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**Inner-sphere reorganisation energy
of iron-sulphur clusters
studied by theoretical methods**

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Abstract

Models of several types of iron-sulphur clusters (e.g. $\text{Fe}_4\text{S}_4(\text{SCH}_3)_4)^{-2/-3/-4}$) have been studied with the density functional B3LYP method and medium sized basis sets. In vacuum, the inner-sphere reorganisation energies are 40, 76, 40, 62, 43, and 42 kJ/mole for the rubredoxin, [2Fe-2S] ferredoxin, Rieske, [4Fe-4S] ferredoxin, high-potential iron protein, and desulfoferredoxin models, respectively. The two first types of clusters were also studied in the protein, where the reorganisation energy was approximately halved. This change is caused by the numerous $\text{NH}\cdots\text{S}_{\text{Cys}}$ hydrogen bonds to the negatively charged iron-sulphur cluster, giving rise to a polar local environment. The reorganisation energy of the iron-sulphur clusters is low because the iron ions retain the same geometry and coordination number in both oxidation states. Cysteine ligands give approximately the same reorganisation energy as imidazole, but they have the advantage of stabilising a lower coordination number and giving more covalent bonds and therefore more effective electron-transfer paths.

Introduction

Iron-sulphur clusters are ubiquitous in biology, being present in all types of organisms from bacteria to higher plants and animals ^{1,2}. They consist of iron ions surrounded by four sulphur ions, either cysteine thiolate groups or inorganic sulphide ions. Regular clusters with one (rubredoxins), two, three, or four (ferredoxins) iron ions are known, as well as a number of more irregular clusters (occasionally with other protein ligands than cysteine) ³⁻⁶. The properties of the six types of sites studied in this paper are shortly summarised in Table 1.

The prime function of the iron-sulphur clusters is electron transfer. Together with cytochromes and blue copper proteins they constitute the most important groups of electron carriers in nature. The reduction potentials of iron sulphur clusters (-700 to +400 mV ⁵; all reduction potentials are relative the standard hydrogen electrode) are in general lower than those of the other two types of electron carriers (-300 to +500 mV for the cytochromes ^{6,7} and +180-1000 mV for the blue copper proteins ⁸). However, potentials of the individual sites and proteins vary a lot. During electron transfer, iron alternates between the Fe(II) and Fe(III) states. In general, the iron ions are in the high-spin state, but for the polynuclear sites, they are antiferromagnetically coupled to give a low total spin ^{2,5,6,9,10}.

According to the semiclassical Marcus theory ¹¹, the rate of electron transfer is given by

$$k_{ET} = \frac{2\pi}{\hbar} \frac{H_{DA}^2}{\sqrt{4\pi\lambda RT}} \exp\left(\frac{-(\Delta G^0 + \lambda)^2}{4\lambda RT}\right) \quad (1)$$

Here, H_{DA} is the electronic coupling matrix element, which depends on the overlap of the wavefunctions of the two states involved in the reaction, ΔG_0 is the free energy of the reaction (the change in reduction potential), and λ is the reorganisation energy (the energy associated with relaxing the geometry of the system after electron transfer). The coupling element depends on the localisation of the electron to be transferred and the protein matrix between the two active sites. The reduction potential is a function of the electronic and solvation energies (including both the surrounding protein and solvent) of the two sites. The reorganisation energy is normally divided into two parts: inner-sphere (λ_i) and outer-sphere reorganisation energy (λ_o), depending on which atoms are relaxed. For a metal-containing

protein, the inner-sphere reorganisation energy is associated with the structural change of the first coordination sphere, whereas the outer-sphere reorganisation energy involves structural changes of the remaining protein as well as the solvent.

Several groups have studied the variation of the reduction potential in various iron-sulphur clusters using theoretical methods¹²⁻²⁰. For example, it has been shown that the reduction potential of related proteins and mutants can be predicted with reasonable accuracy (average error 50 mV) if the crystal structure is known¹⁷. If the structure is not known, the accuracy is worse²¹.

However, no direct estimate of the inner-sphere reorganisation energy of iron-sulphur clusters in proteins seems to be available. From a theoretical point of view, the inner-sphere reorganisation energy is interesting because it is the only parameter in Eqn. (1) that does not depend on the detailed structure of the surrounding protein. Therefore accurate calculations of realistic models of the isolated metal centres would allow a general comparison of the various types of electron-transfer proteins and how they have optimised their function.

Recently, we have published a detailed discussion of the inner-sphere reorganisation energy of the blue copper proteins and the related Cu_A dimer (found in cytochrome *c* oxidase and nitrous oxide reductase), both in vacuum and in the protein²²⁻²⁴. In order to put these results in perspective, we calculate in this paper the inner-sphere reorganisation energy of a number of iron-sulphur cluster models. We compare the results with each other and with those of the blue copper proteins, and discuss how the various sites have achieved a low reorganisation energy.

Methods

Quantum chemical geometry optimisations

Quantum chemical geometry optimisations were performed with the density functional method B3LYP (unrestricted formalism), as implemented in the Turbomole software^{25,26}. Hybrid density functional methods have been shown to give as good or better geometries as correlated ab initio methods for first-row transition metal complexes²⁷⁻²⁹, and the B3LYP method in particular seems to give the most reliable results among the widely available density functional methods³⁰. In all calculations, we have used for iron the double- ζ basis set

of Schäfer et al.³¹ (62111111/33111/311), enhanced with one *d*, one *f*, and two *p* functions with exponents 0.1244, 1.339, 0.134915, and 0.041843, respectively. For the other atoms, we have employed the 6-31G* basis set³². Only the five pure *d* and seven pure *f*-type functions were used. Calibrations have shown that geometries obtained with this approach do not change much when the basis set is increased^{33,34}. The full geometry of all models was optimised until the change in energy between two iterations was below 10^{-6} Hartree (2.6 J/mole) and the norm of the internal gradients was below 10^{-3} a.u. (0.053 pm or 0.057°). Several starting structures were tested to reduce the risk of being trapped in local minima. Only the structures with the lowest energy are reported. No symmetry restraints were imposed.

Six models of iron-sulphur clusters were studied, involving 1-4 iron and 0-4 sulphide ions (c.f. Table 1). Cysteine ligands were modelled by SCH_3^- and histidine by an imidazole (Im) group. The rubredoxin and desulfoferrodoxin models were assumed to be in the high-spin state (quintet for Fe^{II} , sextet for Fe^{III}), whereas the other models were studied in the open-shell low-spin states (singlet or doublet), obtained from antiferromagnetically coupling of the high-spin state. This is in accordance with experiments⁵ and earlier calculations of the electronic structures of the complexes^{9,10}. Thus, the oxidised form of the two [2Fe-2S] clusters contained two $\text{Fe}(\text{III})$ ions in their high-spin state, antiferromagnetically coupled to an open-shell singlet^{9,10} and the reduced form contained $\text{Fe}(\text{II})+\text{Fe}(\text{III})$, both in the high spin state coupled to a doublet.

