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Increased C-Telopeptide Cross-linking of Tendon Type I Collagen in Fibromodulin-deficient Mice*

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Keywords: connective tissue; collagen; cross-links; fibromodulin; mass spectrometry; lysyl oxidase; tendon; SLRP; Small Leucine-Rich Proteins

Background: Collagen cross-linking mechanisms must be regulated to obtain tissue-specific collagen fibre properties.

Results: Deficiency in collagen-associated protein fibromodulin leads to excessively cross-linked specific domain of collagen.

Conclusion: Fibromodulin modulates site-specific cross-linking of collagen.

Significance: The first report showing that a collagen-associated protein can modulate cross-linking of specific collagen domains.

ABSTRACT

The controlled assembly of collagen monomers into fibrils, with accompanying intermolecular cross-linking by lysyl oxidase-mediated bonds, is vital to the structural and mechanical integrity of connective tissues. This process is influenced by collagen-associated proteins, including Small Leucine-Rich Proteins (SLRPs), but the regulatory mechanisms are not well understood. Deficiency in fibromodulin, an SLRP, causes

abnormal collagen fibril ultrastructure and decreased mechanical strength in mouse tendons. In this study, fibromodulin deficiency rendered tendon collagen more resistant to non-proteolytic extraction. The collagen had an increased and altered cross-linking pattern at an early stage of fibril formation. Collagen extracts contained a higher proportion of stably cross-linked $\alpha 1(I)$ chains as a result of their C-telopeptide lysines being more completely oxidized to aldehydes. The findings suggest that fibromodulin selectively affects the extent and pattern of lysyl oxidase-mediated collagen cross-linking by sterically hindering access of the enzyme to telopeptides, presumably through binding to the collagen. Such activity implies a broader role for SLRP family members in regulating collagen cross-linking placement and quantity.

Collagen fibrils are essential for the material strength of animal tissues. Their properties vary dependent on the content and quality of covalent

intermolecular cross-links formed as a result of lysyl oxidase activity. The cross-links form progressively as fibrils grow. Lysyl oxidase deaminates specific telopeptide lysine and hydroxylysine residues, creating reactive aldehydes that initiate covalent bonds with lysines or hydroxylysines at specific sites on neighboring collagen monomers (1). Tissue-specific variations in cross-linking chemistry and bond placement are well documented (1-4). We suspect that such variations are influenced by collagen-associated Small Leucine-Rich Proteins - SLRPs¹. Because their expression patterns vary between tissues and at different developmental stages, SLRPs play a role in regulating the diversity in collagen fibril architecture (5). Clues on function come from SLRP knockout mice, in which various tissues – depending on which SLRP is absent – display abnormal collagen fibrils. Thus, decorin-deficient mice have fragile skin, weak tendons, and lower lung airway resistance, all due to altered collagen fibril structure (6-8). Keratocan-deficient mice develop a flattened cornea, which is a phenocopy of human cornea plana caused by *KERA* gene mutations (9,10). Biglycan deficiency causes an osteoporotic phenotype (11), and lumican-deficient mice have opaque corneas and fragile skin (12,13). Lastly, fibromodulin-null mice have thinner and irregularly fused collagen fibrils in tendons (14), and develop osteoarthritis and weak ligaments (15). Absence of one SLRP, therefore, is not compensated by another, which supports the notion of their having specific functions during different stages of collagen fibrillogenesis.

Compound SLRP knockout mice have more severe phenotypes. Lumican- and fibromodulin-deficient mice have joint laxity and very weak tendons with severely disrupted collagen fibrillogenesis (16,17). Decorin- and biglycan-deficient mice corneas also contain severely misshaped collagen fibrils (18). This specificity may depend to some extent on known differences in SLRP binding sites to collagen. Fibromodulin has two collagen-binding domains, one shared by lumican, and the other specific to fibromodulin. They can therefore compete for collagen binding, but fibromodulin has the stronger affinity, suggesting different roles during collagen fibrillogenesis (19-21). Not

all SLRPs can compete with each other, as in the case of decorin being unable to inhibit fibromodulin binding (19,22), but competing with asporin (23).

To date, several reports have shown that SLRPs can act during collagen fibrillogenesis *in vitro* affecting the kinetics and resulting fibril thicknesses (24-28). Such systems are limited in the insights they can provide, however, since they lack cells, procollagen monomers and continuous growth.

Here we studied the biochemical properties of collagen from fibromodulin-deficient *Fmod*^{-/-} mouse tendons. The findings show that in the absence of fibromodulin, lysyl oxidase-mediated cross-linking of tendon type I collagen becomes deregulated.

EXPERIMENTAL PROCEDURES

Sequential extraction and CNBr digestion of collagen - The method has been previously described (29). Briefly, collagen was sequentially extracted from tail tendons of five month-old *Fmod*^{-/-} and wild-type mice for 24 h each in phosphate buffered saline, pH 7.4, then 0.5 M acetic acid, and finally pepsin (1 mg/ml) in 0.5 M acetic acid at 4°C. Extracts were collected as supernatants after centrifugation at 14,000g for 30 min, and pellets were carried over to the next extraction step. Collagen was analyzed by 8% Tris-Glycine reduced SDS-PAGE. All collagen band identities were verified by mass spectrometry. Collagen was quantified by hydroxyproline assay as described (30). For CNBr digests, whole tendons were resuspended in 70% formic acid with 1 mg/ml CNBr, degassed, and digested overnight. Digests were Speedvac-dried for analysis by SDS-PAGE.

P13 antiserum preparation - The P13 rabbit anti-collagen IgG specifically recognizes $\alpha 2(I)$ -collagen chains in immunoblotting². The IgG was purified by protein A-Sepharose, from serum of rabbits immunized with a CNBr-digest of bovine $\alpha 2(I)$ -collagen chains purified by CM-cellulose under denaturing conditions (29). The collagen type I was isolated from calf skin by pepsin cleavage in acetic acid (29).

