CONSTRUCTION OF A FLUORESCENCE BRONCHOSCHOPE FOR

DETECTION OF EARLY LUNG CANCER

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1985

I LUMENS DALAR

Ack du skrämmande djuplila ljus om blott du ville excitera I lysande fiber ett stilla sus när strålar av ljus penetrera

I skimrande grönt och glödande rött vävnaden tindrar och blänker Detta flämtande ljus varför är det så sprött kan undra vad strålarna tänker

Luminiscens!

Från dunkla kamrar, svartaste svart de smyger så stillsamt, förunderligt lätt Vart ska de hän man frågar sig vart? där fåtal blir mångfald och glest det blir tätt

I visdomens mångbenta bräckliga tempel förtäljes om ljuset som glittrande tändes Budbärare sätt nu raskt Eder stämpel Och du klocka ljud! Och det hördes och kändes

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ABSTRACT

The aim of our work was to construct a fluorescence bronchoscope for in vivo detection of small amounts of Hematoporphyrin derivative (HPD). HPD gathers selectively in tumors after intravenous injection.

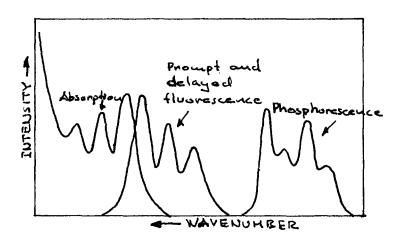
PART 1

THEORY

1. FLUORESCENCE IN MOLECULES

Luminescence is the common name of all light emitting processes, and photoluminescence is the name of light emission caused by absorption of light. Photoluminescence is traditionally divided into phosphorescence and fluorescence.

In a molecule there are more available energy states for the electrons than in an atom, due to the vibrational and rotational movements. As these energy splittings are of lower magnitude than the electronic ones, the vibrational states are given by splitting the electronic states, and the rotational states by splitting the vibrational states. For molecules in solution the rotational levels are broadened and merge together into one continuous band for each vibrational level. This can also be true for vibrational states in complex organic molecules, like Hematoporphyrin, and one obtains energy bands rather than levels. This makes the fluorescence spectrum continuous instead of discrete. Fig. 1.



Multiplicity

A sample of molecules under normal conditions is subject to the Boltzmann distribution:

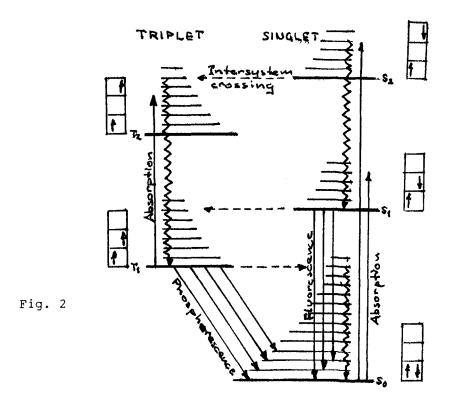
$$N = N_{O} \exp(-E/kT)$$

Here k is the Boltzmann constant, T the temperature, E the energy and N the number of particles. As the energy splitting between vibrational levels is of the order of 0.1 eV and kT at room temperature is about 0.025 eV almost all molecules are in the vibrational ground state.

Nearly all molecules have an even number of electrons, except the free radicals, hence, the resultant spin quantum number must be zero in the ground state, due to the Pauli exclusion principle. If the spin is zero the state is called a singlet state. If an electron is excited to an upper state by interaction with a photon, the spin of the excited electron may be parallel or antiparallel to the electron remaining in the lower level. If they are parallel the resultant spin quantum number is 1/2 + 1/2 = 1. This is a triplet state. The spin vector may have three values -1, 0, 1 when the molecule is situated in an external magnetic field. This gives rise to a small splitting of the energy the excited electron.

Radiative transitions between singlets and triplets are theoretically forbidden in the first order approximation, but the weak spin-orbit coupling gives a small transition propability. The triplet states can

be populated by singlet-triplet transitions. When a triplet state decays radiatively to the ground state the light emission is called phosphorescence and is characterized by a long life time. Fig. 2



Different transitions

A photon is able to interact with an electron if there is an energy interval which corresponds to the energy of the photon. It is known that when an electron is excited to an electronic state higher than the first excited one, the molecule undergoes internal conversions and the molecule goes from a low vibrational level to a high vibrational level in a lower electronic state, fig 2. The molecule will fast lose its vibrational energy by collisions with solvent molecules, Fig. 2. Transitions caused by internal conversions and collisions are radiationless. Some substances may undergo photochemical reactions when raised to upper excited states, for example in the case of photosynthesis.

Conclusion: Most electrons come to the first excited electronic level, where they gather due to a relatively long life time. In that state the molecule can either undergo a chemical reaction or decay to any of the levels in the ground state with light emission. The fluorescence light spectral distribution will depend on the vibrational level distribution. This explains why absorbed photons of different energy often give rise to similar fluorescence.

We now deal with the problem why not all transitions from excited electronic states fall to the lowest vibrational level in the state below. The energy levels of a molecule changes during the vibration. In Fig. 3 is the energy plotted as a function of the distance between the atoms in a diatomic molecule.

