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Visual thresholds for single targets in budgerigars

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2024

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Citation for published version (APA):

Chaib, S. (2024). *Do you see the point? Visual thresholds for single targets in budgerigars*. [Doctoral Thesis (compilation), Department of Biology]. Lund University.

Total number of authors:

1

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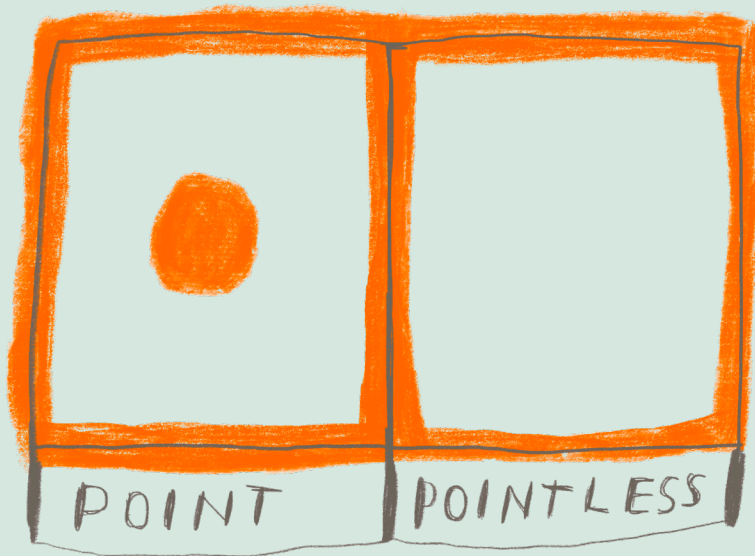
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Do you see the point?

Visual thresholds for single targets in budgerigars

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DEPARTMENT OF BIOLOGY | FACULTY OF SCIENCE | LUND UNIVERSITY



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Visual thresholds for single targets in budgerigars

Sandra Chaib



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DOCTORAL THESIS

Doctoral thesis for the degree of Doctor of Philosophy (PhD) at the Faculty of Science at Lund University to be publicly defended on the 3:rd of October at 09.00 in the Blue Hall, Department of Biology, Sölvegatan 37, Lund

Faculty opponent

Dr Nicholas Roberts
University of Bristol

Organization: LUND UNIVERSITY

Document name: Doctoral thesis

Date of issue: September 2024

Author: Sandra Chaib

Title and subtitle: Do you see the point? Visual thresholds for single targets in budgerigars

Abstract:

The vision of birds has been extensively studied, and a lot is known about what they are able to see. Contrast sensitivity and acuity, in different light intensities, is known. Still, visual perception depends on a combination of many stimulus parameters, which can make it difficult to predict the visibility of ecologically relevant stimuli.

In this doctoral thesis I have investigated visual thresholds of budgerigars (*Melopittacus undulatus*) using stimuli designed to better match visual tasks which birds encounter naturally. Starting from questions regarding the visual thresholds of birds in ecologically relevant tasks, I have used a psychophysical approach in an attempt to reveal their limits of vision.

A number of different species, including humans, are able to detect visual targets below the resolving limit of the retinal mosaic, given they provide enough contrast to the background. In Paper I we tested the detection threshold for circular single dark targets against a brighter background, the single target acuity, of budgerigars. We found that, in contrast to humans, the single target acuity of budgerigars is not higher than their grating acuity. Detection threshold varied with luminance contrast, in a similar way as for gratings, but also with the target luminance profile (0.065° for sinusoidal wave and 0.098° for square-wave shaped target). We concluded that the low contrast sensitivity of budgerigars likely limits their single target acuity.

The single target acuity of budgerigars was further investigated in Paper II where we added a semi-random movement (1.69 degrees s⁻¹) to a "square-wave" single target. Motion can increase the saliency of visual targets through attentional capture, but has also been shown to increase the luminance contrast sensitivity of budgerigars for gratings. Despite this, the single target acuity for moving targets (0.107°) did not differ from the single target acuity for static targets measured in Paper I.

In Paper III we explored the luminance vision of budgerigars immediately after experiencing a decrease in light intensity. Our goal was to simulate the light intensity dynamics experienced by cavity-nesting birds upon nest-entry. We tested the luminance detection, and discrimination, threshold for circular grey targets (9.6 degrees) on a black background as the birds went from a bright environment into a darker facility. The experiments included thresholds measured at illumination drops of ranging between 0.5 and 3.5 log units. Despite having no time limits, the birds made a response within about 1 second after stimulus onset (which was at the same time as the light decrease), and did not wait to adapt to the lower light intensities. The luminance detection threshold was in the same range when the decrease in illumination was 1.7-3.5 log units, while it was considerably higher when the illumination only dropped by 0.5 log units. The birds were able to discriminate between two grey targets with Weber fractions between 0.41 and 0.54 for all light levels. Although the visual performance is inferior to previously measured contrast- or brightness discrimination in fully adapted budgerigars, it is consistent with Weber's law. Thus, our result indicates that budgerigars partially adapt to light drops of at least -3.5 log units within ~1 second.

Key words: Bird vision, Visual ecology, Spatial acuity, Contrast sensitivity, Budgerigar, Cavity nesting. Target detection, Psittaciformes

Language: English

Number of pages: 101

ISBN: 978-91-8104-161-3 (print)

978-91-8104-161-0 (online)

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Visual thresholds for single targets in budgerigars

Sandra Chaib



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Faculty of Science
Department of Biology

ISBN
978-91-8104-161-3 (print)
978-91-8104-161-0 (online)

Printed in Sweden by Media-Tryck, Lund
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Till Flora och Lilja

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List of papers

- I. **Chaib, S.**, Ljungholm, M., Lind, O., & Kelber, A. (2019). Single target acuity is not higher than grating acuity in a bird, the budgerigar. *Vision Research*, 160, 37–42.
- II. **Chaib, S.**, Mussoi, J. G., Lind, O., & Kelber, A. (2021). Visual acuity of budgerigars for moving targets. *Biology Open*, 10(9), Article bio058796.
- III. **Chaib, S.**, Lind, O., & Kelber, A. (2023) Fast visual adaptation to dim light in a cavity-nesting bird. *Proceedings of the Royal Society B*. 290(1998), Article 20230596.

Authors' contributions

- I. S.C.: conceptualization, data curation, formal analysis, investigation, methodology, software, validation, visualization, writing – original draft and editing; M.L.: methodology, software, validation; O.L.: conceptualization, methodology, supervision, writing – review and editing; A.K.: conceptualization, funding acquisition, methodology, project administration, resources, supervision, writing – review and editing.
- II. S.C.: conceptualization, methodology, software, validation, investigation, formal analysis, visualization, writing – original draft and editing; J.G.M.: methodology, investigation, writing – review & editing; O.L.: conceptualization, methodology, software, supervision, writing – review & editing; A.K.: conceptualization, methodology, funding acquisition, project administration, resources, supervision, writing – review & editing.
- III. S.C.: conceptualization, data curation, formal analysis, investigation, methodology, software, validation, visualization, writing – original draft, writing – review and editing; O.L.: conceptualization, methodology, supervision, writing – review and editing; A.K.: conceptualization, funding acquisition, methodology, project administration, resources, supervision, writing – review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Papers not included in this thesis

Lind, O., **Karlsson, S.**, & Kelber, A. (2013). Brightness discrimination in budgerigars (*Melopsittacus undulatus*). *Plos One*, 8(1), Article e54650.

Mitkus, M., **Chaib, S.**, Lind, O., & Kelber, A. (2014). Retinal ganglion cell topography and spatial resolution of two parrot species: budgerigar (*Melopsittacus undulatus*) and Bourke's parrot (*Neopsephotus bourkii*). *Journal of Comparative Physiology A*, 200(5), 371-384.

Chaib, S., Dacke, M., Wcislo, W., & Warrant, E. (2021). Dorsal landmark navigation in a Neotropical nocturnal bee. *Current Biology*, 31(16), 3601-3605.

“I have no idea where this will lead us. But I have a definite feeling it will be a place both wonderful and strange”

Dale Cooper

Populärvetenskaplig sammanfattning

Fåglar uppfattar världen i högre upplösning, både vad gäller tid och rum, jämfört med andra ryggradsdjur. Detta gör det möjligt för dem att manövrera snabbt i flykt, men även att upptäcka fara eller föda från långa avstånd. Fåglar har dessutom ett mer avancerat färgseende och kan därför uppfatta färgnyanser som är osynliga för till exempel oss människor. Jag har i min forskning använt mig av beteendeeexperiment för att ta reda på hur bra fåglar är på att urskilja enskilda föremål under olika förutsättningar. Även om mina frågeställningar har varit relativt allmänna för fåglar, har jag använt mig av undulaten (*Melopsittacus undulatus*) som min modellorganism. Undulater är en liten fröätande papegoja (Psittaciformes) med naturlig hemvist i Australiens inland. Eftersom de är lätttränade och vanliga att ha som husdjur har de emellertid använts flitigt inom forskning.

När forskare mäter synförmågan hos fåglar (och många andra djur) genom beteendeeexperiment är det vanligt att de använder sig av standardmässiga synstimuli (bilder) med mönster av lika breda ljusa och mörka ränder. Genom att presentera bilder med olika kontrast och bredd på ränderna kan man uppskatta fågelns kontrastkänslighet – det vill säga förmågan att se skillnader på olika nyanser av grå – och synskärpa. Resultaten av sådana tester är användbara då de visar hur kontrastkänsligheten varierar med detaljstorlek (representerat av bredden på ränderna), vilket ger en helhetsbild av vad en fågel kan se. Ett ögas upplösningsförmåga begränsas av ögats storlek samt tätheten av nervceller i näthinnan, där varje nervcell (förenklat) utgör en ”pixel” i synfältet. Detta anatomiska mått på synskärpa stämmer i regel överens med måttet på det allra finaste randiga mönster som en fågel kan urskilja. Synsinnet är dock komplext och gränsen för vad en individ kan uppfatta i en given situation påverkas även av faktorer som färg, form, rörelse och ljusintensitet.

Även om vi vet en del om hur olika parametrar påverkar fåglars synförmåga så finns det fortfarande mycket som är okänt. Människor kan uppfatta en enskild linje, mot en i övrigt slät bakgrund, som är smalare än någon av linjerna i det finaste svart och vit-randiga mönstret vi kan se. Samma sak gäller för enskilda punkter, vilket innebär att vi kan uppfatta individuella stjärnor på natthimlen trots att de befinner sig tusentals ljusår bort. Givet att ett föremål har tillräckligt hög kontrast gentemot bakgrunden kan vi människor alltså uppfatta det på ett längre avstånd än vad upplösningsförmågan hos vårt synsystem egentligen tillåter. Den här förmågan är inte unik för människan utan har påvisats hos flera andra djurarter, till exempel ödlor, trollsländor och bin.