Immunoblots – Acetic acid-extracted collagen was run on 4-20% Bis-Tris SDS-PAGE and transferred to a nitrocellulose membrane.

The membrane was blocked with 5% milk in TBS, incubated with P13 anti-serum at 1 µg/ml in 0.5% milk in TBST (TBS with 0.1% Tween-20), washed with TBST, incubated with anti-rabbit HRP-conjugated 1:1000 (Dako), washed with TBST, and developed using SuperSignal West Dura ECL kit (Pierce).

Differential scanning calorimetry (DSC)

- DSC measurements were performed with 0.5 mg/ml acetic acid-extracted collagen, with or without pepsin treatment, followed by precipitation by 0.7 M NaCl, redissolving in 0.5 M acetic acid and dialysis against 0.2 M sodium phosphate, 0.5 M glycerol, pH 7.4 (glycerol prevents new collagen fibril formation). The DSC thermograms were recorded in VP-DSC (MicroCal, USA) using temperature gradient from 25 to 70°C, scan rate 0.25°C/min, and medium feedback. Each thermogram was corrected by subtraction of a linear baseline based on a blank buffer sample, and normalized for collagen concentration.

Mass spectrometry - Tail tendon tissue was digested with bacterial collagenase as described (3). Collagenase-generated peptides were separated by reverse-phase HPLC (C8, Brownlee Aquapore RP-300, 4.6 mm x 25 cm) with a linear gradient of acetonitrile:n-propanol (3:1 v/v) in aqueous 0.1% (v/v) trifluoroacetic acid (2). Individual fractions were analyzed by LC-MS. Collagen chains were also directly extracted from tendon by heat denaturation (90 degrees C) in SDS sample buffer and run on SDS-6% PAGE. Collagen chains and CB-peptides resolved by SDS-PAGE were cut out, digested with trypsin and analysed by LC-MS. Peptides were resolved and analyzed by electrospray LC/MS using an LTQ XL ion-trap mass spectrometer (Thermo Scientific) equipped with in-line liquid chromatography using a C4 5µm capillary column (300µm x 150mm; Higgins Analytical RS-15M3-W045) eluted at 4.5µl min. The LC mobile phase consisted of buffer A (0.1% formic acid in MilliQ water) and buffer B (0.1% formic acid in 3:1 acetonitrile:n-propanol v/v). An electrospray ionization source (ESI) introduced the LC sample stream into the mass spectrometer with a spray voltage of 3kV. Linear and cross-linked peptides were identified based on their known elution positions and MSMS fragmentation patterns (31).

Amino acid analysis - Acetic-acid extracted collagen was hydrolyzed in 6 M HCl, lyophilized, redissolved in water with EDTA, and derivatized with phenylisothiocyanate. Amino acids were then applied on a narrow bore HPLC system for separation on a reverse-phase C18 silica column, and detected for phenylthiocarbonyl chromophore at 254 nm.

Trypsin sensitivity assay - Acid-extracted tail tendon collagen from wild-type and *Fmod*^{-/-} mice was dialysed into 50 mM Tris, pH 8.0, and incubated with trypsin at 1:100 (w/w) ratio. Aliquots of 5 µg collagen were removed after 5, 10, 20, 30 and 60 min, and trypsin activity was quenched by SDS-PAGE loading buffer. Samples were run on 8% Tris-Glycine SDS-PAGE and analysed for the amount of proteolysis by Coomassie staining.

Procollagen processing assay - Smooth muscle cells were isolated from aorta of wild-type and *Fmod*^{-/-} mice into α MEM medium using collagenase. After reaching confluency, ascorbate was added at 50 µg/mL and the cells were pulsed for 10 min with 2.5 µCi/ml of ¹⁴C-Proline and chased from 0 to 2 hours. Procollagen was acetone precipitated from cell culture medium, run on SDS-PAGE, and analysed by autoradiography.

RESULTS

Fmod^{-/-} mouse tendon collagen is abnormally cross-linked - To assess the overall cross-linking, collagen was serially extracted from dissected tail tendons. While PBS extracts only trace amounts of collagen, the subsequent 0.5 M acetic acid extraction solubilizes molecules linked by acid-labile aldimine cross-links, and finally pepsin extraction cleaves collagen telopeptides and releases molecules polymerized by acid-stable cross-links (e.g aldol, see Discussion). The results show that only 20% of total collagen could be extracted with PBS plus acetic acid from *Fmod*^{-/-} mice, compared with 70% from wild-type mice. Thus, a large fraction of collagen in *Fmod*^{-/-} mouse tendon required pepsin to extract it, whereas most collagen in wild-type tendon was solubilized by dilute acetic acid (Fig. 1A).

The acetic acid extracts were run on SDS-PAGE to assess the collagen cross-linking pattern. The α -, β - and γ -components were

quantified (Fig. 1B). Compared to wild-type, *Fmod*^{-/-} extracts had a higher ratio of $\alpha 2(I)$ to $\alpha 1(I)$ chains, one-fourth the amount of the $\beta 12$ dimer ($\beta 12$), and three times the amount of the $\gamma 112$ trimer. Analyses of tendons from over thirty mice revealed that the $\alpha 2(I)$ to $\alpha 1(I)$ ratio could vary somewhat between mice; however, the overall trend of less $\beta 12$ and more $\gamma 112$ in *Fmod*^{-/-} extracts was reproducible. This shift could also be seen on an immunoblot of the extracts using an antiserum specific for the $\alpha 2(I)$ chain (Fig. 2A,B). A CNBr digest of whole *Fmod*^{-/-} tendon contained less of the linear non-cross-linked CB6 peptide than wild-type tendon (Fig 2C). Peptide CB6 comes from the C-terminal sequence including the cross-linking C-telopeptide of the $\alpha 1(I)$ chain. These data are consistent with a marked increase in stable cross-linking through the C-terminal telopeptide of the $\alpha 1(I)$ -chain in the *Fmod*^{-/-} tendon collagen.