Similarly, the [4Fe-4S] clusters were treated as two pairs of ferromagnetically coupled high-spin iron ions^{2,5,9}, coupled antiferromagnetically together to give a low total spin quantum number. Yet, it should be noted that especially for the reduced cluster, there are several competing electronic states^{9,10}. Our study is not intended to exhaustively investigate the electronic structures of these clusters, but to give a picture of their reorganisation energies compared to those of other iron-sulphur clusters. Therefore, we have only studied one electronic state at each oxidation level, the lowest open-shell singlet or doublet obtained by an antiferromagnetic coupling of the high-spin states. This gives states that are similar (in terms of geometry, spin densities, and charges) to those studied in detail by Noodleman and coworkers^{9,10}. More precisely, our $\text{Fe}^{\text{II}}_3\text{Fe}^{\text{III}}$ state closely corresponds to their OC1 state (i.e. the highest occupied orbital has a σ rather than δ interactions between the reduced iron pair,

even if the interaction is far from pure), whereas our $\text{Fe}^{\text{II}}\text{Fe}^{\text{III}}_3$ state is close to their OS3 state (i.e. the lowest unoccupied minority-spin orbital is has iron, rather than sulphur, character).

Reorganisation energies

The inner-sphere reorganisation energy was estimated in the same way as for the blue copper proteins ²². The reorganisation energy for the oxidised complex (λ_{ox}) was calculated as the difference in energy between the oxidised complex at its optimal geometry and at the optimal geometry of the reduced complex. Likewise λ_{red} was calculated as the energy of the reduced complex at its optimal geometry minus the energy of the reduced complex calculated at the geometry optimal for the oxidised complex. The total inner-sphere reorganisation energy for a self-exchange reaction (λ_i) is the sum of λ_{ox} and λ_{red} . In variance to the outer-sphere reorganisation energy, the inner-sphere reorganisation energy does not change significantly when a protein docks with its donor or acceptor protein ³⁵. Therefore, it makes sense to study the inner-sphere reorganisation energy of isolated sites without considering their redox partners.

Frequencies were calculated for some of the optimised complexes with the Gaussian-98 software ³⁶. They were scaled by a factor of 0.963 ³⁷. Force constants for the various bonds, angles, and torsions were calculated from the Hessian matrix using the method of Seminario ³⁸, which has the advantage of being invariant to the choice of internal coordinates. The force constants were used to calculate approximate contributions to the reorganisation energy from various internal distortions:

$$\lambda_j = \sum_b k_b^j (b_{\text{ox}} - b_{\text{red}})^2 + \sum_{\theta} k_{\theta}^j (\theta_{\text{ox}} - \theta_{\text{red}})^2 + \sum_{\varphi} \frac{k_{\varphi}^j}{2} (1 - \cos(n(\varphi_{\text{ox}} - \varphi_{\text{red}}))) \quad (7)$$

where the three terms represent the energies of bond stretching, angle bending, and dihedral torsions, respectively. b_{ox} , b_{red} , θ_{ox} , θ_{red} , φ_{ox} and φ_{red} are the bond lengths, angles, and dihedral angles in the optimal oxidised or reduced geometry, respectively, and k_b^j , k_{θ}^j , and k_{φ}^j are the corresponding force constant, applying for the state j , where j is either the oxidised or reduced state.

Spin-coupling effects

For the [2Fe-2S] site, we examined the effect of spin coupling on the reorganisation energy (the low-spin state is not a pure spin state) using the methods developed by Noodleman and coworkers^{9,13}. In essence, the energy of the pure low-spin ground state (E_{ox} and E_{red} in Eqns. 2-6) is obtained from the low-spin ("broken symmetry") and high-spin energies (E_{BS} and E_{HS}) together with the Heisenberg spin coupling (J_{ox} and J_{red}) and electron delocalisation parameters (B), the latter estimated from the energy difference of the orbitals corresponding to the u and g components of the metal d orbitals split by resonance delocalisation ($e_{HS,u}$, $e_{HS,g}$)¹³. It was hard to identify the correct orbitals in the present systems without symmetry, where the orbitals are far from pure. Therefore, we simply used the HOMO-LUMO gap of the beta high-spin orbitals. This gives the smallest possible B factor, which however still was too large compared to experiments (see below). The high-spin wavefunction was calculated at the same geometry as the low-spin solution. However, we also tested two other conceivable geometries, the optimal high-spin geometry and a symmetrised low-spin geometry (with all Fe-S_i and Fe-S_{Cys} equal to their average in the low-spin geometry)¹³. This changed the orbital energy difference by less than 5%. The following relations were used¹⁵:

$$J_{ox} = 2(E_{HS} - E_{BS})/25 \quad (2)$$

$$E_{ox} = E_{BS} - 5J_{ox}/2 \quad (3)$$

$$B = (e_{HS,u} - e_{HS,g})/10 \quad (4)$$

$$J_{red} = (E_{HS} - E_{BS})/10 + B/2 \quad (5)$$

$$E_{red} = E_{BS} - 2J_{red} \quad (6)$$

Geometry optimisations in the protein

For the rubredoxin and [2Fe-2S] ferredoxin sites, geometry optimisations were also performed inside the proteins. These were run with the COMQUM-00 program^{24,39}, which combines the quantum chemical software Turbomole²⁵ with molecular mechanical minimisations performed by Amber 5.0⁴⁰. COMQUM, divides the protein (including solvent) into three subsystems. The central system 1 (Fe(SCH₃)₄ or (SCH₃)₂FeS₂Fe(SCH₃)₂) was optimised by the B3LYP method. System 2 consisted of all atoms in all amino acids (or solvent molecules) within 1.0 nm of any atom in the quantum system. It was optimised by

classical methods. Finally, system 3 comprised the rest of the protein and a cap of water molecules with a radius of 2.4-2.6 nm, centred on the geometric centre of the protein. System 3 was included in all calculations, but was kept fixed at the crystal geometry (an equilibrated conformation for the water cap; crystal waters were kept at the crystal geometry).

In the quantum chemical calculations, system 1 was represented by a wave function, whereas systems 2 and 3 were modelled by an array of point charges, one for each atom, taken from the Amber force-field libraries ⁴¹. Therefore, the polarisation of the quantum system by the protein is considered in a self-consistent way. In the classical energy and force calculations, systems 1-3 were represented by the Amber force field ⁴¹, but without any electrostatic interactions (which are already treated by quantum mechanics). Special action was taken when there is a bond between the classical and quantum chemical systems (a junction) ³⁹. The quantum chemical system was truncated by hydrogen atoms at the junctions (the C^α of the cysteine ligands), the positions of which were linearly related to the corresponding heavy (typically carbon) atoms in the full system.

The total energy was calculated as:

$$E_{tot} = E_{QC} + E_{MM123} - E_{MM1} \quad (8)$$

Here, E_{QC} is the quantum chemical energy of system 1 with H junction atoms, including all the electrostatic interactions. Similarly, E_{MM1} is the classical energy of system 1, still with H junction atoms, but without any electrostatic interactions. Finally, E_{MM123} is the classical energy of systems 1-3 with C junction atoms and no electrostatics. This approach is similar to the one used in the Oniom method ⁴². The calculated forces are the gradient of this energy, taking into account the variation in the junction atoms ²⁴.

Optionally, system 2 was optimised by a full molecular mechanics optimisation within each cycle of the quantum chemical geometry optimisation. In these calculations, systems 1-3 were represented by standard parameters from the classical force field. Electrostatic interactions were included, using Amber charges for systems 2 and 3 ⁴¹ and quantum chemical charges for system 1 ²⁴.

The reorganisation energy in the protein was calculated from the energies of the isolated quantum system only. For calculations with a fixed protein, we could have included also the electrostatic interaction with the protein in the reorganisation energies. This changes the

individual values of λ_{ox} and λ_{red} , but not their sum by more than a few kJ/mole ³⁹. However, for the flexible proteins, this procedure would include a significant amount of the outer-sphere contribution to the reorganisation energy and it would give unstable energies. This is caused by the local minima problem. We use a single minimised structure for the calculation of the reorganisation energy. However, differences in the hydrogen-bond structure or the positions of the water molecules strongly affect the estimates, and it is impossible to ensure that we have found the global minimum. Moreover, the reorganisation energies should be free energies, so they should reflect the dynamics of the system, especially for the solvent molecules. Therefore, direct estimates of the outer-sphere reorganisation energy from the COMQUM data are not reliable.

