Terms, definitions and measurements to describe sonographic features of myometrium and uterine masses: a consensus opinion from the Morphological Uterus Sonographic Assessment (MUSA) group


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TERMS AND DEFINITIONS FOR DESCRIBING MYOMETRIAL PATHOLOGY USING ULTRASONOGRAPHY

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ABSTRACT:

This paper is a consensus statement on terms, definitions and measurements to describe and report the sonographic features of the myometrium using grayscale sonography, color/power Doppler, and three-dimensional ultrasound imaging.

These terms and definitions may be relevant both for the clinician when reporting ultrasound examinations in daily practice and for clinical research. The use of the terminology when describing the two most common myometrial lesions (fibroids and adenomyosis) is presented.
Abbreviations:

Ultrasonography (US),
Transvaginal ultrasonography (TVS),
Two-dimensional (2D),
Three-dimensional (3D),
Volume Contrast Imaging (VCI),
Endomyometrial junctional zone (JZ),
Total myometrial wall thickness (TWT)
Maximum thickness of the junctional zone ($JZ_{\text{max}}$)
Minimum thickness of the junctional zone ($JZ_{\text{min}}$)
Difference between maximal and minimal JZ thickness ($JZ_{\text{dif}}$)
Penetration (P)
INTRODUCTION

A variety of disorders or pathology in the uterus gives rise to different manifestations and clinical signs.

Ultrasonography (US) is a first-stage imaging technique for assessing the myometrium and requires findings to be reported consistently. Recently the FIGO PALM-COEIN system (polyp; adenomyosis; leiomyoma; malignancy and hyperplasia; coagulopathy; ovulatory dysfunction; endometrial; iatrogenic; not yet classified)\(^1\)\(^2\) has been published, classifying the etiology of abnormal uterine bleeding, including the myometrial pathologies adenomyosis and fibroids. However, the implementation of this classification in daily clinical practice is hampered by the lack of standardization of terms and definitions used to describe ultrasound findings. Standardized terms to be used when describing ultrasound images of the endometrium and uterine cavity have been suggested by the IETA group\(^3\).

However, there is still no standardized terminology for describing ultrasound images of normal and pathological myometrium as well as uterine masses\(^4\).

In clinical practice and research, standardized reporting of ultrasound findings with regard to the myometrium is essential to reduce intra- and inter-observer variability in the evaluation of pathology, to assess the effect of medical or surgical treatment, and to compare ultrasound imaging with other imaging techniques. Moreover, a common terminology is necessary for comparison between studies and when combining data in meta-analyses. Reliable predictors of benign pathology is essential clinically to allow safe use of minimally invasive techniques for the treatment of uterine myomas such as selective uterine artery embolization, fibroid ablation or laparoscopic morcellation\(^5\).

The primary aim of this paper is to present a consensus opinion on the terminology to be used when describing the ultrasonographic features of the myometrium and of myometrial lesions. These terms and definitions should be relevant both for clinicians reporting ultrasound examinations in day to day practice and for clinical research. The secondary aim is to illustrate the use of the terminology when describing the two most common myometrial lesions: fibroids and adenomyosis.
**Methodology**

This consensus paper is based on the opinion of a panel of clinicians (MUSA, Morphological Uterus Sonographic Assessment) with expertise that includes gynecological ultrasonography, fertility treatment, hysteroscopy, general gynecology and clinical research. Amongst the authors are members from the IOTA (International Ovarian Tumor Analysis) and IETA (International Endometrial Tumor Analysis) groups and in order to produce a consensus paper that includes opinions from both ultrasound and endoscopic interest groups, members of the ESGE (European Society of Gynaecological Endoscopy) are also included. A first draft was written in April 2014 by the two first authors and sent to all co-authors. All co-authors had the opportunity to comment within a fixed time limit. Reply was mandatory for co-authorship. After taking all comments into account a revised draft was sent to all co-authors. In the event of conflicting opinions a consensus was proposed after discussion between the two first authors and the last author. This pathway was repeated until a consensus between all authors was reached. After seven revisions the manuscript was deemed ready for submission.

**Scanning the Myometrium (Detailed Description)**

Ultrasound examination of the myometrium may be performed using transabdominal or transvaginal scanning. Although transvaginal ultrasonography (TVS) is generally preferred, transabdominal ultrasonography may be necessary for imaging beyond the small pelvis. For adequate visualization of the uterus, some bladder filling is required to displace the small bowel from the field of view. Image quality during transabdominal ultrasonography may also be hampered by adipositas, scar tissue or uterine retroversion. High-resolution TVS allows for detailed assessment of the myometrium within a limited depth of view. Transabdominal ultrasonography may be necessary for imaging beyond the small pelvis. A TVS starts with a dynamic two-dimensional (2D) scan of the uterus in two perpendicular planes. Some gentle pressure may need to be applied with the probe or the free hand to assess uterine mobility and to screen for site specific tenderness.

On a sonographic cross-section through the uterus, the arcuate venous and arterial vessels can be seen close to the outer myometrial border. The junctional zone (JZ) (also referred to as inner myometrium, archimyometrium or
stratum subvasculare) is visible as a hypoechogenic subendometrial halo. This layer is composed of longitudinal and circular closely packed smooth muscle fibers. Three dimensional (3D) ultrasonography makes off-line examination and manipulation of ultrasound images possible. This may facilitate accessing a second opinion by an expert examiner in difficult cases. 3D volume acquisition of the uterus starts with an adequately enlarged mid-sagittal or transverse section of the uterine body. In optimal conditions the mid-sagittal plane allows the visualization of the entire length of the endometrium as well as the endocervical canal. The acquisition angle is chosen to include the entire uterine volume of interest. Once the 3D volume has been acquired, examination of the volume is performed in the multiplanar view by scrolling in each sectional plane separately.

Coronal sections provide information on the external contour of the uterus and cavity shape. Different features for image optimization and post-processing are used. For example, rendering and Volume Contrast Imaging (VCI) modes deliver details on the continuity and thickness of the junctional zone (JZ). Other post processing modalities such as the use of tomographic ultrasound imaging (also called multi-slice imaging) may also be helpful.

**Uterine Measurements, Shape and External Contour.**

Measuring the corpus of the uterus is performed as shown in Figure 1. If the aim of the ultrasound scan is to evaluate the myometrium (e.g. in the diagnosis of adenomyosis), then measurement of the uterine volume should exclude the cervix. If the length of the entire uterus (including the cervix) is required (e.g. at preoperative evaluation), the sum of the corpus length and the cervical length should be reported.

The corpus length (d1) is the sum of the fundus length (from the fundal serosal surface of the uterus to the fundal tip of the endometrial cavity) and the endometrial cavity length (from the fundal tip of the endometrial cavity to the internal os of the cervix). Both should preferentially be measured separately in the longitudinal plane of the uterus. The largest antero-posterior diameter (d2) is also measured in the sagittal plane. The largest transverse diameter is measured in the transverse plane of the uterus. The formula for the volume calculation based on these
measurements is displayed in Table 1 and in Figure 1. If the length of the entire uterus is required, the sum of the total length of the uterus (d1) and the cervical length should be reported.

The serosal contour of the uterus is reported as regular or lobulated (Figure 2).

The anterior and posterior myometrial walls are measured from the external uterine serosa to the external endometrial contour and should include the JZ, but not the endometrium. The myometrial walls are measured in the sagittal plane perpendicular to the endometrium. Both measurements are taken on the same image, and the measurements are taken where the myometrial wall appears to be at its thickest. The ratio between the anterior and posterior wall thickness is calculated. A ratio between the anterior and posterior wall thickness of around 1 means that the myometrial walls are symmetrical. A ratio well above or below 1 indicates asymmetry, although this may also be estimated subjectively (Figure 3). The myometrial walls can also be measured in the transverse or coronal planes if felt to be necessary.