Fmod^{-/-} tendon type I collagen has increased $\alpha 1(I)$ C-telopeptide aldehyde formation - Type I collagen prepared by bacterial collagenase digestion of whole *Fmod*^{-/-} tendons contained less $\alpha 1(I)$ C-telopeptide with an unmodified lysine, and more lysyl-oxidase-mediated aldehyde plus aldol dimer forms. In Fig. 3 the black peak indicates the C-telopeptide with an unmodified lysine. The grey peaks are the lysine aldehyde-containing linear C-telopeptide (-CHO) and two cleavage variants of the C-telopeptide dimer linked by an allysine aldol cross-link. The present and other unpublished results (Weis and Eyre) indicate that the latter aldol is primarily intermolecular whereas the well-established N-telopeptide aldol dimer formed between N-telopeptides is intramolecular in origin (1). The bacterial collagenase digest also showed that only the lysine form of N-telopeptides and the $\alpha 1(I)$ C-telopeptide were present in the collagen with little or no lysine hydroxylation in either *Fmod*^{-/-} or wild-type tendon type I collagen. Cross-links therefore appeared to be formed exclusively on the lysine aldehyde pathway in both *Fmod*^{-/-} and wild-type tail tendons (2).

Mass spectral analysis of the excised, trypsin-digested $\alpha 1(I)$ chains (Fig. 4) confirmed the higher degree of lysyl oxidase-mediated C-telopeptide aldehyde formation in the *Fmod*^{-/-}

tendon. Trypsin does not cleave after the cross-linking lysine when converted to an aldehyde and produces a 6-residue longer telopeptide cleaving after arginine. This tryptic peptide includes the (GPP)_n sequence from the end of the triple-helix, which is a unique site of prolyl 3-hydroxylation in tendon type I collagen (4) explaining the +16 Dalton mass ladder evident for the molecular ions in Fig. 4. The degree of prolyl 3-hydroxylation was found to be similar in *Fmod*^{-/-} and wild-type collagen.

Differential scanning calorimetry indicates increased cross-linking in Fmod^{-/-} collagen - Differential scanning calorimetry revealed different thermograms during denaturation of acid-extracted wild-type and *Fmod*^{-/-} collagen (Fig. 5A). (Denaturation curve interpretation is described in reference (32)). While both extracts generated a 41.8 °C peak corresponding to monomeric collagen, this peak was smaller for the *Fmod*^{-/-} collagen. Shift in heat capacity from monomer to cross-linked forms (i.e. peaks between 45-52 °C) was prominent in *Fmod*^{-/-} extracts, indicating excessive and dysregulated cross-linking. In contrast, denaturation of pepsin-treated collagen (in which cross-links are in effect removed by pepsin cleaving the telopeptides internal to the cross-linking residues) yielded a single denaturation peak at 41°C in both samples (Fig. 5B). These data support the interpretation that fibromodulin deficiency in tendon results in excessive collagen cross-linking.

Fibromodulin deficiency does not influence procollagen processing, denaturation or the hydroxylation of lysines or prolines - We also considered other mechanisms by which fibromodulin deficiency could impede collagen fibril assembly. The presence of denatured collagen in *Fmod*^{-/-} tendons was ruled out by finding no difference in the time course of degradation in a trypsin sensitivity assay (Fig. 6A). Procollagen processing was unchanged in cultures of *Fmod*^{-/-} vascular smooth muscle cells. (These cells, and not tendon fibroblasts, were used because we've found that the latter lose their fibromodulin expression when cultured in flasks²) (Fig. 6B). Post-translational hydroxylation of prolines and lysines was also unaffected – hydroxyproline comprised 8.2%,

and hydroxylysine 0.7% of total amino acid content in both wild-type and *Fmod*^{-/-} mice.

DISCUSSION

The most prominent tissue phenotype of fibromodulin-null mice is abnormal tendon. The tendons have reduced tensile strength and the animals develop secondary osteoarthritis (14,15). Collagen fibrils in tendons and ligaments appear morphologically abnormal on electron microscopy. Carcinomas induced in *Fmod*^{-/-} mice also have a less dense collagenous stroma (33). Since fibromodulin can inhibit collagen fibril formation at relatively low molar concentrations (26), one of the functions of fibromodulin in a developing connective tissue matrix appears to be in modulating the assembly of collagen monomers into fibrils. The mechanism is not understood. Here we report findings showing that *Fmod*^{-/-} tendons differ from wild-type in having more lysyl oxidase-mediated stable cross-links formed through their type I collagen C-telopeptide domains.

We focused the study on tail tendon collagen. On sequential extraction, acetic acid solubilized only 20% of the total collagen from *Fmod*^{-/-} tendons, compared with 75% of the total collagen from wild-type tendons. This difference suggested altered intermolecular cross-linking. Differences in cross-linking were revealed by SDS-PAGE (Fig. 1). The *Fmod*^{-/-} collagen had a higher ratio of $\alpha 2(I)$ to $\alpha 1(I)$ chains, a deficiency of $\beta 12$ dimer and a higher proportion of $\alpha 1(I)$ chains in stably cross-linked oligomers on SDS-PAGE. Also, a CNBr digest of the *Fmod*^{-/-} tendons contained less $\alpha 1(I)$ CB6 monomer containing an unmodified C-telopeptide lysine (Fig. 2C). This can best be explained by more complete cross-linking of the $\alpha 1(I)$ chains due to a higher proportion having their C-telopeptide lysines converted to aldehydes by lysyl oxidase. This results in more acid-stable aldols being formed between two $\alpha 1(I)$ C-telopeptides, which can account for the observed increase in the $\gamma 112$ trimer (and other acid-stable oligomers involving $\alpha 1(I)$). Therefore the apparent increase in uncross-linked $\alpha 2(I)$ chains is in fact due to a decrease in $\alpha 1(I)$ monomers. The mass spectral analyses of bacterial collagenase-digested whole tendon established that indeed a higher proportion of

total $\alpha 1(I)$ C-telopeptides were present as the lysine aldehyde in *Fmod*^{-/-} versus wild-type collagen (Figs. 3 and 4). The results suggest that fibromodulin binds to the surface molecules of growing fibrils and inhibits lysyl oxidase activity in a significant proportion of their $\alpha 1(I)$ C-telopeptide domains. The increased C-telopeptide lysine oxidation in *Fmod*^{-/-} mice can explain the decrease in acid-soluble collagen and the increase in the pepsin-soluble pool if the resulting aldol cross-links are intermolecular.

