Adult Metachronous Soft Tissue Sarcoma

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Purpose. Patients with soft tissue sarcomas (STS) are at increased risk of second primary malignancies, including a second STS, but distinction between metastases and a second primary STS is difficult. Patients and Methods. Array-based comparative genomic hybridization (aCGH) was applied to 30 multiple STS of the extremities and the trunk wall from 13 patients. Different histotypes were present with malignant fibrous histiocytomas/undifferentiated pleomorphic sarcomas being the predominant subtype. Results. aCGH profiling revealed genetic complexity with multiple gains and losses in all tumors. In an unsupervised hierarchical cluster analysis, similar genomic profiles and close clustering between the first and subsequent STS were identified in 5 cases, suggesting metastatic disease, whereas the tumors from the remaining 8 patients did not cluster and showed only weak pairwise correlation, suggesting development of second primary STS. Discussion. The similarities and dissimilarities identified in the first and second STS suggest that genetic profiles can be used to distinguish soft tissue metastases from second primary STS. The demonstration of genetically different soft tissue sarcomas in the same patient suggests independent tumor origin and serves as a reminder to consider development of second primary STS, which has prognostic and therapeutic implications.

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1. INTRODUCTION

The first observations of an increased risk of secondary sarcoma development came from individuals treated for childhood cancers such as retinoblastoma, leukemia, Wilms’ tumor, Hodgkin’s lymphoma, and sarcoma [1–13]. However, also adult soft tissue sarcoma (STS) patients have been shown to be at higher risk of a second primary malignancy, with a particularly increased risk of a second primary STS [14–17]. STS have been associated with several hereditary syndromes, the most common being neurofibromatosis and the Li-Fraumeni syndrome [18]. Treatment-induced sarcomas include sarcomas associated with Stewart-Treves syndrome and radiation-induced angiosarcomas, which develop median 10 years after radiotherapy [19]. However, after exclusion of STS in neurofibromatosis patients and STS that developed in irradiated fields, an increased risk of a second primary STS remains [17]. Metachronous STS has been described in 1% of sarcoma patients [16, 17, 20] and this observation constitutes the basis for our study on similarities/differences in the genetic profiles of tumors from patients with multiple STS. We applied array-based comparative genomic hybridization (aCGH) that utilizes BAC clones with tiling coverage of the whole genome and allows detailed copy-number analysis, to a series of 30 metachronous STS of different histopathological subtypes from 13 patients.

2. PATIENTS AND METHODS

2.1. Patients

Adult patients (≥16 years of age) who developed two or more STS at different anatomical sites before development of any detectable pulmonary metastases were eligible for the study. Patients with neurofibromatosis type I and the Li-Fraumeni syndrome were excluded. In the southern Swedish
cancer registry, 20 patients who fulfilled these criteria were identified. The tumors had been operated either at the musculoskeletal tumor center in Lund \( n = 24 \) or at local hospitals in the southern Swedish health care region \( n = 6 \). The clinicopathological reports were reviewed to confirm tumor location and to rule out that the second primary STS represented a local recurrence and the histopathological slides were reviewed by a sarcoma pathologist (H.D.) to confirm the diagnoses.

Frozen tumor tissue was available from 16 tumors and paraffin-embedded tissue was used from 28 tumors without systematic differences related to tumor source. After DNA extraction, 7 individuals (14 tumors) were excluded because of poor DNA quality in at least one of the tumors from the same patient. High-quality aCGH data were obtained from 30 tumors (in 15 of which DNA was extracted from frozen tissue) from 13 patients (Table 1). These patients contributed with two to four STS and were mean 73 (28–83) years at diagnosis of the first STS. The second STS developed median 3 (1–7) years after the first STS. No neoadjuvant chemotherapy was given, and only one patient (case 5) had received postoperative chemotherapy after the first STS. Radiotherapy had been administered to four patients (postoperatively in case 5, 6, and 8, and preoperatively in case 1), but none of the second STS developed within the irradiated field.

Clinical data for the 13 cases are presented in Table 1. The lower extremity was the most common tumor site (16 tumors) and 28 tumors were high-grade (grades 3 and 4 on a 4-tiered scale). The first STS included eight malignant fibrous histiocytomas/undifferentiated pleomorphic sarcomas (MFH/UPS), two leiomyosarcomas, two malignant peripheral nerve sheath tumors (MPNST), and one pleomorphic liposarcoma. The histopathological diagnosis of the second STS differed from the first in two patients; a leiomyosarcoma was diagnosed in a patient with two prior MFH/UPS and a leiomyosarcoma was diagnosed in a patient previously operated on for an MPNST. In the remaining patients, including the three cases from which three or four distinct tumors were analyzed, multiple STS of the same histopathological type were diagnosed.

