1 Systematic microbial production of optically active dissolved organic

2 matter in subarctic lake water

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ABSTRACT

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The ecology and biogeochemistry of lakes in the subarctic region are particularly sensitive to 24 changes in the abundance and optical properties of dissolved organic matter (DOM). External 25 input of colored DOM to these lakes is an extensively researched topic, but little is known 26 about potential reciprocal feedbacks between the optical properties of DOM and internal 27 microbial processes in the water. We performed 28-d dark laboratory incubation trials on 28 29 water from 101 subarctic tundra lakes in northern Sweden, measuring the microbial decay of DOM and the resulting dynamics in colored (CDOM) and fluorescent (FDOM) DOM 30 31 components. While losses in dissolved oxygen during the incubations corresponded to a 20% decrease in mean DOM, conversely the mean CDOM and total FDOM increased by 22% and 32 30%, respectively. However, the patterns in microbial transformation of the DOM were not 33 the same in all lakes. Notably, along the gradient of increasing ambient CDOM (water 34 brownness), the lakes showed decreased microbial production of protein-like fluorescence, 35 lowered DOM turnover rates and decreasing bacterial growth per unit of DOM. These trends 36 indicate that browning of subarctic lakes systematically change the way that bacteria interact 37 with the ambient DOM pool. Our study underscores that there is no unidirectional causal link 38 between microbial processes and DOM optical properties, but rather reciprocal dependence 39 between the two. 40

41 INTRODUCTION

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Subarctic regions undergo climate related transitions, evident from thawing permafrost (Åkerman and Johansson 2008; Lawrence et al. 2012), changes in treeline limits (Kullman 2002) and vegetation succession distributions (Callaghan et al. 2002; Kullman 2014). Many lakes in these regions are at risk of transitioning from a clear-water into a brown-water state because of hydrologically activated soil and peat horizons that leach brown-pigmented dissolved organic matter (DOM) (Jansson et al. 2010; Wauthy et al. 2018). In the aquatic environment, this allochthonous DOM tends to reduce productivity of benthic habitats (Karlsson et al. 2009; Urtizberea et al. 2013), change nutrient supply and food web structure (Berggren et al. 2015; Creed et al. 2018; Jansson et al. 2007), and cause CO₂ outgassing to the atmosphere (Lapierre et al. 2013). The extent of these effects are however difficult to predict due to the complex interactions between DOM and lake ecosystems that tend to result in non-linear responses (Solomon et al. 2015). Nonetheless, it is clear that DOM plays a central role in the climate change impact on lake ecosystem functioning, especially in neararctic environments which are particularly sensitive to climate-induced changes in the carbon cycle (Vincent et al. 2012). The fractions of DOM that are optically active, i.e. the light-absorbing 'colored' (CDOM) and the 'fluorescent' (FDOM components) fractions are relatively persistent in the environment (Berggren et al. 2018; Kellerman et al. 2015), compared to the smaller pool of fast cycled non-colored DOM in freshwaters (Berggren et al. 2010). However, due to their commonly large concentrations, CDOM and FDOM are associated with significant quantities of reactive compounds (Lapierre and del Giorgio 2014; Wetzel 1995), including microbially degradable DOM fractions of importance for the landscape carbon cycling (Lapierre et al. 2013). Research on the dynamics of optically active DOM in the arctic and subarctic tundra

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regions is limited, but results from boreal and temperate lakes suggest that increasing loading of terrestrially derived CDOM generally causes a shift from fast-cycled low DOM concentrations characterized by protein-like fluorescence (Koehler et al. 2012), to larger and more slowly cycled humic-like DOM pools maintained by hydrological input (Guillemette and del Giorgio 2011). However, total amounts of reactive and protein-like DOM may increase with CDOM on large scales, because even if such fractions may represent relatively small parts of the terrestrially derived DOM, this is eventually outweighed by the high total DOM concentrations in brown lakes (Lapierre and del Giorgio 2014). Optically active organic constituents are not only imported from the catchment, but also produced by microbial processing of autochthonous (i.e. algal derived) DOM (Romera-Castillo et al. 2011). Aquatic microbial production of CDOM is evident from laboratory (Shimotori et al. 2009; Tranvik 1993) and marine (Rowe et al. 2018) studies, but corresponding observations in natural freshwaters are scarce. In brown-water lakes, CDOM tends to be consumed by bacteria faster that it is produced (Berggren et al. 2018). Regarding FDOM, a study of lakes and streams in southern Québec (Guillemette and del Giorgio 2012) found that most fluorescence components were net produced in clear-water systems where significant fractions of the DOM were of autochthonous origin, whereas in systems dominated by allochthonous DOM the FDOM tended to be net consumed. In clear-water alpine lakes, production of protein-like fluorescence has been observed after algal blooms (Miller and McKnight 2010), attributed to nitrogen accumulation in the DOM pool (Goldberg et al. 2015). This agrees with experimental results confirming FDOM production, especially protein-like fluorescence, in response to bacterioplankton use and transformation of algal DOM (Fox et al. 2017). However, patterns in microbial net production or consumption of CDOM and FDOM in subarctic lakes remain unexplored.

