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# Tetraisopropyldisiloxane-1,3-diyl as a Versatile Protecting Group for Pentopyranosides

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## Abstract

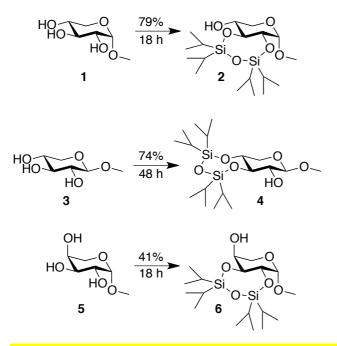
The protecting group tetraisopropyldisiloxane-1,3-yl has been investigated for simultaneous protection of two hydroxyls on pentopyranosides. Methyl  $\alpha$ -D-xylopyranoside is protected in excellent regioselectivity and high yield to form the 2,3-protected xylopyranoside whereas methyl  $\beta$ -D-xylopyranoside gives the 3,4-protected product also with excellent regioselectivity.

Pentopyranosides have recently received attention in medicinal chemistry and for example, simple xylosides have shown to be interesting in cancer therapeutics.<sup>1</sup> Selective protection of xylose is complicated since all three hydroxyls are secondary and equatorial.<sup>2</sup> Several methods have been evaluated for regioselective protection, which can be achieved by stoichiometric benzoylation<sup>3</sup>, benzylation<sup>4</sup> and tosylation<sup>5</sup>. Other methods for regioselective synthesis include phenylborate esters<sup>6</sup>, isopropylidene acetals<sup>4,7,8</sup>, butane-2,3-diacetals<sup>9,10</sup>, cyclohexylidene acetals<sup>4,11,12</sup>, tin acetals<sup>13</sup> and enzymatic deacetylation<sup>14</sup>. However, most of these methods give low selectivity, include toxic reagents or troublesome purifications. To find a versatile method for selective protection of pentopyranosides we decided to introduce tetraisopropyldisiloxane-1,3-diyl (TiPDS) to protect two hydroxyls simultaneously.

TiPDS is a cyclic protecting group that was introduced by Markiewicz in 1979 for protection of ribonucleosides.<sup>15,16</sup> The method gives a clean conversion to the 3',5' protected ribonucleoside, since the primary HO-5' reacts faster followed by the formation of the 8-membered ring. The method is still one of the preferred methods in nucleoside chemistry for modification on HO-2'.<sup>17-19</sup> One year later van Boom and co-workers introduced the TiPDS protection to hexopyranosides, showing that it simultaneously protected HO-4 and HO-6 and concluding that the protecting group rearranges by treatment with acid in DMF to generate the 3,4-protected glucoside.<sup>20</sup>

To investigate the use of TiPDS for pentopyranosides, methyl  $\alpha$ -D-xylopyranoside (1) was dissolved in pyridine and 1,3-dichloro-1,1,3,3-tetraisopropyldisiloxane (TiPDSCl<sub>2</sub>) was added and the reaction was followed by TLC. After 18 h, methanol was added to quench the excess of TiPDSCl<sub>2</sub> and the mixture was concentrated and chromatographed to give the 2,3-protected methyl  $\alpha$ -D-xylopyranoside (2) in 79% yield (Scheme 1).

Methyl  $\beta$ -D-xylopyranoside (**3**) was subjected to the same reaction conditions and after 18 h reaction time the 3,4-protected methyl  $\beta$ -D-xylopyranoside was isolated in 59% yield. However, when the reaction time was increased to 48 h, **4** was isolated in 74% yield, indicating that the  $\beta$ -anomer reacts at a lower rate. In nucleoside chemistry the reaction proceeds at a higher rate if pyridine is exchanged for DMF using imidazole as base.<sup>16</sup> When methyl  $\beta$ -D-xylopyranoside was reacted under these conditions the starting material was consumed in just a couple of hours. However, the isolated yield of the desired product did not increase compared to the reaction in pyridine (59%) and the higher reaction rate also diminished the regioselectivity for the reaction (Scheme 1).