Fåglars skarpa syn tillskrivs ofta deras behov av att kunna upptäcka farliga rovdjur eller bytesdjur på långt håll. I den första artikeln undersöker vi därför undulators synskärpa när det gäller att uppfatta mörka punkter mot en ljus bakgrund. Vi tränade fåglar till att skilja mellan två bilder – en med en punkt och en utan en punkt – för att få en matbelöning. Vi använde oss av olika typer av punkter för att undersöka hur synskärpan förändras med egenskaper som till exempel kontrast. Undulators förmåga att urskilja enskilda punkter överstiger inte de mått på synskärpa som tidigare gjorts med randiga mönster. Snarare kan synskärpan för enskilda punkter betecknas som något sämre, beroende på om punkten har skarpa eller suddiga kanter. Undulators relativt låga synskärpa för enskilda mörka punkter tror vi främst beror på deras låga kontrastkänslighet, en egenskap som de delar med andra fågelarter.

I den andra artikeln visar vi att rörelse inte påverkar undulators förmåga att uppfatta enskilda punkter. Då ett annalkande rovdjur ofta rör sig så förväntade vi oss att rörelse skulle underlätta upptäckten av enskilda punkter. Tidigare forskning har visat att undulater har högre kontrastkänslighet för randiga mönster som rör sig horisontellt jämfört med om de är stilla, men detta verkar inte påverka synskärpan för enskilda punkter.

Fokus i den tredje artikeln är på undulators synförmåga under plötsliga minskningar i ljusintensitet. Många fåglar häckar i trädhålor, liksom undulater som i sin naturliga miljö bygger bon i ihåliga gamla eukalyptusträd. Ljusskillnaden mellan den mörka bohålan och den soliga utsidan är troligtvis hög, vilket påverkar deras synförmåga. Liksom för människor så tar det tid för fåglar att helt anpassa sina ögon till mörker – upp till 45 minuter. Ändå spenderar hålhäckande fåglar oftast bara några sekunder åt gången i boet när de matar sina ungar. Vi ville veta hur bra hålhäckande fåglar kan se när de precis kommit in i sitt bo efter att ha vistats i dagsljus. För att ta reda på detta behövde vi testa synförmågan hos fåglar under ljusförhållanden som efterliknar dem som de naturligt möter i denna situation. Vi tränade undulater till att, från en ljus bur, flyga in i en mörkare låda. Väl inne i lådan fick de välja mellan två olika bilder (större punkter med olika grå nyanser på en svart bakgrund) i utbyte mot en matbelöning. Våra resultat visar att undulater delvis anpassar sin synförmåga till den mörkare miljön i lådan redan inom en sekund. Undulater är lika bra på att se skillnad på större punkter med olika grå nyanser oavsett om ljusintensiteten i lådan är mycket eller bara lite lägre jämfört med utanför. Även om synförmågan försämras vid en hastig minskning av ljuset, så sker en viss anpassning nästan med en gång. Detta innebär att fåglar troligtvis kan se sina ägg och ungar även i en mörk bohåla.

Sammantaget visar mina studier att synförmågan hos fåglar är ett område som kräver fortsatt forskning, inte minst om hur den påverkas av dynamiska ljusförhållanden. Studier inom beteendekologi, fysiologi och anatomi är nödvändiga för att förstå synens funktionella betydelse samt hur detta avspeglar sig i både fysiska och beteendemässiga anpassningar. Att mäta fåglars synförmåga genom kontrollerade beteendexperiment ger dock direkt vetskap om vad de kan uppfatta, något som ofta behövs för korrekta tolkningar av synrelaterade beteenden och anpassningar.

Introduction

For the majority of bird species, vision is the primary sense (Martin, 2017a). It allows instant gathering of information about remote objects and events, making it especially useful when moving in mid-air. Indeed, birds depend more on vision than any other vertebrate class (Hodos, 1993; Walls, 1942). The visual system of birds allows them to experience their surroundings both fast (Boström et al., 2016) and in great spatial detail, the latter reflected in some species of raptors having the highest spatial resolving power measured in an animal (Potier, Mitkus, et al., 2020). In addition, birds have highly advanced colour vision (Kelber, 2019). However, even though birds are visual champions the interspecific variation is great and some visual aspects, such as contrast sensitivity, are comparatively poor in all birds (Ghim & Hodos, 2006; Potier et al., 2018).

Visual thresholds in animals are commonly measured under controlled conditions using standard stimuli. Experiments performed in this way are needed to compare different species and make deductions based on previous knowledge. However, if one is interested in what an animal can perceive during specific tasks in its behavioural repertoire, the standard measurements do not always suffice. Visual thresholds are often influenced by context, and different dimensions of visual perception might affect each other (e.g., Haller et al., 2014; Lind, 2016; van den Berg et al., 2020).

The aim of this thesis is to investigate visual thresholds of birds using stimuli designed to better match visual tasks which birds encounter naturally. Starting from questions regarding the visual thresholds of birds in ecologically relevant tasks, I have used a psychophysical approach to investigate their limits of vision.

Even though my questions apply to many species, I have used the budgerigar (*Melopsittacus undulatus*) as a model throughout the papers included in this thesis. The budgerigar is commonly known as a sociable, affectionate, and easily trained pet bird. Indeed, it is probably the most common pet bird in the world. Many of the same qualities which make it appreciated as a pet also make it the perfect bird for behavioural experiments. The budgerigar has been studied quite extensively regarding vision (e.g., Bhagavatula et al., 2009; Goldsmith & Butler, 2003; Haller et al., 2014; Lind et al., 2014; Lind et al., 2013; Lind et al., 2012; Mitkus et al., 2014), but also behaviour (Brockway, 1964a, 1964b; Stamps et al., 1985, 1989; Stamps et al., 1987), providing me with a stable ground of knowledge for asking further questions.

Besides being a popular pet and a model animal in science, the budgerigar is native to the inland of Australia. The budgerigar belongs to the psittacines (parrots), and wild birds are small (20-40 g), mostly bright green with a yellow face and black and yellow wings (Menkhorst et al., 2017). Preferentially they inhabit arid and semi-arid open grasslands with few trees where they move around in large flocks feeding on grass-seeds (Menkhorst et al., 2017; Wyndham, 1980a, 1980b). While hawks (Accipitridae) and falcons (Falconidae) belong to the natural threats of budgerigars (Cowie, 2014; Wyndham, 1980a), their open foraging habitat enables detection of predators at a long distance. But at what distance would a budgerigar be able to detect a potential aerial threat? In Papers I and II, we explored the visual acuity and contrast sensitivity of budgerigars for single targets in an attempt to answer such questions.

In Paper III we asked what birds nesting in dark cavities are able to see when they enter the nest to feed their chicks. Having growing offspring, cavity nesting birds are obliged to move repeatedly in and out of the nest to provide the young with food. Visits to the nest are often quick, and the light intensity difference between the inside and outside can be substantial (Maziarz & Wesolowski, 2014; Reynolds et al., 2009; Wesolowski & Maziarz, 2012). To be able to use vision during these circumstances the visual system would need to adapt rapidly. While feeding in cavity nesting birds likely involves more than one sensory modality, many studies show that visual cues play a role (e.g., Dugas, 2015; Heeb et al., 2003; Podkowa et al., 2019; Podkowa & Surmacki, 2017). Budgerigars typically nest in old hollowed out eucalyptus trees (Higgins, 1999; Wyndham, 1981). The nest entrance hole is small (3-6 cm) and the eggs may be laid up to several metres below (Higgins, 1999; Schrader, 1975), likely out of reach of much illumination. In paper III we explored whether it would be possible for budgerigars to use visual cues when feeding their nestlings.

The outcome of our studies will be further discussed in the last chapter, "Spatial vision in birds", where I also summarize current knowledge on bird spatial vision and visual ecology. The papers can be found in full length at the end of this thesis. In the chapter following this introduction, "The vertebrate eye", I present the main structure and building blocks of the vertebrate eye. Next, in "Spatial vision" I briefly discuss some of the basic principles of luminance mediated vision with an emphasis on vertebrates in general. The following chapter, "Measuring spatial vision", introduces methods for the quantification of stimuli parameters and spatial visual abilities.

The vertebrate eye

The general structure of the eye

Vertebrates have *camera-type* eyes, in which all entering light is refracted through a single optical unit (the *lens* and the *cornea*) and focused on the light sensitive inner surface of the eye (Cronin et al., 2014) (fig. 1a). While the lens accounts for all refraction in aquatic vertebrates, most of the refraction in terrestrial vertebrates is caused by the cornea, which is the curved outer surface at the front of the eye (Land & Nilsson, 2012). In front of the lens is the *iris*, a pigment-containing thin structure, with an aperture, the *pupil*, which regulates the amount of incoming light (Douglas, 2018). The space between the cornea and the lens is filled with a clear liquid (*aqueous humour*).

The back of the eye, the “eye cup”, has a roughly hemispherical or tubular (in owls and some fish) shape. Its inside, the *vitreous body*, is filled with a transparent gel-like substance (*vitreous humour*). The eye cup itself consists of several layers of tissue including sturdy connective tissue (the *sclera*), thin blood vessels (the *choroid*), and a layer of dark melanin containing cells (the *retinal pigment epithelium*). At the innermost lining of the eye cup is the *retina*, a sheet of specialized neurons, whose purpose is to turn light into a visual signal and transport it via the optic nerve to the brain. In mammals, thin blood-vessels running across retina provide it with necessary nutrients. Birds lack these vessels but instead have a pleated pigmented vascular structure, called the *pecten oculi*, which protrudes into the vitreous body where it emits nutrients into the vitreous humour (Pettigrew et al., 1990).

The organization of retinal neurons

The vertebrate retina contains five main types of neurons whose cell bodies and intricate synaptic network are arranged in distinct layers. The eye-cup of vertebrate eyes has evolved from evaginations of the frontal parts of the brain and the organization of the retinal layers is therefore “inverted” (Lamb et al., 2007). As a consequence, the *photoreceptors* initiating the visual pathway are situated in the outermost retinal neuronal layer, and the visual signal while downstream retinal neurons are positioned further in (fig. 1b).

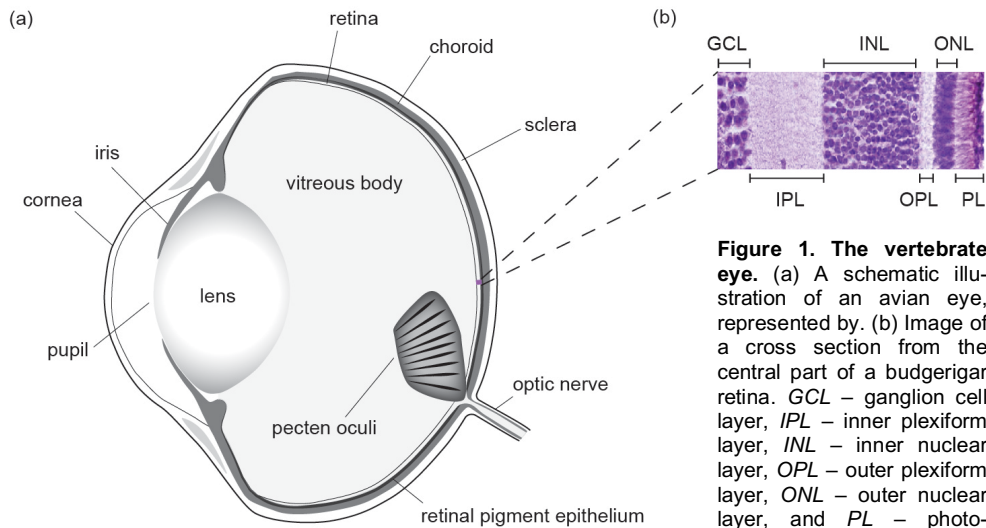


Figure 1. The vertebrate eye. (a) A schematic illustration of an avian eye, represented by. (b) Image of a cross section from the central part of a budgerigar retina. GCL – ganglion cell layer, IPL – inner plexiform layer, INL – inner nuclear layer, OPL – outer plexiform layer, ONL – outer nuclear layer, and PL – photoreceptor layer. Adapted from Mitkus et al. (2014).