We excluded the unlikely possibility that $\alpha 2(I)$ chains had formed triple helical molecules other than [$\alpha 1\alpha 1\alpha 2$] in the null mice because the melting temperature of monomeric collagen molecules was unchanged (Fig. 5). Fibromodulin deficiency also did not appear to affect post-translational modifications of collagen in the triple-helix since the amount of 4-hydroxyproline and hydroxylysine was similar in *Fmod*^{-/-} and wild-type mouse tendon on amino acid analysis.

Mass spectral analysis of trypsin or bacterial collagenase-generated peptides was less informative on the cross-linking status of the N-telopeptides. Aldol intramolecular cross-links between $\alpha 1-\alpha 1$ and $\alpha 1-\alpha 2$ N-telopeptides are known to be the main source of the β dimers seen on electrophoresis of skin and tendon type I collagen. SDS-PAGE indicated normal amounts of $\beta 11$ and evidence that $\beta 12$ was shifted to γ and higher cross-linked oligomers in *Fmod*^{-/-} collagen (Figs. 1 and 2). The latter shift is explained by excessive C-telopeptide cross-linking, which we confirmed by mass spectral analyses. In interpreting the tissue extraction and gel electrophoresis patterns, note that aldol cross-links are stable to low pH and denaturation, whereas aldimine (Schiff's base) cross-links formed between lysine aldehyde and hydroxylysine or lysine are not. This can explain the differences in chain and CNBr-peptide profiles seen on electrophoresis (Figs. 1 and 2).

The collagen assembled without fibromodulin also had altered physical properties, as revealed by differential scanning calorimetry. Thermal denaturation of collagen includes an initial melting of less cross-linked triple-helical monomers at lower temperature, followed by denaturation of cross-linked collagen molecules; whereas any inherently

denatured collagen cannot refold at physiological temperatures and so does not contribute to a thermogram (32,34). In addition, the denaturation peak pattern of a DSC thermogram correlates with the degree of intermolecular cross-linking (35), supported by the similar melting points and enthalpies of the pepsin-treated (i.e. non-cross-linked) collagens. The altered physical properties of collagen in *Fmod*^{-/-} tendons may regulate cellular behaviour, including cell migration, survival, and matrix protein synthesis (36,37).

In summary, we propose that fibromodulin modifies covalent cross-linking of type I collagen during fibril assembly by modulating lysyl oxidase action on the C-telopeptide lysines. It is clear that other SLRPs such as lumican (which is elevated in content (14)) do not replace fibromodulin in this activity. It implies that individual SLRPs may have evolved distinct activities affecting cross-linking during fibril growth and maturation. Absence of one constraining protein (in this case fibromodulin) may allow premature assembly of intermediates and their cross-linking through $\alpha 1(I)$ C-telopeptide aldehydes, which results in deregulated growth of fibrils and of the tendon. Collectively, our findings suggest that collagen, despite its self-assembling properties, has evolved to be finely regulated by many fibril-associating proteins along its surface that tailor it for specialized tissue functions.

It is also intriguing that increased lysyl oxidase-mediated cross-linking leads to mechanically weaker tendons. This finding may seem counter-intuitive but it emphasizes the importance of understanding not just the total number and stability of covalent cross-links but their placement, for example whether they are solely intrafibrillar or can also form interfibrillar bonds as fibrils grow and interact. One possible explanation for decreased tissue tensile strength in *Fmod*^{-/-} is loss of latent fibril-surface C-telopeptide lysines to allow fibril-to-fibril bonding during tissue growth. Clearly, collagen-binding members of the SLRP family have the potential to play a major role in such processes. To what degree SLRPs other than fibromodulin have evolved to directly modulate access to lysyl oxidase(s) during collagen fibrillogenesis by related mechanisms deserves attention.

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FOOTNOTES

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¹The abbreviations used are: SLRP, small leucine-rich repeat protein; LRR, leucine-rich repeat; PBS, phosphate-buffered saline; HAc, acetic acid (CH₃COOH); DMEM, Dulbecco's modified Eagle's medium.

²Unpublished observation.

FIGURE LEGENDS

FIGURE 1. Altered extractability and electrophoretic pattern of *Fmod*^{-/-} tail tendon collagen. A. Collagen yields on sequential extraction (PBS, 0.5M acetic acid, and pepsin) of tail tendons from *Fmod*^{-/-} and wild-type mice. *Filled bars*, wild type mice (n=5), *open bars*, *Fmod*^{-/-} mice (n=5). B. 8% SDS-PAGE of acetic acid-extracted collagen from wild type (n=3) and *Fmod*^{-/-} (n=3) mice stained with Coomassie Blue. Relative amounts of α -, β -, and γ -components of collagen were quantified with ImageJ software (NIH), and are listed as percentage of total collagen \pm SD.