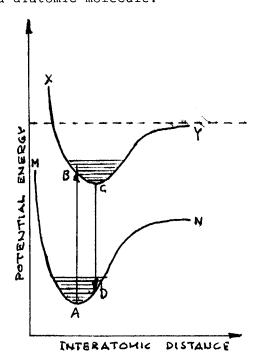


Fig. 3

The Frank-Condon principle is stated as follows. The interatomic distance does not change during a light emission or absorption, i.e. a transition is vertical in the diagram above. If the molecule absorbs a photon when the atoms are at a distance r_1 the electron makes a transition from A \rightarrow B. After some collisions the electron will be

radiationless transitions the interatomic distance has changed to r₂ and correspondingly the energy of the lowest level in the ground state has increased, i.e. the emitted photons have less energy than those absorbed, if excited from the lowest level (Stokes' law). The number of emitted photons is proportional to the number of absorbed ones.

At last the fluorescence quantum efficiency function $\Phi_{\mbox{$\lambda$}}$ ($\mbox{$\lambda$})$ is defined

$$\Phi_{\lambda_{\rm exc}}(\lambda) = \frac{\text{photons emitted at wavelength } \lambda}{\text{photons absorbed at exc. wavelength } \lambda_{\rm exc}}$$

hence, the fluorescence quantum efficiancy, Y.

$$\Psi_{\lambda_{\text{exc}}}(\lambda) = \int_{\lambda_{\text{exc}}}^{\infty} d\lambda \cdot \Phi_{\lambda_{\text{exc}}}(\lambda)$$

Ref. 1.

2. HEMATOPORPHYRIN DERIVATIVE, HPD

Porphyrins are naturally occuring dyes which consists of chained pyrol rings, Fig. 4. Porphyrins are able to bind small molecules like Oxygen, Nitrogen, Carbon dioxid, Nitrogen monoxide etc. They are important in biological systems and are among other things included in enzymes, chlorophyll and haemoglobin. Porphyrins are often photodynamically active in vitro.

Fig. 4

Hematoporphyrin (HP) is a metabolic product from haemoglobin. Its empirical formula is C $_{3\,4}$ H $_{3\,8}$ O $_{6}$ N $_{4}$ and its molecular weight is 598.7, Fig. 5.

Fig. 5

By a special treatment of HP with acetic acid one obtains Hematoporphyrin derivative (HPD). This agent has proved to be much more photodynamically efficient than HP, in vivo. Pure HP is probably photodynamically inactive in vivo.

HPD is not a pure chemical substance. Its components have been investigated, but not all are indentified. In vitro Hematoporphyrin-diacetate is the photodynamically most active component. When injecting HPD in humans, the substance is solved in a solvent in which acetates do not survive. Thus in vivo there must be other, still unknown, active components. Recent investigations indicate that dihematoporphyrin ether is the most active component.

HPD has three important properties:

Ιt

- 1) accumulates in malignant tissue
- 2) causes necrosis of the tissue when irradiated with light at a proper wavelength
- 3) Fluoresces characteristically when excited.
- 1) HPD has a different excretion in malignant vs normal tissue. After the HPD is injected intravenously it will be taken up by all tissue. The normal tissue excrete the HPD faster than malignant tissue. The origin of the selective retention has been suggested to be due to the high vascular permeability together with the lack of adequate lymphatic drainage of tumors. After 2-3 days the ratio between the HPD concentration in tumor vs in normal tissue is about 10 or even higher. It takes about one month until the HPD has been totally excreted. (During this period the patient must stay out of sunlight).

† According to several papers there is also a difference in the uptake of HPD for normal vs malignant tissue.

HPD is also accumulated in areas of moderate and marked squamos cellular atypia. Thus diagnostic methods using HPD are fully reliable only if combined with biopsy and subsequent pathological investigation.

- 2) When light is absorbed in a HPD-molecule it is excited to a higher energy state (¹HPD*). The excited HPD reacts with oxygen in the tissue. This reaction produces aggressive singlet oxygen, which causes necrosis, Eq. 1 and Fig. 6.
- * HPD + $h\nu \rightarrow {}^{1}\text{HPD}^{*}$ abs. \rightarrow exc. molecule
- * $^{1}\text{HPD}^{*} \rightarrow ^{3}\text{HPD}^{*}$ singlet \rightarrow triplet transition (radiationless)
- * $^{3}\text{HPD}^{*}$ + $^{3}\text{O}_{2}$ \rightarrow $^{1}\text{O}_{2}$ + HPD exchange reaction with "normal" oxygen
- * $^{1}\mathrm{O}_{2}$ + substrate \rightarrow oxidation the singlet oxygen affects the mitochondria and the cell membranes.

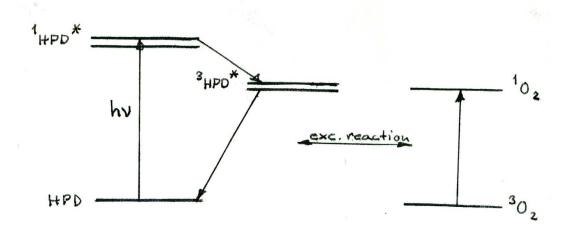


Fig. 6

3) When HPD is irradiated with light of a proper wavelength it will fluoresce with a characteristic spectrum in the red region. Absorption and emission spectra are shown in Fig. 7. The fluorescence quantum efficiency, Ψ , in HPD is Ψ = 0.01 (defined on page 10).