The proteins

The calculations on rubredoxin (from *Desulfovibrio vulgaris*) were based on two crystal structures, the reduced protein at 92 pm resolution and the oxidised protein at 200 pm resolution (Brookhaven protein databank files 1rb9 and 1rdv, respectively) ^{43,44}. The calculations on [2Fe-2S] ferredoxin were based on the oxidised and reduced crystal structures from *Anabaena* PCC7119 (130 and 117 pm resolution; PDB files 1qt9 and 1czp, respectively) ⁴⁵. The proteins were protonated and equilibrated (keeping heavy atoms fixed at the crystal position) together with a water cap (2.4 nm for rubredoxin, 2.6 nm for ferredoxin) in the same way as for the blue copper proteins ^{46,47}. For residues with multiple conformations, the one with the lower occupancy was ignored, except for residues 45-48 in reduced ferredoxin, where the conformation representative of the reduced site was used (0.4 occupancy). Some water molecules far from the proteins or with low occupancy and close to other atoms were also removed.

Based on the solvent accessibility and hydrogen-bond pattern, the two histidine residues in rubredoxin were assumed to be doubly protonated (positively charged). No histidines are present in the ferredoxin. The other amino acids were assumed to be in their normal protonation state at neutral pH, except for the cysteine ligands of the iron sites, which were considered to be deprotonated and negatively charged. The charges of all atoms were taken from the Amber libraries ⁴¹, except for the iron site, which were obtained by quantum

chemical calculations. Those of ferredoxin were taken from Noodleman et al. ⁹, assuming that the most solvent-exposed iron ion (the one bound to Cys-41 and 46) is the reduced ion. Those of the rubredoxin site were taken from Merz-Kollman calculations ⁴⁸ on $\text{Fe}(\text{SCH}_3)_4$, using a high number of potential points (~40 000). They were 1.209, -0.8434, 0.2200, -0.0594, and 0.0041 e for the Fe, S $^\gamma$, C $^\beta$, H $^\beta$, and C $^\alpha$ atoms in the reduced site and 0.8144, -0.5422, 0.0353, 0.0178, and -0.1687 e in the oxidised site. The numbers of amino acids, water molecules, and atoms in each protein are given in Table 2. All calculations were run on IBM SP2, SGI Origin 2000 or Octane workstations.

Results and Discussion

The rubredoxin site

First, we studied $\text{Fe}(\text{SCH}_3)_4^{-2-}$ as a model of the rubredoxin site. The optimised geometries are set out in Table 3. In the reduced state, all four Fe-S distances are 242 pm. This is longer than the values from experimental structure determinations. For the three most accurate (lowest resolution) crystal structures of reduced proteins in the Brookhaven protein data bank, the average Fe-S distance is 229 pm ⁴⁹. Extended X-ray absorption fine structure (EXAFS) measurements give a slightly longer Fe-S bond length, 232 pm ⁵⁰. Interestingly, small inorganic model complexes have an even longer Fe-S bond length, 236 pm ⁵¹.

When our $\text{Fe}(\text{SCH}_3)_4$ model is oxidised, the Fe-S bond lengths decrease to 232 pm (Figure 1). Again, this is longer than those obtained from experiments. The average Fe-S distance in the six most accurate structures of oxidised proteins is 229 pm ⁵². This is slightly longer than the EXAFS result of 226 pm ⁵⁰ and the average Fe-S distance in model compounds, 227 pm ⁵¹.

Thus, we can conclude that our vacuum optimisations give Fe-S bonds that are 5-6 pm longer than those of accurately determined model compounds. However, the *difference* in the Fe-S bond length between the reduced and oxidised structure is well described in our calculations: Our estimate, 10 pm, is very close to the result obtained for inorganic models, 9 pm ⁵¹. Therefore, the calculated reorganisation energy, 40 kJ/mole, should be quite accurate.

However, there is a clear difference between the Fe-S_{Cys} bond lengths in inorganic models and in proteins, especially for the reduced model. This is probably an effect of polar

interactions in the protein. In the rubredoxins, there are at least six hydrogen bonds between backbone amide groups and the S_{Cys} atoms⁵³. A detailed study of the analogous ZnCys₄²⁻ cluster in alcohol dehydrogenase showed that such hydrogen bonds decrease the Zn-S distances by ~7 pm⁵⁴.

Therefore, we ran a series of geometry optimisations of the rubredoxin cluster in the protein, using the combined quantum chemical and molecular mechanical program COMQUM-00^{24,39}. Two crystal structures were used (one oxidised, one reduced)^{43,44}. For each protein, two calculations at each oxidation state were run, one with the protein fixed at the crystal structure, and one where the protein was allowed to relax. The results in Table 3 show that the average Fe-S_{Cys} bond length in the protein (237 pm) is 5 pm shorter than in vacuum for the reduced complex. This is 5 pm longer than the EXAFS value⁵⁰, reflecting a systematic error of the B3LYP method^{33,34}. The variation among the four calculations is quite small, ~1 pm, showing that the results are reliable.

For the oxidised complex, the Fe-S_{Cys} bonds are only 2 pm shorter in the protein (231 pm) than in vacuum. Again, this is 6 pm longer than the EXAFS results^{5,50,55}. Therefore, the change in the Fe-S_{Cys} bond length upon oxidation is very well reproduced, giving confidence in the calculated reorganisation energies, presented in Table 4. It can be seen that these energies vary quite substantially among the various calculations (11-35 kJ/mole, average 22 kJ/mole), but all are smaller than the vacuum value (40 kJ/mole). Thus, we can conclude that the dielectric surroundings of the iron site reduce the reorganisation energy by almost a factor of two. The reason for this effect is that the isolated iron site has a high charge, -2 in the reduced state and -1 in the oxidised state. NH-S_{Cys} hydrogen bonds in the protein solvate this charge, but more for the reduced than the oxidised state. This explains why the change is larger for the reduced state.

The inner-sphere reorganisation energy of rubredoxin has been estimated to be 22-26 kJ/mole from the change in the Fe-S_{Cys} distances in EXAFS experiments (6 pm) combined with the corresponding vibrational frequency^{56,57}. This is close to our estimates. In fact, a harmonic analysis based on the calculated Hessian matrix and the geometry change during reduction in vacuum shows that the Fe-S_{Cys} bonds give the major contribution to the reorganisation energy, in total ~20 kJ/mole. Other bond lengths contribute by less than

1 kJ/mole and the angles by less than 6 kJ/mole (dominated by the S-Fe-S angles). The contribution from the dihedral angles is larger, but for these, the simple approximation in Eqn. 7 breaks down (their sum is much larger than the total reorganisation energy). However, it is clear that our calculated reorganisation energy is more accurate than estimates from vibrational frequencies, since we consider all degrees of freedom and not only the four Fe-S_{Cys} bonds.

Desulforedoxins and the S-Fe-S angle

The S-Fe-S angles in our optimised models are 109-110° for the reduced and 106-112° in the oxidised model. The larger variation in the latter model is due to stronger hydrogen bonds between the methyl groups and the thiolate ions (c.f. Figure 1). The angle is 104-115° in inorganic models⁵¹. Most rubredoxins have a similar variation in the S-Fe-S angles, 104-117°.