Vision starts when incoming photons are absorbed by photosensitive pigments in the photoreceptors, initiating an electric response in a process called photo-transduction. The photoreceptors forward the signal to the *outer plexiform layer* (OPL), which is the first synaptic layer, where they contact *horizontal cells* (HCs) and *bipolar cells* (BC). BCs connect the OPL to the *inner plexiform layer* (IPL), the second synaptic layer, where they make connections with *amacrine cells* (ACs) and *retinal ganglion cells* (RGCs) (Baden et al., 2020). In between the OPL and the IPL is the *inner nuclear layer*, housing the cell bodies of BCs, HCs and ACs. The innermost layer of the retina contains the cell bodies of RGCs and is referred to as the *ganglion cell layer*. The axons of the RGCs carry the integrated visual signal, via the optic nerve, to the visual centres in the brain.

In the OPL, each photoreceptor commonly synapses with several BCs. Different BCs have distinct response characteristics, thereby creating several parallel information channels from the output of the same photoreceptors (Masland, 2012). Classically, BCs are divided into “ON” BCs cells, responding to light onset (bright stimuli), and “OFF” BCs, which respond to light off-set (dark stimuli). The temporal characteristic of their response further divides them into “transient” or “sustained” BCs (Masland, 2012).

HCs connect laterally to photoreceptors and BCs, where they provide both feedback, as well as feedforward information. The lateral connections of horizontal cells typically organise the bipolar cells in *centre-surround* structures, where the surrounding BCs typically respond in an antagonistic manner to the centre BC. This type of lateral organization of neurons is also referred to as *surround suppression* and is present at several levels in the visual pathway. A classic example of surround

suppression is the ON-OFF centre-surround organization, where a light stimulus will make the bipolar cell in the centre respond maximally, while the surround will suppress this response (Barlow, 1953; Kuffler, 1953).

Bipolar cells of different types carry their information to specific levels of the IPL where they synapse with ganglion cells and amacrine cells (Masland, 2001; Masland, 2012). Amacrine cells work laterally in a similar way as horizontal cells do in the outer OPL although their function is more multifaceted, and they build more complex networks (Masland, 2012). They modify the output of bipolar cells to ganglion cells, but they also connect directly to ganglion cells as well as other amacrine cells. The function of amacrine cells are often refined to code intricate visual features. Some amacrine cells have large axonal arbores enabling wide-field computations of visual input; others are sensitive to motion in specific directions (Berson, 2020; Masland, 2012).

Input from several bipolar and amacrine cells are typically combined to create the receptive fields of ganglion cells. Like the neurons in the OPL, the receptive fields of retinal ganglion cells almost always have a centre-surround organization, although their feature selectivity is typically more complex. Different types of retinal ganglion cells often selective to specific spatio-temporal features and send their output along parallel pathways to different brain regions (Ibbotson & Meffin, 2020; Schwartz & Swygart, 2020). Example of feature selectivity of ganglion cells are movement direction, orientation, and object motion (Schwartz & Swygart, 2020).

Spatial vision

Vision, a bit simplified, is the sampling of light reflected or emitted from structures in the environment. Light reaching an eye has a number of different properties which can be used to extract information: its spatial origin, intensity (luminance), spectral composition, polarization, and temporal properties. The most basic form of true vision involves the simultaneous sampling of luminance from different directions (Land & Nilsson, 2012), information which can be used to create a spatial representation of the surroundings and guide behaviour. This is what is commonly referred to as spatial vision. However, spatial information is not only extracted from the variation of light intensity across space, but also from its change over time. The retinal image is almost never completely still and even when fixating targets, most vertebrates make small involuntary eye movements (Martinez-Conde & Macknik, 2008). Image motion is integrated with spatial perception already at the level of retinal processing and has an impact, for example, on object saliency, depth vision, spatial resolution, and contrast sensitivity. Although luminance, spatial resolution, and motion are greatly entangled and inter-dependent, this chapter is divided into separate sections which are primarily dedicated to each of these properties separately.

Luminance and contrast

Objects and structures are visible to the eye because they emit or reflect light. Perceiving spatial differences in the intensity of this light is a fundamental visual ability, which can be used to extract information about, for example, texture, form, and depth. The amount of light reflected from a surface (the luminance) is proportional to the intensity of the incident light (the illuminance) (Shapley & Enroth-Cugell, 1984). Since the ambient illumination changes by more than 9 log units over a 24-hour period (Rieke & Rudd, 2009) spatial luminance differences in absolute values are most often not reliable visual cues. Thus, the visual system strives to keep its response invariant to the ambient light conditions to be able to extract useful information from its surroundings (Shapley & Enroth-Cugell, 1984). This is achieved by scaling the response to the overall luminance in the scene, thereby measuring proportional rather than absolute differences. As a result, visual stimuli will convey

information about the characteristics of the reflecting surfaces, rather than of the ambient light level.

Proportional processing is present in many different sensory modalities and can be described by *Weber's law* (Akre & Johnsen, 2014). Weber's law states that the minimum perceptible change in a stimulus is proportional to the stimulus magnitude. Applied to spatial luminance vision, Weber's law predicts that the minimum luminance difference ΔI needed for an object to be visible against its background, is proportional to the absolute luminance of the background I :

$$\Delta I \sim I \quad (0.1)$$

In other words, the smallest detectable luminance difference on a light background is larger than on a dark background (fig. 2). However, the ratio of the smallest detectable luminance difference to the background luminance is the same, and is commonly referred to as the *Weber fraction* (ω):

$$\frac{\Delta I}{I} = \omega \quad (0.2)$$

Weber's law holds well for large, long duration stimuli and over a wide range of intensities (Perlman & Normann, 1998). At very high light levels Weber's law fails due to photoreceptor response saturation, while quantal fluctuations – also called photon shot noise – limit visual sensitivity at low light levels (Shapley & Enroth-Cugell, 1984). The absorption of photons is stochastic and follows Poisson statistics, which means that the photon shot noise (the “uncertainty”) in a signal of N photons is \sqrt{N} . The reliability of the signal, expressed as the signal to noise ratio N/\sqrt{N} , thus decreases with light intensity (Cronin et al., 2014; Land & Nilsson, 2012). The *DeVries-Rose law* (or the square root law) tells us that the minimum detectable luminance difference, ΔI , at low light levels is proportional to the square root of the background intensity I :

$$\Delta I \sim \sqrt{I} \quad (0.3)$$

At even lower light intensities an additional source of noise, *dark light*, is noticeable (fig. 2). Dark light originates from spontaneous thermal activation in the photoreceptors and is what ultimately sets the limit to vision (Barlow, 1957; Warrant, 1999).

Luminance contrast is the physical measure of relative luminance variation in a visual stimulus. Although luminance contrast can be calculated in a few different ways, depending on stimulus type, it always describes the magnitude of luminance variation in relation to the average luminance (see “Measuring spatial vision”).

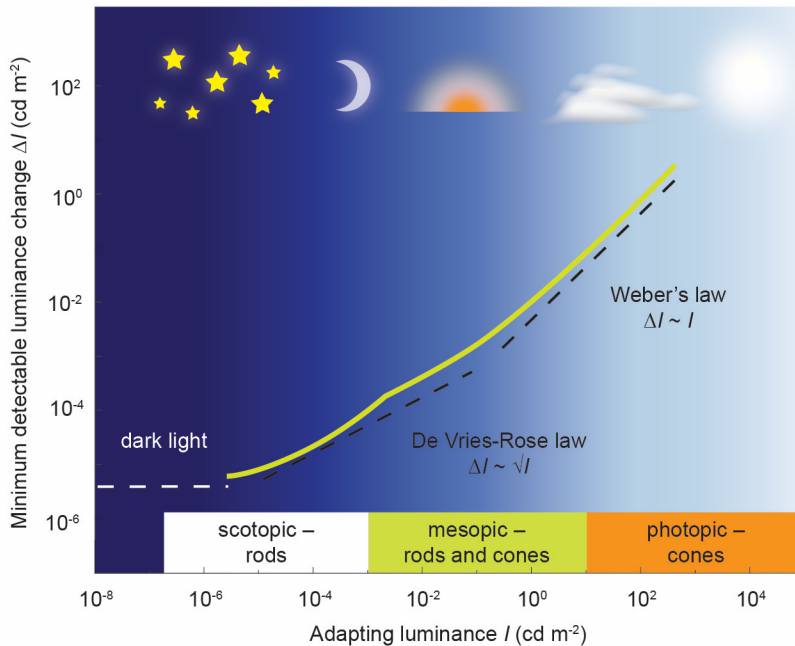


Figure 2. Visual sensitivity of humans at different light intensities. The yellow line shows the minimum detectable luminance change (ΔI) for different adapting luminances (I). The dashed lines illustrate the effect of noise and adaptation on ΔI in different regions. Adapted from (Cronin et al., 2014). The sensitivity ranges of rods and cones are noted in the bottom of the figure.

Luminance adaptation

The visual system responds to relative luminance differences by adapting to the prevailing luminance. Luminance adaptation includes numerous mechanisms which are active at different light intensity ranges, and which work at different retinal processing levels (Rieke & Rudd, 2009). The timeframes for the different adaptation processes are also diverse, suiting the array of different instances in which light intensity might vary throughout the active hours of an animal (Schwartz & Levine, 2021). Some mechanisms are slow and suite the larger cyclic changes in light availability between day and night. Others are fast and operate in the millisecond range and therefore work well for the rapid luminance changes that occur when moving the gaze (Dunn et al., 2007; Rieke & Rudd, 2009) or moving rapidly between different light environments.

The pupillary light response

The most distal luminance adapting mechanism is the pupillary light response, which controls the amount of light reaching the retina by contraction or dilation of the iris muscles. In most animals, pupil movement only has a marginal effect on

luminance adaptation and is therefore believed to have primarily other functions (i.e., enhancing spatial acuity by preventing optical aberrations; Douglas, 2018; Lind et al., 2008).

Retinal duplicity

In contrast, the “duplex retina” of vertebrates contributes greatly to the adaptability of their luminance sensitivity. Most vertebrates, including humans and birds, have two major classes of photoreceptors, rods, which dominate vision at low light intensities, and cones, which are the primary photoreceptors at high light intensities. Based on the human visual system, the light intensities in which only rods are active are referred to as *scotopic*, whereas the ones in which only cones are active are called *photopic* (fig. 2). The working ranges of rods and cones overlap in the *mesopic* light intensity range (Barbur & Stockman, 2010).