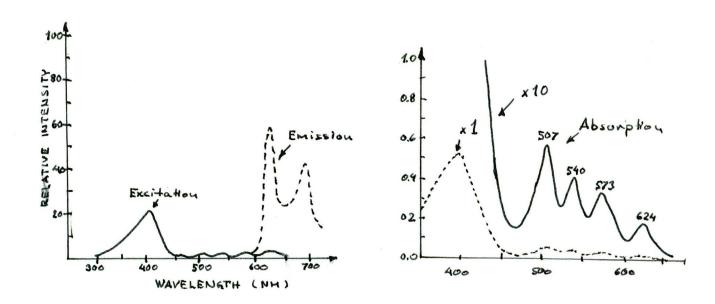


Fig. 7a (Ref. 2)

Fig. 7b (Ref. 3)

The fluorescence component in HPD is not the same as the one which

causes necrosis. It would be a great advantage if one could isolate them from each other. Then it would be possible to increase the diagnostic sensitivity, using more light. At present the diagnostic light dose has to be kept far below the treatment dose which is $10-20 \text{ J/cm}^3$.

Wanting to find the characteristic red fluorescence from HPD in vivo one must separate it from the fluorescence of the tissue. Fig. 8 shows a typical spectrum from tissue, excited with a N_2 -laser (337 nm).

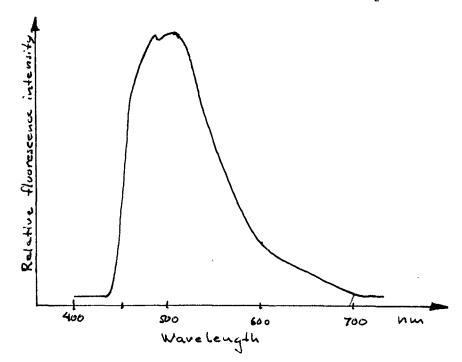


Fig. 8

Malignant tissue fluoresces less than normal, except in the red region, Ref. 4.

PART 2

THE APPARATUS

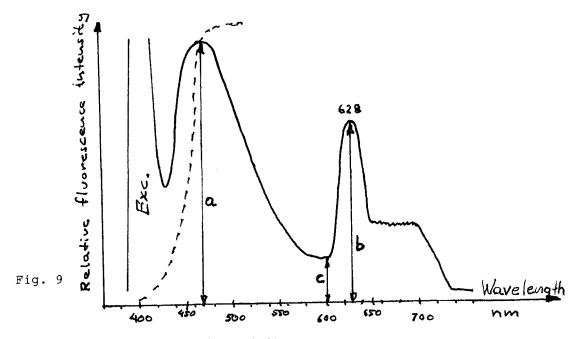
3. THE OPTICAL SYSTEM

When we began designing the equipment, which was to be used for the localization of occult lung tumours, we had as a starting point the apparatus built of Kinsey, Cortese and Sanderson, Ref. 5. In the following sections we discuss the choice of every important part in the equipment.

- 1) Choice of function of measure.
- 2) Light source.
- 3) Light transportation.

1) Choice of function of measure

We wanted to locate the two most significant wavelengths in the fluorescence spectrum, Fig. 9.



Kinsey et.al. measured the red fluorescence.

We considered two alternatives:

- * The fluorescence at b and c is measured using two narrrow interference filters, calculate b-c.
- * The fluorescence around a and b respectively is measured, using two broadband filters, calculate b/a

We chose the latter alternative because b/a is independent of the distance between the bronchoscope and the searched area in the lung. A third alternative, and possibly an improvement, would be to choose (b-c)/a as the measured quantity.

2) Light source

We need a light source with the following properties:

- * High intensity at λ =400 nm for excitation
- * A small emitting area so we could focus it on an optical fiber bundle
- * Emitting white light, for the ocular inspection through the bronchoscope.

Although the HPD excitation spectrum has some peaks at wavelenghts longer than 400 nm, we decided to use the latter absorption peak because it is stronger than the others and we have no use for the larger penetration depth of the longwave light since lung tumors are

normally situated at the surface of the bronchius.

We investigated the following light sources: Halogen lamps, lasers, high pressure lamps, light emitting diodes.

A halogen lamp will not be suitable as an excitation source, but may be used for illumination.

Some lasers are excellent as excitation source but they are very expensive.

A light emitting diode has many advantages. It is cheap, easy to pulse and only demands a low voltage source. After a slight market research we realized that light emitting diodes at 400 nm do not exist. We also tried to find a diode which fitted with some of the lower peaks in Fig. 7, but none of them had the neccessary emittance (light power/-unit area).

We investigated the possibility of focusing the light from many diodes. After a theoretical consideration we realized that this was not possible for fundamental physical reasons as the area of the P-N transition in the diode is of the same size as the fiberbundle. One soon reaches the limit of the number of emitting areas which can be superimposed. So, we had to abandon the idea of using light emitting diodes for the excitation. The diode we searched for will maybe exist in a couple of years.