THE JUNCTIONAL ZONE

Although the JZ can often be visualized on 2D ultrasound, acquisition of a 3D-volume enables a more complete assessment in the sagittal, transverse and coronal plane as shown in a standardized multi-planar view (Figure 4). Using the standardized multi-planar view reduces inter-observer variation in measurements, is used in general clinical practice for evaluation of the coronal view, and may be obtained by the z-rotation technique. Imaging the JZ may be optimized by using a post-processing rendering mode, for example Volume Contrast Imaging (VCI). The thickness of the slices or render box may be selected between 1 and 4 mm.

The JZ (Table 2 and Figure 5) may be regular, irregular, interrupted, not visible, not assessable or may manifest more than one feature (e.g. irregular and interrupted). For research purposes, any irregularity in the JZ may be described (e.g. cystic areas, hyperechogenic dots, hyperechogenic buds and lines) in each location (anterior, posterior, lateral left, lateral right, fundus) according to the specific research protocol.

Detailed morphological assessment and measurement of the JZ is generally only currently relevant in the context of research protocols. The JZ and the total myometrial wall thickness (TWT) are measured perpendicular to the
endometrium on the same section through the uterus. The maximum thickness of the junctional zone ($JZ_{\text{max}}$) is measured at the area where the JZ appears to be at its thickest, and the minimum thickness $JZ_{\text{min}}$ where it appears to be at its thinnest, after evaluation of the total three-dimensional volume of the uterus (Figure 6). To define the ratio between the JZ and the total uterine wall thickness, both the JZ and the total uterine wall thickness should be measured on the same image. Where to take the measurement(s) to calculate this ratio depends on the research protocol. If the JZ is ill-defined or not visible, it should be reported as ‘non-measurable’.

The magnitude and the extent of any irregularity of the JZ may be reported and the location of the JZ irregularity (anterior, posterior, lateral left, lateral right, fundus) specified according to the research protocol. The magnitude of a JZ irregularity is expressed as the difference between the maximal and minimal JZ thickness: $(JZ_{\text{max}}) - (JZ_{\text{min}}) = JZ_{\text{dif}}$.

The extent of JZ irregularity is reported as the subjective estimation of the percentage of the JZ that is irregular (<50% or ≥50%). This estimation can be made for the uterus as a whole or for each location. Interruption of the JZ may be caused by focal infiltration of the JZ by endometrial tissue, but contractions and changes within the JZ may also give rise to apparent JZ irregularities or influence JZ thickness. The extent of interruptions are recorded as a subjective estimation of the percentage of the JZ that is interrupted (<50% or ≥50%). Again this may be for the uterus as a whole or in each specific location.

**DESCRIPTION OF MYOMETRIAL PATHOLOGY** (Table 1)

The overall echogenicity of the myometrium is reported as homogeneous or heterogeneous. The reason for the heterogeneity (e.g. cysts, shadowing) should be specified as outlined below.

Myometrial pathology may be localized (one or more ‘lesions’) or diffuse. A myometrial lesion may be well-defined as typically seen in fibroids or ill-defined as typically seen in adenomyosis. Each lesion should be described according to its location, size, and site (Table 1 and Figures 7-9). The site and size may be impossible to describe for some ill-defined lesions.
The lesion location within the myometrium may be anterior or posterior, fundal, right lateral or left lateral. A lesion is global if the pathology diffusely involves the whole myometrium.

The site of a well-defined lesion should be reported using the FIGO classification for fibroids: 0 = pedunculated intracavitary; 1 = submucosal <50% intramural; 2 = submucosal ≥50% intramural; 3 = 100% intramural, but contacts the endometrium; 4 = intramural; 5 = subserosal ≥50% intramural; 6 = subserosal <50% intramural; 7 = subserosal pedunculated; 8 = other (e.g. cervical, parasitic)\(^1\)\(^2\)\(^3\)\(^4\) (Figure 7). Lesion size is estimated by measuring the three largest orthogonal diameters. The minimal distance from the lesion to the endometrium (inner lesion free margin, IFM) and serosal surface (outer lesion free margin, OFM) of the uterus\(^5\)\(^6\)\(^7\)\(^8\)\(^9\)\(^10\)\(^11\)\(^12\) is measured as described in Figure 8.

Ill-defined lesions are by definition difficult to delineate and measurements may be inaccurate. The extent of an ill-defined lesion can be estimated subjectively as the percentage of the myometrium of the whole uterine volume that is involved. If less than 50% of the total myometrium is involved, the lesion is reported as localized, if at least 50% of the myometrium is involved, it is reported as diffuse. For research purposes or in a preoperative setting, the percentage involved in each location may need to be recorded. For ill-defined lesions, the penetration (P) is defined as the ratio between the maximal thickness of the lesion and the total uterine wall thickness. The penetration is measured where the lesion appears to be at its largest as shown in Figure 9.

The echogenicity of a lesion is reported as uniform (homogeneous and/or symmetrical pattern of echogenicity) or non-uniform (heterogeneous) (Figure 10a). A uniform lesion may be hypo-, iso- or hyperechogenic.

For research purposes, the echogenicity of the lesion may be compared to the adjacent myometrium and semi-quantified as shown in Figure 10b (very hypoechogenic−, hypoechogenic−, isoechogenic, hyperechogenic+ or very hyperechogenic++).

A lesion may have non-uniform echogenicity because of mixed echogenicity, the presence of echogenic areas or of cystic areas (regular or irregular). If present, cyst contents may be anechoic, of low-level echogenicity, of ground glass appearance or of mixed echogenicity\(^13\)\(^14\). Anechoic areas can be differentiated from large vessels by using power Doppler to confirm the absence of blood flow.
The rim of a lesion may be ill-defined, hypo-, or hyper-echogenic in comparison to the myometrium (Figure 11), and the shape of a lesion may be round or not-round. A lesion that is not-round may be oval, lobulated or irregular (Figure 11).

Shadowing (Figure 12a) may arise from the edge of a lesion in which case they are reported as edge shadows, or from areas within the lesion when they are termed internal shadows. The degree of shadowing is reported subjectively as slight, moderate or strong.

Fan-shaped shadowing (Figure 12b). Fan-shaped shadowing is defined as the presence of hypoechochogenic linear stripes, sometimes alternating with linear hyperechogenic stripes. Fan-shaped shadowing may be caused by overlying (micro)cystic structure(s). The degree of shadowing is subjectively recorded as slight, moderate or strong.

Cysts (Figure 13a). Myometrial cysts are rounded lesions within the myometrium. The cyst contents may be anechoic, of low-level echogenicity, ground glass appearance or mixed echogenicity. A cyst may be surrounded by a hyperechogenic rim. In the context of research studies, the number of cysts and the largest diameter of the largest cyst or of a specified number of cysts, as well as the echogenicity of the cyst fluid may be reported. Some cysts are not measurable individually and may form aggregates of tiny, hypoechogenic microcysts (anechoic lacunae) within the myometrium. There are often several aggregates of microcysts in an area.

Hyperechogenic islands (Figure 13b). These are hyperechogenic areas within the myometrium and they may be regular, irregular or ill-defined. The number and the maximum diameter of the largest hyperechogenic island (or, if applicable, for example as part of a research protocol, of a specified number of hyperechogenic islands) may be reported. Hyperechogenic islands should be distinguished from small hyperechogenic spots seen in the subendometrium (Figure 13c).