Pigment bleaching

In addition to the shift between different types of photoreceptors, luminance adaptational mechanisms also operate at the level of the individual receptors. For example, the sensitivity of both rods and cones is partly regulated by the concentration of the light sensitive visual pigments. Visual pigment molecules consist of an opsin molecule which is bound to a chromophore. The absorption of a photon by a pigment changes the shape of the chromophore, transforming the pigment from an inactive form to an active form, an event which is the start of the visual process (Cronin, 2020). The active form of the visual pigment is said to be “bleached” and must be regenerated into its inactive form before it can absorb another photon (Perlman & Normann, 1998). At higher light intensities a larger proportion of pigment in the receptor cell is bleached, which makes the photoreceptor less likely to absorb photons. Rods are more sensitive than cones and bleach at lower light intensities.

The recovery from full bleaching is commonly referred to as “dark adaptation”. In cones this process is limited by pigment regeneration and usually takes around 5 minutes (Jiang & Mahroo, 2022). Dark adaptation in rods is slower, likely as a consequence of local photoproduct concentrations which hampers the regeneration process (Hecht et al., 1937; Lamb & Pugh, 2004). Full dark adaptation of rods takes between 15-40 minutes, depending on degree of bleaching (Hecht et al., 1937; Lamb & Pugh, 2004).

Spatial integration

At low light levels luminance sensitivity is increased by integration of visual signals across both space and time. Partly this is a consequence of the transition from cones to rods, since rods have a wider receptive field size and a longer integration time, but the cone and rod pathways are also individually adjusted. The reliability of visual signals is increased at low light levels by averaging the signals of adjacent retinal

neurons. The ambient light level regulates the production of “gap-junctions” which mediate the electric coupling between neurons (Schwartz & Levine, 2021). Increased neuronal coupling may, for example, result in a weakening of the surround suppression mechanism (see “The organization of retinal neurons”), whereupon the receptive field of retinal ganglion cells becomes dominated by its centre (Barlow, 1953; Kuffler, 1953; but see: Warwick et al., 2023). The functional consequence of this adjustment is an increased sensitivity at the expense of spatial resolution (Barlow, 1958; Barlow et al., 1957).

Background adaptation

The luminance range encountered by an eye in just a single visual scene is wide and often changes abruptly by a shift of gaze (Frazor & Geisler, 2006). To keep up with rapid light fluctuations, the retina uses adaptation mechanisms that work in less than a second (Fain et al., 2001). These mechanisms are often referred to as *background adaptation* and they modify both the gain (response magnitude for a fixed signal input) and the speed of signal integration, and operate at several retinal levels (Dunn et al., 2007; Rider et al., 2019). Furthermore, different adaptational mechanisms work at different light levels. As a general rule, mechanisms working early in the visual pathway (e.g. in the phototransduction cascade) are active at higher light levels, while those working at later stages, where the signal convergence rate is high (e.g. at the synapses between bipolar cells and ganglion cells), are active at lower light levels (Dunn et al., 2007; Schwartz & Levine, 2021).

Contrast adaptation

The visual system does not only adapt to the average luminance but also to the average amount of luminance contrast. Like luminance adaptation, *contrast adaptation* involves several mechanisms which act at both different stages in the visual pathway and different timeframes (Baccus & Meister, 2002; Kaplan, 2020).

Other factors affecting contrast sensitivity

As previously mentioned, Weber’s law works best for luminance differences in stimuli with large spatial extent and long temporal duration. Both the receptive field size and integration time of the retinal pathways are affected by luminance adaptation, which in turn may affect the processing of fine or fast-moving stimuli.

Retinal processing mechanisms which are independent on the general light level may also affect the perception of spatial luminance differences. Lateral inhibition between retinal neurons (see previous chapter) can enhance luminance differences at sharp transitions while they are reduced at gradual changes (Enroth-Cugell & Robson, 1966; Kuffler, 1953). Other factors that may affect perceived luminance difference are stimulus area (Campbell & Robson, 1968; Robson & Graham, 1981),

spatio-temporal characteristics (Burr, 1981; Haller et al., 2014; Robson, 1966) and luminance polarity (Adrian, 1989; Lu & Sperling, 2012; van den Berg et al., 2020).

Spatial acuity

Spatial acuity is the maximum fineness with which the visual system can resolve an image. It can vary a lot between different species, but specific behaviours or contexts also require more or less detailed visual information. Many visually controlled behaviours, such as movement control or obstacle avoidance, need only a rough representation of the environment; other behaviours, such as prey identification, or communication, require detailed visual information (Land & Nilsson, 2012). The limit to the spatial acuity of an eye depends primarily on the optical quality and the sampling frequency of the retina.

Optical factors affecting spatial acuity

The optical unit of the eye strives to focus the incoming light onto the retina to create a sharp image. However, imperfections of the optical unit and the physical properties of light cause the image to lose some of its sharpness in this process. Typically, smaller details (higher spatial frequencies) are blurred more than larger details (lower spatial frequencies). The loss of image quality caused by the passage through an optical device is usually described by the modulation transfer function, which is an expression of the decrease in contrast as a function of spatial frequency.

Diffraction

When passing an edge or an opening a flat wavefront will “curve”, causing the part of the wave closest to the obstacle to be out of phase with the rest of the wavefront (fig. 3a). The same thing happens to light when passing the pupil, which cause a delay to some parts of the wave fronts. When reaching the retina, those parts of the wavefront that are in phase will reinforce while those that are out of phase will cancel out, giving rise to a diffraction pattern. The diffraction pattern leads to a “blurring” of the image, which is more prominent for finer spatial details and for smaller pupil sizes (Cronin et al., 2014; Land & Nilsson, 2012). The finest details of an image passing the pupil will be completely filtered out.

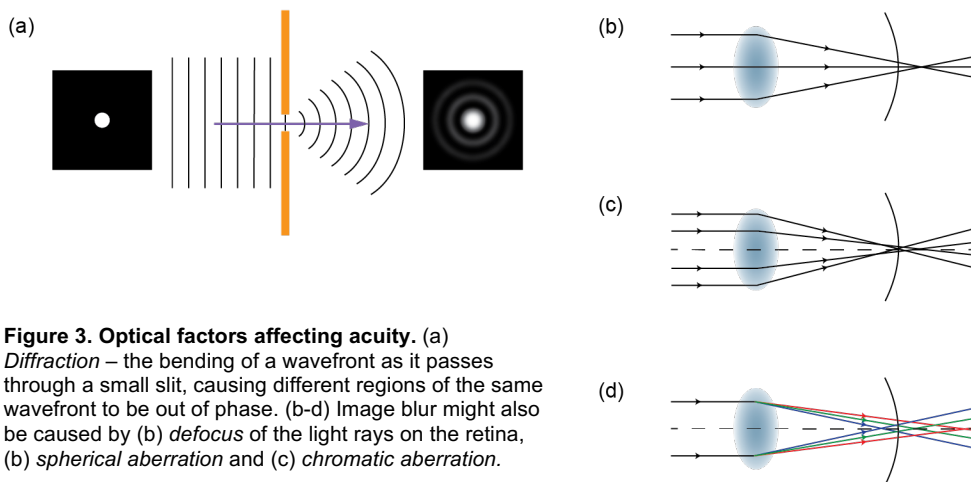


Figure 3. Optical factors affecting acuity. (a)

Diffraction – the bending of a wavefront as it passes through a small slit, causing different regions of the same wavefront to be out of phase. (b-d) Image blur might also be caused by (b) *defocus* of the light rays on the retina, (b) *spherical aberration* and (c) *chromatic aberration*.

Defocus

Although diffraction sets the ultimate limit to image resolution, other optical phenomena can also contribute to decreased image sharpness. One obvious cause of image blur is defocus – that is, when the image is focused in front or behind the retinal plane (fig. 3b). Nearby objects are brought to focus further away from the lens than more distant objects, creating difficulties in maintaining a sharp image in a three-dimensional world. Among vertebrates there are different solutions to this problem. Fishes move their lens back and forth, which changes the distance between the lens and retina, allowing them to keep the desired object in focus. Mammals, birds, and reptiles change the curvature of their optic unit, they accommodate, which alters its focal length (Land & Nilsson, 2012; Ott, 2006). Some cartilaginous fishes (i.e. bluntnose stingray [*Hypanus say*], Atlantic stingray [*H. sabinus*] and smooth butterfly ray [*Gymnura micrura*]) have so-called “ramp retina”, where the dorsal and ventral parts of the retina have different distances to the lens. Thus, the viewing distance at which an object is in focus differs for different areas of their field of view (Ott, 2006; Sivak, 1976; Walls, 1942). In a similar fashion, some animals that forage on the ground (a few species of bird included) instead have a variable state of refraction across the lens (Vietnamese leaf turtle [*Geoemyda spengleri*]: Henze et al., 2004; Hodos & Erichsen, 1990; rock pigeon [*Columba livia*] and domestic chicken [*Gallus domesticus*]: Millodot & Blough, 1971; Rounsley & McFadden, 2005; northern leopard frog [*Rana pipiens*] and Common frog [*R. temporaria*]: Schaeffel et al., 1994). The lower and frontal visual field of these species are myopic, making it possible to keep the nearby ground in focus while at the same time looking out for more distant objects in the rest of the visual field (Hodos & Erichsen, 1990; Millodot & Blough, 1971).

Aberration

Spherical aberration is an additional phenomenon which may cause image blur. Light which enters the eye at the periphery of the lens comes to focus closer to the lens than light entering through the center or the lens. The focus plane of parallel light rays will thus differ depending on where they pass the lens, and the result is a decrease in sharpness of the image (fig. 3c). Many animals, such as fishes and humans, compensate for spherical aberration by having a lower refractive index at the edges of the lens (Cronin et al., 2014; Land & Nilsson, 2012).

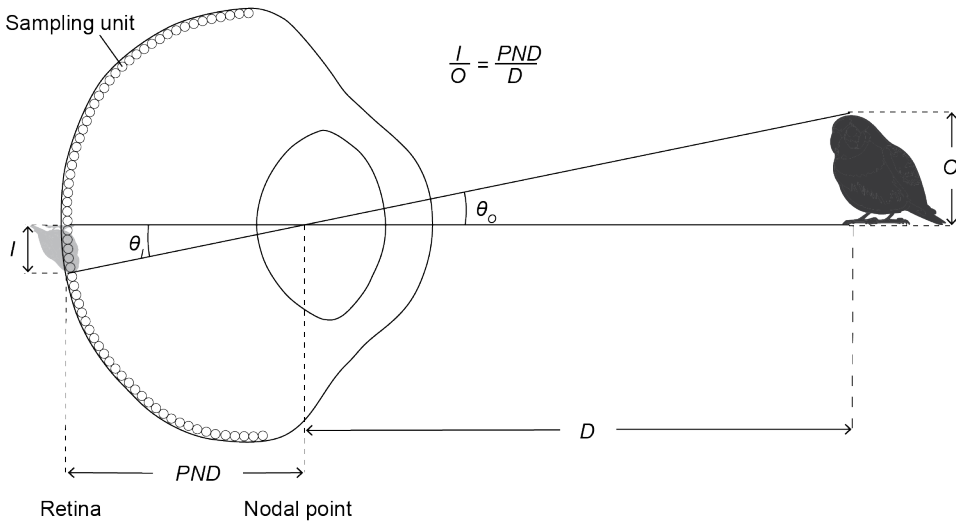


Figure 4. Retinal sampling frequency. The resolution in which an image is seen depends on how many retinal units that samples it. Light passing through the nodal point is refracted minimally, and the angular subtense of an object (θ_o) in the visual field thus corresponds to the angular subtense of the retinal image (θ). The size of the retinal image (I) depends further on the posterial nodal distance (PND) of the eye.