A dramatic increase in allochthonous DOM (i.e. lake browning) has recently been observed in many arctic lakes (Wauthy et al. 2018), with anticipated impacts on lake carbon balances (Lapierre et al. 2013) as well as food webs (Creed et al. 2018; Williamson et al. 2015). However, little is known about the interaction between lake browning and the balance between internal microbial production and degradation of optically active constituents. Such interactions, which are non-linear by nature (Berggren et al. 2018), can potentially exacerbate or dampen the terrestrial influence on the optical environment in lakes with even small changes, thus influence a wealth of ecological and biogeochemical processes.

Here we hypothesized that due to the natural clear-water state of subarctic tundra lakes, with relatively small inputs of CDOM from terrestrial sources, FDOM is net produced by bacterioplankton, similar to what has been observed in temperate and alpine clear-waters based on algal-derived DOM (Guillemette and del Giorgio 2012; Miller and McKnight 2010). However, we expected that with increasing ambient CDOM the balance between production and consumption of FDOM could switch to net consumption. Moreover, we expected a shift from highly reactive protein-like DOM to more recalcitrant and humic-like DOM along the gradient of increasing CDOM. We tested these ideas by combining a large survey sampling of 101 subarctic lakes in northern Sweden with 28-d laboratory incubation experiments, during which we measured bacterial production and respiration rates and followed changes in the microbial production or consumption of optically active compounds.

METHODS

Study area, sampling and site selection

Sampling was carried out west of Abisko, Northern Sweden, in a subarctic mountainous plateau (Fig. S1). The area is characterized by the transition between tundra (Virtanen et al. 2016) and Nordic mountain birch forest (Wielgolaski 2005). A map and information on the relative contribution of the land use types in the area is available in Fig. S2. The mean annual temperature and precipitation for the 1970-2000 period (Hijmans et al. 2005) for the study lakes ranged between -0.1 and -1.4 °C, and from 477 to 873 mm respectively along a gradient from east to west. The permafrost conditions of the region is characterised by sporadic and discontinuous permafrost (Brown et al. 1997), with isolated pockets of permafrost in the study catchments (Gisnås et al. 2017).

In autumn (September 20th, 2017) we carried out a helicopter survey of 149 lakes sampled within 5 h. We used the Eurocopter EC-120B helicopter equipped with special floats, and modified to allow sampling through a trap in the floor within seconds upon landing on the lake surface. This aircraft is the same as that used to carry out the Swedish national lake monitoring program, which samples ca 1000 lakes per year for analysis of water chemical and optical characteristics. Lake water was collected from 1 m depth using a 1.5 L water sampler (UWITEC) pre-rinsed with 10% HCl and Milli-Q water. The turbulence caused by the helicopter upon landing may have induced water mixing to a degree. However, the impact of such mixing on the samples was assumed to be negligible because the lakes were already well mixed, without thermal stratification, on the sampling date. Moreover, the sampling point beneath the helicopter's body is the place with the least turbulence within the helicopter's footprint. Recorded video footage, showing the relatively limited rotor downwash effects around the helicopter, will be shared with readers upon request.

Samples were transported cold to the laboratory in acid-washed 700 ml polycarbonate bottles, and then further subsampled. Subsamples of water for decay experiments and analyses of optical characterizations were filtered using pre-combusted (4h at 500°C) glass fibre filters (Whatman GF/F), and stored cold in 250 ml acid-washed high-density polyethylene (Nalgene) bottles in a cold room at a constant temperature of 2 °C for a total of 40 days (since sampling) before initiation of the experiments. Preservation of the DOC during storage was optimized by the combination of using burnt GF/F filters, which have less than half pore-size diameter compared with non-combusted GF/Fs thus removing more of the bacteria (Nayar and Chou 2003), and using a storage temperature close to the freezing point. Unfiltered water subsamples for total phosphorus (TP) and total nitrogen (TN) were frozen at -20°C until analysis, while samples for dissolved organic carbon (DOC) were filtered (Whatman GF/F) and acidified with HCl to pH 2. The preserved DOC and nutrient samples were analyzed during spring 2018.

In total 101 of the sampled sites were selected for this study (Table 1). We selected only sites with *in situ* DOC concentrations above 1.0 mg L⁻¹, due to the general difficulty of measuring the turnover of small carbon pools with acceptable accuracy using standard techniques available to us (Jennings et al. 2018). Additionally, sites were excluded if data records were incomplete or if the specific ultraviolet absorption of DOC at 254 nm (SUVA₂₅₄; decadic absorbance coefficient divided by DOC concentration) was anomalously high indicating that constituents other than DOC, although not measured (e.g. iron), contributed to the optical characteristics of the water (Poulin et al. 2014; Weishaar et al. 2003). We chose the threshold value for accepted *in situ* SUVA₂₅₄ of 6 L mg⁻¹ m⁻¹. However, since it is in the nature of SUVA₂₅₄ to have relatively large error variability at low DOC concentrations, the higher

threshold of 8 L mg $^{-1}$ m $^{-1}$ was used for SUVA $_{254}$ values calculated at the end of the decay experiment when DOC had decreased and thus SUVA $_{254}$ uncertainty had increased.