However, in the desulforedoxins, a group of rubredoxins from the bacteria *Desulfovibrio*, the variation is appreciably larger, 103-122°⁵⁸. The reason for this is that two of the cysteine ligands come directly after each other in the sequence (Cys-28 and 29), which for steric reasons gives rise to the largest angle. This has been taken as evidence that the site is in an entatic state⁵⁸, i.e. a state strained by the protein to give a catalytic advantage.

In order to test this suggestion, we optimised the geometry of the rubredoxin model with one of the S-Fe-S angles constrained to 121.8° as in desulforedoxin⁵⁸. This increased the energy of the complex, but only by 3 kJ/mole in both the reduced and oxidised states. Thus, the strain is very small in energy terms. Moreover, the calculated reorganisation energy of the constrained complex is 4 kJ/mole *higher* than for the unconstrained complex, so the constraint does not enhance the rate of electron transfer. Similarly, the reduction potential is unlikely to change significantly, since the energy of reduced and oxidised forms increased by the same amount when constrained. Therefore, it seems that this larger angle, which undoubtedly is caused by the folding of the protein, does not affect the properties of the iron-sulphur site in any significant way.

This analysis shows that a large difference in a geometric parameter between two protein structures does not necessarily imply any relationship to the function of the site; instead, it may reflect a small force constant (the bond or angle is flexible) so that the difference in

energy terms is small and therefore of minor functional importance. Similar results have been obtained for axial interactions in blue copper proteins and Cu_A ^{23,59,60}.

The [2Fe-2S] ferredoxin site

Next, we studied $(\text{SCH}_3)_2\text{FeS}_2\text{Fe}(\text{SCH}_3)_2$ as a model of the [2Fe-2S] ferredoxins. The optimised structures are described in Table 5. The Fe-Fe distance in the oxidised site is 285 pm, while the Fe- S_i distances are 226-227 pm and the Fe- S_{Cys} distances 235-237 pm (S_i denotes an inorganic sulphide ion). The variation in the Fe-S distances is caused by hydrogen bonds between the methyl groups and the sulphur ions (c.f. Figure 2). The Fe-S distances are 4-7 pm longer than in model compounds and protein crystal structures⁶¹⁻⁶³. Similarly, the calculated Fe-Fe distances seem to be ~15 pm too long. However, as can be seen in Figure 3, the Fe-Fe potential energy surface is quite flat: A 15-pm difference corresponds to a change in energy of only ~5 kJ/mole. Moreover, spin coupling effects (see below) tend to shorten this distance by at least 4 pm.

When the model is reduced, the Fe- S_{Cys} and Fe-Fe distances become 9-12 and 14 pm longer, respectively (Figure 2). The Fe- S_i distances to one of the iron ions increase by 15 pm, whereas those to the other ion decrease by 2 pm. Thus, they become quite dissimilar, 241 and ~225 pm. This shows that the added electron is localised on one iron atom (the one with the longer bonds). However, it is also strongly delocalised over the neighbouring sulphur ions.

No binuclear inorganic models in the reduced state seem to exist⁵⁵. However, the first crystal structure of a reduced [2Fe-2S] site in a protein was recently published⁴⁵. It does not show the same asymmetry as our structures. This is because the unpaired electron alternates between the two iron sites during the data collection⁵⁵. EXAFS data are also available⁶⁴, but they do not distinguish between the two types of Fe-S bonds. Yet, it is clear that our calculated Fe-S and Fe-Fe distances are longer than those in the proteins. Moreover, the experimental structures show a smaller decrease in the Fe-S distance upon reduction in the protein (1-3 pm) than in our calculations (9 pm). This is not surprising, since ~60 % of the iron-sulphur sites in the crystal are oxidised⁴⁵, giving rise to multiple conformations that are hard to discern.

The calculated reorganisation energy of our model in vacuum is quite large, 76 kJ/mole, almost twice as much as for the rubredoxin model. This is in accordance with a lower rate of electron transfer for the [2Fe-2S] sites in proteins as well as in model systems^{55,61}. The increase can be rationalised by a detailed study of the change in bond lengths and force constants of the two models. In the dimer, the reduction is mainly confined to one of the iron ions. The changes in the bond lengths upon reduction of this iron ion are slightly larger than for the rubredoxin monomer (on average 13 pm compared to 9 pm). However, there are also appreciable changes around the other iron ion (6 pm on average). The Fe-S_{Cys} force constants are slightly lower in the dimeric site (0.017 compared to ~0.015 kJ/mole/pm² in the reduced site and 0.032 compared to ~0.026 kJ/mole/pm² in the oxidised site), but those of the Fe-S_i bonds (which are not present in the monomer) are larger (0.027 and 0.039 kJ/mole/pm²). Therefore, the total reorganisation energy of the ferredoxin model is almost doubled.

At first, the increase in reorganisation energy for the dimeric iron-sulphur clusters may seem strange, considering that the reorganisation energy of the dimeric Cu_A site was found to be appreciably *lower* than for the blue-copper monomer (43 kJ/mole compared to 62 kJ/mole). However, the reason for this difference is that the unpaired electron in the mixed-valence Cu_A site is delocalised over the two copper ions^{23,65,66}. As suggested by Larsson and coworkers⁶⁷ and confirmed by detailed calculations²³, such delocalisation of an electron over two monomers reduces both the bond length change and the force constants of the site, leading to a reduction of the reorganisation energy.

It is clear that there are significant differences between the calculated and experimental structure of the reduced site. Considering the results for the rubredoxins, an important reason for this discrepancy may be solvation effects in the protein. The isolated site has a charge of -3, which is larger than for the reduced rubredoxin site (-2). Therefore, we ran a series of COMQUM calculations also of the [2Fe-2S] ferredoxins in the protein. Again, we used crystal structures for both reduced and oxidised sites, but the protein was kept rigid in all calculations. The results of these calculations are gathered in Table 5. As for rubredoxin, the Fe-S_{Cys} bond lengths shorten in the protein, by 9 pm in the reduced site and by 3 pm in the oxidised site. These changes are larger than for the rubredoxins, reflecting the larger net charge of the binuclear sites. Interestingly, the Fe-S_i bond lengths do not change significantly

in the protein, but the variation increases slightly. The Fe-Fe distance varies quite substantially in the calculations but without any clear trend. This probably reflects the flexibility of the bond (c.f. Figure 3).

Consequently, our optimisations in the protein predict that the average Fe-S_{Cys} and Fe-S_i bond lengths should decrease by ~5 pm upon oxidation. This leads to a reduced reorganisation energy inside the protein (Table 6). As for the rubredoxins, the individual values vary quite a lot, 37-52 kJ/mole, but both estimates are appreciably lower than in vacuum (75 kJ/mole). On average (44 kJ/mole), the reorganisation energy is almost halved in the protein. It is also notable that λ_{ox} is appreciably larger than λ_{red} . This is most likely caused by the large variation in the Fe-S bonds in the reduced site.

Finally, it should be noted that the ΔE_1 values (also included in Table 5), i.e. the cost of changing the energy of the iron site from the vacuum geometry to the one in the protein, are quite large for the [2Fe-2S] ferredoxin models, 43-110 kJ/mole. This is larger than the corresponding values for rubredoxin, 14-38 kJ/mole (Table 3), and also for the blue copper proteins and alcohol dehydrogenase (30-70 kJ/mole)^{24,39,68-70}. This is probably an effect of the large charge of the cluster and the many polar interactions in the protein. ΔE_1 can be interpreted as the strain energy of the cluster when it is bound to the protein. It includes, however, several terms normally not considered as protein strain⁵⁹. Interestingly, ΔE_1 is larger for the native protein structure (i.e. the oxidised site in the oxidised structure and vice versa); the opposite is normally found²⁴. ΔE_1 can be expected to decrease if the protein is allowed to relax, typically by 5-20 kJ/mole²⁴.