Hyperechogenic subendometrial lines or buds (Figure 14). The JZ may be disrupted by hyperechogenic subendometrial lines or buds. Hyperechogenic subendometrial lines are (almost) perpendicular to the endometrial cavity and are in continuum with the endometrium. These buds and lines should be distinguished from small hyperechogenic spots seen in the subendometrium (Figure 13c). For research purposes, the number and location of the subendometrial lines or buds may be reported.
Vascularization of the myometrium and myometrial lesions

When using color or power Doppler the arcuate vessels of the uterus are often visible at the periphery of the myometrium running parallel to the uterine serosa. Perpendicular to the arcuate vessels, the radial arteries and veins flowing throughout the myometrium are usually detectable (Figure 15).

Power Doppler is preferred to color Doppler because in general it is superior for the detection of small vessels with low blood flow velocities. Color Doppler is used to assess the direction of blood flow. Depending on the area of interest, the color or power Doppler box should include the whole or a specific part of the uterus, or be focused on a myometrial lesion. Magnification and settings should be adjusted to ensure maximum sensitivity, and the Doppler gain should be reduced until all colour artefacts disappear. Usually settings allowing the detection of blood flow velocities of 3-9 cm/sec are optimal, but this may vary from one ultrasound machine to another.

The vascular pattern within the myometrium may be uniform or non-uniform (Figure 16).

The vascular pattern of a myometrial lesion may be circumferential, intra-lesional or both circumferential and intra-lesional (Figure 16).

Some lesions are associated with disruption of the normal uterine vasculature, while others are not. Trans-lesional vascularity (Figure 17) is characterized by the presence of vessels, perpendicular to the uterine cavity/serosa crossing the lesion.

The degree of vascularization should be reported using a subjective color score (color score 1 representing no color and color score 4 abundant color signals). The color score is based on the subjective evaluation of both the percentage of the lesion being vascularized and the color hue. The color score is assigned taking into account the lesion as a whole, but in lesions with uneven internal vascularization (e.g. because of cystic areas or central necrosis) the score reflects the degree of vascularization in the solid parts of the lesion. If there is an uneven spread of vascularization in the solid components of the lesion, the score for the most vascularized solid component and the
percentage of the solid components with color signals may be recorded. A color score may be assigned separately to circumferential and intra-lesional vascularity (Figure 18).

When carrying out research studies, the vascularity of lesions may be reported as iso-, hypo- or hyper-vascular compared to the vascularity of the surrounding myometrium. Reporting a lesion’s vascularity may include the number of vessels (single or multiple), vessel size (small and equal, large and equal, unequal; or the vessel diameter may be measured), the direction of vessels (perpendicular or not perpendicular to endometrium), the vessel branching pattern (no branching, regular branching, irregular branching) and may be further specified as outlined in Table 3 and in Figure 19. Irregular branching vessels may be defined as abnormal tortuous vessels, irregular caliber vessels, a lack of hierarchy in branching with varying branching angles, vessels sprouts and an overall impression of a chaotic vessel pattern. The term circumferential vessels relates to vessels that surround a lesion, whereas vessels located inside a lesion are called intra-lesional.

Again within the context of research, color flow within a lesion may be quantified using 3D ultrasound with virtual organ computer-aided analysis (VOCAL) in order to calculate 3D power Doppler indices: the vascularity index (VI: number of color voxels in the volume expressed as a percentage of the total number of voxels in the volume), the flow index (FI: mean color value in the color voxels expressed as a number from 0-100) and the vascularization-flow index (VI multiplied with FI: VFI reflects the mean color value in all the volume’s voxels expressed as a number from 0-100) potentially reflecting vascularity, flow velocity and tissue perfusion, respectively. However, because 3D vascular indices depend on machine settings, there remains doubt about their reproducibility and their clinical use has yet to be adequately explored. Until the pitfalls with these indices have been resolved, we recommend not using them outside the context of a specific research project.

Scanning the Myometrium in General Clinical Practice

In general clinical practice reporting on the myometrium may be more succinct, as summarized in Table 4.

The uterine corpus is measured, the symmetry of the myometrial walls is estimated and the overall echogenicity of the myometrium is reported as homogenous or heterogeneous.
In the presence of a myometrial lesion, it is specified if the lesion is well defined or ill defined. The number (or the estimated number if > 4 lesions) of lesions is reported as well as the location, the site and maximal diameter of the clinically relevant lesion(s).

The presence of shadowing, myometrial cysts, hyperechogenic islands or subendometrial echogenic lines and buds is reported.

The junctional zone is reported as ‘regular’ or ‘poorly defined’ (if irregular, interrupted, not visible or not assessable).

When it is clinically relevant to evaluate vascularity, the overall vessel pattern within the myometrium of the whole uterus is reported as uniform or non-uniform. The amount of color within a lesion is reported using the color score (1 = no color; 2 = minimal color; 3 = moderate color; 4 = abundant color).

We propose to include the following ultrasound pictures when reporting on the myometrium (ultrasound images of the endometrium should be described using the IETA terminology):

- at least one mid-sagittal section of the uterus (gray scale and with power Doppler)
- preferably also a transverse section and/or a coronal 3D-reconstruction of the uterus
- if a lesion is seen, at least one section of the uterus including the lesion
- preferably also a detailed (zoomed) image of the lesion (in gray scale and with power Doppler)
- for the mapping of fibroids 3D-imaging showing the three standard orthogonal planes through the uterus or tomographic ultrasound images (TUI) may sometimes be illustrative.
ULTRASOUND FINDINGS ASSOCIATED WITH PATHOLOGY

In this section we describe ultrasound features that in the opinion of the authors and on the basis of reports in the literature, are thought to be associated with pathology and in particular with fibroids and adenomyosis (see summary in table 5). Further research should validate the importance of each of these features.

ADENOMYOSIS

Adenomyosis is caused by a proliferation of endometrial glands and stroma leading to ill-defined lesions within the myometrium. Adenomyosis may be present on one or more sites within the uterine wall or involve most of the myometrium and may often be dispersed within the myometrium rather than forming a confined lesion: i.e. diffuse adenomyosis. On the other hand if adenomyosis is present in only one part of the myometrium, it is called focal adenomyosis. In rare cases it may present as a large cyst (an adenomyotic cyst or cystic adenomyoma)\textsuperscript{20-24}. On histological examination, adenomyosis is classified as diffuse when endometrial glands or stroma are distributed diffusely in the myometrium, and focal when circumscribed nodular aggregates are seen. Focal adenomyosis is not the same as an adenomyoma. These are defined by pathologists as focal adenomyosis with additional compensatory hypertrophy of the surrounding myometrium\textsuperscript{25}. The ultrasound features of adenomyosis (Figure 20) should be reported and quantified (Table 1-3). The ultrasound features of a globular uterus with ill-defined adenomyotic lesions may be explained by direct invasion of endometrial tissue from the endometrium as seen in “classic adenomyosis”, or invasion from endometriotic implants on the serosal surface of the uterus\textsuperscript{26}. More seldom diffuse adenomyosis may be localized as a solitary finding without direct continuation with the serosa or the endometrium\textsuperscript{22}. The proportion of endometrial glandular structures, endometrial stroma and hypertrophic muscle elements within a lesion probably explains the different ultrasound features reported to be typical of adenomyosis. The link between the ultrasound features and histopathology has to be demonstrated and specified\textsuperscript{27}. 
FIBROID (LEIOMYOMA)

A uterine fibroid is typically seen on ultrasound as a well-defined, round lesion within the myometrium or attached to it, often showing shadows at the edge of the lesion and/or internal fan-shaped shadowing (Figure 21). The echogenicity varies and some internal high echogenicity may be present. At color or power Doppler imaging, circumferential flow around the lesion is often visible. However some fibroids do not exhibit such typical features. We suggest that such fibroids are labelled sonographically atypical fibroids (Figure 22).