In 9/13 patients the STS developed at different anatomical locations, for example, different extremities or extremity and trunk wall. Three patients developed metachronous STS in the same extremity but at different locations, for example, lower leg and thigh (cases 8, 12, and 13) and one patient (no. 7) developed two STS in the same extremity; a deep-seated leiomyosarcoma in the medial thigh and five years later a subcutaneous leiomyosarcoma in the lateral part of the thigh (Table 1). Clinical follow-up was complete for a minimum of 8 years for the survivors. During follow-up, two patients (cases 12 and 13) developed local recurrences, 1 and 10 years after primary surgery. Lung metastases developed in 5/13 patients, median 50 (range 15–51) months after diagnosis of the primary tumor. Apart from the metachronous STS, two patients (cases 4 and 7) developed adenocarcinomas of the breast and the colon, respectively. Ethical permission for the study was granted from the Lund University ethics committee.

2.2. DNA extraction and array-based comparative genomic hybridization

Genomic DNA from frozen \( n = 15 \) and paraffin-embedded \( n = 15 \) tumors was extracted using the Wizard Genomic DNA Purification kit (Promega, Madison, WI) and overnight proteinase-K digestion treatment followed by phenol-chloroform purification. When paraffin-embedded tissue was used, a fresh-µm section was obtained, stained with hematoxylin & eosin and a representative tumor area was chosen. Thereafter, 1-mm tissue cores were obtained and used for DNA extraction. The tissue cores were pretreated in xylene before proteinase-K treatment and phenol-chloroform purification. DNA quality was checked using a Ready-To-Go RAPD analysis kit (Amersham Biosciences, Buckinghamshire, UK), and the concentration was measured using a Nano drop (NanoDrop Technologies, Wilmington, Del, USA). Commercial genomic male DNA, derived from

![Figure 1: Unsupervised hierarchical cluster analysis of all 30 soft tissue sarcomas from 13 patients. The analysis was based on the ~17,000 clones that survived the preprocessing filters, and close clustering of the metachronous STS was demonstrated in 5 patients (case 1, 2, 6, 9, and 12 marked with different colours), whereas the STS from the remaining 8 patients were scatter in the analysis. Clustering was done using the TMeV application from the TM4 microarray software suit.](image)
Table 1: Summary of clinical data from the 30 STS analyzed from 13 patients.