High pressure lamps comply with our demands. We decided to use a 200 W Mercury lamp. This lamp has the following benefits:

* Plenty of light in the vicinity of 400 nm, due to a strong line in the Hg-spectrum. Fig. 10.

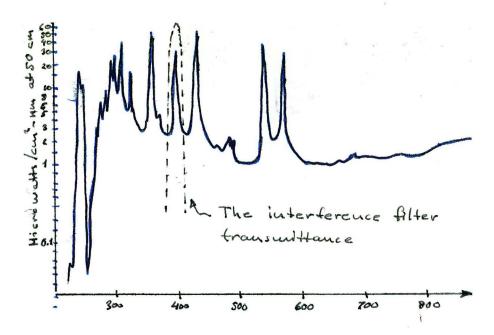


Fig. 10

- * Due to pressure broadening it has a quasi-continuous spectral distribution. This quasicontinuum is our white light utilized in the ocular inspection phase.
- * Long life-time
- * The light-arc between the electrodes has the neccessary high emittance.

Drawbacks:

- * It must operate in a lamp housing due to the explosion hazard.
- * Due to the radiation below 200 nm, it produces ozone, which is toxic.

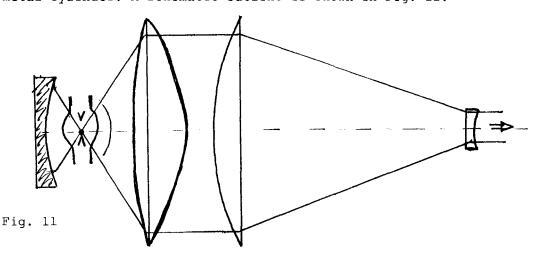
* Rather expensive as it demands housing and a high voltage support.

In order not to obscure the weak fluorescence light the excitation line is separated from the remaining lamp spectrum with an interference filter, Fig. 10.

3) Light transportation

The Lens tube.

The chosen light source radiates isotropically and we want to focus as much as possible of the light into the acceptance cone of the fibre optic bundle. We chose a positive lens having a short focal length and fixed it on the lamp housing with the light arc in the focal plane. A fused silica lens of aspherical type was selected because spherical abberation would be substantial for a spherical lens when operating at such low F-numbers. The collected solid angle is doubled, by placing a spherical mirror on the opposing side of the arc with this set-up the resulting collection efficiency is estimated to be 29%. For practical reasons the light beam is compressed down to a diameter of less than one inch, using two additional lenses. The lens system is placed in a metal cylinder. A schematic outline is shown in Fig. 11.



Combination of illumination and fluorescence measurement

Our aim is to use the white light from the lens tube partly to excite and partly for illumination. As we cannot measure the faint fluorescence and at the same time illuminate with the white light, we have to use a chopping technique. Following Ref. 4 a chopper wheel was used. The chopper wheel was driven by a strong one pole asynchronous electrical motor. As we wanted a rotating frequency which was not a multiple of the mains system, we added a gear and got a rotating frequency at 16 2/3 Hz. The wheel was mounted in a housing. The chosen frequency was expected to be high enough not to cause visual flicker. Unfortunately ones see a slight flicker. After passing the wheel, the incoming light is focused on the entrance of the fiber optic probe.

Travelling in the opposite direction, the collected fluorescence light is transformed to parallel light. On the other side of the chopper wheel a photomultiplayer tube (PMT) is mounted, transforming the fluorescence light into an electric current.

Since we want to measure the red and the green fluorescence while simultaneously exciting with the violet light as well as illuminating with white light at alternate cycles the chopper wheel was configured as shown in Fig. 12. The filter are fastened on the wheel.

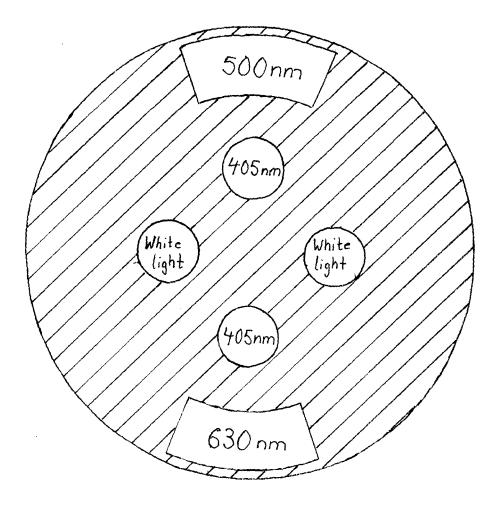


Fig. 12

As the white * light path from the lens tube to the fiber optic probe is opened the fluorescence light path to the PMT is blocked. This is the visual inspection phase. When the wheel has rotated a quarter of a revolution the excitation light path opens and simultaneously the fluorescence is transmitted to the PMT through the red or the green filter.

* The white light passes a cut-off filter blocking radiation below 435 nm, in order to minimize the damage of the examined tissue.

Also, the eye's sensitivity is low at these wavelenghths.