On histological examination, fibroids are composed of smooth muscle cells and connective tissue in densely packed whorls. Acoustic shadows may arise from the interface between smooth muscle bundles, hyalinized connective tissue and normal myometrium. The ultrasound appearances of a fibroid may depend on the proportion of muscle cells and fibrous stroma within the lesion.

VARIANTS OF FIBROIDS AND OTHER UTERINE SMOOTH MUSCLE TUMORS

VARIANTS OF FIBROIDS

Fibroids may undergo degeneration. This may be spontaneous or a result of induced infarction following uterine artery embolization. Coagulate necrosis is induced after high-intensity ultrasound or radio frequency ablation. Types of degeneration are: a) red, b) hyalin and c) cystic / myxoid (myxoid leiomyoma) or d) hydropic. Spontaneous degeneration may occur in pregnancy, and red degeneration is an initial manifestation within days after infarction. The sonographic appearance of red degeneration may be unremarkable, although some cases of red degeneration have been reported as homogenous lesions with low echogenicity, a hyperechogenic rim and absent internal vascularity. Hemorrhage and edema in these fibroids may give rise to tumors of mixed echogenicity. Late manifestations after infarction are most commonly hyaline degeneration. Fibroids after induced infarction are often uniform, hypoechoic, with a hyperechogenic rim with acoustic shadows. There is usually no internal vascularity or at most a few disparate vessels. After spontaneous hyaline infarction fibroids may show mixed echogenicity or hypoechoic cystic areas. Cystic or myxoid degeneration may
develop, resulting in regular hypoechoic cystic areas with fluid or myxoid content. Degeneration may also occur in malignant uterine smooth muscle tumors.

**Uterine Sarcomas and Other Uterine Smooth Muscle Tumors:**

The prediction of malignancy is of utmost importance. However, data on the prediction of uterine sarcoma by ultrasound examination are scarce and mainly based on small retrospective case series, precluding definitive guidelines.

There are many rare uterine smooth muscle tumors other than benign leiomyomas, but only limited information on their ultrasound features has been reported to date. This issue has become increasingly important in view of the debate about when, or if, fibroids may be morcellated during laparoscopic surgery.

**Malignant sarcomas** comprise leiomyosarcoma (Figure 23), endometrial stromal sarcoma, adenosarcoma and undifferentiated sarcoma. Uterine sarcomas present as purely myometrial lesions and are typically single, large tumors. Their ultrasound features may be indistinct from ordinary fibroids or they may appear as an irregularly vascularized mass, with a regular or irregular outline often with irregular anechoic areas due to necrosis.

**Uterine smooth muscle tumor of uncertain malignant potential (STUMP).**

There are no specific ultrasound features described for STUMP. Intravenous leiomyomatosis, disseminated peritoneal leiomyomatosis and benign metastasizing leiomyoma have the same ultrasound features as ordinary fibroids. There are often multiple fibroid and they may be recognized by their location outside uterine borders. These multiple fibroid should be distinguished from “diffuse leiomyomatosis”.

**Fibroids with little or no recurrent and/or metastatic potential:**

Ultrasound features of leiomyoma with bizarre nuclei (bizarre/symplastic/typical leiomyoma); mitotically active leiomyoma; cellular and highly cellular leiomyoma; dissecting leiomyoma; leiomyoma with increased cellularity, no atypia nor mitotic figures and increased vascularity may have the same macroscopically pathologic features as fibroids and may have increased vascularity, as this feature seems to be related to cellularity.
A cotyledonoid leiomyoma or cotelleydonoid dissecting leiomyoma\textsuperscript{56-58} is a nodular tumor with placenta-like echogenicity at ultrasound, but it may also be cystic. Ultrasonographic features of lipoleiomyoma\textsuperscript{59} comprise a hyperechogenic mass partly encased by a hypoechogetic rim. Ultrasonographic features for epithelioid leiomyoma\textsuperscript{40} and pallisading / neurilemoma-like leiomyoma\textsuperscript{60} have not been described.
DISCUSSION

The terms and definitions presented in this paper aim to facilitate consistent reporting of myometrial lesions when using ultrasonography in both daily clinical practice and for research purposes. Clearly the clinical relevance of some of the terms that have been proposed have not yet been evaluated in prospective clinical studies. We acknowledge that some aspects of the systematic reporting we have suggested may require a relatively high level of ultrasound training. We also acknowledge that some of the proposed terms and definitions are too detailed for use in general clinical practice and will initially be suitable only for use in research settings. Future research should focus on the ability to predict specific pathologies and on the clinical relevance of the ultrasound features described in this paper. Although the members of panel involved in the writing of this consensus have different fields of expertise including gynecological ultrasonography, fertility treatment, hysteroscopy, general gynecology and clinical research, we acknowledge that they all come from Europe and the USA, leaving most of the areas of the world unrepresented.

The recent controversy about the safety of morcellation of lesions thought to be benign fibroids, but turning out to be malignant\(^5\), highlights the importance of the reliable preoperative characterization of myometrial lesions.

Although recognizing a typical fibroid on ultrasound is usually straightforward, differentiating between an atypical fibroid and a uterine sarcoma remains challenging. The establishment of an international database of ultrasound and Magnetic Resonance Imaging (MRI) images of uterine sarcomas and rare uterine tumors would be of great clinical value.

Adenomyosis may be difficult to diagnose with ultrasound. Different ultrasound features have been suggested to be associated with adenomyosis but at present it is not clear which are the minimal ultrasound criteria for diagnosis. Some features may carry a greater diagnostic weight than others\(^6\) and the presence of more than one ultrasound feature associated with adenomyosis might increase the likelihood of the diagnosis\(^6\). We did not include the so called \(s\)-sign suggested to be typical of adenomyosis in our consensus statement, because this sign occurs when there is also deep infiltrative endometriosis in the posterior compartment\(^6\).
The terms that we suggest to characterize the JZ are derived from MRI studies. The JZ is better visualized by 3D than by 2D ultrasound. The clinical implications of a thickened JZ or of JZ disruption according to ultrasound needs to be established.

The clinical relevance of myometrial lesions for abnormal uterine bleeding, pelvic pain, subfertility and pregnancy outcome is an important topic for research. Certain ultrasound features might prove to be more clinically relevant than others.

The role of a systematic evaluation of the ultrasound features of myometrial lesions when choosing management (expectant management, medical therapy, selective embolization, High-Intensity Focused Ultrasound or surgical treatment) and in the follow-up during or after treatment is another important topic for future research.

To conclude, the terms and definitions in this consensus statement enable clinicians to write a structured report when describing the ultrasound appearance of the myometrium and myometrial lesions and harmonize nomenclature for future research.

**Acknowledgements**

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*Dirk Timmerman is Senior Clinical Investigator of Scientific Research Fund (FWO) Flanders.*


40. Ip PP, Tse KY, Tam KF. Uterine smooth muscle tumors other than the ordinary leiomyomas and leiomyosarcomas: a review of selected variants with emphasis on recent advances and unusual morphology that may cause concern for malignancy. *Adv Anat Pathol* 2010; 17:91-112.