<table>
<thead>
<tr>
<th>Case no.</th>
<th>Sex</th>
<th>Age</th>
<th>Site</th>
<th>Type</th>
<th>Years after first tumor</th>
<th>Site</th>
<th>Type</th>
<th>Years after first tumor</th>
<th>Site</th>
<th>Type</th>
<th>Follow-up status/months</th>
<th>Tumor source frozen/paraffin</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>First tumor</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>M</td>
<td>80</td>
<td>Trunk wall</td>
<td>LMS</td>
<td>3</td>
<td>Left cheek</td>
<td>LMS</td>
<td></td>
<td>MET/50</td>
<td>t/f</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>F</td>
<td>82</td>
<td>Right upper arm</td>
<td>MFH/UPS</td>
<td>2</td>
<td>Left buttock</td>
<td>MFH/UPS</td>
<td>3</td>
<td>Scalp</td>
<td>MET/47</td>
<td>p/f/p</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>F</td>
<td>80</td>
<td>Right thigh</td>
<td>LS</td>
<td>1</td>
<td>Left shoulder</td>
<td>LS</td>
<td>2</td>
<td>Right lower leg</td>
<td>TD/22</td>
<td>f/p/p</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>F</td>
<td>79</td>
<td>Lower leg</td>
<td>MFH/UPS</td>
<td>1</td>
<td>Right thigh</td>
<td>MFH/UPS</td>
<td></td>
<td>NED/36</td>
<td>p/f</td>
<td></td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>M</td>
<td>76</td>
<td>Right lower leg</td>
<td>MFH/UPS</td>
<td>7</td>
<td>Right thigh*</td>
<td>MFH/UPS</td>
<td></td>
<td>TD/164</td>
<td>p/f</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Second tumor</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>F</td>
<td>77</td>
<td>Right shoulder</td>
<td>MFH/UPS</td>
<td>5</td>
<td>Left thigh</td>
<td>MFH/UPS</td>
<td>9</td>
<td>Right lower arm</td>
<td>NED/163</td>
<td>f/f</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>F</td>
<td>75</td>
<td>Left lower arm</td>
<td>MFH/UPS</td>
<td>4</td>
<td>Right lower leg</td>
<td>MFH/UPS</td>
<td></td>
<td>NED/54</td>
<td>f/p</td>
<td></td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>F</td>
<td>60</td>
<td>Right lower arm</td>
<td>MPNST</td>
<td>4</td>
<td>Right thigh</td>
<td>LMS</td>
<td></td>
<td>NED/56</td>
<td>f/p</td>
<td></td>
<td></td>
</tr>
<tr>
<td>7</td>
<td>M</td>
<td>83</td>
<td>Left thigh (laterally)</td>
<td>LMS</td>
<td>4</td>
<td>Left thigh (medially)</td>
<td>LMS</td>
<td></td>
<td>TD/28</td>
<td>p/f</td>
<td></td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>M</td>
<td>73</td>
<td>Left knee</td>
<td>MFH/UPS</td>
<td>1</td>
<td>Left lower leg</td>
<td>MFH/UPS</td>
<td></td>
<td>MET/15</td>
<td>f/p</td>
<td></td>
<td></td>
</tr>
<tr>
<td>10</td>
<td>M</td>
<td>28</td>
<td>Right trunk wall</td>
<td>MPNST</td>
<td>1</td>
<td>Scalp</td>
<td>MPNST</td>
<td></td>
<td>TD/32</td>
<td>f/p</td>
<td></td>
<td></td>
</tr>
<tr>
<td>11</td>
<td>F</td>
<td>77</td>
<td>Right thigh</td>
<td>MFH/UPS</td>
<td>2</td>
<td>Back</td>
<td>MFH/UPS</td>
<td></td>
<td>MET/51</td>
<td>p/f</td>
<td></td>
<td></td>
</tr>
<tr>
<td>13</td>
<td>M</td>
<td>76</td>
<td>Right foot</td>
<td>MFH/UPS</td>
<td>4</td>
<td>Right thigh</td>
<td>MFH/UPS</td>
<td></td>
<td>MET/51</td>
<td>p/f</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

MFH/UPS = malignant fibrous histiocytoma/undifferentiated pleomorphic sarcoma, LMS = leiomyosarcoma, LS = liposarcoma, MPNST = malignant peripheral nerve sheath tumor.

*Located 15 cm from the first tumor.

†Located 15 cm from the first tumor.

The patient developed one additional LS, left thoracic wall.

Table 2: Recurrent high-level amplifications and homozygous deletions.

<table>
<thead>
<tr>
<th>Chromosome</th>
<th>No of tumors</th>
<th>Cytoband</th>
<th>Start position</th>
<th>Size (Mbp)</th>
<th>HLA/Hz del*</th>
<th>Cancer related genes</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>2</td>
<td>1p32.1</td>
<td>58571482</td>
<td>1.2</td>
<td>HLA</td>
<td>JUN</td>
</tr>
<tr>
<td>1</td>
<td>3</td>
<td>1q44</td>
<td>241558190</td>
<td>1.0</td>
<td>Hz del</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>2</td>
<td>3q23</td>
<td>141082484</td>
<td>0.51</td>
<td>Hz del</td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>3</td>
<td>3p12.1–3</td>
<td>78780218</td>
<td>7.0</td>
<td>HLA</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>2</td>
<td>4q12</td>
<td>54016334</td>
<td>1.5</td>
<td>HLA</td>
<td>PDGFRA, KIT, CHIC2</td>
</tr>
<tr>
<td>6</td>
<td>3</td>
<td>6p 12.3–21.2</td>
<td>37360640</td>
<td>11.4</td>
<td>HLA</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>6</td>
<td>9p21.3</td>
<td>21647433</td>
<td>2.4</td>
<td>Hz del</td>
<td>CDKN2A, CDKN2B, MTAP</td>
</tr>
<tr>
<td>10</td>
<td>2</td>
<td>10q23.31</td>
<td>89538420</td>
<td>1.5</td>
<td>Hz del</td>
<td>PTEN</td>
</tr>
<tr>
<td>11</td>
<td>2</td>
<td>11q13.4–5</td>
<td>71730097</td>
<td>5.3</td>
<td>HLA</td>
<td></td>
</tr>
<tr>
<td>13</td>
<td>13</td>
<td>13q14.2</td>
<td>47901160</td>
<td>0.65</td>
<td>Hz del</td>
<td>RB1</td>
</tr>
</tbody>
</table>