Dissolved organic carbon decay

Decay experiments were carried out in standard dark 20°C conditions, during which the DOC decay with microbial re-growth communities (ambient bacteria that passed through the GF/F filtration) was assessed in parallel with bacterial production (BP) measurements. The DOC loss was inferred from dissolved oxygen (DO) loss using a conversion factor (i.e., the respiratory quotient 'RQ') of 1.0 mole of DOC lost per mole of O₂, which is lower than the average RQ reported for net heterotrophic boreal and temperate lakes but a reasonable assumption for lakes with low terrestrial influence (Berggren et al. 2012).

For each lake water sample, simultaneous and continuous analysis of DO was carried out in duplicate or triplicate 5 ml sensor vials with butyl rubber septa screw caps using a SensorDish Reader (SDR; PreSens, Germany) optical oxygen sensing system (Soares et al. 2018). Before incubation start, the sensor vials were top-filled with sample water in a 20°C temperature controlled room and left for two hours with the cap open to equilibrate with the oxygen concentration in the air. The vials were then sealed without headspace or bubbles, inserted into the SDR plates and a first reading was carried out immediately to obtain a 100% DO calibration point. Thus all experiments started with nearly 100% DO saturation at 20°C (8.76 mg L⁻¹), before oxygen was gradually consumed during 28-d incubation in a closed PU-3J high precision temperature chamber (ESPEC, Japan). We used factory calibration data for the 0% DO calibration point, which is stable over time. The dissolved O₂ concentrations decreased in all incubations, but since this loss rate declined gradually over the course of the incubation, the oxygen (> 5.7 mg O₂ L⁻¹) was not exhausted in any of the cases.

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In parallel, the remaining water in the 250 ml Nalgene bottles was incubated in a 20°C constant room for the measurement of BP, absorbance and fluorescence during the incubation. The BP was measured in a total of five 5 tubes per lake (day 1, 3, 7, 14 and 28) using the 3H-leucine incorporation method (Smith and Azam 1992). Aliquots of 1.2 ml were exposed to 40 nM leucine during 1 h, before the incubation was stopped with trichloroacetic acid and the protein content was purified through repeated (three times) centrifugation and removal of supernatant (Karlsson et al. 2001). Uptake of leucine was converted into bacterial carbon (Simon and Azam 1989) applying the standard conversion factor 1.55 kg C mol leu-1 multiplied with an isotopic dilution factor of 2. Total cumulative BP during the incubation was calculated using the trapezoid rule.

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- 199 Reactivity continuum modeling
- The DOC decay was assessed using the reactivity continuum model (Boudreau and Ruddick
- 201 1991; Koehler et al. 2012), where the DOC concentration at time 't' relative to the initial
- DOC concentration at time '0' (theoretical range from 0 to 1) was described as a function of
- 203 the rate parameter 'a' and the shape parameter 'v' (Eq 1).

$$204 \quad \frac{DOC_t}{DOC_0} = \left(\frac{a}{a+t}\right)^{U} \tag{1}$$

- The model was fitted with days as time unit using the nonlinear least squares (nls) function in
- 206 R build 3.4.1 (R Development Core Team 2013). The accuracy of the model was given by the
- 207 root square mean error (RMSE) for deviations between predicted and measured relative DOC
- 208 concentrations. The apparent decay coefficients k for any time point was calculated (Eq 2).

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$$k = v^*(a+t)^{-1}$$
 (2)

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Optical organic matter characterization

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the end of the decay experiment using an Aqualog (Horiba Scientific). Scans were collected 213 in a 1 cm quartz cuvette (2 s integration time) over 5 nm increments with excitation 214 wavelengths ranging from 230 to 800 nm and emission wavelengths from 250 to 800 nm. The 215 EEMs were blank-subtracted (Milli-Q water) daily, corrected for instrument-specific biases 216 (Cory et al. 2010) and inner filter effects (Kothawala et al. 2013), and normalized to the 217 218 Raman area of deionized (Milli-Q) water (Lawaetz and Stedmon 2009). Fluorescent components were identified with parallel factor analysis (PARAFAC) of the corrected 219 220 EEMs using the toolbox and procedure described in Murphy et al. (2013). The PARAFAC model was validated using a split-half validation routine, core consistency diagnostic, and a 221 visual inspection of the residuals to ensure no systematic signal remained (Fig. S3). Six 222 fluorescent components were identified by the PARAFAC analysis: C1 and C3 (visible 223 humic-like components; ex/em: 325/456 and 265-390/512 nm, respectively), C2 and C5 (UV 224 humic-like components; ex/em: <250/446 nm and 255/438 nm, respectively), C4 (microbial 225 humic-like component; ex/em: <250-305/406 nm), and C6 (tryptophan-like component; 226 ex/em: 280/326 nm). All components have been previously listed in the online spectral 227 library OpenFluor [www.openfluor.org; (Murphy et al. 2014)], and our model's emission and 228 excitation loadings are available on the OpenFluor web site under the name "Swedish 229 subarctic lakes" from the time of acceptance of this publication. These components are also 230 shown visually in Fig. S3. 231 232 Colored dissolved organic matter (CDOM) was calculated as the absorbance at 440 nm, 233 corrected for turbidity by subtracting the absorbance at 690 nm, and converted to Naperian 234 units through division by cuvette length in meters and by multiplying with a correction factor 235

of 2.303 (i.e. ln(10) which converts a decadic coefficient into a Naperian).