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Table 3: Reporting the vascularity of the myometrium

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Table 1: Reporting the myometrium

<table>
<thead>
<tr>
<th>FEATURE TO BE DESCRIBED</th>
<th>DESCRIPTION/TERM</th>
<th>QUANTIFICATION/MEASUREMENT</th>
</tr>
</thead>
<tbody>
<tr>
<td>Uterine corpus (Fig. 1)</td>
<td>Length, antero-posterior diameter, transverse diameter, volume</td>
<td>Length = [fundus] + [cavity] (d1), antero-posterior diameter (d2), transverse diameter (d3), ( V ) (cm(^3)) = ( d1 \times d2 \times d3 \times 0.523 )</td>
</tr>
<tr>
<td>Uterus corpus and cervix (Fig. 1)</td>
<td></td>
<td>Total length = [fundus] + [cavity] + [cervix] = ( d1 + c )</td>
</tr>
<tr>
<td>Serosal contour (Fig. 2a+b)</td>
<td>Regular / lobulated</td>
<td></td>
</tr>
</tbody>
</table>

MYOMETRIUM

- **Myometrial walls (Fig. 2b)**
  - Symmetrical / asymmetrical
  - Ratio or subjective impression of asymmetry

**MYOMETRICAL LESIONS**

<table>
<thead>
<tr>
<th>Feature</th>
<th>Description/Term</th>
<th>Quantification/Measurement</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number*</td>
<td></td>
<td>Exact number: n</td>
</tr>
<tr>
<td>Location*</td>
<td>Location: anterior, posterior, fundal, right lateral or left lateral, global</td>
<td></td>
</tr>
<tr>
<td>Site (Fig. 7)*</td>
<td>Site [for well-defined lesions]: FIGO-classification 1-7</td>
<td></td>
</tr>
<tr>
<td>Size*</td>
<td></td>
<td>Three perpendicular diameters (a1,a2,a3), and/or Volume (V) ( V = a1 \times a2 \times a3 \times 0.523 )</td>
</tr>
<tr>
<td>Outer lesion free margin (OFM) (Fig. 8)</td>
<td></td>
<td>The minimal distance between the serosal surface and the outermost border of the lesion</td>
</tr>
<tr>
<td>Inner lesion free margin (IFM) (Fig. 8)</td>
<td></td>
<td>The minimal distance between the endometrium and the inner border of the lesion</td>
</tr>
<tr>
<td>Penetration of ill-defined lesions (Fig. 9)</td>
<td>Ratio between thickness of lesion and the total uterine wall thickness, measured on the same image</td>
<td>Penetration = maximal diameter of the lesion perpendicular to the endometrium / maximal wall thickness perpendicular to the endometrium</td>
</tr>
<tr>
<td>Extent of ill-defined lesions</td>
<td>Localized (&lt; 50% of total uterine volume involved) or diffuse (≥ 50% of total uterine volume involved)</td>
<td>% of myometrium volume involved</td>
</tr>
<tr>
<td>Echogenicity (Fig. 10a, Fig. 10b)</td>
<td>Uniform: hypo-, iso-, hyper-echogenic Non-uniform: mixed echogenicity, cystic areas (regular/irregular); anechogenic, low level, ground glass, mixed echogenicity of cyst fluid</td>
<td>Very hypo-echogenic-, hypo-echogenic-, iso-echogenic, hyperechogenic+, very hyperechogenic++</td>
</tr>
<tr>
<td>Rim (Fig. 11)</td>
<td>Hypo- or hyper-echogenic, or ill-defined</td>
<td></td>
</tr>
<tr>
<td>Shape (Fig. 11)</td>
<td>Round / Not round: oval, lobulated, irregular</td>
<td></td>
</tr>
<tr>
<td>Shadowing (Fig. 12a)</td>
<td>Edge shadows</td>
<td>Degree of shadowing: slight, moderate, strong</td>
</tr>
<tr>
<td>Internal shadows</td>
<td>Present / Absent</td>
<td>Degree of shadowing: slight, moderate, strong</td>
</tr>
<tr>
<td>Fan shaped shadowing (Fig. 12b)</td>
<td>Present / Absent</td>
<td>Degree of shadowing: slight, moderate, strong</td>
</tr>
<tr>
<td>Cysts (Fig. 13a)</td>
<td>Size</td>
<td>Maximum diameter of largest cyst</td>
</tr>
<tr>
<td>Number of cysts</td>
<td>Exact number or (single, 1-5, &gt;5)</td>
<td></td>
</tr>
<tr>
<td>Echogenicity</td>
<td>Cyst fluid: anechogenic, low level, ground glass, mixed echogenicity Hyperechogenic rim: Present/Absent</td>
<td></td>
</tr>
<tr>
<td>Hyperechogenic islands (Fig. 13b)</td>
<td>Outline</td>
<td>Regular, irregular or ill defined</td>
</tr>
<tr>
<td>Size</td>
<td>Maximum diameter</td>
<td></td>
</tr>
<tr>
<td>Number</td>
<td>Exact number or (single, 1-5, &gt;5)</td>
<td></td>
</tr>
<tr>
<td>Subendometrial echogenic lines &amp; buds (Fig. 14)</td>
<td>Number</td>
<td>Exact number or (single, 1-5, &gt;5)</td>
</tr>
</tbody>
</table>

Definitions of the terms and their quantifications are described in the text and illustrated by ultrasound images and schematic drawings. Those with a white background are items of importance in daily clinical practice; while items on a shaded background are of interest for research purposes. Measurements are reported in mm or cm (to tenths of a cm).* if clinically relevant (e.g. preoperative workup before myomectomy)
Table 2: Reporting the Junctional Zone (JZ)

<table>
<thead>
<tr>
<th>STRUCTURE TO BE DESCRIBED</th>
<th>DESCRIPTION</th>
<th>MEASUREMENT</th>
</tr>
</thead>
</table>
| Junctional zone           | Regular / irregular / interrupted / not visible / not assessable | maximum JZ thickness \((\text{JZ}_{\text{max}})\) in mm  
  minimal JZ thickness\((\text{JZ}_{\text{min}})\) in mm  
  or ratio \(\text{JZ} / \text{total myometrial wall thickness}\) |
| Irregular or interrupted  | Location: anterior, posterior, 
  fundus, lateral right, lateral left, or global | Magnitude of irregularity:  
  \((\text{JZ}_{\text{max}}) - (\text{JZ}_{\text{min}}) = \text{JZ}\text{dif}\)  
  Extent of irregularity: % of JZ being irregular  
  \(< 50\% \text{ or } > 50\%\) |
| junctional zone            |             |             |
| Interrupted junctional     | Location: anterior, posterior, 
  fundus, lateral right, lateral left, or global | Interruption of JZ: % of JZ not visualized \(< 50\% \text{ or } > 50\%\) |
| zone                      |             |             |
| Irregularity of JZ         | Cystic areas, hyperechogenic 
  dots, hyperechogenic buds and 
  lines (in each location) |             |

Definition of the terms and their quantifications are described in the text and illustrated by ultrasound images and schematic drawings (Figure 5 and 6). With a white background are items of importance in daily clinical practice, while items on a shaded background are of interest for research purposes.
Table 3: Reporting the vascularity of the myometrium

<table>
<thead>
<tr>
<th>Vascularization to be assessed</th>
<th>Description</th>
<th>Measurement</th>
</tr>
</thead>
<tbody>
<tr>
<td>Whole uterus</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Overall vessel pattern</td>
<td>Uniform / non-uniform</td>
<td></td>
</tr>
<tr>
<td>(Fig. 16)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lesions</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Amount of color (in a lesion)</td>
<td>Color score (both the percentage of the lesion being vascularized and the color hue are taken into account)</td>
<td>no color (1); minimal color (2); moderate color (3); abundant color (4)</td>
</tr>
<tr>
<td>(Fig. 18)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>In case of uneven spread of</td>
<td>Color score in most vascularized part</td>
<td>no color (1); minimal color (2); moderate color (3); abundant color (4)</td>
</tr>
<tr>
<td>vascularization</td>
<td>Percent of solid tissue with color signal</td>
<td>0-100%</td>
</tr>
<tr>
<td>Compared to adjacent myometrium</td>
<td></td>
<td>iso-, hypo-, or hyper-vascularity</td>
</tr>
<tr>
<td>Location of vessels</td>
<td>Circumferential / intra-lesional</td>
<td></td>
</tr>
<tr>
<td>(Fig. 16, 17)</td>
<td>Uniform / not uniform (areas with increased / decreased vascularity)</td>
<td></td>
</tr>
<tr>
<td>Vessel morphology</td>
<td>Number: single / multiple</td>
<td></td>
</tr>
<tr>
<td>(Fig. 16, 19)</td>
<td>Size: large and equal / small and equal / unequal</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Branching: regular, irregular, no branching</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Direction: perpendicular, not perpendicular</td>
<td></td>
</tr>
</tbody>
</table>