All energies presented until now are those directly obtained from the B3LYP low-spin calculations (the "broken-symmetry" solutions,^{9,10}). As discussed in the Methods section, this solution is not a pure spin state. Instead, the energy of the pure-spin ground state can be obtained by an extrapolation from this energy using also the properties of the high-spin wavefunction at the same geometry (Eqns. 2-6)^{9,10,13}. In Figure 3 we compare the Fe-Fe potential surface of the pure-spin and broken-symmetry states. It can be seen that the difference is quite small, less than 4 kJ/mole for all investigated distances (260-310 pm). Yet, spin coupling systematically favours shorter Fe-Fe distances in both oxidation states. Therefore, the optimum Fe-Fe distance is reduced by 4 pm in the oxidised state and by 6 pm

in the mixed-valence state. This makes the distances closer to those found in experiments (c.f. Table 5), even if the discrepancy is still 10-15 pm, corresponding to an energy cost of 1-5 kJ/mole.

Moreover, we have estimated the effect of spin coupling on the reorganisation energies. From Table 6, it can be seen that the effect in most cases is small, 1-3 kJ/mole. However, for the two mixed-valence calculations in the protein, the effect is larger, 6-12 kJ/mole, but with a varying sign. This can be traced back to the electron delocalisation parameter (B). This parameter can be estimated together with the Heisenberg spin coupling parameters (J) from Eqns 2-5. The Heisenberg spin coupling parameters for the oxidised state, J_{ox} , are 252-381 cm^{-1} , in excellent agreement with experimental estimates (298-364 cm^{-1}) but less than half as large than earlier quantum chemical estimates (763-868 cm^{-1}), obtained with other, less accurate, functionals¹⁵. However, the electron delocalisation parameters for the mixed-valence state, 1900-2200 cm^{-1} are much larger than in earlier calculations (394-912 cm^{-1})¹⁵. This overestimation propagates to the Heisenberg spin coupling parameters, J_{red} , which also are too large (1140-1260 cm^{-1}) compared both to experiments (196 cm^{-1}) and to other calculations (514-504 cm^{-1})¹⁵. If the B -factor contribution to J_{red} is ignored, the results are only slightly smaller than experiments, 139-186 cm^{-1} . This indicates that the method to estimate the B -factor from orbital energy differences does not work properly for these systems without symmetry. Therefore, and also because of the small effect of spin coupling observed in most systems, we decided to ignore this effect for the other spin-coupled systems.

Finally, it should be mentioned that we have also studied the fully reduced $(\text{SCH}_3)_2\text{FeS}_2\text{Fe}(\text{SCH}_3)_2^{4+}$ complex. However, the large negative charge of this complex makes it unstable in vacuum: Two of the cysteine models dissociate from the iron ions when it is optimised in vacuum.

The Rieske site

In the electron transport chain of chloroplasts and mitochondria, binuclear iron-sulphur clusters are found with unusually high reduction potentials (c.f. Table 1). These so-called Rieske centres are [2Fe-2S] ferredoxins with the cysteine ligands of one of the iron ions replaced by two histidine ligands. In the reduced state ($\text{Fe}^{\text{II}}\text{Fe}^{\text{III}}$), the extra electron resides on

iron ions with two histidine ligands, whereas it alternates between the two ions, in the normal ferredoxins ⁵. This is reproduced in our calculations. We have modelled these sites by $(\text{SCH}_3)_2\text{FeS}_2\text{Fe}(\text{Im})_2^{0/-}$. The optimised structures are shown in Table 7. All distances around the iron ions are shorter than in $\text{Fe}_2\text{S}_2(\text{SCH}_3)_4$, the Fe-Fe distance by 10 and 28 pm, the Fe-S_{Cys} distances by 2 and 6 pm, and the average Fe-S_i distances by 7 and 9 pm, for the oxidised and mixed-valence states, respectively. This is probably an effect of the smaller net charge (-1 and 0) of the Rieske models. The Fe-N distances are 211 and 218 pm, respectively.

There are two crystal structures available for Rieske centres, one from mitochondria and one from chloroplasts ^{71,72}. Both are in the mixed-valence (reduced) state. As can be seen from Table 7, the calculated Fe-S_{Cys} bond lengths are too long by 8-11 pm (as an effect of deficiencies in the B3LYP method and protein solvation effects), whereas the Fe-S_i and Fe-N bonds lengths are close to the experimental averages. The Fe-Fe bond length is exactly the same as in the two crystal structures, 271 pm, which of course is a lucky coincidence, considering the low resolution of the structures and the flexibility of this bond.

Upon oxidation, the Fe-S_{Cys} and Fe-N distances of the Rieske model decrease by ~7 pm, whereas the Fe-Fe distance increases by 4 pm. As in the [2Fe-2S] ferredoxin model, the Fe-S_i distances decrease slightly for the iron ion that is not oxidised (by 1-2 pm), whereas the distances to the other iron ion increase by 7-8 pm. However, contrary to what was found for the former system, the Fe-S_i distances are quite similar in the mixed-valence state, 225-229, whereas they are distinctly dissimilar in the oxidised state (230 pm for the iron ion bound to the thiolate groups and 219 pm for the other iron ion). This is of course an effect of the ligand substitution.

Interestingly, the inner-sphere reorganisation energy of the Rieske model, 40 kJ/mole, is appreciably lower than for the ferredoxin model and similar to that of the rubredoxin model. The reason for this is that the change in all iron-ligand distances are quite small, <8 pm, i.e. less than in the rubredoxin model.

Recently, it has been suggested that theoretical calculations of antiferromagnetically coupled polynuclear metal complexes often can be done in the ferromagnetically coupled high-spin state (which is computationally more simple) ^{73,74}. Therefore, we have calculated structures and reorganisation energies of the [2Fe-2S] ferredoxin and Rieske models also in

the high-spin states (with nine or ten unpaired electrons). The results in Tables 5-7 show appreciable differences compared to the low-spin calculations. Even if the distances to the terminal ligands do not differ by more than 1 pm, the Fe-S_i bond lengths may change by up to 9 pm and the flexible Fe-Fe distances by as much as 18 pm. For the Rieske site, the Fe-S_i bonds lengthen in the high-spin state, whereas in the ferredoxin model they both increase and decrease, giving a smaller variation in the reduced state, but a larger one in the mixed-valence state.

Most importantly, these changes in geometry have large effects on the reorganisation energy. As can be seen in Table 6 and 7, the reorganisation energy of the ferredoxin model in the high-spin state is decreased by 18 kJ/mole to 59 kJ/mole, whereas it is more than doubled for the Rieske model, to 82 kJ/mole. Thus, as regards geometries and reorganisation energies, it is not a good idea to use high-spin models, at least for iron-sulphur clusters.

The [4Fe-4S] ferredoxin site

Next, we studied three oxidation states of Fe₄S₄(SCH₃)₄ as a model of the [4Fe-4S] ferredoxin site. The optimised structures are described in Table 8. As expected, they are slightly distorted cubes (c.f. Figure 4). Compared to the [2Fe-2S] sites, the Fe-S_{Cys} distances are consistently shorter; the average Fe-S_{Cys} bond lengths in the three oxidation states are 239, 232, and 226 pm, respectively, compared to 247 and 236 pm in the two Fe₂S₂(SCH₃)₄ complexes. The Fe-S_i and Fe-Fe distances are similar to those found in the dimeric sites, but they vary quite a lot within the cuboidal sites.