Absorbance and excitation-emission matrices (EEMs) were collected at the beginning and at

We chose not to include SUVA₂₅₄ and spectral slope in the analysis and presentation, because we did not have access to iron data and thus could not correct these parameters for potential iron interference which is particularly high in the ultraviolet region (Weishaar et al. 2003). As an alternative to SUVA₂₅₄, we used the ratio between CDOM at 440 mm (where iron interference is relatively lower) and DOC as a corresponding indicator. For reference, the ambient SUVA₂₅₄ in the selected study sites was 3.57 ± 0.77 (mean \pm SD). Our analysis thus strongly focus on PARAFAC components, which have been shown to be little effected by iron, while maximizing the extracted information about DOM composition (Poulin et al. 2014). Moreover, we excluded several commonly reported standard spectral indices (Gabor et al. 2014), including the fluorescence index FI, humification index HIX, biological index BIX and the freshness index FRESH, because the change in these indices from the beginning to the end of our experiment was negligible (1-5% on average), and thus they were not informative about the major changes in DOM quality that occurred in response to the microbial processing of the DOM.

Element analyses

Dissolved organic carbon was measured via high temperature catalytic oxidation on a Shimadzu TOC-LCPH analyzer. The DOC concentrations are reported as the mean of three replicate injections for which the coefficient of variation was < 2%. The TP and TN were determined using a QuAAtro39 Continuous Segmented Flow Analyzer (SEAL Analytical), equipped with an auto sampler and ADM-unit (Autoclave Distillation Module). Samples were processed through two-stage on-line digestion with potassium persulfate at 110°C, first under alkaline conditions and then under acidic conditions, both under a pressure of 0.09 MPa. The TN was analyzed as nitrite under chemical reactions to form an azo dye measured

- at 550 nm, while TP was analyzed through a colored reaction by molybdenum blue and
- measured at 880 nm.

RESULTS

The mean O_2 -inferred DOC decreased by 20% during the 28 days of incubation. In spite of the storage time prior to the experiment, there was surprisingly good preservation of the most labile DOC fraction, evident from the distinct high decay rates found during the first days of the bio-decay experiment (Fig. 1). Based on the observed dissolved oxygen changes during incubation (Fig. 1a) and the known initial DOC concentrations, the fraction DOC remaining over time could be calculated and used to fit the reactivity continuum model, yielding low average RMSE values (0.45%) in the range of 0.06-2.8%. The modeled estimates of the apparent decay coefficient k spanned from 0.006 to 0.092 at t = 0 days and from 0.001 to 0.017 at t = 28 days (Fig. 1b). The bacterial production showed a similar pattern of highly variable values between sites, and a slightly decreasing trend over time (Fig. 1c). The total bacterial production during the incubation period amounted to 0.2-11.5% of the initial DOC, with a mean of 3.6%.

While the average DOC decreased (paired t < -76.3), from 3.5 mg L⁻¹ initially to 2.8 mg L⁻¹ at the end of the experiment, CDOM and FDOM increased from 1.7 m⁻¹ to 2.0 m⁻¹ (paired t > 10.7) and from 4.5 R.U. to 5.9 R.U (paired t > 28.4), respectively (2-tail p < 0.001, n = 101; see standard deviation of means in Fig. 2a). As a consequence, the ratio between CDOM and DOC increased 1.7 times and the ratio between FDOM and DOC increased 1.8 times (Fig. 2a). Among the six components of FDOM identified using PARAFAC (see description in methods), all were produced (1-sample t > 9.1) except for one of the humic-like components (C2) which was net consumed (1-sample t < -21.4, 2-tail p < 0.001, n = 101; Fig. 2b). The components showing the largest production rates were the protein-like component C6 (mean 0.007 R.U. d⁻¹) and, especially, the humic-like component C5 (mean 0.048 R.U. d⁻¹).

Bacterial production rates at the end of the incubation showed weak positive correlations with TN and TP (Table S1), but there were no clear indications of nutrient limitation in the DO consumption experiments used to model the DOC decay. On the contrary, initial and final decay coefficients as well as the total DOC loss were negatively correlated to nutrient concentrations. Among the site characteristics, lake area, elevation and longitude showed relatively few and weak correlations with decay variables. However, decay (k) coefficients, total DOC loss and carbon use for bacterial production correlated positively with latitude and negatively with CDOM and FDOM (Table S1).