Items with a white background are of importance in daily clinical practice; while items on a shaded background are of interest for research purposes.
### Table 4: Reporting the myometrium in general clinical practice

<table>
<thead>
<tr>
<th>Feature to be described</th>
<th>Description/term</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Uterine Corpus</strong></td>
<td>Length, antero-posterior diameter, transverse diameter</td>
</tr>
<tr>
<td>Myometrial walls</td>
<td>Symmetrical / asymmetrical</td>
</tr>
<tr>
<td>Overall echogenicity</td>
<td>Homogeneous / heterogeneous</td>
</tr>
<tr>
<td><strong>Myometrial Lesions</strong></td>
<td>Well defined / ill defined</td>
</tr>
<tr>
<td>Number</td>
<td>Number (1, 2, 3 or estimation in case &gt; 4 lesions)</td>
</tr>
<tr>
<td>Location</td>
<td>Location of the largest/clinically relevant lesion(s): anterior, posterior, fundal, right lateral or left lateral, global</td>
</tr>
<tr>
<td>Site</td>
<td>Site (for well-defined lesions) of the largest/clinically relevant lesion(s): FIGO-classification 1-7</td>
</tr>
<tr>
<td>Size</td>
<td>The maximal diameter of the largest/clinically relevant lesion(s)</td>
</tr>
<tr>
<td>Shadowing</td>
<td>Edge shadows: Present / Absent</td>
</tr>
<tr>
<td></td>
<td>Internal shadows: Present / Absent</td>
</tr>
<tr>
<td></td>
<td>Fan shaped shadowing: Present / Absent</td>
</tr>
<tr>
<td>Cysts</td>
<td>Present / Absent</td>
</tr>
<tr>
<td>Hyperechogenic islands</td>
<td>Present / Absent</td>
</tr>
<tr>
<td>Subendometrial echogenic lines &amp; buds</td>
<td>Present / Absent</td>
</tr>
<tr>
<td><strong>Junctional Zone</strong></td>
<td>Regular / Poorly defined</td>
</tr>
<tr>
<td><strong>Vascularity of the Myometrium</strong></td>
<td></td>
</tr>
<tr>
<td>Overall vessel pattern (in the whole uterus)</td>
<td>Uniform / non-uniform</td>
</tr>
<tr>
<td>Amount of color (in a lesion)</td>
<td>Color score: (1) no color; (2) minimal color; (3) moderate color; (4) abundant color</td>
</tr>
</tbody>
</table>
Table 5: Features considered important in the diagnosis of fibroids and adenomyosis.

<table>
<thead>
<tr>
<th>FEATURE</th>
<th>TYPICAL FIBROID</th>
<th>ADENOMYOSIS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Serosal contour of the uterus</td>
<td>lobulated or regular</td>
<td>often globally enlarged uterus</td>
</tr>
<tr>
<td>Definition of lesion</td>
<td>well-defined</td>
<td>ill-defined in diffuse adenomyosis (adenomyoma may be well-defined)</td>
</tr>
<tr>
<td>Symmetry of uterine walls</td>
<td>asymmetrical in the presence of well-defined lesion(s)</td>
<td>myometrial anterior-posterior asymmetry</td>
</tr>
<tr>
<td><strong>LESION</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Outline</td>
<td>well-defined</td>
<td>ill-defined</td>
</tr>
<tr>
<td>Shape</td>
<td>round, oval, lobulated</td>
<td>ill-defined</td>
</tr>
<tr>
<td>Contour</td>
<td>smooth</td>
<td>irregular or ill-defined</td>
</tr>
<tr>
<td>Rim</td>
<td>hypo- or hyper-echogenic</td>
<td>no rim</td>
</tr>
<tr>
<td>Shadowing</td>
<td>edge shadows</td>
<td>no edge shadows</td>
</tr>
<tr>
<td></td>
<td>internal shadows (often fan shaped</td>
<td>fan shaped shadowing</td>
</tr>
<tr>
<td></td>
<td>shadowing)</td>
<td></td>
</tr>
<tr>
<td>Echogenicity</td>
<td>uniform: hyper- / iso- / hypo-echogenic</td>
<td>non-uniform: mixed echogenicity&lt;sup&gt;67,68&lt;/sup&gt; cysts&lt;sup&gt;20,24,62&lt;/sup&gt;, hyper-echogenic islands, subendometrial lines and buds&lt;sup&gt;20,63&lt;/sup&gt;</td>
</tr>
<tr>
<td>Vascularity</td>
<td>circumferential flow</td>
<td>translesional flow&lt;sup&gt;69&lt;/sup&gt;</td>
</tr>
<tr>
<td>JUNCTIONAL ZONE (JZ)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>JZ-thickness, regularity</td>
<td>not-thickened, regular or not visible</td>
<td>thickened</td>
</tr>
<tr>
<td></td>
<td></td>
<td>irregular or ill-defined&lt;sup&gt;6,61-63&lt;/sup&gt;</td>
</tr>
<tr>
<td>JZ interruption</td>
<td>interrupted or overstretched junctional zone in areas with lesions of FIGO type 1-3 (Fig. 7)</td>
<td>interrupted junctional zone (even in absence of localized lesions)&lt;sup&gt;9&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>67,68</sup>
Myometrium paper: FIGURES

Figure 1: Measurement of the uterus.

Figure 1. Schematic drawings showing how to measure the uterus. The total length of the uterine corpus (d1) is the sum of the fundus length (a) and the cavity length (b): \[d1 = a + b\]; AP is the largest antero-posterior diameter (d2); T is the largest transverse diameter (d3). The total length of the uterus is the sum of the corpus (d1) and the cervix length (c). The volume (cm³) of the corpus uteri is calculated as \[d1(cm) \times d2(cm) \times d3(cm) \times 0.523\].

Figure 2: The serosal contour of the uterus

Figure 2. Schematic drawings illustrating how to describe the serosal contour of the uterus. The serosal contour of the uterus may be regular (upper row) or lobulated (lower row). The definition of a regular uterus is a normal uterus without any myometrial lesions, a uterus with an intramural lesion that does not alter the normal smooth serosal contour of the uterus (yellow arrow). Regular uterus includes for example an asymmetrical uterus (green arrows highlight the thickness of the anterior and posterior myometrial walls) or a globally enlarged uterus (illustrated by dotted line arrows in the upper row of images).

Figure 3: Symmetry of the uterine walls

Figure 3. Schematic drawings illustrating the symmetry of the uterine walls. Symmetrical walls are shown in the upper row and asymmetrical walls in the lower row.

Figure 4: Junctional zone on 3D-ultrasound imaging

Figure 4. Multiplanar view of the uterine corpus obtained by three-dimensional ultrasound. The junctional zone (JZ) can be seen as a dark line just beneath the endometrium (green arrow, and yellow dotted line). The JZ of the anterior and posterior wall is visualized in the A and B plane (upper row left and right), the JZ of the left and right lateral wall and of the fundus in the C plane (lower row).