FIGURE 2. Enhanced stable cross-linking of α 1(I) chains. Acid-extracted collagen was run on 4-20% SDS-PAGE gradient gels and either stained with Coomassie blue (A) or transferred to nitrocellulose membrane and immunoblotted with an antiserum specific to the collagen α 2(I) chain (B). Whole tendons were digested with CNBr and the resulting collagen fragments were resolved on 12.5% SDS-PAGE staining with Coomassie Blue (C). The indicated α 1(I)CB6-containing monomer and dimer bands were identified by in-gel trypsin digestion and mass spectroscopy. The prominent α 1(I)CB6 monomer from wild type tendon gave a C-terminal tryptic peptide cleaved after its C-terminal unmodified lysine residue. From *Fmod*^{-/-} tendon this component was largely missing and the

retarded, broad $\alpha 1(I)CB6$ monomer band gave a C-terminal tryptic peptide cleaved at arginine, six residues more C-terminal. Sequences of the latter same two tryptic peptides are shown later in Fig 4.

FIGURE 3. Mass spectrometric identification of the collagen type I C-telopeptide cross-linking domain. Bacterial collagenase-digested tendon collagen peptides from *Fmod*^{-/-} and wild-type mice were resolved by HPLC (A) and identified by tandem mass spectrometry from their MSMS fragments (B and C). Consistently the ratio of unmodified lysine-containing C-telopeptide to aldehyde- plus aldol-containing C-telopeptides was about 2:1 from wild-type compared with 1:2 from mutant tendon. The small peaks eluting from 25-27 min in wild type tendon are amino- and aldehyde- versions of the C-telopeptide missing one or two Tyr residues from the C-terminus. The structure of the aldol cross-link from *Fmod*^{-/-} tendon was identified by MS (B) and MSMS fragmentation of the 801.25+ion as shown (C). M4+-b4-b2 is a fragment of the parent ion with 4 charges that lost b2 from one arm and b4 from the other arm of the cross-linked structure.

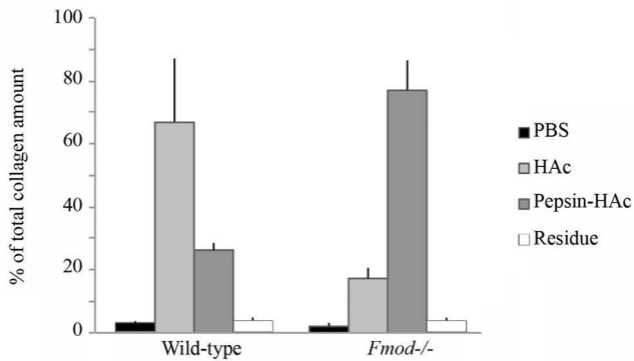
FIGURE 4. Mass spectral analysis of the C-terminal domain of collagen $\alpha 1(I)$ chains extracted from *Fmod*^{-/-} and wild-type tendons. Heat denatured extract of tendon collagen was run on 6% SDS-PAGE and excised $\alpha 1(I)$ chains were digested in-gel with trypsin and the peptides profiled by LC-tandem mass spectrometry (A). The C-terminal tryptic peptide domain (underlined in total ion current (TIC) profile), identified by MSMS fragmentation (D), showed more of the longer aldehyde-containing peptide from *Fmod*^{-/-} tissue (C) than from wild-type (B). Note that the (GPP)₅ C-terminus of the triple-helix is equally modified with up to four 3-hydroxyproline residues per chain in both *Fmod*^{-/-} and wild-type tendon collagen, a post-translational modification that is peculiar to tendon (4).

FIGURE 5. Differential scanning calorimetry of collagen. A. Acid-extracted collagen in phosphate-glycerol pH 7.4 buffer was melted in a differential scanning calorimeter heating at 0.25°C/min. Solid line is wild-type collagen, dotted line is *Fmod*^{-/-} collagen. B. As in A, but prior to analysis the acid-extracted collagen was pepsin-treated, precipitated with salt, resuspended in acetic acid, and dialyzed against phosphate-glycerol buffer.

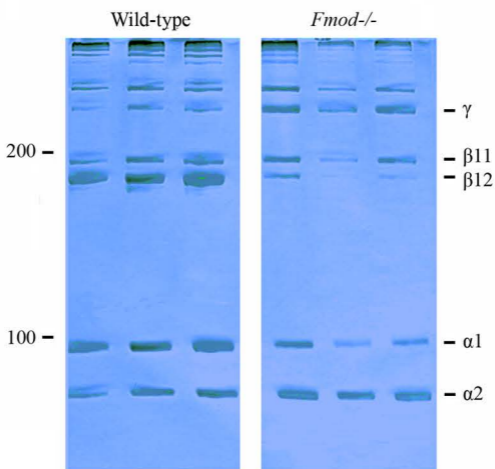
FIGURE 6. A. Collagen trypsin sensitivity assay. Acid-extracted collagen was neutralized and incubated with trypsin (1:100 w/w). Aliquots (5 μ g) of collagen were removed at indicated time points and trypsin activity was quenched by adding SDS loading buffer. Amount of proteolysis was assessed by SDS-PAGE staining with Coomassie blue. B. Procollagen processing assay. Primary cultures of smooth muscle cells from *Fmod*^{-/-} and wild type aortas were pulsed with 2.5 μ Ci/ml of ¹⁴C-proline and chased from 0 to 2 hours. Procollagen was acetone-precipitated from the medium, run on SDS-PAGE, and analysed by autoradiography.