Scheme 1: TiPDSCl<sub>2</sub> protection of methyl D-glycosides. Reaction conditions: TiPDSCl<sub>2</sub> 1.1 eq. in pyridine 0.1 M.

The difference in regioselectivity between the  $\alpha$ - and  $\beta$ -anomer was expected based on previous literature. The reactivity for the secondary hydroxyls in methyl  $\alpha$ -Dglucopyranoside towards benzoyl chloride was investigated by Williams *et. al.* and they concluded that the reactivity is HO-2 > HO-4 > HO-3.<sup>23</sup> The higher reactivity of HO-2 was reasoned to be due to activation by the anomeric substituent, probably through a hydrogen bond to the anomeric oxygen. In addition gauche effects between HO-2 and HO-3 as well as steric effects cause HO-4 to be more reactive than HO-3. Sivakumaran *et. al.* investigated benzoylation on benzyl  $\alpha$ -D-xylopyranosides and concluded that the order of reactivity was the same as for methyl  $\alpha$ -Dglucopyranoside. <sup>3</sup> The reactivity order for methyl  $\beta$ -D-xylopyranoside has been previously established to be HO-4 > HO-3 > HO-2.<sup>24</sup> The results from this study support these observations since methyl  $\alpha$ -D-xylopyranoside forms the 2,3-cyclic product and methyl  $\beta$ -D-xylopyranoside forms the 3,4-cyclic product. See Table 1 for comparison of different cyclic protection groups on D-xylopyranosides.

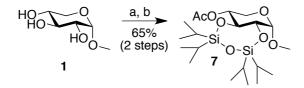
To investigate the difference in reaction rate between the  $\alpha$ - and  $\beta$ -anomer, the consumption of the starting material was followed by NMR. The reaction was hence run over 10 h in an NMR-tube in pyridine-d<sub>5</sub> with 1 equivalent of toluene as internal

standard. The progress of the reaction was monitored by the disappearance of H-1. As expected, methyl  $\alpha$ -D-xylopyranoside was consumed at a higher rate, in comparison to methyl  $\beta$ -D-xylopyranoside (Figure 1).

Figure 1: The consumption of methyl  $\alpha$ -D-xylopyranoside (circles,  $\bullet$ ) and methyl  $\beta$ -D-xylopyranoside (squares,  $\blacksquare$ ) as a function over time. The disappearance of H-1 is followed by NMR.

The reducing form of xylose was also subjected to the reaction conditions. Unfortunately multiple products were formed and xylose is not suitable for this method.

Next, methyl  $\beta$ -L-arabinopyranoside (**5**) was also reacted under the same conditions but did not proceed as cleanly and several products were observed on TLC. However, the major product was the 2,3-protected methyl  $\beta$ -L-arabinopyranoside (**6**) that was isolated in 41% yield (Scheme 1). The reactivity of the hydroxyls of methyl  $\beta$ -Larabinopyranoside has been suggested to be HO-2, HO-3 > HO-4, where the relative reactivity of HO-2, HO-3 is uncertain, and this reactivity is also supported by the silylation experiments in this study.<sup>3,13,25</sup>



Scheme 2: The selective acetylation of HO-4: Reaction conditions: a) TiPDSCl<sub>2</sub> 1.1 eq. in pyridine 0.1 M. 18 h b) Ac<sub>2</sub>O/Pyridine 4:5 v:v 18h.

To confirm the usability of this protecting group, methyl  $\alpha$ -D-xylopyranoside was protected with TiPDSCl<sub>2</sub> and with a short work-up, without column chromatography. The crude was treated with acetic anhydride in pyridine to give methyl 2,3-*O*-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)- $\alpha$ -D-xylopyranoside (7) in 65% yield over two steps (Scheme 2).