Crystallographically characterised inorganic models and protein crystal structures are available for all three oxidation states^{61,62,64,75-83}. Their structures are included in Table 8. It can be seen that our calculated models have 4-12 pm too long Fe-S bonds and 6-41 pm too long Fe-Fe bonds. Probably, all distances will be shorter in the protein, as an effect of the hydrogen bonds present there (typically eight to the S_{Cys} and three to the S_i atoms⁸¹).

As in the inorganic model system, the calculated distances can be divided into two or three groups representing distances within or between Fe₂S₂ dimer planes (with the same or different oxidation state). For example, in the Fe^{II}₂Fe^{III}₂ state, the eight Fe-S_i distances within the two mixed-valence Fe₂S₂ pairs are longer, ~237 pm, than those between the two planes,

234 pm. In the model systems, the difference is similar, 3-6 pm ⁷⁶. The same is true for the Fe-Fe distances; the four between the planes are shorter than the two within the planes, 282 and 293 pm, respectively. In the models, the same trend is present, but the difference is smaller ^{76,82}.

For the reduced cluster, model compounds show large variation in the structure, ranging from compressed to elongated sites ⁷⁵⁻⁷⁸. This indicates that it is highly plastic ^{5,55}. Our optimised structure is similar to two of the observed structures, with four short bonds within the mixed-valence Fe₂S₂ pair, four long bonds within the reduced Fe₂S₂ pair, and two short and two long distances between the two pairs. Of course, such a clear distinction between the two Fe₂S₂ pairs can only be seen if the extra electron is localised on one pair, which is unlikely in models and proteins ^{84,85}. This may explain why our structures show a larger variation in the bond lengths than the experimental ones.

When the Fe^{II}₃Fe^{III} site is oxidised, several changes are seen (c.f. Figure 4a). The Fe-S_{Cys} bonds contract by 5-8 pm (most for the reduced Fe₂S₂ pair) and the Fe-S_i distances within the reduced Fe₂S₂ pair decrease by 9 pm, whereas those of the mixed-valence pair hardly change at all. Two of the Fe-S_i bonds between the pairs decrease by 8 pm whereas the other two increase by 1 pm. Thus, six Fe-S bonds change by the same amount as in the rubredoxin site (where there are only four bonds). Therefore, the reorganisation energy is slightly larger than this site, 62 kJ/mole.

When the complex is further oxidised to the Fe^{II}Fe^{III}₃ state, changes in the geometry are smaller (see Figure 4b). All Fe-S_{Cys} bonds decrease by 6 pm, whereas most Fe-S_i bonds lengths change by less than 1 pm. Only two bonds within a pair increase by 3 pm and two bonds between the pairs decrease by 7 pm. All these changes are less than in the rubredoxin site. Therefore, it is somewhat unexpected that the reorganisation energy is slightly larger than for that site, 43 kJ/mole (even considering that there are four times as many Fe-S bonds in the [4Fe-4S] site). The reason is probably that there are quite substantial changes in the Fe₄S₄ core structure, as is flagged by a 28-pm increase in two of the Fe-Fe bonds. Compared to inorganic models, the calculated changes seem to be reasonable for the average Fe-S_i distances (2 pm), but slightly too large for the Fe-S_{Cys} bonds (4 pm) ⁸⁰.

Holm and coworkers have estimated the total self-exchange reorganisation energy ($\lambda_i + \lambda_o$) for the $\text{Fe}^{\text{II}}_3\text{Fe}^{\text{III}} - \text{Fe}^{\text{II}}_2\text{Fe}^{\text{III}}_2$ couple of the $\text{Fe}_4\text{S}_4(\text{SC}_6\text{H}_4\text{CH}_3)_4$ model compound to 146 kJ/mole ⁵⁷. They also estimated the inner-sphere part from the observed changes in the Fe-S bonds and the corresponding frequencies to 32 kJ/mole. The latter estimate is half as large as ours, probably because we have not included the protein in our calculations.

Similarly, the total self-exchange reorganisation energy of a high potential iron protein has been estimated to 70-90 kJ/mole ⁸⁶. This result is in accordance with our calculations, provided that the outer-sphere contribution of this protein is similar to that of plastocyanin (a protein of a similar size), ~40 kJ/mole ⁸⁷, and that the reorganisation energy of the HiPIP model inside the protein is approximately half of the vacuum estimate, as was observed for the rubredoxin and [2Fe-2S] ferredoxin models, as well as for blue copper proteins ²⁴.

The desulfoferredoxin site

Recently, it has been shown that desulfoferredoxin contains an iron site with only one cysteine and four histidine ligands ⁸⁸. We have modelled this site by $\text{Fe}(\text{Im})_4(\text{SCH}_3)^{+/2+}$. As can be seen in Table 9, the optimised structures have two short and two long Fe-N_{His} bonds, due to $\text{CH}\cdots\text{S}_{\text{Cys}}$ hydrogen bonds. The bond lengths in the oxidised structure are close to those in the crystal structure of desulfoferredoxin; the average Fe-N distance is 215 pm in both structures, whereas our calculated Fe-S_{Cys} distance is 3 pm shorter than in the crystal structure. The reorganisation energy of the desulfoferredoxin model is close to that of rubredoxin, 42 kJ/mole.

This indicates that imidazole and cysteine give rise to sites with similar electron-transfer properties. Therefore, we studied the five models of the type $\text{Fe}(\text{Im})_n(\text{SCH}_3)_{4-n}$, with $n = 0-4$ ($n = 0$ gives the rubredoxin site). The results are also included in Table 9 and show that the reorganisation energy varies between 40 and 49 kJ/mole. It is lowest for the rubredoxin site, whereas the other four sites all have a reorganisation energy around 47 kJ/mole. Interestingly, the Fe-S_{Cys} bonds change more during oxidation than the Fe-N_{His} bonds. This indicates that the latter bond is stiffer, although the Fe-S_{Cys} bond involve an interaction between two oppositely charged ions. This is confirmed by direct calculations of the corresponding force constants,

which are 0.027 and 0.038 kJ/mole/pm² for the Fe-N bonds in Fe(Im)₄^{2+/3+}, but 0.017 and 0.032 kJ/mole/pm² for the Fe-S bonds in Fe(SCH₃)₄^{2-/1-}.

Thus, in terms of reorganisation energy, cysteine and histidine can be used almost interchangeably. However, cysteine has the advantage of stabilising a low coordination number for both oxidation states (due to its negative charge and bulkiness), whereas higher coordination numbers are preferred with the uncharged histidine ligands. Moreover, cysteine forms more covalent bonds and therefore a better path for electron transfer. This can be seen from the higher spin densities on the cysteine models than on the imidazoles (e.g. 0.10 *e* compared to 0.05 *e* for the reduced Fe(SCH₃)₄ and Fe(Im)₄ complexes and 0.30 *e* compared to 0.20 *e* for the oxidised complexes).

Comparison with the blue copper proteins

We have shown that the inner-sphere reorganisation energy of six types of iron-sulphur clusters is similar to those found for the blue copper proteins and the binuclear Cu_A site, both in vacuum (43-90 kJ/mole²²⁻²⁴) and in the proteins (27-38 kJ/mole²⁴). It is instructive to compare how the two types of sites have achieved such a low reorganisation energy. The blue copper proteins reduce their reorganisation energy by using ligands with a small force constant for the bond to copper (methionine) and ligands which give structures that are similar for Cu(I) and Cu(II) (cysteine)²². The Cu_A site has a delocalised electron in the mixed-valence state, which reduces the force constants and the change in the metal-ligand bond lengths upon reduction²³.