Figure 1

A



B



Distribution of collagen components (% of total collagen amount)		
	Wild-type	<i>Fmod</i> ^{-/-}
γ	8 \pm 1	23 \pm 2
β 11	14 \pm 1	15 \pm 2
β 12	31 \pm 2	7 \pm 2
α 1	28 \pm 3	19 \pm 0
α 2	19 \pm 2	36 \pm 6

Figure 2

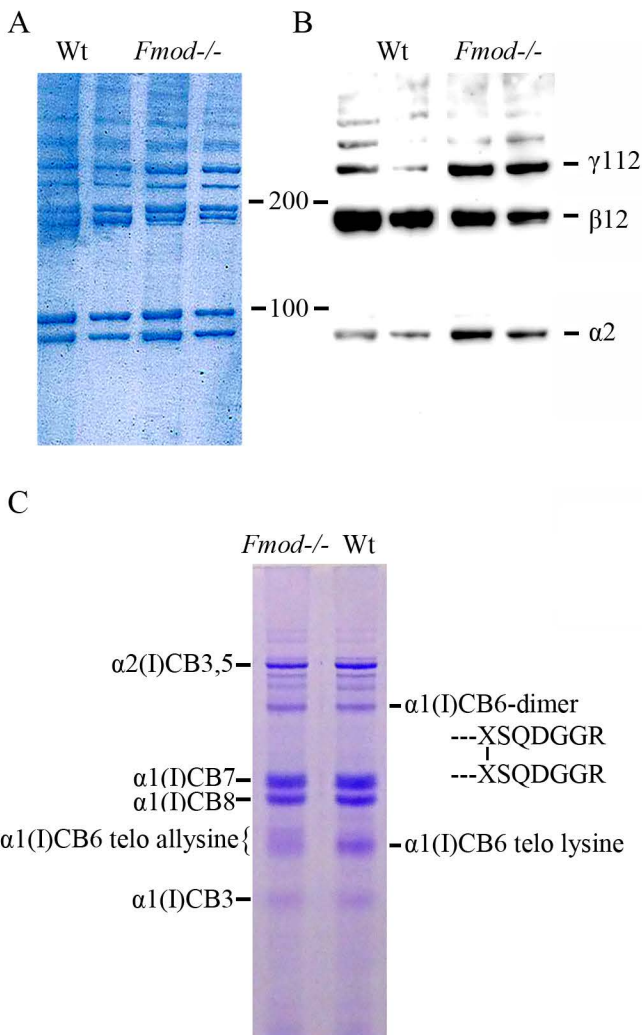


Figure 3

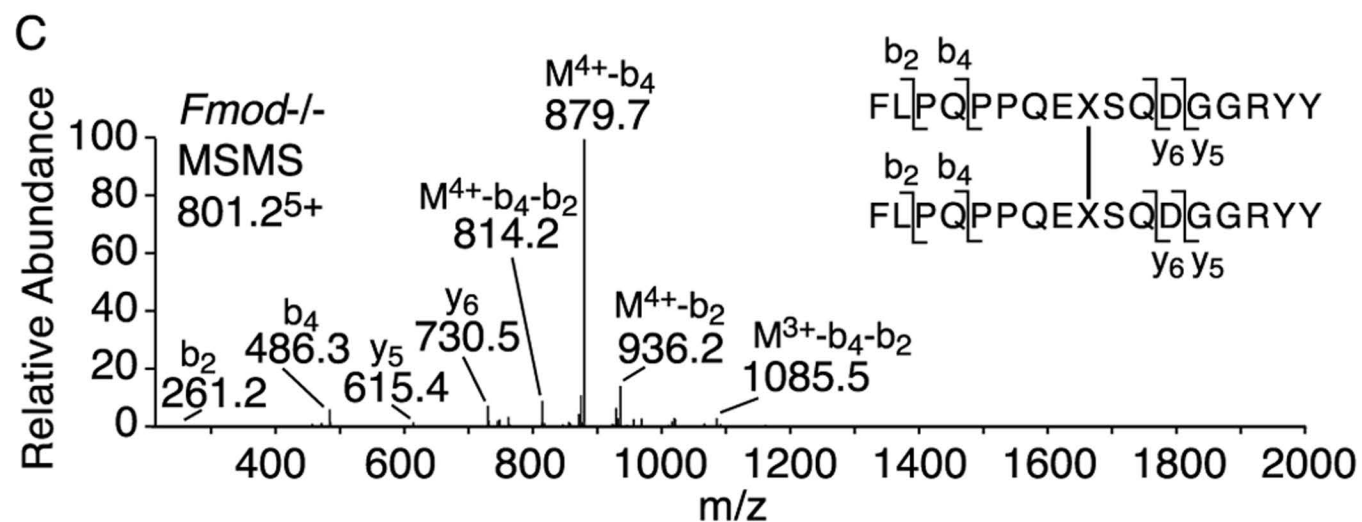
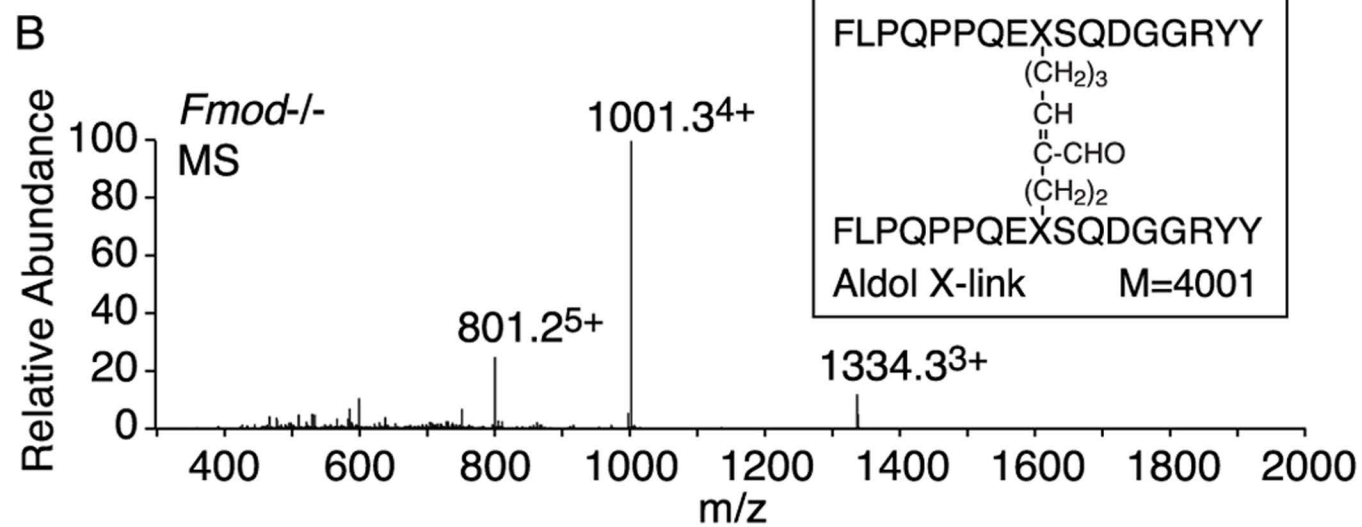
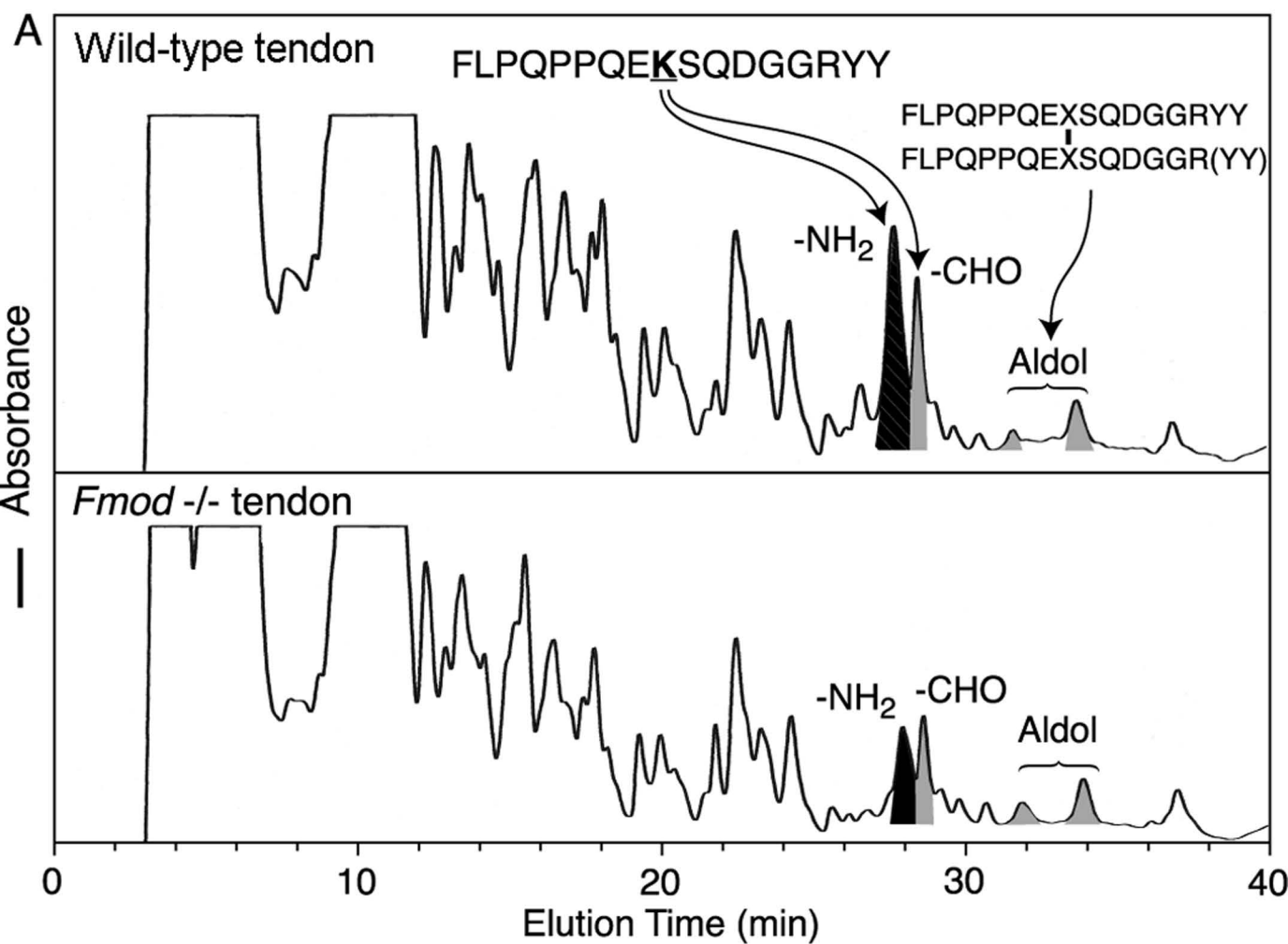