#### Table 1: Comparison of yield and selectivity for cyclic protective groups on D-

Entry	Anomeric	Protective group	2,3-	3,4-	Ref.
	configuration		protected	protected	, , , , , , , , , , , , , , , , , , ,
1	α-OMe	TiPDS	79%	_	This work
2	α-OMe	Isopropylidene	39%	13%	4
		acetal			
3	α-OAll	Isopropylidene	70%	-	8
		acetal			
4	α-OMe	Cyclohexylidene	63%	13%	4
		acetal			
5	α-OBn	Cyclohexylidene	42%	11%	12
		acetal			
6	β-OMe	TiPDS	-	74%	This work
7	β-OMe	Isopropylidene	72%	-	7
		acetal			
8	β-OAll	Isopropylidene	77%	6%	21
		acetal			
9	β-OBn	Isopropylidene	78%	14%	22
		acetal			
10	β-OAll	Butane-2,3-	47%	47%	9
		diacetal			

#### xylopyranosides.

To summarize, we have developed a new methodology for regioselective protection of xylopyranosides to simultaneously protect HO-2 and HO-3 on methyl  $\alpha$ -D-xylopyranosides and methyl  $\beta$ -L-arabinopyranosides as well as protection of HO-3 and HO-4 on methyl  $\beta$ -D-xylopyranosides by using TiPDSCl<sub>2</sub>. The reaction proceeds cleanly and in high yield for the xylopyranosides although a lower yield was observed for the arabinopyranoside.

## 1. Experimental

#### **1.1 General experimental details**

NMR spectra were recorded with a Bruker Avance II 400 MHz and Bruker Avance 500 MHz. <sup>1</sup>H-NMR spectra were assigned using 2D-methods (COSY, HMQC). Chemical shifts are given in ppm downfield from the signal for  $Me_4Si$ , with reference to residual  $C_6D_5H$ . Reactions were monitored by TLC using alumina plates coated with silica gel and visualized using either UV light or by charring with *para*-anisaldehyde. Preparative chromatography was performed with silica gel (35-70 µm,

60 Å). DMF was distilled prior to use; pyridine (extra dry) and all other reagents were used as supplied from manufacturer.

#### 1.2 General experimental for the 1,1,3,3-tetraisopropyldisiloxane protections.

Methyl glycoside (56-116 mg, 0.34-0.71 mmol) was dissolved in pyridine (0.1 M) and stirred at r.t. under N<sub>2</sub>. TiPDSCl<sub>2</sub> (1.1 eq.) was added dropwise during 5-10 min. Upon completion the reaction was quenched by addition of MeOH (1-2 mL) and the mixture was concentration to dryness by co-evaporation with toluene. Purified by column chromatography (SiO<sub>2</sub> heptane/EtOAc 6:1) to give the product as an amorphous white solid.

#### 1.3 Methyl 2,3-O-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)-α-D-xylopyranoside

(2). Yield 79%.  $[\alpha]_D^{20}$  59.6 (*c* 0.8, C<sub>6</sub>H<sub>6</sub>). <sup>1</sup>H-NMR (C<sub>6</sub>D<sub>6</sub>):  $\delta$  4.60 (d, 1 H, *J* 3.6 Hz, H-1), 4.14 (t, 1 H, *J* 8.3 Hz, H-3), 3.76 (dd, 1 H, *J* 9.1, 3.6 Hz, H-2), 3.68-3.70 (m, 2 H, H-5, H-5'), 3.61 (dt, 1 H, *J* 8.1, 2.7 Hz, H-4), 3.12 (s, 3 H, OMe), 1.97 (d, 1 H, *J* 2.8 Hz, OH-2), 1.11-1.21 (m, 28 H, Silyl-H). <sup>13</sup>C-NMR (C<sub>6</sub>D<sub>6</sub>):  $\delta$  100.9, 77.9, 75.6, 71.6, 61.2, 55.2, 17.8, 17.73, 17.67, 17.61, 17.57, 17.54, 17.51, 17.48, 13.3, 13.2, 12.9, 12.6. HRMS calcd for C<sub>18</sub>H<sub>38</sub>O<sub>6</sub>Si<sub>2</sub>Na (M+Na): 429.2105, found: 429.2136.