A PMT was used instead of a photodiode, since the former yields a superior signal to noise ratio for low light levels. A photodiode has an about twice as high responsive quantum efficiency as a PMT but in the latter, the signal is amplified about a million times without much deterioration of the S/N in the incoming light signal. In a photodiode the noise mechanisms significantly degrades the S/N already before amplification. A red (and green)-sensitive PM tube type Hamamatsu R 928 was finally chosen.

The used fibre-optic cable consists of two fibre-bundles merging into one. It is enclosed in an opaque metal envelope, and has the neccessary high transmission at 400 nm and for our purpose does not give any significant fluorescence.

The PMT signal is fed to the micro-computerized signal conditioning unit.

4. ELECTRONIC SIGNAL CONDITIONING

The PMT delivers current pulses from the alternating red or green signals. The desired function is the ratio between the amplitudes the "red" and "green" pulse.

This function can be obtained by using sample-and-hold circuits and an analogue divider but we considered this to be a too restricted solution. For greater flexibility we chose to solve the problem with a digital system. For this purpose an 8-bit EPROM one-chip computer (MC 68705R3) equipped with an A/D-converter was procured. By using a computer many changes and improvements in the signal conditioning are turned into a software problem.

The pulse amplitudes vary between 1 and 50 μ A, due to the variations in distance between the optic probe and the examined area. To obtain high resolution in case of small signals and to be able to measure large signals as well, a preamplification-unit controlled by the micro-computer is necessary.

Since a voltage level on an output pin is hard to detect with the human senses, we decided to transform this output signal. According to the doctors concerned the best way to present the result is in form of an audible tone, the ears being of no use anyway during the examination.

Preamplification

The signal from the PMT is fed into a current-to-voltage converter, consisting of a resistor and a low-noise operational amplifier, followed by a non-inverting follower. Fig. 13.

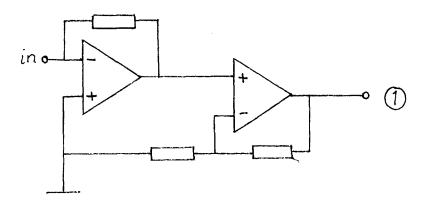


Fig. 13

The next step is an amplifier designed to amplify the red pulse only, this to make the quotient span between 0 and 1 and thereby obtain a sufficiently high resolution. The FET is functioning as a switch controlled by the microcomputer. The inverting amplifier is there only to provide the FET-gate with the necessary negative voltage. When the green pulse is measured the switch is open for unity gain. When the red pulse is put in, the switch is closed and a variable resistor determines the gain. With this resistor the quotient span is adjusted. This means that the maximum amplitude of the red pulse will be amplified to approximately the same level as the green pulse. Fig. 14.

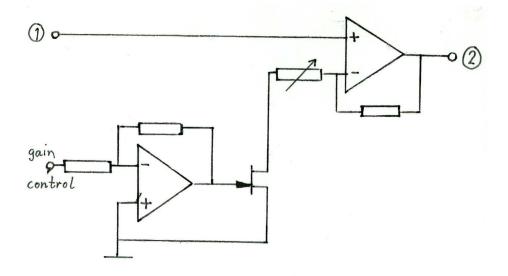


Fig. 14

Next step yields overall amplification, to optimize the use of the A/D-converter. The FET is here performing as a voltage-controlled resistor. When the gain is too high the gain-control output from the microcomputer goes high, and the capacitor is charged. This increases the FET-resistance, which, in turn decreases the gain until it becomes too low and the control output goes low and accordingly the gain increases. As a result the gain will slowly oscillate around an equilibrium. The zener-diode has a purely protective function.

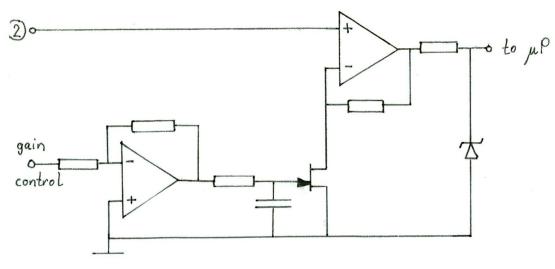


Fig. 15

Software

The main programme consists of two infinite loops which are interrupted in two different ways:

- 1. By a hardware interrupt input connected to the reference-signal, which is synchronous with the green signal and is derived from a reflective sensor inside the chopperwheel-housing.
- 2. By a software interrupt (described on page 25) when the red pulse arrives at the input.

When the reference signal arrives the following functions are performed.

- The software interrupt is adjusted to the actual chopper wheel frequency (see page 25).
- 2. The pulse is sampled 128 times during the duty cycle, the intervals between samples being obtained by software time loops. The mean value is calculated by add- and shift operations from these readings. This leads to low-pass filtering, unless the noise signal has the same frequency as the sample frequency or its overtones. In this way high-frequency noise is eliminated. Fig. 16.