Figure 4

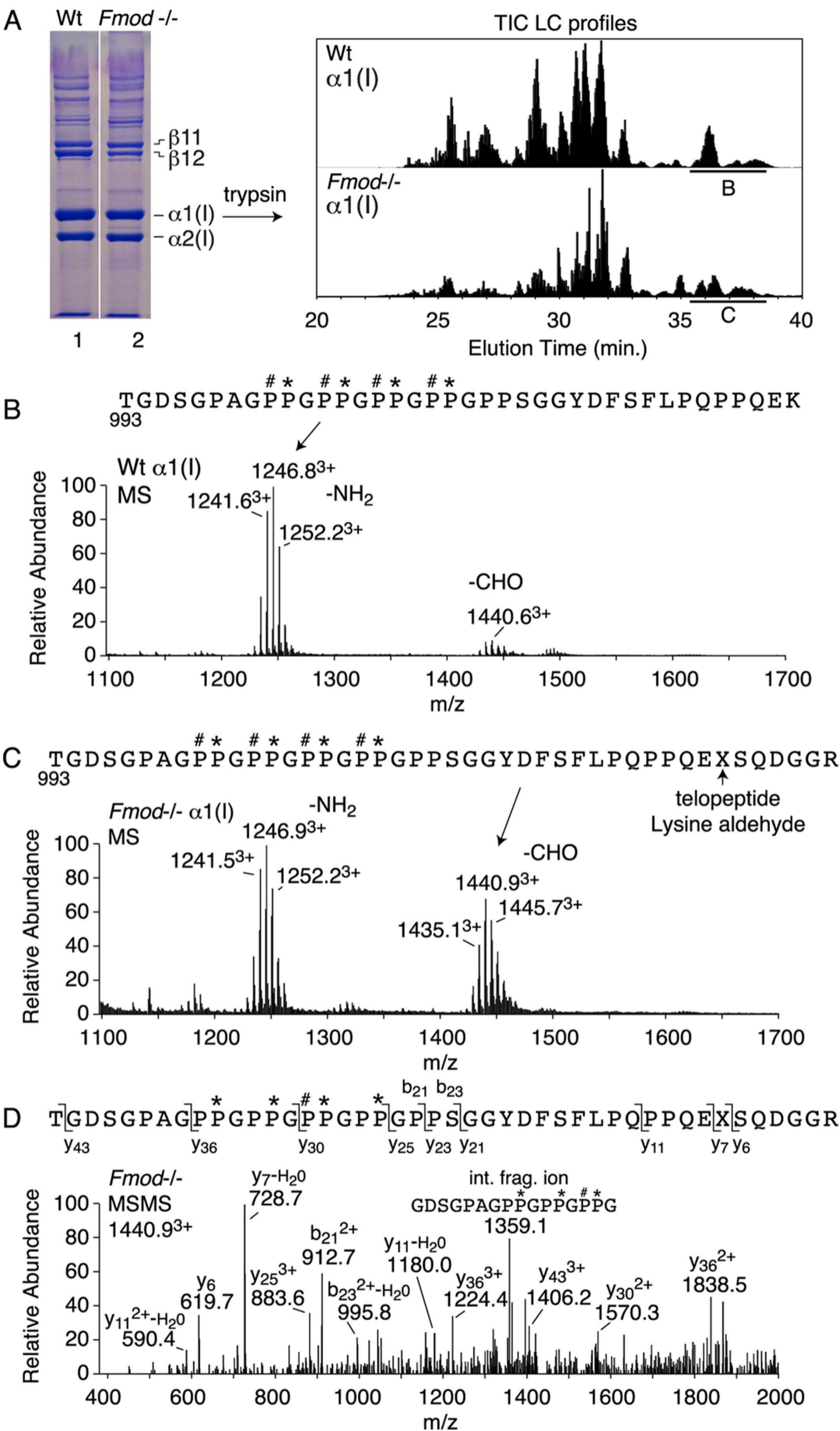
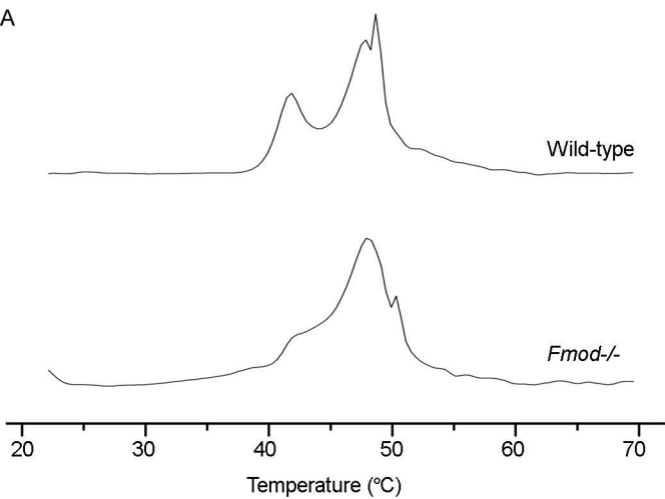


Figure 5

A



B

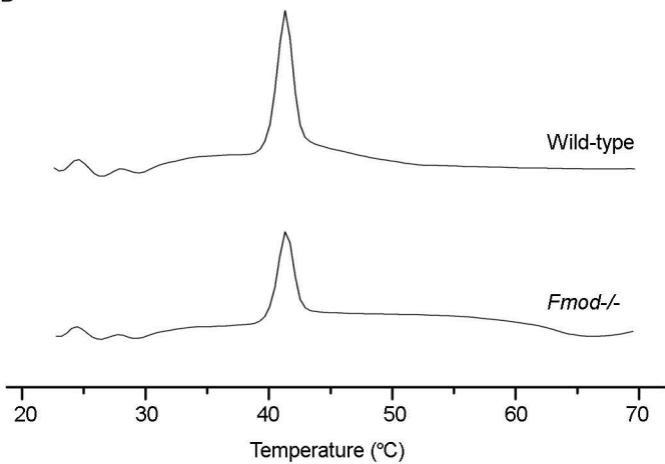


Figure 6

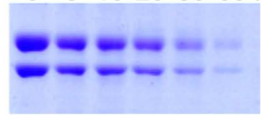
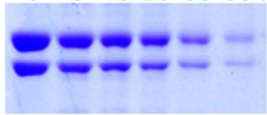
A

Wild-type

Fmod^{-/-}

0 5 10 20 30 60 min

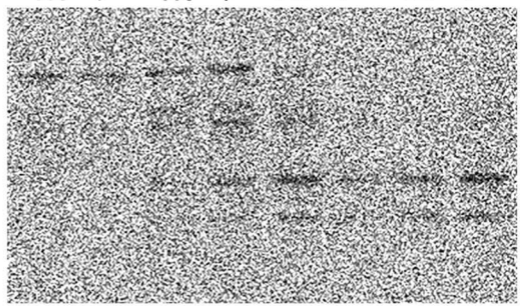
0 5 10 20 30 60 min



B

Wt -/- Wt -/- Wt -/- Wt -/-

Procollagen
pC-collagen
pN-collagen
alpha1
alpha2



0 h 1/2 h 1 h 2 h