## 1.4 Methyl 3,4-O-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)-β-D-xylopyranoside

(4). Yield 59% (18 h reaction time), 74% (48 h reaction time).  $[α]_D^{20}$  -5.5 (*c* 0.8, C<sub>6</sub>H<sub>6</sub>). <sup>1</sup>H-NMR (C<sub>6</sub>D<sub>6</sub>): δ 3.98 (d, 1 H, *J* 7.6 Hz, H-1), 3.92 (dd, 1 H, *J* 11.5, 5.6 Hz, H-5), 3.79-3.85 (m, 1 H, H-4), 3.69 (t, 1 H, *J* 8.8 Hz, H-3), 3.55 (ddd, 1 H, *J* 8.8, 7.7, 2.4 Hz, H-2), 3.29 (s, 3 H, OMe), 3.12 (dd, 1 H, *J* 11.5, 10.0 Hz, H-5'), 2.17 (d, 1 H, *J* 2.4 Hz, OH-2), 0.97-1.26 (m, 28 H, Silyl-H). <sup>13</sup>C-NMR (C<sub>6</sub>D<sub>6</sub>): δ 104.9, 80.3, 74.6, 73.4, 66.2, 56.6, 17.7, 17.63, 17.60, 17.56, 17.5, 17.4, 13.4, 13.3, 12.60, 12.57. HRMS calcd for C<sub>18</sub>H<sub>38</sub>O<sub>6</sub>Si<sub>2</sub>Na (M+Na): 429.2105, found: 429.2107. *DMF/Imidazole method:* Methyl β-D-xylopyranoside (57 mg, 0.35 mmol) was dissolved in DMF (3.5 mL) and stirred at r.t. under N<sub>2</sub> and imidazole (110 mg, 1.62 mmol) was added. TiPDSCl<sub>2</sub> (0.13 mL, 0.40 mmol) was added dropwise during 10 min. After 7 h the reaction was quenched by addition of MeOH (1 mL) and concentration to dryness by co-evaporation with toluene. Purified by column chromatography (SiO<sub>2</sub> heptane/EtOAc 4:1) to give **4** (84 mg, 59%) as an amorphous white solid.

#### 1.5 Methyl 2,3-O-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)-β-L-

arabinopyranoside (6). Yield 41%.  $[\alpha]_D^{20}$  88.0 (*c* 0.6, C<sub>6</sub>H<sub>6</sub>). <sup>1</sup>H-NMR (C<sub>6</sub>D<sub>6</sub>):  $\delta$  4.80 (d, 1 H, *J* 3.5 Hz, H-1), 4.28 (dd, 1 H, *J* 9.2, 3.5 Hz, H-2), 4.17 (dd, 1 H, *J* 9.2, 3.8 Hz, H-3), 3.82 (dd, 1 H, *J* 12.4, 1.6 Hz, H-5), 3.75-3.76 (m, 1 H, H-4), 3.59 (bd, 1 H, *J* 12.4 Hz, H-5'), 3.15 (s, 3 H, OMe), 2.72 (d, 1 H, *J* 1.7 Hz, OH-4), 1.00-1.19 (m, 28 H, Silyl-H). <sup>13</sup>C-NMR (C<sub>6</sub>D<sub>6</sub>):  $\delta$  101.2, 73.2, 72.6, 70.3, 61.6, 55.3, 17.71, 17.70, 17.64, 17.61, 17.57, 17.5, 17.4, 13.29, 13.28, 12.9, 12.5. HRMS calcd for C<sub>18</sub>H<sub>38</sub>O<sub>6</sub>Si<sub>2</sub>Na (M+Na): 429.2105, found: 429.2119.