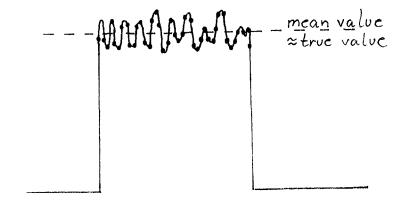
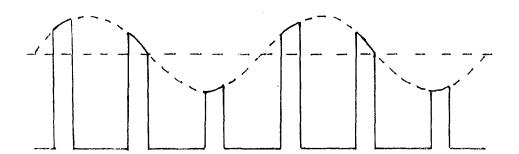


Fig. 16

- 3. The red pulse gain control pin is set since the next pulse will be a red one.
- 4. A mean value of previous green pulses is multiplied with seven and added to the new pulse. This sum is divided by eight by three shifts and stored as the new mean value. By doing this low-frequency noise components are surpressed. Fig. 17.

Fig. 17



- 5. The last pulse and the previous pulse is compared with the wanted level and the overall gain control pin is set or reset accordingly.
- 6. The last red pulse is divided with the new green one using a standard division routine.
- 7. Waits until the red pulse is expected to come.
- 8. Samples the red pulse and calculates the mean value.
- 9. The red pulse gain control pin is reset.
- 10. The red pulse is highpass filtered in the same way as the green one.
- 11. The overall amplification is checked and adjusted.
- 12. The new red pulse is divided with the last green one
- 13. Returns to the main loop and waits for the next reference signal.

See Appendix 1.

Frequency measurement and interrupt generation

Since the reference signal is used to "label" the green signal the programme is designed to calculate when to expect the red signal, in

case there are problems with the frequency stability. When the reference signal arrives the timer/counter is loaded with hexnumber FF and is counted down with a prescaling factor of F. A loop is entered in which the counter state is continously read and compared with a stored value. When the values are equal an interrupt is generated and the red signal is dealt with as described earlier. The timer continues to count down and does not stop until the next arrival of the reference signal. The readout is compared with an expected rest stored in the memory. If these values are not equal the frequency is not the expected one and therefore the interrupt value is changed to compensate for this error.

Error functions

An output pin on the computer is connected to a disabling input on the sound generating VCO. This output will become active in two cases:

1) If one of the pulses is greater than 5 volts. This can happen if the overall amplification is high, due to small signals and the probe is moved too fast to an area where the fluorescence is much stronger. This disabling function will also be activated if the signals are so small that the large gain makes the ratio inreliable.

Digital output-to sound-transformation

The digital output from the microcomputer is transformed by a D/A-converter. This analogue signal is allowed to control a voltage controlled oscillator, producing a square-wave signal.

This signal is amplified by a push-pull power amplifier of utmost simplicity, since we have no need for high fidelity. The oscillations

are transformed to sound by a small loudspeaker and a rather nasty little sound can be heard, due to the square wave. Fig. 18.

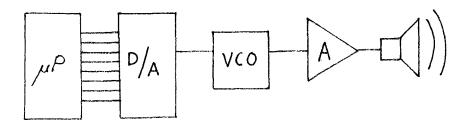
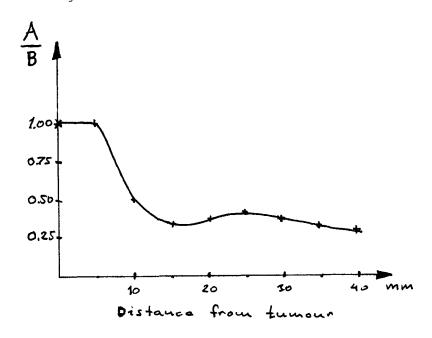
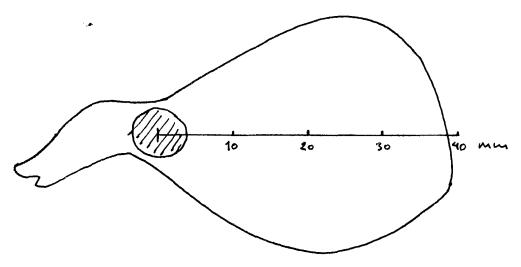