In general, relative (%) abundances of the fluorescence components classically interpreted as terrestrially-derived humic (C1-C3) were negatively related to different aspects of DOC reactivity and use by bacteria, while the microbially derived humic C4, the humic-like C5, and the protein-like (C6) components were positively related to these decay variables (Table S2). Component C6 was the strongest positive indicator of both total DOC loss (Pearson r = 0.56, $r^2 = 0.32$, n = 101, p < 0.001; Fig. 3a) and total share of DOC used for bacterial production (Pearson r = 0.55, $r^2 = 0.31$, n = 101, p < 0.001; Fig. 3b). The corresponding relationships for components C1, C2 and C3 had similar strengths, but these correlations were negative (Table S2).

Although most lakes of the study area can be characterized as clear-water oligotrophic, there was a 27-fold variability in ambient CDOM, with a few values spanning up toward 8 m⁻¹. The patterns of change in concentration and optical properties of DOC that we observed during the incubation experiment were not randomly distributed along this gradient of increasing CDOM. For example, the rate of change in the humic-like C2 (R.U. d⁻¹) followed a logarithmic function of CDOM m⁻¹ (y = -0.0066*ln(x) + 0.014, $R^2 = 0.30$, n = 101, p < 0.001;

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Fig. 4a) implying that more of C2 was consumed with increasing brownness of the water, but in contrast the combined change in C1, C3, and C5 followed a logarithmic curve that increased along the CDOM gradient (y = 0.0094*ln(x) + 0.051, $R^2 = 0.16$, n = 101, p < 0.001; Fig. 4a). Production rates of the microbial-like component C4 were unrelated to the CDOM gradient. Noteworthy, production of protein-like fluorescence (C6) was characteristic of lakes with low ambient CDOM, and C6 production rates decreased with the CDOM gradient toward zero in the brownest lakes $(y = -0.0029*ln(x) + 0.0082, R^2 = 0.20, n = 101, p < 0.001;$ Fig. 4a). Moreover the reactivity of the DOC (initial k) decreased as a power function of increasing in situ CDOM (0.024* $x^{-0.372}$, $R^2 = 0.25$, n = 101, p < 0.001; Fig. 4c). The pattern of accumulating protein-like fluorescence together with strongly decreasing k during the degradation experiments created a paradoxical negative relationship between k and protein-like fluorescence in absolute units in all of the data combined (Fig. 5). This reverse relationship was enhanced by the fact that C6 occurred in high absolute concentrations (but low relative concentrations) in some of the browner lakes that were also characterized by low k values. Thus, although the relative (%) abundance of C6 was a strong positive indicator of different aspects of DOC reactivity (Fig. 3), the relationship between k and C6 in absolute units surprisingly was negative (Fig. 5).

DISCUSSION

Our survey of subarctic lakes confirms the existing knowledge on how increased CDOM in freshwaters is associated to changes in the composition and biological reactivity of DOM (Fasching et al. 2014; Hopkinson et al. 1998; Koehler et al. 2012). However, for the first time we also show that in these subarctic lakes there is a systematic microbial production of CDOM and FDOM that, in turn, shows patterns across the gradient of ambient water brownness. Thus, browning of subarctic tundra lakes may not only affect the concentration, composition and reactivity of DOM, but it also impacts on interactions between bacteria and DOM that further modifies the pool of optically active DOM. Hence, our laboratory results suggest a previously unrecognized feedback mechanism of relevance for assessing the changes in subarctic lakes due to climate-induced increases in CDOM loading from the catchment. However, further research is needed to validate the representativeness of these laboratory results to field conditions.

There was an expected pattern of decreasing initial k values with increasing CDOM (Fig. 4c), in parallel with decreasing percentage protein-like fluorescence. These results agree with the pattern that Koehler et al. (2012) found in a comparison between clear-water and brownwater boreal lakes, caused by a decreasing share of labile algal DOM with increasing CDOM. In our study we did not investigate the origin of the DOM in the lakes, but previous studies have shown that low CDOM values combined with large fractions of protein-like fluorescence are indicative of major algal contributions to the DOM pool (Guillemette and del Giorgio 2011; Stedmon and Markager 2005) which could explain the high k values and large proportion of DOC used for bacterial production in lakes with low CDOM (Table S1) and high percentage protein-like fluorescence (Fig. 3). Interestingly, in terms of the absolute (mg L⁻¹) DOC decay, the decreasing k values along the CDOM gradient were compensated

by increasing DOC concentrations, such that there was no correlation between the total absolute DOC loss (final minus initial values in mg L^{-1}) and increasing CDOM (p = 0.86, not shown). Thus, contrary to what Lapierre et al. (2013) found for boreal lakes, increased CDOM in our study does not appear to imply that the total concentration of labile carbon increases in the water column of lakes, at least not during the autumn (and low discharge) conditions when our measurements were performed. However, during other seasons such as the snowmelt season (spring-midsummer) when fresh colored terrestrial DOC is entering the lakes, previous studies in our study area indicate the respiration is boosted also in absolute carbon units, leading to substantial CO_2 emissions (Jonsson et al. 2007; Jonsson et al. 2003).