Chromatic aberration is caused by the different refractive index of light of different wavelengths. Short wavelength light (“blue light”) refracts stronger than long wavelength light (“red light”), in the same medium, and will consequently come to focus closer to the lens (fig. 3d; Land & Nilsson, 2012). To work around chromatic aberration some vertebrates have developed “multi-focal lenses” which have concentric zones with different refractive indices, allowing a part of the light from all visible wavelengths to be focused on the retinal plane (Kröger et al., 1999).

Retinal factors affecting spatial acuity

There is little use to pass high-quality images through the optics of an eye unless the retina can sample it. The resolution in which the retina can sample an image, the *retinal sampling frequency*, depends on two main factors: the size of the image projected onto the retina, and the density of retinal units which sample the image (Land & Nilsson, 2012).

The size of the image reaching the retina is decided by the *retinal magnification factor* (RMF), which is a measure of the retinal distance covered by 1° of the visual field (Pettigrew et al., 1988). The RMF depends on the *posterior nodal distance* (PND), which is the distance between the nodal point (center of curvature of the lens) and the back of the eye (fig. 4). A large eye (with a large PND) generally has a high RMF, which can create large retinal images.

The resolving power of the eye further depends on the density of retinal sampling units. One sampling unit may correspond to one single photoreceptor, but more often several, if their signals converge onto the same ganglion cell. (See “The organization of retinal neurons”).

Other factors affecting spatial acuity

Not all images are perceived with the highest spatial acuity, but the resolving power of the visual system varies with several parameters. One example is the luminance contrast of the image. Since the contrast of small details is attenuated by passing the optics (but also other tissue), only high contrast images can be perceived at the highest resolution (De Valois & De Valois, 1991). Luminance intensity also has a profound effect on spatial acuity because of the increased spatial pooling with adaptation to lower light levels (Barlow et al., 1957; Lind et al., 2012).

Center-surround mechanisms are known to increase the luminance contrast of small spatial details and thus improve their sharpness. However, the receptive fields of these units are too large to have an effect at the spatial acuity limit (Westheimer, 2009b).

Feature detection below the theoretical resolution limit

Predicted acuity limits based on optical quality and retinal sampling frequency generally agrees well with the behavioral ability to visually resolve gratings and conventional optotypes (e.g. tumbling E or Landholt C; Crossland, 2010; Rossi & Roorda 2010; Williams and Coletta 1987). For some visual tasks, however, the ability to perceive spatial detail may exceed the resolution limit. For example, some vertebrates, including humans, are better at detecting small single objects or targets against a uniform background, compared to resolving fine details in a pattern

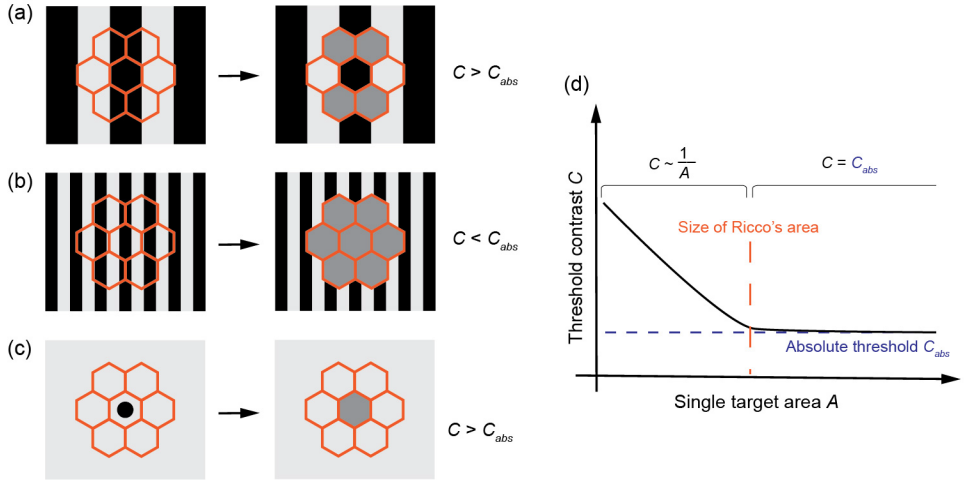


Figure 5. Ricco's law of complete spatial summation. (a-c) The red hexagons represent *Ricco detectors*, which are hypothetical receptive field units the size of Ricco's area. The figures in the left column illustrate the image before spatial summation and the figures in the right column after spatial summation. (a) A grating is visible if the contrast between adjacent Ricco detectors (C) exceeds the threshold contrast (C_{abs}). (b) Single stripes in a grating cannot be perceived below the retinal sampling limit, (c) unlike single targets provided the contrast is high enough. (d) Below the size of Ricco's area, the threshold contrast is inversely proportional to the target area, while above it is constant.

(Ehrenhardt, 1937; Hecht et al., 1947; Sandow & Hanke, 2024). A uniform target which is too small to be fully resolved by the retinal mosaic, can still be detected if it has enough contrast to the background (O'Carroll & Wiederman, 2014; Thibos et al., 2019). For such small targets, the detection threshold contrast is inversely proportional to the target area (fig. 5b). This relation is known as *Ricco's law of complete spatial summation*¹. Ricco's law is valid for uniform targets below a critical angular size, "Ricco's area", within which visual signals are spatially summed (Crume, 2014; Thibos et al., 2019). For targets exceeding the size of Ricco's area, the detection threshold approaches an asymptote of the absolute contrast threshold (Blackwell, 1946; Crume, 2014). The size of Ricco's area depends on factors like retinal locus and the adaptational state of the eye. In humans, the Ricco's area is smallest in the center of the eye while it increases in size towards the periphery. The size of Ricco's area also expands as the eye adapts to lower light levels (Barlow et al., 1957). The exact anatomical and physiological basis for Ricco's law is debated but it is commonly assumed that the size of Ricco's area corresponds to the receptive

¹ Ricco's law of complete spatial summation: $C=A*k$, where C is the threshold contrast, A is target area and k is a constant. Annibale Riccò (1844-1919), Italian astronomer.

field center of retinal ganglion cells, or is related to ganglion cell density (Glezer, 1965; Lie, 1980; Thibos et al., 2019; Volbrecht et al., 2000).

Another example where visual perception can exceed the limits of retinal sampling frequency is the detection of small displacements of borders and lines, a phenomenon known as *hyperacuity* (Westheimer, 1975). Humans have been shown to be able to see misalignments that are 5 to 10 times finer than what the retinal sampling rate would predict (Westheimer & McKee, 1977; Westheimer, 2009a).

Motion vision

As mentioned in the first paragraph of this chapter, motion vision is often an integral part of spatial visual perception. The retinal image is in constant change due to movement of external objects or to movement by the eyes of the animal itself. Many times the retinal motion per se carries important information, like the sudden movement of a prey animal, while at other times, the motion is rather a “side-effect” of a behaviour, like the motion of the background during visual tracking of a prey.

Retinal image motion is broadly divided into two classes: 1) *self-induced motion*, and 2) *object motion* (Frost, 2010). How the motion signal is interpreted and what type of action (if any) it will invoke, usually depends on which of these two categories it belongs to. The division between self-induced and object motion signals is thus often made already at the retinal level, and the information is processed along separate visual pathways (Wurtz, 1998; Wylie, 2013).

Self-induced motion

The most common cause of retinal image motion is movement of the eyes of the viewer itself (Cronin et al., 2014; Frost, 2010). Self-induced motion, also referred to as “global motion”, typically covers the entire, or a large part of the visual field. The pattern of retinal motion created by a viewer moving relative a static environment is called an *optic flow field*. The optic flow field varies in a predictable way with the viewer’s direction, speed, and type of movement, but also with the distance to objects in the environment (Gibson, 2015). Optic flow can thus be used to derive information both about one’s own movement and the spatial construction of the environment.

Translational optic flow is caused by a spatial displacement of the viewer relative to its surroundings, for example forward locomotion. Perpendicular to the direction of heading, the optic flow field moves in a single direction, the opposite direction of the translation of the viewer (fig. 6a). The strength of the optic flow depends on the speed of the viewer, but also on the distance to the objects and structures which

are imaged on the retina, where objects close by move faster than objects far away (Lee & Kalmus, 1980). The use of optic flow for distance assessment is also called motion parallax and is thought to be utilized by several animal species which during visual fixation move their heads repeatedly from side to side (Kral, 2003).

In the direction of heading, the optic flow field moves radially outward or expands (fig. 6b). In the focus of expansion, which indicates the direct heading, the retinal image is completely still, while the strength of the optic flow increase with increasing distance to this point. This information can thus provide useful information on the heading of translation (Warren Jr et al., 1988). The expanding flow field can also be used to assess the "time to contact" with external objects; the rate of expansion of the image of an object at the focus of expansion increases when one approaches it (Lee & Kalmus, 1980).