In order to examine how the iron-sulphur clusters have achieved a low reorganisation energy, we start to examine an iron ion in aqueous solution, modelled by Fe(H₂O)₆. As can be seen in Table 10, this complex has an inner-sphere reorganisation energy of 65 kJ/mole. This is actually slightly less than for the [2Fe-2S] site. However, the outer-sphere component is large in aqueous solution, giving a total reorganisation energy of ~250 kJ/mole^{89,90}. The reorganisation energy of the corresponding copper complex is more than five times larger (336 kJ/mole)²², because Cu(I) assumes a lower coordination number than Cu(II). Yet, even if Cu(I) is forced to have the same coordination number and geometry as Cu(II), the reorganisation energy is still twice as large as for iron (112 kJ/mole)²², since the changes in

the metal-ligand bond distances are larger: 24 pm for four bond and 3 pm for two bonds for copper (Cu^{2+} is Jahn-Teller distorted) compared to 10 pm for all bonds in the iron complex.

Both the iron-sulphur clusters and the blue copper proteins are four-coordinate. Therefore, we studied the $\text{Fe}(\text{H}_2\text{O})_4$ system. The reorganisation energy of this complex is almost identical to that of the six-coordinate complex, 66 kJ/mole. This is an effect of a smaller change in the bond lengths (8 pm) and a lower number of bonds combined with more stiff bonds. The geometry is tetrahedral for both oxidation states. This is in strong contrast to copper, for which $\text{Cu}(\text{I})$ forms a tetrahedral complex, whereas $\text{Cu}(\text{II})$ assumes a square-planar structure. Therefore, the reorganisation energy of copper (136 kJ/mole) is almost three times higher than for iron ²².

The situation is similar for $\text{Fe}(\text{NH}_3)_4$ complex. It has a very low reorganisation energy of only 18 kJ/mole, which is more than seven times lower than for copper (122 kJ/mole). Thus, such a complex would constitute a excellent electron-transfer site. However, four neutral ligands do not form stable complexes with $\text{Fe}(\text{III})$ and amine ligands are not available in most proteins (the functional group of lysine has a too large acid constant and is therefore protonated at normal pH values). Instead, cysteine ligands are needed to lower the coordination number and give a favourable electron-transfer path, whereas histidine could be used instead of ammonia (but with higher reorganisation energies, as we saw in the preceding section).

Thus, we can conclude that iron a priori is a much better electron-transfer ion than copper, since the two oxidation states of iron have the same preferences in coordination number and geometry. Moreover, the iron-sulphur clusters, like the blue copper proteins, employ nitrogen and sulphur ligands rather than oxygen ligands, in order to reduce the metal-ligand force constants and therefore the reorganisation energy. This is a general rule that has also been observed experimentally ⁹¹. However, after this, it seems that the actual choice of iron ligands is determined more by practical reasons (i.e. that the metal should bind strongly to the protein) and by the need of obtaining a proper electronic path, than by considerations of the reorganisation energy. Similarly, the choice of the type of the iron-sulphur cluster (i.e. the number of iron ions, and the type of ligands), seems to be mainly a question of stability and reduction potential. A large cluster may decrease the distance of electron transfer ⁶⁵, allow the

site to be more buried in the protein, and give a directionality for electron transfer^{56,92}. This may explain why the polynuclear iron-sulphur clusters are more common than the rubredoxin site.

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Legends to the Figures

Figure 1. The difference in geometry between the reduced and oxidised (shaded) forms of the rubredoxin model $\text{Fe}(\text{SCH}_3)_4$.

Figure 2. The difference in geometry between the mixed-valence and oxidised (shaded) forms of the [2Fe-2S] ferredoxin model $(\text{SCH}_3)_2\text{FeS}_2\text{Fe}(\text{SCH}_3)_2$.

Figure 3. The potential surface of the Fe-Fe interaction in the mixed-valence (Mv) and oxidised (Ox) state of $(\text{SCH}_3)_2\text{FeS}_2\text{Fe}(\text{SCH}_3)_2$. Energies are given both for the broken-symmetry solution (for which the geometry was fully optimised in each step) and the corrected pure-spin ground state (Corr).

Figure 4. The difference in geometry (a) between the $\text{Fe}^{\text{II}}_3\text{Fe}^{\text{III}}$ and $\text{Fe}^{\text{II}}_2\text{Fe}^{\text{III}}_2$ (shaded) forms and (b) between the $\text{Fe}^{\text{II}}_2\text{Fe}^{\text{III}}_2$ (shaded) and $\text{Fe}^{\text{II}}\text{Fe}^{\text{III}}_3$ forms of the [4Fe-4S] ferredoxin model $\text{Fe}_4\text{S}_4(\text{SCH}_3)_4$.

Table 1. Chemical structure, formal oxidation states, and reduction potential of the six iron-sulphur clusters included in this investigation ⁵. The reduction potentials are relative the standard hydrogen electrode.

Site	Structure	Reduced state	Oxidised state	Reduction potential (mV)
Rubredoxin	FeCys ₄	Fe ^{II}	Fe ^{III}	-100 to 0
Desulfoferrodoxin	FeCysHis ₄	Fe ^{II}	Fe ^{III}	240
[2Fe-2S] ferredoxin	Cys ₂ FeS ₂ FeCys ₂	Fe ^{II} Fe ^{III}	Fe ^{III} ₂	-450 to -150
Rieske site	Cys ₂ FeS ₂ FeHis ₂	Fe ^{II} Fe ^{III}	Fe ^{III} ₂	-100 to +400
[4Fe-4S] ferredoxin	Cys ₄ Fe ₄ S ₄	Fe ^{II} ₃ Fe ^{III}	Fe ^{II} ₂ Fe ^{III} ₂	-700 to -300
High-potential iron protein	Cys ₄ Fe ₄ S ₄	Fe ^{II} ₂ Fe ^{III} ₂	Fe ^{II} Fe ^{III} ₃	+100 to +400

Table 2. The number of amino acids, water molecules, and atoms in systems 2 and 3 in the COMQUM calculations of rubredoxin and ferredoxin.

Protein	System	Residues	Water	Atoms
Rubredoxin, oxidised	2	36	252	1268
	3	16	1340	4237
Rubredoxin, reduced	2	36	257	1284
	3	16	1367	4311
Ferredoxin	2	54	222	1465
	3	44	1664	5618

Table 3. Optimised Fe-S_{Cys} distances for the rubredoxin model (Fe(SCH₃)₄) in vacuum and calculated in the protein with ComQUM^{24,39}. For the latter calculations, it is indicated what protein structure has been used (oxidised or reduced^{43,44}) and whether system 2 was allowed to relax or not (flexible or fixed). ΔE_1 is the energy difference (calculated in vacuum) of system 1 between the vacuum and ComQUM geometries. Experimental data from model compounds⁵¹ and proteins (both crystal structures^{52,87} and EXAFS data^{5,50,55}) are also included.