Our initial apparent k values were considerably higher than those found in other lake studies using 20°C dark incubations (Koehler et al. 2012; Mostovaya et al. 2016). A possible reason for this difference is that the previous studies measured DOC concentrations with low temporal resolution (several days per measurement) while we used a high temporal resolution optical sensor approach, allowing us to detect some of the most labile DOC fractions used in the very beginning of the decay (Guillemette and del Giorgio 2011; Pollard 2013). After a few days of incubation, our k values had decreased to a range that represent the reactivity of a wide range of typical DOC molecules found in lakes (Mostovaya et al. 2017), which together with low RMSE gives credibility to the modeled DOC decay.

Due to logistic challenges with measuring the DOC decay directly in a large number of samples, we were limited to indirect DOC assessment based on an oxygen sensing system. A weakness with this approach is the uncertainty related to the RQ value used for unit conversion to carbon. Studies in unproductive lakes in different boreal regions have reported average bacterioplankton RQs of 1.2 (Berggren et al. 2012) or ~2 (Cimbleris and Kalff 1998),

and RQs as high as ~3 have been experimentally achieved by feeding bacteria with UV light treated partially oxidized DOC (Allesson et al. 2016). Considering that partial UV oxidation followed by microbial decay could be a major pathway of organic carbon decay in tundra region freshwaters (Cory et al. 2014), it is possible that the RQ and thus also the DOC decay rates in this study were underestimated. However, this should be particularly true for the clearest and thus most UV light exposed lakes, which would further amplify the pattern of high k values in lakes with low CDOM (Fig. 4c). Therefore, if the RQ value biased our results it would not necessarily mean that any conclusion from the study is wrong, but rather that the strength in some of the reported patterns might be underestimated.

We found support for the hypothesis that bulk FDOM and its sub-components (except C2) are net produced in clear-water subarctic tundra lakes, as expected due to low terrestrial influence relative to that of algal derived DOM, which can lead to FDOM production within the lakes (Fox et al. 2017). This agrees with the temperate lake study by Guillemette and del Giorgio (2012), where both protein- and humic-like FDOM were microbially net produced in clear lakes (CDOM < 2) that had significant algal DOM fractions. However, while their study suggested a generic shift to net FDOM production at a certain level of terrestrial influence (CDOM > 2, terrigenous DOM > ~85%) (Guillemette and del Giorgio 2012), we did not reach a corresponding threshold that shifted the production-consumption balances of all FDOM components, but instead there were variable responses among the FDOM components to the increasing CDOM (Fig. 4). In our case, the protein-like fluorescence was produced the most in the clearest lakes, but even if the production decreased with increasing CDOM the values remained positive throughout the color gradient Fig. 4b. The humic-like components, on the other hand, were in our study processed to a larger degree with higher CDOM, either through increased production (C1, C3 and C5) or increased consumption (C2). Thus the

microbial-induced dynamics of FDOM appear to be remarkably different in subarctic lakes compared to boreal and temperate lakes (Guillemette and del Giorgio 2012).

Our study demonstrates counter-intuitive aspects of protein-like fluorescence dynamics in lakes. First, while the relative C6 *in situ* abundance (%) in the study lakes was negatively correlated to CDOM, there was an increase in the absolute (R.U.) C6 concentration along the CDOM gradient because of the associated increases in total FDOM (Table S2). This reflects that protein-like FDOM may arrive in high absolute (but low relative) concentrations with terrestrial drainage water (Lapierre and del Giorgio 2014). Second, our incubations showed that C6 was produced in parallel with microbial consumption of labile DOC, but the protein-like FDOM itself was not very reactive as it tended to accumulate during the incubations. These two aspects together shaped an unexpected reversed (negative) relationship between reactivity (k) and the absolute concentration of protein-like fluorescence (Fig. 5), adding to the emerging view that predictions of reactivity based on FDOM must be made with great caution because the relationships are indirect and their causality is not straightforward (Fox et al. 2017). Our findings also help explain why protein-like DOM appears as persistent (refractory) in nature along spatial gradients of increasing water residence times (Kellerman et al. 2015) or over time in slow-turnover lakes (Goldberg et al. 2015).

Another surprising result was that not only FDOM, but also CDOM, increased systematically in our incubation experiments (Fig. 1). Such observations of increasing CDOM in dark bacterial incubations have only rarely been observed before (Berggren et al. 2018). However, it has been long known that bacterioplankton can excrete or by other means produce CDOM (Shimotori et al. 2009; Tranvik 1993), at the same time as they tend to prefer consuming non-colored DOM fractions (Asmala et al. 2014; Berggren et al. 2018; Hansen et al. 2016).

Theoretically, this makes it possible for bacteria to make a net contribution to CDOM, especially in lakes where non-colored algal DOC may be a main carbon source. However, whether or not this CDOM production alongside with FDOM production that we observed is representative to *in situ* conditions can merely be speculated on, given the artificial dark environment in our experiment and the preceding 40-d refrigerated sample storage time that likely altered the composition of the microbial community (Calvo-Diaz et al. 2011).