Rotational optic flow is experienced by an animal as it rotates around its own axis. In contrast to translational optic flow, rotational optic flow does not contain information about the distance to external objects since the entire surroundings will move

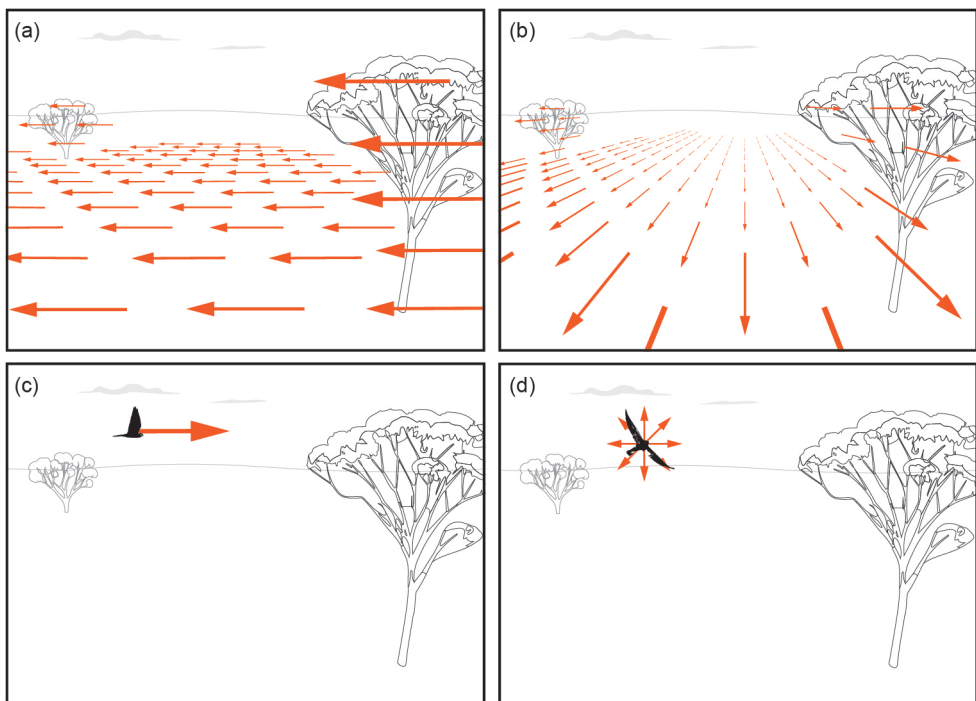


Figure 6. Retinal image motion. The direction and speed of image motion is indicated by the direction and size of the arrows. (a,b) Illustrations of self-induced translational image motion. (a) The left lateral field of view of someone moving "to the right" relative the image, and (b) the field of view in the direction of travel when moving "into" the image. Inspired by illustrations in Gibson (2015) (c,d) Illustrations of objects motion. (c) A raptor passing by the viewer, and (d) a raptor approaching the viewer ("looming motion").

at the same angular speed. Usually, animals strive to separate the translational components of the optical flow field from the rotational components. This can partly be achieved by making compensatory movements eye- or head movements (e.g. the optokinetic- and optomotor reflexes) when experiencing rotation (Land, 1999). These movements typically consist of a slow stabilizing phase in which the animal fixates its gaze at a point in the moving surrounding followed by a fast saccade directing the eyes a new fixating point (Land, 2014). During the slow phase, the rotational optic flow is minimized, whereas other visual information becomes more conspicuous.

Most vertebrate species do saccadic eye and head movements also during other types of visual behaviour, such as visual search and target tracking. The fast gaze shift in between fixations is thought to minimize image smear (Land, 2014). During the fixation phase of the saccades, the eyes of many vertebrates are counterintuitively not still, but make small fixational eye movements (e.g. microsaccades, ocular drift; Martinez-Conde & Macknik, 2008). Fixational eye movements have been found to prevent image fading (Riggs et al., 1953) but have also been suggested to have a function in perception of spatial information through dynamic visual sampling (Ahissar & Arieli, 2001; Ehud & Amos, 2012; Rucci et al., 2018).

Object motion

Object motion, or “local motion”, is retinal image motion that is restricted to a smaller area of the visual field (fig. 6c-d). It is important to most species since it often involves the presence of other animals (Frost, 2010). The detection of a predator, prey, or conspecifics, may cause for immediate action (Franconeri & Simons, 2003) and needs to be discriminated from other motion input at an early stage. Thus, object motion, in particular if it has a sudden onset or expands, is effective at catching the viewer attention (Abrams & Christ, 2003; Christ & Abrams, 2008; Pratt et al., 2010). Indeed, locally moving objects will “pop out” even against a background of optic flow (Rushton et al., 2007).

Effect of motion on contrast sensitivity and spatial acuity

Motion can have a considerable effect on some aspects of the visual image. Image motion may increase the sensitivity for luminance contrast, in particular for larger spatial structures (low spatial frequencies), while it typically decreases for finer structures (high spatial frequencies; Burr, 1981; Burr & Ross, 1982; Robson, 1966). At high velocities the finite integration time of photoreceptors can cause motion blur, which most strongly impacts small spatial details, while large structures become more conspicuous due to impaired lateral inhibition (Burr, 1981; Land & Nilsson, 2012; Lewis et al., 2011).

“I am not crazy; my reality is just different from yours”

Cheshire cat

Measuring spatial vision

The world surrounding an animal is often highly complex and can contain an endless amount of information. Even excluding parameters like wavelength composition and polarization of light, the visual information brought by spatial and temporal intensity changes is substantial within just a single field of view (Frazor & Geisler, 2006). The photoreceptors of any species samples only a fraction of the available light, which provide information that is further filtered and processed along the visual pathway before providing the animal with relevant information (Douglas & Cronin, 2016). Although a lot of image processing takes place already in the retina, the brain continues the analysis through many parallel pathways, integrating information from different locations in the visual field, but also from other sensory modalities and previous knowledge (Isa et al., 2021).

A species' natural environment and behaviour can provide insight into how it uses vision in different contexts and which stimulus parameters are most relevant to them. Furthermore, morphological traits, for example the size and placement of their eyes (e.g. if at the side of their head or at the front), often offer cues on sensory adaptation (Martin, 2017a). However, to find out the limits to what an animal can or cannot see, behavioural experiments are usually needed. Linking a visual stimulus to a behavioural (or sometimes physiological) response provides a robust indication that the animal can perceive the stimulus.

Quantification of visual stimuli

When measuring visual capacity, quantification of the physical components that make up the visual stimulus is required. For these parameters to accurately reflect the visual ability being tested, it is important that they are measured from the subject's point of view. For example, spatial distance is better measured by the angular subtense from the subject's field of view, rather than by absolute distance, since this is the information that reaches its eyes. Furthermore, quantification with objective units enables comparisons between species, but also with the physical characteristics of the habitat of the study species. In the next section I will introduce some of the more common ways of quantifying visual stimuli in animal visual research.

Light intensity

Depending on the purpose, light intensity is commonly measured in two functionally different ways. For example, if one aims to measure the ambient light intensity in a specific habitat, *illuminance* is a suitable measure. Illuminance is the luminous flux (amount of light per time unit) received by a surface, per unit of area (BIPM, 2019). The SI (Système international d'unités) unit for illuminance is *lux* (or $\text{candela}\cdot\text{sr}\cdot\text{m}^{-2}$)².

If one is interested in the light intensity of a visual stimulus, *luminance* is a suitable measure. Luminance signifies the amount of light, which is reflected from, or emitted by, surface and that reaches an observer from a specific viewing angle. The SI unit for luminance is $\text{candela}\cdot\text{m}^{-2}$, and it is defined as the amount of luminous flux per unit area which falls within a given solid angle (BIPM, 2019).

Illuminance and luminance are based on the *candela* (luminous intensity), which historically refers to the amount of light produced by a pure spermaceti³ candle (Johnsen, 2012). The candela, and units derived from it, are *photometric* units which are weighted for the spectral sensitivity of the human visual system. Other photometric units used in vision research include lamberts (Adler & Dalland, 1959; Blough, 1956), footcandles (Hersloff et al., 1974; Wells et al., 1975), and footlamberts (Blackwell, 1946), which can all easily be converted into $\text{candela}\cdot\text{m}^{-2}$ or lux.

An alternative to measuring light in photometric units, is to use *radiometric* units. Radiometric units are either based on the number of photons or the energy content of light and is in contrast to photometric units independent on the spectral sensitivity of the human eye (Johnsen, 2012; Land & Nilsson, 2012). In radiometric units irradiance ($\text{photons}\cdot\text{s}^{-1}\cdot\text{m}^{-2}$ or $\text{watts}\cdot\text{m}^{-2}$) is analogous to illuminance and radiance ($\text{photons}\cdot\text{s}^{-1}\cdot\text{sr}^{-1}\cdot\text{m}^{-2}$ or $\text{watts}\cdot\text{sr}^{-1}\cdot\text{m}^{-2}$) to luminance.

Since the spectral sensitivity differs between various animal species, a unit based on the spectral sensitivity of humans is not ideal. In the experiments included in this thesis we anyway chose to do measurements in photometric units. The main reason for this approach was to simplify comparison with the plethora of literature involving bird vision where light intensities are given in photometric units (e.g. Blough, 1956; Donner, 1951; Heeb et al., 2003; Hodos et al., 1976; Lind et al., 2012; Martin, 1977; Wesolowski & Maziarz, 2012). Furthermore, all stimuli in our experiments vary only in intensity and have the same overall broad spectral composition.

² Sr, steradian, is the unit of a solid angle subtended at the centre of a sphere, with the radius r , to a circular surface area r^2 .

³ Spermaceti is a waxlike substance found in the head of toothed whales (Odontoceti), especially the sperm whale (*Physeter macrocephalus*).

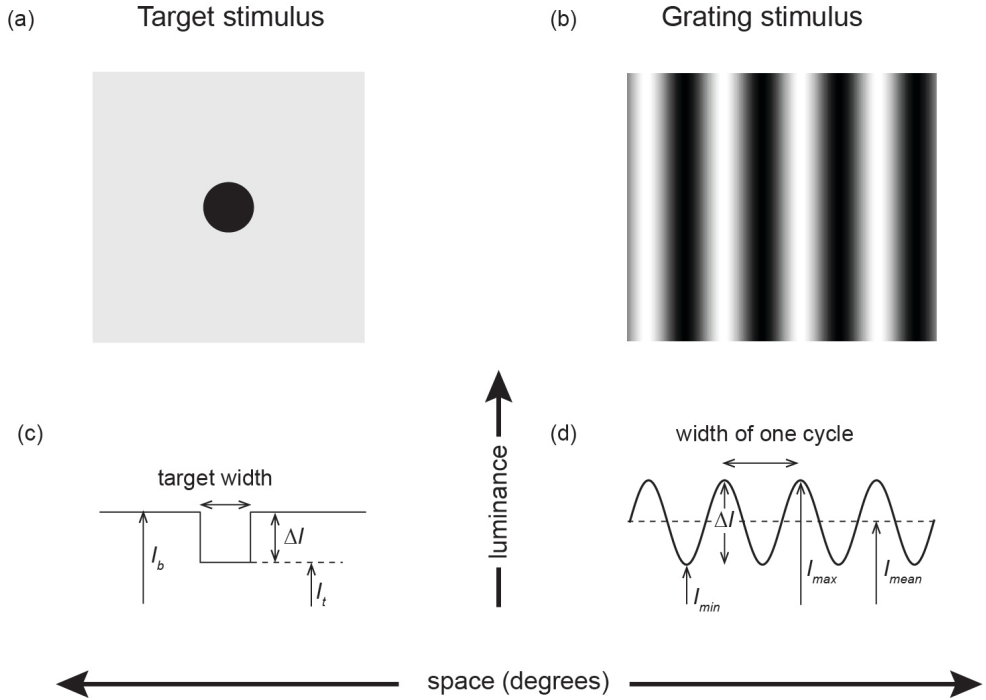


Figure 7. Luminance distribution of visual stimuli. Example of (a) a target (aperiodic) stimulus and (b) a sinusoidal grating (periodic) stimulus. The luminance profile of (c) the target stimulus and (d) the grating stimulus, illustrating how their luminance contrast and spatial extent is quantified.