* HLA = high level amplification, Hz del = homozygous deletion.

a pool of healthy individuals, was used as a reference (Promega, Madison, Wiss, USA). CyDye coupling/labeling was carried out using a random labeling kit (Invitrogen Life Technologies, Carlsbad, Calif, USA) according to the manufacturer’s recommendations. In short, 2-µg genomic tumor DNA and reference DNA were differentially labeled with fluorescent dyes, Cy3 for tumor tissue, and Cy5 for reference DNA. After a purification step, these were pooled, mixed with COT-1 DNA to block repetitive DNA sequences, dehydrated and resuspended in a formamide-based buffer (Invitrogen). The labeled DNA was then applied to arrays pretreated in washing solutions (Pronto! Microarray Reagent System, Corning Labsystems, Corning, NY, USA) and hybridization was performed for 48–72 hours at 37°C. The incubation was performed under cover slips for the DNA isolated from the frozen tumor material whereas the MAUI hybridization System (BioMicro systems Inc., Salt Lake City, Utah, USA) was used for the DNA derived from paraffin-embedded tumors. Dye-swaps (i.e., complementary hybridization in which Cy5 was used for tumor tissue and Cy3 for reference DNA) were used in three cases and allowed subtraction of dye-related noise. The slides were treated in post-hybridization washing solutions and finally scanned using an Agilent Microarray scanner (Agilent Technologies, Palo Alto, Calif, USA).

2.3. BAC array platform

The BAC array slides were produced at the Swegene DNA Microarray Resource Center, Department of Oncology, Lund University. These have an average resolution of 80 kb and contain a total of 32 433 BAC clones from the 32k human genome high-resolution BAC arrayed clone set, version 1.0 from the BACPAC Resource Center at Children’s Hospital Oakland Research Institute (Oakland, Calif, USA) (http://bacpac.chori.org/). The clones provide >99% coverage of the fingerprint map and current sequence assembly with a resolution of 100 kb.

2.4. Data analysis

Image analysis and data extraction were carried out using GenePix Pro 4.1.1.4 version (Axon Instruments Inc., Foster City, CA, USA) and the quantified data matrix was then uploaded into the web-based BioArray Software Environment (BASE; http://gothmog.thep.lu.se/int/index) [21], where all data management and analysis were carried out. The background correction and intensities of Cy3 and Cy5 were calculated using the median feature and median-local background intensities of the uploaded files, and the intensity ratios were calculated using the background corrected spot intensities by calculating the log2 ratios of tumor to reference intensity. In BASE a preliminary filter, based on the flagging in the image analysis, was applied, and spots with a diameter <55 µm and a signal to noise (SNR) ratio ≤3 in the tumor or reference channel were flagged as “bad” and filtered away from further analysis. The intensity-dependent LOWESS algorithm [22] was used to normalize the data within individual arrays. To correct for spatial bias, the data were normalized within groups of 8 print-tip blocks. A moving average smoothing algorithm with a 250 kbp sliding window was then applied, and a BASE-adapted CGH-plotter software was used to identify regions of gains and losses [23]. In the CGH-plotter, each clone was assigned a calculated level log2 ratio value, corresponding to the level that the clone belongs to, in order to reduce the noise. Hereafter an unsupervised hierarchical cluster analysis, using the Pearson correlation distance metric and the average linkage method, was applied to the data derived from the CGH-plotter (the TMeV application from the TM4 microarray software suite was used; http://www.tm4.org/mev.html). The CGH-plotter was also used to generate a ternary scale, where all clones were designated gained, lost or unchanged. These values were used to calculate the percentage of altered clones in each assay and the mean number of altered clones in tumor subgroups. Pearson correlation was used to determine the correlation between tumors within the same individual, based on the number of altered clones. Gains and losses were defined as a
log$_2$ ratio $\pm 0.2$. Amplifications were defined as clones with a log$_2$ ratio $\geq 0.5$, whereas high-level amplifications were defined as a log$_2$ ratio $\geq 1.5$. Homozygous deletions were suspected when the log$_2$ ratio was $\leq 1.5$.