Figure 5: Junctional zone

Figure 5. These schematic drawings illustrate regular, irregular, interrupted and not visible junctional zone (JZ) displayed in the coronal plane (images to the left) and in the sagittal plane (images to the right).
Figure 6: **Measurement of the junctional zone** (for research purposes)

**Figure 6:** Schematic drawings and ultrasound images illustrating measurement of junctional zone (JZ) thickness. Measurement of $JZ_{\text{max}}$ and $JZ_{\text{min}}$ is illustrated in the schematic drawings (first row, red arrows). $JZ_{\text{max}}$ is the thickness of the JZ where it appears to be at its thickest and $JZ_{\text{min}}$ is where the JZ appears to be at its thinnest after evaluation of the total three-dimensional volume of the uterus. The total wall thickness (yellow arrow) is measured at the site where the JZ is measured to calculate the ratio $JZ$/total wall thickness. The total wall thickness includes the JZ.

In the two-dimensional image (lower row left) the JZ is measured where it appears at its thickest (red arrows). In the coronal view (C-plane) of the uterus (lower row right) the $JZ_{\text{max}}$ in the right lateral wall is marked (yellow calipers).

Figure 7: **The FIGO classification of myomas**

**Figure 7:** The FIGO classification of myomas (Adapted from: Munro MG et al1) should be used to report the site of well-defined localized lesions: 0 = pedunculated intracavitary; 1 = submucosal <50% intramural; 2 = submucosal ≥50% intramural; 3 = 100% intramural, but contacts the endometrium; 4 = intramural; 5 = subserosal ≥50% intramural; 6 = subserosal <50% intramural; 7 = subserosal pedunculated; 8 = other (e.g. cervical, parasitic)1,15

Figure 8: Schematic drawings illustrating the measurement of the inner lesion free margin (IFM) (green arrow and dotted line) and of the outer lesion free margin (OFM) (yellow arrow and dotted line) of a lesion

Figure 9: **Penetration** (for research purposes)

**Figure 9:** Penetration is defined as the ratio between the maximal thickness of the lesion (yellow arrow) and the total uterine wall thickness (red arrow). Both measurements are taken perpendicular to the endometrium on the same ultrasound image. The penetration is measured where the lesion appears to be at its largest. Although the images illustrate sagittal sections through the uterus the measurements to calculate penetration can be taken in any plane.
Figure 10a: Lesion echogenicity

Figure 10a. Schematic drawings and ultrasound images illustrating different types of lesion echogenicity. The echogenicity of a lesion may be uniform (hypo-, iso- or hyper-echogenic) or non-uniform with mixed echogenicity, echogenic areas, or cystic areas.

Figure 10b. Echogenicity of a uniform lesion (for research purposes)

Figure 10b. The mean grey-tone of a lesion may be compared to the adjacent myometrium and classified as hypo-, iso- or hyper-echogenic. Echogenicity may be scored as very hypoechogenic (- -), hypoechogenic (-), isoechogenic, hyperechogenic (+) or very hyperechogenic (++).

Figure 11. Rim and shape of myometrial lesions

Figure 11. Schematic drawings illustrating the rim and shape of myometrial lesions. The rim of a myometrial lesion may be hypo- or hyper-echogenic or ill-defined and its shape may be round or not round, e.g. oval shaped, lobulated or irregular.

Figure 12: Shadowing

Figure 12a. Schematic (upper row) and ultrasound images (lower row) illustrating edge shadowing and internal shadowing. Ultrasound pictures illustrate edge shadows (yellow arrow) and internal shadows (green arrow).

Figure 12b. All ultrasound images illustrating fan shaped shadowing. The image in the lower row also shows an anechogenic myometrial cyst with a hyperechogenic rim surrounding the cyst and acoustic enhancement posterior to the cyst.

Figure 13a-c: Ultrasound images showing a) myometrial cysts (yellow arrow) b) hyperechogenic islands (surrounded by green dotted lines), and c) echogenic spots (yellow arrows).

Figure 14: Ultrasound images illustrating echogenic subendometrial lines and buds. The lines and buds are encircled by green dotted lines.

Figure 15: Normal vascular pattern of the myometrium

Figure 15: Schematic drawings (upper row) and ultrasound images (lower rows) illustrating the normal vascular pattern of the myometrium. The arcuate vessels of the uterus are visible at the periphery of the myometrium, parallel to the uterine serosa. Perpendicular to the arcuate vessels, the radial arteries and veins flowing throughout the myometrium are usually detectable.

Figure 16: Vascular pattern within the myometrium
Figure 16. Schematic and ultrasound images illustrating the vascular pattern within the myometrium and in myometrial lesions. The vascular pattern of the myometrium may be uniform (upper row) or non-uniform (middle row). The vascular pattern of a myometrial lesion may be circumferential, intra-lesional or both circumferential and intra-lesional (lower row).

Figure 17: Translesional vascularity

Figure 17. Ultrasound images and a schematic drawing illustrating translesional vascularity which is defined as vessels perpendicular to the endometrium crossing the lesion.

Figure 18: Color score of circumferential and intra-lesional vascularity.

Figure 18. Schematic images illustrating the color score (amount of color Doppler signals) in the circumference of and inside myometrial lesions. The amount of color is estimated subjectively. The color score is based on the subjective evaluation of both the percentage of the lesion being vascularized and the color hue. A color score of 1 represents no color, 2 minimal amount of color, 3 moderate and 4 abundant amount of color.

Figure 19: Vascularization of a myometrial lesion: vessel number, size, branching and direction (for research purposes)

Figure 19. Schematic drawings illustrating how to describe the vascularization of a myometrial lesion in clinical research, in terms of vessel number, vessel size (depending on the research protocol the vessel diameter may be measured), vessel branching and vessel direction. Circumferential vessels are the vessels surrounding a lesion.

Figure 20: Schematic drawings illustrating the ultrasound features currently considered to be typical of adenomyosis.

Figure 21: Schematic drawings illustrating the ultrasound features currently considered to be typical of uterine fibroids.

Figure 22: Ultrasound images showing fibroids with atypical sonographic features. These fibroid have a non-uniform echogenicity, intralesional anechoic cysts and some have areas with hyperechogenicity. There is an irregular outline of the FIGO type1 fibroid (lower right).

Figure 23: Gray scale and color Doppler images of a sarcoma in the anterior wall of the uterus. The uterine corpus (yellow arrows) is located posteriorly and contains clear fluid (green arrow).
Myometrium paper: FIGURES

Figure 1: Measurement of the uterus.

**Corpus length** (d1)
- a = fundus
- b = cavity

**Corpus anteroposterior (AP) diameter** (d2)

**Corpus transverse (T) diameter** (d3)

**Total length of the uterus**

**Figure 1.** Schematic drawings showing how to measure the uterus. The total length of the uterine corpus (d1) is the sum of the fundus length (a) and the cavity length (b): \[d1 = a + b\]; AP is the largest antero-posterior diameter (d2); T is the largest transverse diameter (d3). The total length of the uterus is the sum of the corpus (d1) and the cervix length (c). The volume (cm³) of the corpus uteri is calculated as \(d1(cm) \times d2(cm) \times d3(cm) \times 0.523\)
**Figure 2: The serosal contour of the uterus**

**Figure 2.** Schematic drawings illustrating how to describe the serosal contour of the uterus. The serosal contour of the uterus may be regular (upper row) or lobulated (lower row). The definition of a regular uterus is a normal uterus without any myometrial lesions, a uterus with an intramural lesion that does not alter the normal smooth serosal contour of the uterus (yellow arrow). Regular uterus includes for example an asymmetrical uterus (green arrows highlight the thickness of the anterior and posterior myometrial walls) or a globally enlarged uterus (illustrated by dotted line arrows in the upper row of images).
Figure 3: Symmetry of the uterine walls