Luminance contrast

The visual system is tuned to detect relative, rather than absolute, differences in light intensity (see “Luminance and contrast”). The luminance difference of visual stimuli is quantified in a similar way. Depending on the spatial distribution of light intensities in the stimulus, contrast can be defined as either *Weber contrast* or *Michelson contrast*. They both describe the magnitude of luminance variation relative to the overall luminance (Shapley & Enroth-Cugell, 1984). Weber contrast C_W , which is typically applied to the contrast between a smaller target and a uniform background (fig. 7a,c), is defined as:

$$C_W = \frac{I_t - I_b}{I_b} = \frac{\Delta I}{I_b} \quad (4.1)$$

where I_t is the luminance of the target and I_b is the luminance of the background. The definition of Weber contrast is based on Weber’s law (eq. 2.2), where $|C_W|$ is

equivalent to ω (Weber fraction). Weber contrast is applicable to stimuli where the background luminance have the main influence of the adaptive state of the eye.

If the dark and light areas occupy equal parts of the stimulus, they are assumed to affect the adaptive state of the eye to the same extent. The contrast of such stimuli are best represented by the Michelson contrast C_M :

$$C_M = \frac{I_{\max} - I_{\min}}{I_{\max} + I_{\min}} = \frac{\Delta I}{2I_{\text{mean}}} \quad (4.2)$$

where I_{\max} and I_{\min} are the maximum and minimum luminance values (fig. 7d).

Spatial structure of visual stimuli

The size of the retinal image of an object does not reflect its absolute size but rather the angle of visual space that it subtends. Spatial measures of visual stimuli, such as distance and resolution, is thus best described in angular subtense from the point of view of the test subject.

Periodic visual stimuli – grating stimuli

One of the most commonly used stimulus types when measuring the resolving power of the visual system is a grating stimulus (fig. 7b). The luminance of such stimuli varies periodically (i.e. according to a sinusoid or a square-wave) between a maximum and a minimum value, forming the light and dark bars in a grating. The use of grating stimuli facilitates the analysis of vision as a linear system (De Valois & De Valois, 1991). Through Fourier transformation, any visual stimulus can be decomposed into a combination of different sinusoidal wave functions with different amplitude (luminance difference), frequency (size) and phase (position in space). In the realm of linear systems analysis, the response to any visual stimulus is equal to the sum of the responses to each of its wave components. Similarly, it is possible to predict the response to any visual stimulus, if the response to each of its components is known. Since the basic components of Fourier transformation are sinusoidal waves, the simplest visual stimulus is a grating composed of a single frequency.

The resolution of a grating stimulus is quantified in *spatial frequencies*, which have the unit cycles degree⁻¹, where one cycle corresponds to one period of the fundamental wave function (one dark and one light bar in a grating; fig. 7b,d). It is assumed that a grating stimulus can be resolved as long as adjacent dark and bright stripes are sampled by the receptive field centres of separate retinal ganglion cells.

Grating stimuli are also used to measure the *contrast sensitivity function* (CSF), which describes the contrast sensitivity of the visual system as a function of spatial

frequency (De Valois & De Valois, 1991). The CSF typically has a band-pass shape, which means that contrast sensitivity is highest for intermediate spatial frequencies (fig. 8). Contrast sensitivity falls slowly for low frequencies, while the drop is comparably sharp for high frequencies. The function reaches the baseline at the cut-off frequency, which corresponds to the acuity limit. The general shape of the CSF for all animals tested is similar, although the position on the frequency axis, contrast sensitivity peak and cutoff frequency may vary (De Valois & De Valois, 1991; Souza et al., 2011).

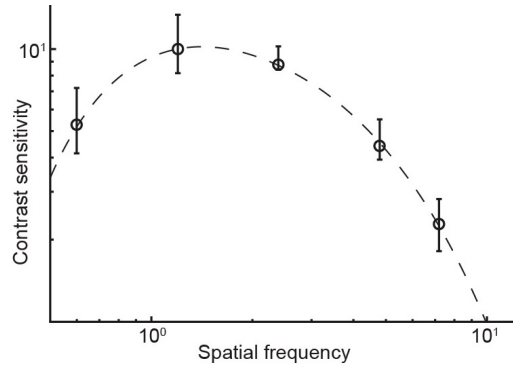


Figure 8. The contrast sensitivity function (CSF). The contrast sensitivity (in Michelson contrast⁻¹) of budgerigars as a function of the spatial frequency (cycles degree⁻¹) of a square-wave grating (cycles degree⁻¹) of a grating. Adapted from Lind and Kelber (2011).

Aperiodic visual stimuli – target stimuli

Aperiodic visual stimuli, or *target stimuli*, does not have a repeating pattern but instead one or several targets, for example dots, lines, or circles. The spatial properties of target stimuli are often quantified by their angular subtense (in degrees), because they constitute a discrete event. Spatial frequencies (cycles degree⁻¹) can also be an appropriate measure, for example when using targets that have been constructed from a discrete piece of a wavefunction.

Many classical studies on the interaction between area, luminance contrast, exposure time, and adaptational state on visual thresholds were conducted with target stimuli (Barlow, 1957, 1958; Blackwell, 1946; Blough, 1956; Hecht et al., 1947). The detection threshold for small uniform targets of high contrast, *single target acuity*⁴, can be utilised for making estimates of detection distances of ecologically relevant targets (Adrian, 1989; Champ et al., 2014; Hecht et al., 1947; Sandow & Hanke, 2024; Spratte et al., 2021), but also for studying the receptive field properties (i.e. spatial summation) of retinal neurons (e.g. Donner, 1987; Tuten et al., 2018; Volbrecht et al., 2000).

Although the detection threshold for uniform single targets is limited by contrast sensitivity (see: “Feature detection below the theoretical resolution limit”), it is possible to sidestep luminance cues by using isoluminant targets, which have the same overall luminance as the background. For such targets to be visible, the dark and

⁴ This measure is also known as single object threshold (Land, 1997), single object resolution, single target detection (Spaethe and Chittka, 2003) and minimum visible (e.g., Lythgoe, 1932; Donner 1951).

light regions need to be differentially sampled (i.e. sampled by both ON- and OFF receptive fields of retinal neurons). Examples of isoluminant targets are wavelet bar stimuli, composed of a single period of a wave function (difference of Gaussians: Kirwan, Bok, et al., 2018; Haar and piecewise sine: Kirwan, Graf, et al., 2018; Kirwan & Nilsson, 2019; Sumner-Rooney et al., 2020), and vanishing optotypes, which are constructed by providing a contrasting core to classic ophthalmological targets like tumbling E or Landholt C (Demirel et al., 2012).

Temporal structure of visual stimuli

Motion is spatial displacement in time, which means that moving visual stimuli have both spatial and temporal components. The temporal resolving power depends on the *critical duration time*, which is the time-frame over which incoming photons can be summed to create a visual signal (Donner, 2021). Temporal resolution is usually estimated using a light source with a periodically modulated intensity (Barten, 1999). Below the temporal frequency threshold, the visual system perceives the light as flickering, while above, the light is perceived as continuous (Donner, 2021). The frequency at which the light goes from flickering to continuous is referred to as the *critical flicker-fusion frequency* (CFF) and is measured in hertz or cycles second⁻¹.

The CFF is often used as a proxy for motion vision (Donner, 2021). Still, motion vision is not simply a sum of temporal and spatial vision but involves intricate retinal computations where these properties are entangled (Murphy-Baum et al., 2021; Schwartz & Swygart, 2021). Thus, stimuli for motion vision experiments most often have both spatial and temporal characteristics. A common stimulus type used for assessing motion vision involves drifting gratings or targets. The temporal aspect of target stimuli is typically quantified by angular velocity (degrees s⁻¹) while grating stimuli in addition can be quantified by temporal frequency (cycles s⁻¹).

Methods for measuring visual capacity

Visual capacity is commonly measured by the minimum perceptible stimulus intensity, the *absolute threshold*, or the minimum perceptible difference in stimulus intensity, the *difference threshold*. Visual thresholds can be assessed with behavioural experiments (*psychophysics*), or with electrophysiological measurements. Luminance contrast sensitivity and spatial acuity are also possible to estimate through modelling or calculations if specific physiological and anatomical parameters are known. Below, I summarise some of the most frequently used approaches to study luminance vision in vertebrates.

Modelling the perception of luminance contrast with the receptor noise limited model

Luminance contrast thresholds can be estimated based on photoreceptor noise and spectral sensitivity. The *receptor noise limited model* (RNL model) was originally developed for estimating colour vision thresholds (Vorobyev et al., 2001; Vorobyev & Osorio, 1998) but has since been adapted to work also for luminance vision (Siddiqi et al., 2004; Olsson et al., 2018).

In the RNL model, the discriminability between a target on a background is described by the *perceptual distance* ΔS defined as:

$$\Delta S = \left| \frac{\Delta f}{e} \right| \quad (4.3)$$

Δf is the *receptor contrast*, which is derived from the relative *quantum catch* (the number of photons absorbed by each photoreceptor) between the target and the background, and e is the receptor noise of the luminance channel. ΔS is described in terms of *just noticeable differences* JNDs, and $\Delta S=1$ JND at the visual threshold.

Estimating spatial acuity from the retinal mosaic

The resolving power of an eye depends largely on the retinal sampling density and the posterior nodal distance (PND; see “Retinal factors affecting spatial acuity”). Thus, these measures can be used to make an estimate of the spatial acuity of an eye.

Generally, retinal ganglion cell (RGC) density is used as a proxy for retinal sampling density. The signal from several photoreceptors often converges on the same ganglion cell, whose axon forms the only connection between the retina and the brain (Pettigrew et al., 1988). In cases where RGCs outnumber photoreceptors, or there is a 1:1 relationship, photoreceptor density may be used instead. An additional exception is for species with a *fovea* (retinal invagination: see “Retinal topography”), where RGCs are “displaced” making it difficult to estimate their local density (Coimbra et al., 2015).

RGC and photoreceptor densities are estimated from cell counts in selected retinal areas. Cell counts are done either on retinal wholemounts, or on a combination of wholemounts and cross-sections, for regions in which RGCs are organized in many layers (Mitkus et al., 2014). Typically the spatial resolving power is calculated from the region(s) with the highest density of sampling units.

The retinal magnification factor (RMF) is defined as the retinal distance corresponding to 1° of the visual field, and is calculated as

$$RMF = \frac{2\pi PND}{360} \quad (4.5)$$

The maximum resolving power (F) of the eye can then be estimated by:

$$F = \frac{RMF}{2} \times \sqrt{\frac{2D}{\sqrt{3}}} \quad (4.6)$$

where D is the peak density of sampling units (cells mm^{-2}), and F is expressed in $\text{cycles degree}^{-1}$ (Snyder & Miller, 1977; Williams & Coletta, 1987).

Electroretinogram

Electrophysiological measurement of visual thresholds can be made at different processing levels along the visual pathway. However, responses to basic physical stimulus parameters, like acuity and contrast, are usually measured at the retinal level using a method called the *electroretinogram* (ERG). Using this method, a small electrode, in contact with the cornea, measures the electric activity generated by the retinal neurons as the subject is presented with a visual stimulus. The ERG amplitude is plotted as a function of stimulus intensity, and the threshold is obtained by extrapolating the function down to the “noise level” (= electric potential recorded in the absence of stimuli; Hodos, 2012).