Fig. 18

5. THE FIRST TEST

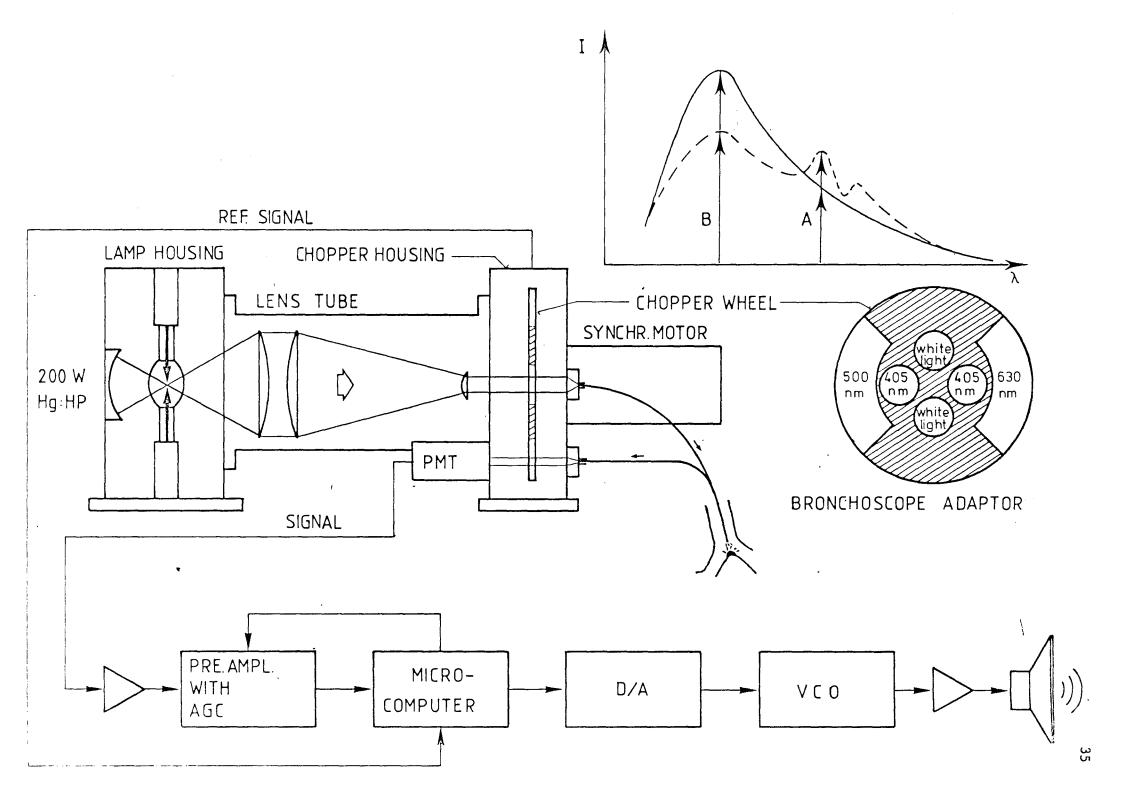
Due to initial problems with the electronic circuitry the optical system was tested by measuring the current pulses from the PMT directly with an oscilloscope. The object examined was a rat with a tumour situated in one of the legs. The rat was injected with a HPD solution (5 µg/ml) two days before the test. The skin was removed from the leg and it was scanned with the fibre-optic bundle. The measured signals were read, the quotients calculated and the result is presented in the diagram below.





Later the equipment was tested a second time on another rat with the microcomputer connected and the output from the D/A- converter connected to an X-Y plotter and a similar result was obtained.

Needless to say, the equipment has to be tested further to determine for example lower detection limits and reliability.