Oxidation state	System	ΔE_1 kJ/mole	Distance to Fe (pm)				
			S ₁	S ₂	S ₃	S ₄	S _{average}
II	Vacuum		242	242	242	242	242.3
	Reduced protein fixed	20.6	234	238	236	238	236.2
	Reduced protein flexible	18.1	236	238	238	236	236.7
	Oxidised protein fixed	27.4	234	241	237	239	237.7
	Oxidised protein flexible	37.2	237	238	239	240	238.5
	Models			232-238			236
	Protein crystals			224-236			229
	Protein EXAFS						232
III	Vacuum		232	232	232	232	232.4
	Reduced protein fixed	25.0	229	233	228	232	231.0
	Reduced protein flexible	36.6	228	232	231	233	230.9
	Oxidised protein fixed	15.9	229	231	229	231	230.1
	Oxidised protein flexible	14.1	231	229	230	230	230.2
	Models			225-228			227
	Protein crystals			223-233			229
	Protein EXAFS						226

Table 4. Inner-sphere reorganisation energies (kJ/mole) for the rubredoxin model ($\text{Fe}(\text{SCH}_3)_4$) calculated in vacuum and in the protein with COMQUM. For the latter calculations, it is indicated what protein structure has been used (oxidised or reduced ^{43,44}) and whether system 2 was allowed to relax or not (flexible or fixed).

System	λ_{red}	λ_{ox}	λ_i
Vacuum	21.4	18.3	39.7
Reduced protein fixed	15.9	8.2	24.1
Reduced protein flexible	6.7	4	10.7
Oxidised protein fixed	14.4	3.2	17.6
Oxidised protein flexible	19	16.1	35.1

Table 5. Optimised geometries for the [2Fe-2S] ferredoxin model (SCH₃)₂FeS₂Fe(SCH₃)₂ in vacuum and calculated in the protein with the COMQUM program ^{24,39}. For the latter calculations, it is indicated what protein structure has been used (oxidised or reduced ⁴⁵). All calculations were run with a fixed system 2. ΔE_1 is the energy difference (calculated in vacuum) of system 1 between the vacuum and COMQUM geometries. In vacuum, a set of calculations of the high-spin state is also presented. Experimental data from model compounds ^{61,62} and proteins (both crystal structures ^{45,63} and EXAFS data ⁶⁴) are also included.

Oxidation state	System	ΔE_1 kJ/mole	Distance to Fe (pm)				
			Fe-S _{Cys}		Fe-S _i		Fe-Fe
			range	av.	range	av.	
II+III	Vacuum		245-249	247.4	225-241	233.0	299
	Reduced protein	92.5	234-243	239.0	225-241	233.0	301
	Oxidised protein	54.6	234-240	237.5	221-246	231.6	289
	Protein crystals		230-236	232	218-229	224	275
	Protein EXAFS			224			276
	Vacuum, high spin		247-248	248	230-237	233.2	280
III+III	Vacuum		235-237	236.4	226-227	226.5	285
	Reduced protein	42.6	230-234	232.1	223-231	226.5	283
	Oxidised protein	110.5	228-238	233.8	226-233	228.4	302
	Models		230-233	232	219-223	221	267-270
	Protein crystals		222-237	229	211-228	221	260-278
	Protein EXAFS			223			270-273
	Vacuum, high spin		235-237	236.2	230-233	231.6	304

Table 6. Inner-sphere reorganisation energies (kJ/mole) for the [2Fe-2S] ferredoxin model $(\text{SCH}_3)_2\text{FeS}_2\text{Fe}(\text{SCH}_3)_2$ calculated in vacuum and in the protein with COMQUM. For the latter calculations, it is indicated what protein structure has been used (oxidised or reduced ⁴⁵). Reorganisation energies have been calculated both directly from the low-spin ("broken-symmetry") energies and from the energies of the pure spin state extrapolated from the broken-symmetry and high-spin solutions (the latter in brackets). Data for the high-spin vacuum structure are also given.

Protein	λ_{red}	λ_{ox}	λ_i
Vacuum	34.3 (35.5)	41.5 (44.7)	75.8 (80.2)
Reduced protein	44.7 (50.6)	6.9 (7.9)	51.9 (58.5)
Oxidised protein	29.0 (17.2)	7.8 (9.8)	36.8 (27.0)
Vacuum, high spin	25.5	33.5	59.0

Table 7. Geometries and inner-sphere reorganisation energies (kJ/mole) for the Rieske $(\text{SCH}_3)_2\text{FeS}_2\text{Fe}(\text{Im})_2$ models calculated by the B3LYP method, both at the low-spin (in accordance with experiments) and high-spin states. Experimental results have also been included ^{71,72}.

Oxidation state	System	λ	Distance to Fe (pm)			
		kJ/mole	S _{Cys}	S _i	N _{His}	Fe
II+III	Vacuum	18.3	233-239	225-229	216-220	271
	Protein crystals		222-231	223-235	213-223	271
	Vacuum, high spin	39.1	234-240	229-233	216-219	261
III+III	Vacuum	21.8	227-232	219-230	210-212	275
	Vacuum, high spin	43.0	227-231	222-239	211-212	293

Table 8. Geometries and inner-sphere reorganisation energies (λ) for the $\text{Fe}_4\text{S}_4(\text{SCH}_3)_4$ ferredoxin models calculated by the B3LYP method. Experimental results for model complexes ^{61,62,75-80} as well as proteins (both crystal structures ^{81,83} and EXAFS data ⁶⁴) have also been included when available.

Oxidation state	System	λ kJ/mole	Distance to Fe (pm)		
			S _{Cys}	S _i	Fe
II ₃ III ₁	Vacuum	32.7	237-241	233-246	277-322
	Models		228-231	227-237	271-281
	Protein crystals		228-232	226-232	254-270
	Protein EXAFS		226		271
II ₂ III ₂	Vacuum	λ_{red} 29.2	232-233	234-237	282-293
		λ_{ox} 18.3			
	Models		224-229	223-233	270-279
	Protein crystals		213-233	214-240	256-279
II ₁ III ₃	Protein EXAFS		225		266-273
	Vacuum	24.8	226-227	227-240	284-310
	Models		220-221	223-228	274 ^a
	Protein crystals		218-226	215-237	267
	Protein EXAFS		226		274

^a Average distance.

Table 9. Geometries and inner-sphere reorganisation energies for some iron models related to desulfoferrodoxin calculated by the B3LYP method. All complexes were studied in the high-spin state. The order of the L₁-L₄ ligands is the same as in the model formula.

Model	Oxidation state	λ_i kJ/mole	Distance to Fe (pm)				
			L ₁	L ₂	L ₃	L ₄	L ₅
Fe(Im) ₄ (SCH ₃)	II	21.7	218	218	229	230	234
	III	20.1	213	213	217	217	226
Protein crystal ⁸⁸	III		206	211	220	221	229
Fe(Im) ₄	II	23.1	208	208	208	208	
	III	23.8	200	200	200	200	
Fe(Im) ₃ (SCH ₃)	II	24.0	204	204	205	223	
	III	25.5	211	211	211	228	
Fe(Im) ₂ (SCH ₃) ₂	II	22.9	216	217	232	232	
	III	24.1	207	208	224	224	
Fe(Im)(SCH ₃) ₃	II	21.5	221	236	236	239	
	III	25.3	212	227	228	228	

Table 10. Geometries and inner-sphere reorganisation energies (kJ/mole) for some four- and six-coordinate iron models calculated by the B3LYP method. All complexes were assumed to be high spin. The order of the L₁-L₄ ligands is the same as in the chemical formula for the model. For Fe(H₂O)₆, all six distances are the same so only four are given.

Model	Oxidation state	λ_i kJ/mole	Distance to Fe (pm)			
			L ₁	L ₂	L ₃	L ₄
Fe(H ₂ O) ₆	II	30.5	215	215	215	215
	III	34.9	205	205	205	205
Fe(H ₂ O) ₄	II	35.9	204	204	204	204
	III	29.9	196	196	196	196
Fe(NH ₃) ₄	II	8.0	214	214	214	214
	III	9.5	209	209	209	209