3. RESULTS

Genomic profiles from the 30 metachronous STS of five different histopathological subtypes revealed multiple gains and losses and identified several high-level amplifications (HLA) and homozygous deletions (Table 2). The alterations affected mean 39 (9–70)% of the whole genome with 19% amplifications and 20% deletions. When the first STS ($n = 13$) were compared to the subsequent STS ($n = 17$) a small difference in the total number of alterations was found with 35 (16–54)% and, 42 (9–70)% of the genome altered, respectively. Several recurrent aberrations were identified with the most frequent changes (present in $>60\%$ of the tumors) being deletions of 10q24.3–25.2, 13q12.1–12.2, 13q21.1–21.2, 16q13–23.2, 18q12.2–12.3, and amplifications of 1q21.3–23.1 and 19p13.3.

Unsupervised hierarchical cluster analysis, based on the $\sim 27000$ clones that survived the filters in BASE, revealed close clustering of the tumors from five individuals without significant differences between the first and subsequent STS (43% and 41% of the genome altered) (Figure 1 and Table 1, cases 1, 2, 6, 9, and 12). These tumor pairs showed strong similarities between the genomic plots (Figure 2(a)) with a mean correlation of 0.7 (0.5–0.9). The many shared alterations outnumbered the few differences in all five cases and deletions identified in the first tumor were always present in the second STS. The median time interval between the first and second STS in these five patients was 1 (1–7) year, and two of these patients subsequently developed lung metastases. In the remaining eight cases STS from the same individual did not cluster together and showed a significantly weaker correlation, mean 0.1 (0–0.4). These tumors had pronounced intertumor variability (30% of the genome being altered in the first tumor compared to 42% in the subsequent tumors), which was comparable to the interpatient variability, which had a mean correlation of 0.1 (0.04–0.2). In four of these cases, deletions present in the first tumor were not present in the subsequent tumor, which supports independent tumor origin. The second STS in these eight cases developed median 4 (1–5) years after the first STS and three of the patients later developed pulmonary metastases.

4. DISCUSSION

Despite multidisciplinary and multimodality treatment, distant metastases develop in about 30% of STS patients. Hematogenous, pulmonary metastases predominate, whereas lymphatic spread occurs in $<5\%$ of the patients [19, 24]. Soft tissue metastases are rare and have mainly been reported in liposarcomas [16, 25]. Development of synchronous or metachronous STS has been described in several case studies, but it remains a rare clinical presentation and the interpretations hereof have varied [25–29]. Epidemiological data support an increased risk of a secondary primary sarcoma among adult STS patients [14, 17]. In order to reduce bias from inclusion of familial sarcoma syndromes, we excluded multiple sarcoma patients diagnosed with neurofibromatosis or the Li-Fraumeni syndrome. The only patient who developed more than one MPNST was carefully examined without any sign of neurofibromatosis (until death 3 years later). Since only one patient had received adjuvant chemotherapy after the first STS and none of the second STS developed in irradiated fields, the second STS studied are unlikely to represent treatment-induced secondary tumors.

Application of CGH in STS has mainly involved leiomyosarcoma and MFH/UPS and has in these highly malignant and pleomorphic STS subtypes demonstrated extensive genomic complexity with recurrent copy number changes, including losses of 2p, 2q, 10q, 11q, and 13q and

![Figure 2](image-url): Superposition of whole genome plots from two different tumors from the same patient showing (a) similarities in gene copy number changes in two liposarcomas (case 6). (b) Differences in gene copy number changes in two malignant fibrous histiocytomas (case 3).
gains of 1q, 5p, 8q, and 17p [30–36]. Several of these recurrent changes were also identified among the 30 STS in this study with the most frequent being deletions of 10q24.3–25.2, 13q12.1–12.2, 13q21.1–21.2, 16q13–23.2, 18q12.2–12.3, and amplifications of 1q21.3–23.1 and 19p13.3. In order to obtain as many tumor pairs as possible, frozen as well as paraffin-embedded tumor tissue was used. Among the tumors from which high-quality DNA was obtained, no differences were identified related to tumor source (Table 1). When the genomic profiles from the different tumor pairs were compared, five pairs showed highly correlated genomic profiles suggestive of metastatic disease, whereas eight cases showed different profiles suggestive of distinct primary STS. In the latter STS, the differences by far outnumbered the similarities, which resulted in a weak correlation, which was comparable to the interpatient variability. Importantly, there was no systematic difference in sample preparation methodology between the two groups suggestive of metastatic disease and distinct primary STS, respectively. The similarities and differences were evident in an unsupervised hierarchical cluster analysis (Figure 1) in which the five former tumor pairs clustered closely, whereas the latter eight did not.