#### 1.6 Methyl 4-O-acetyl-2,3-O-(1,1,3,3-tetraisopropyldisiloxane-1,3-diyl)-α-D-

xylopyranoside (7). Methyl  $\alpha$ -D-xylopyranoside (60 mg, 0.33 mmol) was dissolved in pyridine (3.5 mL) and stirred at r.t. under N<sub>2</sub>. TiPDSCl<sub>2</sub> (0.12 mL, 0.37 mmol) was added dropwise during 5 min. After 18 h the reaction was quenched by addition of MeOH (1 mL) and concentration to dryness by co-evaporation with toluene. The residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub> and washed twice with brine. The water phase was extracted twice with CH<sub>2</sub>Cl<sub>2</sub> and the combined organic phase was dried with MgSO<sub>4</sub> and concentrated to give 2. Compound 2 was dissolved in pyridine (2.5 mL) and Ac<sub>2</sub>O (2.0 mL) was added and the mixture was stirred at r.t. After 18 h the mixture was concentrated to dryness and purified by column chromatography (SiO<sub>2</sub>, heptane/EtOAc 10:1) to give 7 (97 mg, 65%, 2 steps) as an amorphous white solid.  $[\alpha]_{D}^{20}$  81.9 (c 0.8, C<sub>6</sub>H<sub>6</sub>). <sup>1</sup>H-NMR (C<sub>6</sub>D<sub>6</sub>):  $\delta$  5.23 (ddd, 1 H, J 15.0, 9.0, 6.1 Hz, H-4), 4.64 (d, 1 H, J 3.6 Hz, H-1), 4.27 (t, 1 H, J 9.0 Hz, H-3), 3.74-3.78 (m, 2 H, H-2, H-5), 3.55 (t, 1 H, J 10.8 Hz, H-5'), 3.10 (s, 3 H, OMe), 1.73 (s, 3 H, OAc), 1.03-1.21 (m, 28 H, Silyl-H). <sup>13</sup>C-NMR (C<sub>6</sub>D<sub>6</sub>): δ 169.2, 100.5, 75.8, 74.5, 71.8, 58.6, 55.3, 20.3, 17.7, 17.63, 17.61, 17.58, 17.53, 17.48, 17.4, 17.3, 13.2, 12.8, 12.7. HRMS calcd for  $C_{20}H_{40}O_7Si_2Na$  (M+Na): 471.2210, found: 471.2216.

#### **1.7** Kinetics study, Methyl α-D-xylopyranoside (1).

Methyl  $\alpha$ -D-xylopyranoside (1) (13 mg, 0.077 mmol) was dissolved in pyridine-d<sub>5</sub> (0.6 mL) and toluene (0.008 mL, 0.075 mmol) was added as an internal standard in an NMR tube. TiPDSCl<sub>2</sub> (0.028 mL, 0.086 mmol) was added. NMR spectra were taken immediately after addition of TiPDSCl<sub>2</sub> (t=0) and once every 30 min for 10 h.

# 1.8 Kinetics study, Methyl $\beta$ -D-xylopyranoside (3).

Methyl  $\beta$ -D-xylopyranoside (**3**) (12 mg, 0.071 mmol) was dissolved in pyridine-d<sub>5</sub> (0.6 mL) and toluene (0.008 mL, 0.075 mmol) was added as an internal standard in an NMR tube. TiPDSCl<sub>2</sub> (0.026 mL, 0.080 mmol) was added. NMR spectra were taken immediately after addition of TiPDSCl<sub>2</sub> (t=0) and once every 60 min for 10 h.