**Figure 3.** Schematic drawings illustrating the symmetry of the uterine walls. Symmetrical walls are shown in the upper row and asymmetrical walls in the lower row.
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**Figure 4.** Multiplanar view of the uterine corpus obtained by three-dimensional ultrasound. The junctional zone (JZ) can be seen as a dark line just beneath the endometrium (green arrow, and yellow dotted line). The JZ of the anterior and posterior wall is visualized in the A and B plane (upper row left and right), the JZ of the left and right lateral wall and of the fundus in the C plane (lower row).
Figure 5: Junctional zone

Figure 5. These schematic drawings illustrate regular, irregular, interrupted and not visible junctional zone (JZ) displayed in the coronal plane (images to the left) and in the sagittal plane (images to the right).
Figure 6: Measurement of the junctional zone (for research purposes)

Figure 6: Schematic drawings and ultrasound images illustrating measurement of junctional zone (JZ) thickness. Measurement of JZ$_{\text{max}}$ and JZ$_{\text{min}}$ is illustrated in the schematic drawings (first row, red arrows). JZ$_{\text{max}}$ is the thickness of the JZ where it appears to be at its thickest and JZ$_{\text{min}}$ is where the JZ appears to be at its thinnest after evaluation of the total three-dimensional volume of the uterus. The total wall thickness (yellow arrow) is measured at the site where the JZ is measured to calculate the ratio JZ/total wall thickness. The total wall thickness includes the JZ.

In the two-dimensional image (lower row left) the JZ is measured where it appears at its thickest (red arrows). In the coronal view (C-plane) of the uterus (lower row right) the JZ$_{\text{max}}$ in the right lateral wall is marked (yellow calipers).
**Figure 7:** The FIGO classification of myomas

**Figure 7:** The FIGO classification of myomas (Adapted from: Munro MG et al²) should be used to report the site of well-defined localized lesions: 0 = pedunculated intracavitory; 1 = submucosal <50% intramural; 2 = submucosal ≥50% intramural; 3 = 100% intramural, but contacts the endometrium; 4 = intramural; 5 = subserosal ≥50% intramural; 6 = subserosal <50% intramural; 7 = subserosal pedunculated; 8 = other (e.g. cervical, parasitic)¹;¹⁵
Figure 8: Schematic drawings illustrating the measurement of the inner lesion free margin (IFM) (green arrow and dotted line) and of the outer lesion free margin (OFM) (yellow arrow and dotted line) of a lesion.
Figure 9: Penetration (for research purposes)

**Figure 9: Penetration is defined as the ratio between the maximal thickness of the lesion (yellow arrow) and the total uterine wall thickness (red arrow). Both measurements are taken perpendicular to the endometrium on the same ultrasound image. The penetration is measured where the lesion appears to be at its largest. Although the images illustrate sagittal sections through the uterus the measurements to calculate penetration can be taken in any plane.**
Figure 10a: Lesion echogenicity

**Uniform Echogenicity**
- Hypo-echogenic
- Iso-echogenic
- Hyper-echogenic

**Non-uniform Echogenicity**
- Mixed echogenicity
- Echogenic areas
- Cystic areas

**Figure 10a.** Schematic drawings and ultrasound images illustrating different types of lesion echogenicity. The echogenicity of a lesion may be uniform (hypo-, iso- or hyper-echogenic) or non-uniform with mixed echogenicity, echogenic areas, or cystic areas.
**Figure 10b.** Echogenicity of a uniform lesion (for research purposes)

- **Hypoechogenic**
- **Hypoechogenic**
- **Isoechogenic**
- **Hyperechogenic**
- **Hyperechogenic**

**Figure 10b.** The mean grey-tone of a lesion may be compared to the adjacent myometrium and classified as hypo-, iso- or hyper-echogenic. Echogenicity may be scored as very hypoechogenic (--), hypoechogenic (-), isoechogenic, hyperechogenic (+) or very hyperechogenic (++).
Figure 11. Rim and shape of myometrial lesions

**RIM**

Hypo-echogenic

Hyper-echogenic

**SHAPE**

Round

Not round

Oval shaped

Lobulated

Irregular

Figure 11. Schematic drawings illustrating the rim and shape of myometrial lesions. The rim of a myometrial lesion may be hypo- or hyper-echogenic or ill-defined and its shape may be round or not round, e.g. oval shaped, lobulated or irregular.
**Figure 12a: Shadowing**

**Figure 12a.** Schematic (upper row) and ultrasound images (lower row) illustrating edge shadowing and internal shadowing. Ultrasound pictures illustrate edge shadows (yellow arrow) and internal shadows (green arrow).
**Figure 12b.** All ultrasound images illustrating fan shaped shadowing. The image in the lower row also shows an anechoic myometrial cyst with a hyperechogenic rim surrounding the cyst and acoustic enhancement posterior to the cyst.
Figure 13a-c: Ultrasound images showing a) myometrial cysts (yellow arrow) b) hyperechogenic islands (surrounded by green dotted lines), and c) echogenic spots (yellow arrows).
Figure 14: Ultrasound images illustrating echogenic subendometrial lines and buds. The lines and buds are encircled by green dotted lines.
Figure 15: Normal vascular pattern of the myometrium

**Figure 15**: Schematic drawings (upper row) and ultrasound images (lower rows) illustrating the normal vascular pattern of the myometrium. The arcuate vessels of the uterus are visible at the periphery of the myometrium, parallel to the uterine serosa. Perpendicular to the arcuate vessels, the radial arteries and veins flowing throughout the myometrium are usually detectable.
Figure 16: Vascular pattern within the myometrium

Figure 16. Schematic and ultrasound images illustrating the vascular pattern within the myometrium and in myometrial lesions. The vascular pattern of the myometrium may be uniform (upper row) or non-uniform (middle row). The vascular pattern of a myometrial lesion may be circumferential, intra-lesional or both circumferential and intra-lesional (lower row).
Figure 17: Translesional vascularity

Figure 17. Ultrasound images and a schematic drawing illustrating translesional vascularity which is defined as vessels perpendicular to the endometrium crossing the lesion.
Figure 18: Color score of circumferential and intra-lesional vascularity.

Figure 18. Schematic images illustrating the color score (amount of color Doppler signals) in the circumference of and inside myometrial lesions. The amount of color is estimated subjectively. The color score is based on the subjective evaluation of both the percentage of the lesion being vascularized and the color hue. A color score of 1 represents no color, 2 minimal amount of color, 3 moderate and 4 abundant amount of color.
Figure 19: Vascularization of a myometrial lesion: vessel number, size, branching and direction (for research purposes)

<table>
<thead>
<tr>
<th>Number</th>
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<th>Multiple</th>
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</thead>
<tbody>
<tr>
<td>Size</td>
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<td>Small</td>
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<tr>
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<td>Regular</td>
<td>Irregular</td>
</tr>
<tr>
<td>Direction</td>
<td>Perpendicular</td>
<td>Not perpendicular</td>
</tr>
</tbody>
</table>

**Figure 19.** Schematic drawings illustrating how to describe the vascularization of a myometrial lesion in clinical research, in terms of vessel number, vessel size (depending on the research protocol the vessel diameter may be measured), vessel branching and vessel direction. Circumferential vessels are the vessels surrounding a lesion.
Figure 20: Schematic drawings illustrating the ultrasound features currently considered to be typical of adenomyosis.
Figure 21: Schematic drawings illustrating the ultrasound features currently considered to be typical of uterine fibroids.
Figure 22: Ultrasound images showing fibroids with atypical sonographic features. These fibroid have a non-uniform echogenicity, intrallesional anechoic cysts and some have areas with hyperechogenicity. There is an irregular outline of the FIGO type1 fibroid (lower right).
**Figure 23:** Gray scale and color Doppler images of a sarcoma in the anterior wall of the uterus. The uterine corpus (yellow arrows) is located posteriorly and contains clear fluid (green arrow).