Nonetheless, if assuming representativeness, then the measured bulk CDOM and FDOM production would be sufficient for renewing the ambient respective pools in 104 d and 82 d, respectively (median for all lakes) under steady-state conditions. This highlights the need to perform further research on the origin of optically active compounds in lakes, as the commonly assumed terrestrial origin of colored DOM constituents in lakes may not always be true.

The average CDOM production (0.37 m⁻¹ in total during 28 d) was evenly distributed across the CDOM gradient (0.29-7.8 m⁻¹), but its relative importance was largest in the clearest lakes. In fact, the measured microbial CDOM and FDOM production rates would be theoretically sufficient (again, assuming steady-state conditions and data representativeness) to renew the bulk CDOM and FDOM pools in a many of the clearest lakes during e.g. a 2-month summer period with stagnant water. In lakes with long water residence times (a few years) it could be hypothesized that the internal production of CDOM and FDOM is always larger than the input of these components from the catchment. Thus, in such lakes it can be expected that CDOM is mainly controlled by the balance between microbial CDOM production and photochemical CDOM removal by sunlight (Cory et al. 2014).

In summary, this study shows that clear-water subarctic lakes in northern Sweden had high DOC reactivity (high k values), high bacterial production per unit DOC, and FDOM production characterized by increases in protein-like fluorescence. Relatively browner lakes had lower k values and relatively less production of biomass and protein-like fluorescence. Our study is the first to suggest that DOM in subarctic clear waters may derive its characteristically high protein-like fluorescence from internal microbial production, but this effect appears to be limited to lakes with low CDOM. Thus, water 'browning' can be expected to impact the internal processes that control the balance between renewal and turnover of optically active organic matter in the water, potentially with fundamental consequences for the functioning of these ecosystems.

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TABLES

Table 1. Ambient nutrient concentrations, geographical settings and optical characteristics of 101 subarctic Swedish lakes. Variables represent dissolved organic carbon (DOC), total nitrogen (TN), total phosphorus (TP), lake area/location, colored dissolved organic matter at 440 nm wavelength (CDOM), total fluorescent dissolved organic matter (FDOM) and relative contributions from the six different components of FDOM (C1-C6). The three respective data columns show overall range, median (interquartile range in brackets) and mean (standard deviation in brackets).

Variable	Range	Median [IQR]	Mean [SD]
DOC (mg L ⁻¹)	1.6-9.6	3.1 [2.5-4.2]	3.5 [1.6]
TN (mg L ⁻¹)	0.06-0.39	0.16 [0.11-0.20]	0.16 [0.06]
TP (μg L ⁻¹)	1.1-22.6	3.8 [2.8-5.9]	4.8 [3.4]
Area (ha)	0.1-112.8	1.0 [0.7-2.1]	3.2 [11.4]
Elevation (m a.s.l.)	326-587	444 [409-491]	447 [59]
Latitude (DD)	68.44-68.50	68.46 [68.45-68.48]	68.47 [0.02]
Longitude (DD)	18.19-18.65	18.43 [18.31-18.54]	18.43 [0.13]
CDOM (m ⁻¹)	0.3-7.8	1.3 [0.8-2.1]	1.7 [1.2]
FDOM (R.U.)	1.2-13.4	4.1 [3.1-5.8]	4.5 [2.1]
C1 (%)	13.9-24.9	19.7 [18.1-21]	19.6 [2.2]
C2 (%)	26.9-43.2	37.3 [35.1-38.4]	36.6 [2.7]
C3 (%)	7.9-14.0	11.1 [10.4-12.1]	11.2 [1.3]
C4 (%)	15.7-21.6	18.1 [17.3-18.9]	18.1 [1.2]
C5 (%)	0.0-5.4	2.3 [1.5-3.3]	2.3 [1.3]
C6 (%)	4.7-32.4	11.4 [8.9-14.7]	12.2 [4.6]

648 FIGURES

visually separate the lines.

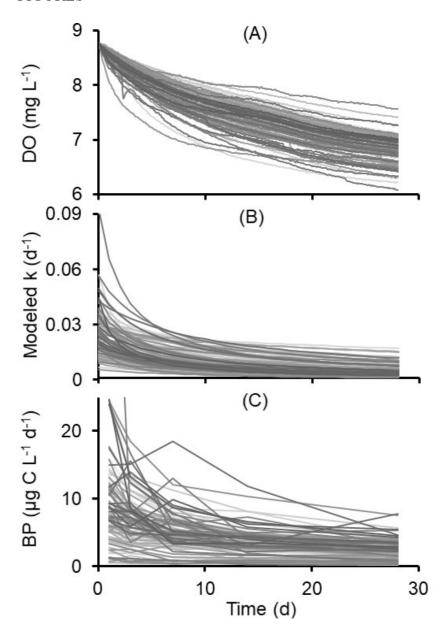


Figure 1. Visual presentation of the variability in in measured and models variables during 28-day dark incubations of water from 101 lakes in a Swedish tundra landscape. Charts show (A) observed dissolved oxygen, (B) decay coefficient k for the calculated (based on dissolved oxygen changes) degradation of dissolved organic carbon, and (C) measured bacterial production rates. The y axis of panel C is limited to 25 μ g L⁻¹, which excludes one high BP value of 72 μ g C L⁻¹ at t = 1. The grey scale coloring of the curves (A-C) is random, to