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# Supplementary data

Supplementary data associated with this article can be found, in the online version, at

# References

- 1. Nilsson, U.; Johnsson, R.; Fransson, L.-Å.; Ellervik, U.; Mani, K. *Cancer Res.* **2010**, *70*, 3771-3779.
- 2. Siegbahn, A.; Aili, U.; Ochocinska, A.; Olofsson, M.; Rönnols, J.; Mani, K.; Widmalm, G.; Ellervik, U. *Bioorg. Med. Chem.* **2011**, *19*, 4114-4126.
- 3. Sivakumaran, T.; Jones, J. K. N. *Can. J. Chem.* **1967**, *45*, 2493-2500.
- 4. Morishima, N.; Koto, S.; Kusuhara, C.; Zen, S. *Bull. Chem. Soc. Jpn.* **1982**, *55*, 631-632.
- 5. Buchanan, J. G.; Fletcher, R. J. Chem. Soc., C **1966**, 1926-1931.
- 6. Ferrier, R. J.; Prasad, D.; Rudowski, A.; Sangster, I. *J. Chem. Soc.* **1964**, 3330-3334.
- 7. Naleway, J. J.; Raetz, C. R. H.; Anderson, L. *Carbohydr. Res.* **1988**, *179*, 199-209.
- 8. Goodby, J. W.; Haley, J. A.; Watson, M. J.; Mackenzie, G.; Kelly, S. M.; Letellier, P.; Gode, P.; Goethals, G.; Ronco, G.; Harmouch, B.; Martin, P.; Villa, P. *Liq. Cryst.* **1997**, *22*, 497-508.
- 9. Jenkins, D. J.; Potter, B. V. L. J. Chem. Soc., Perkin Trans. 1 1998, 41-50.
- 10. Oscarson, S.; Svahnberg, P. J. Chem. Soc., Perkin Trans. 1 2001, 873-879.
- 11. Bissett, F. H.; Evans, M. E.; Parrish, F. W. *Carbohydr. Res.* **1967**, *5*, 184-193.
- 12. Koto, S.; Morishima, N.; Takenaka, K.; Uchida, C.; Zen, S. *Bull. Chem. Soc. Jpn.* **1985,** *58*, 1464-1468.
- 13. Tsuda, Y.; Haque, M. E.; Yoshimoto, K. *Chem. Pharm. Bull.* **1983**, *31*, 1612-1624.
- 14. Marek, M.; Medonos, I.; Kefurt, K.; Jary, J. *Biocatalysis* **1989**, *2*, 235-238.
- 15. Markiewicz, W. T. J. Chem. Soc. (S) **1979**, 24-25.
- 16. Markiewicz, W. T. J. Chem. Soc. (M) **1979**, 178-194.
- 17. Johnsson, R.; Lackey, J. G.; Bogojeski, J. J.; Damha, M. J. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 3721-3725.

- 18. Dellinger, D. J.; Timár, Z.; Myerson, J.; Sierzchala, A. B.; Turner, J.; Ferreira, F.; Kupihár, Z.; Dellinger, G.; Hill, K. W.; Powell, J. A.; Sampson, J. R.; Caruthers, M. H. *J. Am. Chem. Soc.* **2011**, *133*, 11540-11556.
- 19. Hermann, C.; Kvassiouk, E.; Pfleiderer, W. *Helv. Chim. Acta* **2011**, *94*, 362-370.
- 20. Verdegaal, C. H. M.; Jansse, P. L.; de Rooij, J. F. M.; van Boom, J. H. *Tetrahedron Lett.* **1980**, *21*, 1571-1574.
- 21. Best, W. M.; Macdonald, J. M.; Skelton, B. W.; Stick, R. V.; Tilbrook, D. M. G.; White, A. H. *Can. J. Chem.* **2002**, *80*, 857-865.
- 22. Rio, S.; Beau, J.-M.; Jacquinet, J.-C. *Carbohyd. Res.* **1991**, *219*, 71-90.
- 23. Williams, J. M.; Richardson, A. C. *Tetrahedron* **1967**, *23*, 1369-1378.
- 24. Chalk, R. C.; Ball, D. H. *Carbohyd. Res.* **1973**, *28*, 313-325.
- 25. Reist, E. J.; Fisher, L. V.; Goodman, L. *J. Org. Chem.* **1967**, *32*, 2541-2544.