Previous reports of multiple STS have predominantly involved liposarcomas. In a mixed series of nine STS patients who developed synchronous and metachronous STS, Grobemyer et al. identified liposarcomas \((n = 5)\) and gastrointestinal stromal tumors \((n = 5)\) as the most frequent subtypes whereas Blair et al. reported 16 patients with multiple STS of whom nine had liposarcomas \([20, 37]\). Our study only included successfully analyzed liposarcomas from one patient (case no. 6) who developed four liposarcomas in four years without signs of lung metastases and died from locally advanced tumor masses 22 months later. Antonescu et al. applied Southern blot analysis to tumors from six patients who developed recurrent changes among the 30 STS in our series present in metachronous STS of the same histopathological types; three MFH/UPS, one liposarcoma, and one leiomyosarcoma, which developed with median 1 year interval. Subsequent development of lung metastases occurred in two of these five patients (Table 1). Previous studies that have examined genetic differences between primary and recurrent STS have been carried out using conventional CGH and have demonstrated increasing genetic complexity from primary STS to a local recurrence \([38–40]\). However, the issue of development of second primary STS has to our knowledge not been addressed using genetic profiling. We demonstrate strikingly similar genetic profiles in the five STS likely representing soft tissue metastases with mean 43% and 41% of the genome altered and several shared deletions identified. This stands in contrast to the eight STS patients from which the metachronous STS showed different genetic profiles. Among these, multiple histologic subtypes (i.e., MFH/UPS, MPNST, and leiomyosarcomas) were present and four of the deletions identified in the primary tumors were not found in the second STS. In summary, the clinical presentation, histopathology, and the genetic profile support independent sarcoma origin in 8 of the 13 patients (Table 1). Although development of metachronous STS is rare, our demonstration of different genetic profiles in the majority of these cases serves as a reminder to consider independent tumor origin, which has implications for the choice of therapy, for example, use of adjuvant chemo- and/or radiotherapy after surgery for a second STS that should not per se be perceived to represent metastatic disease.

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**REFERENCES**


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Parasitic Diseases, Diagnostic Approaches, and Therapies

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Over the entire course of human evolution, parasites likely inhabited us as they went through their own evolutionary processes. In recent centuries, environmental and social changes often have disrupted the traditional cycles of transmission of parasites to and among humans. In many instances, this has resulted in improvements in the health of affected populations, but in some cases new modes of transmission have arisen, thus resulting in overall negative impacts. New diagnostic methods make it possible to explore these complex and dynamic relationships. This special issue will familiarize readers with:

- Rapid diagnostic methods based on immunologic, molecular, and other formats
- Transmission of parasites by blood transfusion and organ transplantation
- Potential therapeutic uses of parasites for noninfectious diseases and other controversial issues related to these clever creatures

Authors should follow the Interdisciplinary Perspectives on Infectious Diseases manuscript format described at the journal site http://www.hindawi.com/journals/hipid/. Prospective authors should submit an electronic copy of their complete manuscript through the journal Manuscript Tracking System at http://mts.hindawi.com/, according to the following timetable:

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Interdisciplinary Perspectives on Infectious Diseases

Special Issue on
Role of Infection in Neurologic and Psychiatric Diseases

Call for Papers

Infectious agents have been changing and challenging us in ways unprecedented in scope. There have been discoveries of new agents, recognition of heretofore unknown links between pathogens and disease, and documentation of changes in virulence and geography. Some of the best examples of these trends are found in examinations of the role of infectious agents in neurological and psychiatric diseases.

In this special issue, the editors of Interdisciplinary Perspectives on Infectious Diseases are soliciting articles to cover current investigations on discovery, epidemiology, clinical pathophysiology, and pathogenesis of infectious agents linked to neurological and psychiatric diseases. The topics, listed below, were chosen in an effort to show the diverse routes leading to increased understanding of the etiology, treatment, and prevention of these CNS diseases. Authors are invited to submit under the following topics:

- New encephalitic agents/or new to man
- Global spread and burden of encephalitic agents
- Diagnostic methods and pathogen discovery
- Coinfections/comorbidities
- Pathogens and chronic or long-term neurologic diseases
- Pathogens and chronic psychiatric diseases
- Viruses and neuropharmacology/signaling/signal transduction
- Animal models for study of encephalitic agents

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