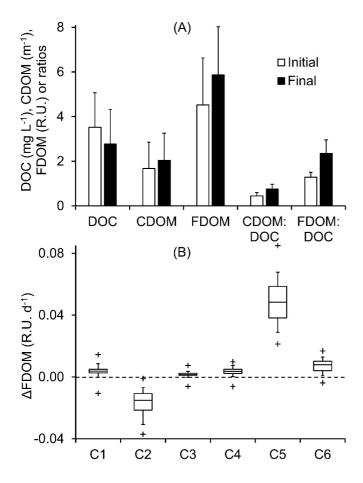


Figure 2. (A) Initial and final incubation values for dissolved organic carbon (DOC; measured initial values and final values inferred from dissolved oxygen change), colored dissolved organic matter (CDOM), fluorescent dissolved organic matter (FDOM), and ratios between these variables during 28-day dark laboratory incubations of water from 101 subarctic tundra lakes. Bars show mean values and error bars standard deviations. Note that while the overall distributions overlap for each variable, the paired differences between initial and final values for each respective variable are highly significant (t > 10.7 or t < -76.3 or, n = 101, 2-tail p < 0.001, paired t-test). (B) Box and whisker plots for production rates of six PARAFAC-derived subcomponents of FDOM during incubation. Boxes show quartiles two and three, lower and upper whiskers the 5th and 95th percentiles, respectively, and plus symbols minimum/maximum values. The mean Δ vales of all components are significantly different from zero (t > 9.1 or t < -21.4, n = 101, 2-tail p < 0.001, 1-sample t-test).

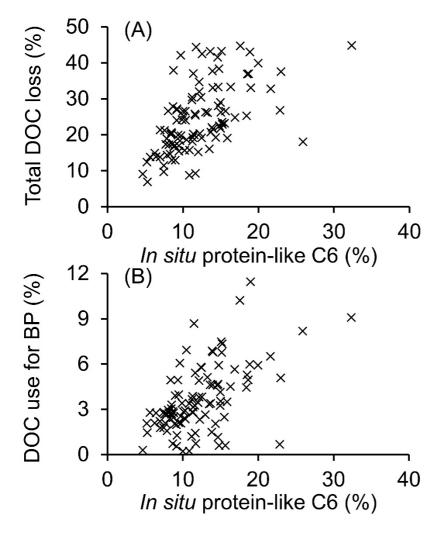
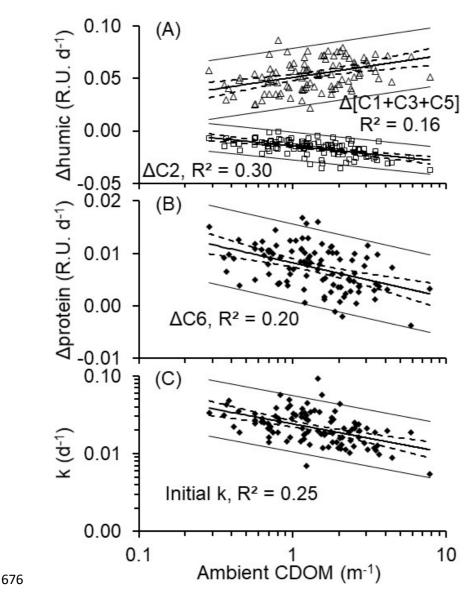


Figure 3. Microbial use of dissolved organic (DOC) during 28-d dark laboratory incubations of water from 101 lakes in a subarctic tundra landscape in northern Sweden. Percentages of (A) total loss of DOC and (B) use of DOC for bacterial production are plotted against the *in situ* share (%) of protein-like fluorescent dissolved organic matter (component 6).



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Figure 4. (A-B) Incubation changes in fluorescence components and in (C) initial decay coefficients k regressed from colored dissolved organic matter at 440 nm (CDOM). Dashed lines show 95% CI of the regressions. Observations are 95% likely to fall between the outer upper and lower lines running parallel with the regression lines.

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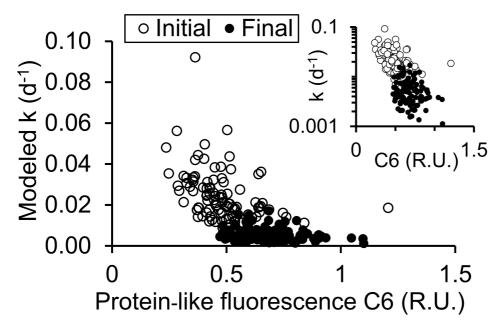


Figure 5. Modeled decay coefficient k plotted against the absolute concentration of proteinlike fluorescence C6 at the initial and final stage of 28-day dark laboratory incubations of water from 101 Swedish tundra lakes. The insert figure shows the same data, but on a logarithmic y-axis scale.