APPENDIX 1

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| 0708 A6 00 | 66 | LÜÄ | #0000000B | |
| 07CA 87 06 | 67 | STA | DDRC | |
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| 07D8 A6 DF | 76 | LDA | #110111118 | |
| 07DA B7 01 | 77 | STA | PORTB | |
| | 78 | | | |
| 07DC A6 36 | 79 | LDA | #5 4 | |
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| 07E6 AE 48 | 35 | LDX | #72 | |
| 07E8 A6 49 | 86 L4 | LDA | #73 | |
| 07EA 4A | 87 L3 | DECA | | |
| 07EB 26 FD | 35 | BHE | L3 | |
| OZED 5A | 89 | DECX | | |
| 07EE 26 F8 | 9.0 | BME | <u>L</u> .4 | |
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| 07F0 0102 FD | | BRCLR | 0.PORTC,LL3 | |
| 07F0 0102 FD | 93 LL3 94 | BRCLR | 0,PORTC,LL3 | |
| 07F0 0102 FD | 93 LL3 94 | | 0,PORTC,LL3 ********* | (水水水水水水 |
| 07F0 0102 FD | 93 LL3 94 95 **** | | | : ************************************ |
| 07F0 0102 FD | 93 LL3 94 95 **** 96 * | ****** | *************** | |
| 07F0 0102 FD | 93 LL3 94 95 **** 96 * 97 * | | *************** | * |
| 07F0 0102 FD | 93 LL3 94 95 **** 96 * 97 * 98 * | ************ | «************************************ | * * * |
| 07F0 0102 FD | 93 LL3 94 95 **** 96 * 97 * 98 * | ************ | *************** | * * * |
| | 93 LL3 94 95 **** 96 * 97 * 98 * 99 **** | ************************************** | (жиникижиники жини R O G R A M (миники жиники жиники) | * * * |
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| 07F3 CD 08ED 07F6 A6 FF 07F8 B7 08 | 93 LL3 94 95 **** 96 * 97 * 98 * 99 **** 100 101 STAR 102 103 Z1 | ************* H U V U D P ************************************ | ************************************** | * * * |
| 07F3 CD 08ED 07F6 A6 FF 07F8 B7 08 07FA A6 3F | 93 LL3 94 95 ***** 96 * 97 * 98 * 100 101 STAR 102 104 105 | ************************************** | ************************************** | * * * |
| 07F3 CD 08ED 07F6 A6 FF 07F8 B7 08 07FA A6 3F 07FC B7 09 | 93 LL3 94 95 ***** 96 * 97 * 98 * 100 101 STAR 102 103 Z1 104 105 | ************************************** | ************************************** | * * * |
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| 07F3 CD 08ED 07F6 A6 FF 07F8 B7 08 07FA A6 3F 07FC B7 09 07FE 9A | 93 LL3 94 95 **** 96 * 97 * 98 * 99 **** 100 101 STAR 102 103 Z1 104 105 106 107 108 | ************************************** | ************************************** | * * * |
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| 07F3 CD 08ED 07F6 A6 FF 07F8 B7 08 07FA A6 3F 07FC B7 09 07FE 9A 07FF 1D 01 0801 B6 40 0803 B7 45 0805 B7 46 | 93 LL3 94 95 ***** 96 * 97 * 98 * 99 **** 100 STAR 102 Z1 104 105 106 107 106 107 108 110 1112 113 | ************************************** | ************************************** | * * * |
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| 07F3 CD 08ED 07F6 A6 FF 07F8 B7 08 07FA A6 3F 07FC B7 09 07FE 9A 07FF 1D 01 0801 B6 40 0803 B7 45 0805 B7 46 0807 B6 48 | 93 LL3 94 95 ***** 96 * 97 * 98 * 99 **** 100 STAR 102 Z1 104 105 106 107 108 109 111 112 113 114 115 | ************************************** | ************************************** | * * * |
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| 0811 E | | 121 | 79 45 21 52 | LDA STA | PULS UT1 |
|------------------------|---------------|------------|----------------|------------|------------------------|
| 0815 6 | | 123 | | LDA | SUM |
| 0817 E | | 124 | | STA | SUM1 |
| 0819 | | 125 | | LDA | SUM+1 |
| 0818 | | 126 | | | |
| | | 127 | | STA | SUM1+1 |
| 881D 0 | | 128 129 | | JSR | GAIN,E |
| 0820 0 | | 130 131 | 23 | JSR | DIV,E |
| 0823 8 | 96 4F | 132 | Z 4 | LDA | KVOT |
| 0825 A | 37 00 | 133 | | STA | PORTA |
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| 0827 B | 36 08 | 137 | LS | LDA | TDR |
| 0829 B | | 138 | | SUB | REST |
| 0828 3 | | 139 | | BHE | L5 |
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| 082D A | | 141 | | LDA | #DISINT |
| 082F E | | 142 143 | | STA | TCR |
| 0831 (| DD 08ED | 144 145 | | JSR | PULSIN, E |
| 0834 6 | 46 FF | 146 | Z5 | L.DA | #OFFH |
| 0836 B | 37 08 | 147 | | STA | TDR |
| 0838 A | 96 3F | 148 | | LDA | #ENINT |
| 083A B | 37 09 | 149 | | STA | TCR |
| 0830 9 | | 150 | | CLI | |
| 083D 1 | 1C 01 | 151 152 | | BSET | 6,PORTB |
| | | 153 | | | |
| 083F E | | 154 | | LDA | SLASKI |
| 0841 E | | 155 | | STA | PULS |
| 0843 B | | 156 | | STA | PULS2 |
| 0845 B | 36 4C | 157 | | LDA | SUM2 |
| 0847 E | 37 43 | 158 | | STA | SUM |
| 0849 E | 96 4 <i>0</i> | 159 | | LDA | SUM2+1 |
| 0848 E | | 160 | | STA | SUM+i |
| 084D A | | 161 162 | | 85R | FILT |
| | | 163 | | | |
| 084F B | 36 45 | 164 | 26 | LDA | PULS |
| 0851 E | 37 48 | 165 | | STA | UT2 |
| 0853 E | | 166 | | LDA | SUM |
| | | 167 | | STA | SUM2 |
| 0857 8 | | 168 | | LDA | SUM+1 |
| 0859 B | | 169 | | STA | SUM2+1 |
| UU | | 170 | | 0/H | 20072.77 |
| 0858 (| DD 0913 | 171 172 | | JSR | GAIH,E |
| 085E 0 | OD 0869 | 173 174 | 27 | JSR | DIV,E |
| 0861 E | | 175 | Z8 | LDA | KVOT |
| 0863 E | | 176 | = = | STA | PORTA |
| contract that that the | - · · · | 177 | | ₩ 111 | 15.111 |
| 0865 (| 0102 FD | 178 179 | L6 | BRCLR | 0,PORTC,L6 |
| 0868 A | 46 7F | 180 | | LDA | #DISINT |

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0860 B6 50
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086E 27 02
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                             BEQ
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0870 IF 01
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                 185
                             BOLR
                 186
0872 86 08
                187 L7
                             LDA
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0874 B0 4E
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                             SUB
                                      REST
9876 28 05
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                                      LNER
0979 3C 4E
                190
                             INC
                                      REST
087A CC 07F3
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                                      START,E
                             JMP
087D 3A 4E
                 192 LHER
                             DEC
                                      REST
087F CC 07F3
                 193
                             JMP
                                      START, E
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                 196 *
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                             SUM EQU SLASKI
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0882 B6 43
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0884 B7 42
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0886 B6 44
                208
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0898 44
                209
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0889 36 42
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0888 44
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0880 36 42
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088F 36 42
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0891 B6 43
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0893 B0 42
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0895 B7 43
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0897 86 44
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0899 A2 00
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089B B7 44
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089D B6 43
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0803 86 40
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0800 B0 47
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0807 87 40
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08F8 B6 0F
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08FE B7 41
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0900 B6 40
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0902 A9 00
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| 0906 | | | 301 | | DECX | | |
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| OFFS | 093F | 362 363 | FDB | INTERR |
| | | 364 365 | EMD | |

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