Flash ERG is generated from the presentation of a spatially homogenous test field that produces flashes of light. This method is often used to measure the absolute sensitivity to light (Hodos, 2012), a periodically modulated flash can also be used to measure the CFF (e.g. Lisney, Ekesten, et al., 2012).

The stimuli used in *pattern ERG* varies in both space and time, many times a counter-phase modulated grating. Pattern ERG can be used for testing spatio-temporal contrast sensitivity.

Psychophysics

Psychophysics is defined as the science of relating physical stimuli to a sensation (Gescheider, 1997). Since a sensation by itself cannot be objectively measured, it needs to be approximated with something which is. If the perception of a sensory stimulus is linked, either via an innate mechanism or associative learning, to a

specific behavioural (or sometimes physiological) response, this response can be used as a proxy for sensation.

Behavioural experiments usually provide the most robust measure of visual perception. The methods described above estimate or measure an upper limit of vision at the retinal level, without considering the processing that occurs further up the visual pathway. Behavioural experiments, in contrast, demonstrates the existence of a link all the way from retinal detection of a visual signal to a behavioural output. Behavioural responses employed in animal visual psychophysics may range from simple innate reflexes, like the visual fixation of new objects, to more elaborate experimentally learned behavioural repertoires.

Innate responses to visual stimuli

Most animal species have innate behavioural responses that can be induced by visual stimulation (e.g. reflexes, taxes, fixed action patterns). Some responses have a long evolutionary history and are present in entire phyla (Land, 2019), while others have developed to suit the specific needs of single species (Tinbergen & Perdeck, 1950; Williams, 2022).

Phototaxis, a directional movement in response to a light stimulus, might be the oldest innate behavioural response to light and is found in unicellular organisms as well as in vertebrates (Jékely, 2009; Land & Nilsson, 2012). The phototactic response has been utilized to measure visual thresholds in a range of species for example the common diving petrel (*Pelecanoides urinatrix*: Brooke, 1989) and frogs (*R. temporaria* and *R. pipiens*: Aho, Donner, & Reuter, 1993).

Moving or looming visual targets tend to capture attention. In species hunted by aerial predators, a target moving or looming can induce an innate defence response (e.g. escape- and freeze response; Carlile et al., 2006; De Franceschi et al., 2016; Hébert et al., 2019; Marquez-Legorreta et al., 2020). For a predatory species, in contrast, a moving target can induce prey-catching response (Bianco et al., 2011; Ewert et al., 2001). Although defence and prey-catching responses often are highly context dependent, they can be used in vision experiments. The spatial acuity of mice have been assessed by their innate defence response to looming target stimuli (Storchi et al., 2019), while the luminance sensitivity of toads (*Bufo bufo*) was measured using their prey-catching response triggered by moving targets (Aho, Donner, Helenius et al., 1993).

In vision research, the most widely used innate response is likely what is referred to as the *optokinetic*, *optocollic*, or *optomotor response* (depending on whether the subject moves its eyes, head, or body; Land, 2019; Wagner et al., 2022). This reflexive response has the function to stabilize vision and can be found in almost all vertebrates. The optokinetic-, optocollic-, or optomotor response and can be induced by rotational optic flow (see “Self-induced motion”). Typically, the subject (if sufficiently small) is placed inside a device called an “optomotor cylinder”, which has

a vertically oriented grating at the inside wall. Rotation of the cylinder around the subject elicits a reflex if the grating can be seen, but not if the cylinder is still or if the stimulus is below the visual threshold.

Other behavioural responses related to self-induced motion can be studied by letting the subject itself move in a stationary experimental arena. Behaviours that rely on cues from retinal image motion is then studied under controlled changes in stimulus parameters such as contrast or spatial resolution. The influence of translational optic flow on locomotion has been investigated in several vertebrate species trained to move through a tunnel with grating stimuli on the walls (Bhagavatula et al., 2011; Dakin et al., 2016; Kugler et al., 2019; Scholtyssek et al., 2014).

Methods using innate responses to visual stimuli allow for comparatively fast collection of data and seldom require training of the subject. However, only a narrow range of visual stimuli elicit innate behaviours, and the threshold for eliciting a behavioural response is not necessarily the same as the sensory threshold. In fact, visual thresholds can be context dependent and differ between different behavioural realms (Yovanovich et al., 2017). Assessment of thresholds for specific parameters might be further complicated if the response depends on a combination of several stimulus parameters, and the change in one parameter might result in a lack of response or even in a different response (Bianco et al., 2011; Carlile et al., 2006; De Franceschi et al., 2016; Ewert et al., 2001; Hébert et al., 2019; Procacci et al., 2020; Solomon et al., 2023). Innate responses might thus not necessarily reveal the absolute sensory threshold of a subject, although they will likely better reflect the sensory constraints met in a specific behavioural context.

Classical and instrumental conditioning of visual stimuli

When testing the threshold for visual stimuli that do not elicit any innate response in the subject, *conditioning* can be an alternative method. *Classical conditioning* (also Pavlovian conditioning, after I.P. Pavlov [1849–1936]) means that a subject is trained to associate one stimulus (the *conditioned stimulus*) with another stimulus (the *unconditioned stimulus*) which naturally triggers an innate reflex (the *conditioned response*). By conditioning the visual stimulus of choice, the presence or absence of the conditioned response can be used to evaluate visual capacity (Blake, 1998). Examples of unconditioned stimuli (and associated conditioned responses) are, brief electric shocks (increased heartrate), air-puffs to the eyes (blinking), or delivery of food item (increased salivation) (Blake, 1998; Haug & Florsheim, 2010).

Although classical conditioning is applied to animal psychophysics, *operant conditioning* (or instrumental conditioning) is a more common approach. In operant conditioning the subject is trained to elicit a specific behaviour (the *response*) when presented with a specific stimulus. A *reinforcer*, which can be a reward (*positive reinforcer*, e.g. food), or absence of aversive stimulation (*negative reinforcer*, e.g. an electric shock), following the response, will increase the prevalence of the

response to the stimulus (Skinner, 1957). This sequence, stimulus → response → reinforcement, was referred to as a “three-term contingency” by behaviourist B.F. Skinner (1903-1993). In contrast to reinforcement, *punishment* will decrease the prevalence of a behaviour. In psychophysical experiments the behaviour and motivation of the subject can be shaped by reinforcing the response to one stimulus, while punishing the response to another stimulus. Common punishments are prolonged waiting time between trials (positive punishment) or simply the absence of a reward (negative punishment; Haug & Florsheim, 2010; Mora et al., 2009).

The test subject can be trained to perform either a *single response* or not, or to make a *choice* between two (or more) responses, when presented with a stimulus. The *go/no-go* method is an example of a single response method. The subject is presented with one stimulus at a time and trained to elicit a response (e.g. pressing a key or make an oriented movement) if it identifies it as the “correct stimulus” (S+) and to withhold the response if it identifies it as the “incorrect stimulus” (S-; Blough & Blough 2022).

In the *yes/no* method, which is a choice method, the subject is also presented with a single stimulus at a time. However, unlike in the *go/no-go* method, the subject in the *yes/no* method is expected to elicit one response (e.g. press the green key) in the presence of S+ and different response (e.g. press the red key) in the presence of S- (Blough, 1956; Hodos et al., 2002).

When applying the *forced-choice* method, several stimuli are presented simultaneously, and the subject is trained to identify which one of them is the S+ and make a response that indicate its choice (Gescheider, 1997). The *two-alternative forced choice* (2AFC) procedure, in which two stimuli (one S+ and one S-) are displayed simultaneously in each trial, is extensively used in animal vision psychophysics (Blough & Blough 2022), including the experiments in this thesis. The S+ and S- is usually displayed side by side, with their relative position varied pseudo randomly between each trial to avoid unwanted cueing. The response indicates the position of the S+ (e.g. pressing the right/left key or make an oriented movement).

Stimulus presentation

In psychophysical vision experiments, the subject is presented with a series of stimuli of varying intensities ranging from well below to well above its visual threshold. The threshold is not considered as a fixed intensity above which all stimuli are correctly identified. Rather, it is the intensity at which the subject can correctly identify a stimulus with a predefined likelihood (usually somewhere between “chance level” and correct identification nearly all the time; Gescheider, 1997). The likelihood of making a correct stimulus identification (for a specific intensity) is estimated from the proportions of correct stimulus identifications made during the experiment.

The stimulus intensity (“the level of difficultness”) can be alternated from trial to trial according to various sequential methods. Those most commonly applied in

animal vision research are based on a few classic methods developed by *G.T. Fechner* (1801-1887) (Gescheider, 1997; Malone, 2017). One of these are the *method of constant stimuli*, where a fixed set of stimulus intensities are repeatedly presented to the subject. The set commonly includes between five and nine stimulus intensities, ranging from just below the sensory threshold to well above it, which are presented in a random or semi-random sequence (Gescheider, 1997). Every intensity is tested many times throughout an experiment to obtain a ratio of correct stimulus identification at each intensity level. A *psychometric function* which expresses the likelihood of correct stimulus identification as a function of stimulus intensity, is then fitted to the data. The shape of the psychometric curve is sigmoid, with the lower asymptote at the ratio correct identifications below threshold by chance, and the higher asymptote at the ratio correct identifications well above threshold. The threshold intensity is usually found at the point of the psychometric curve that is halfway between the two asymptotes (e.g. 0.75 in a 2AFC; Gescheider, 1997).

The *method of limits* starts off with an intensity which is either well above (the “descending series”) or below the sensory threshold (“ascending series”). In the descending series, the stimulus in each successive trial is slightly lower than the previous one until the subject fails to identify the S+, at which point the test is terminated. In the ascending series instead, the stimulus intensity increases with each trial, until the subject can identify the S+ stimulus (Gescheider, 1997). The threshold is usually defined by averaging the stimulus intensity of the last two trials (correct identification ↔ incorrect identification) of a series.

The *staircase method* is a modification of the method of limits (Gescheider, 1997; Levitt, 1971). This method begins as a descending series, only the test is not terminated when the subject fails to identify the S+. Instead, the direction of change in intensity is reversed. In other words, the intensity of the subsequent stimulus will have increased. A correct stimulus identification will again make the series descend, and so it continues throughout the experiment. A change in direction (descending ↔ ascending) is called a reversal, and usually a pre-defined minimum number of reversals must occur before a test sequence is terminated (Levitt, 1971).

The experiments described in the articles that are part of this thesis were performed using the *1-up/2-down staircase method*. This is a variant of the staircase method where the stimulus intensity decreases after *two* consecutive correct responses but increases after only *one* incorrect response (fig. 9; Levitt, 1971). The stimulus intensity will eventually oscillate around the threshold level, where the probability of a descending step (two consecutive correct responses) is the same as that of an ascending step (one incorrect response). The threshold intensity is calculated from the mean value of the intensity at the reversals, which corresponds to the intensity that the subject can identify with 70.7% probability. (Levitt, 1971).

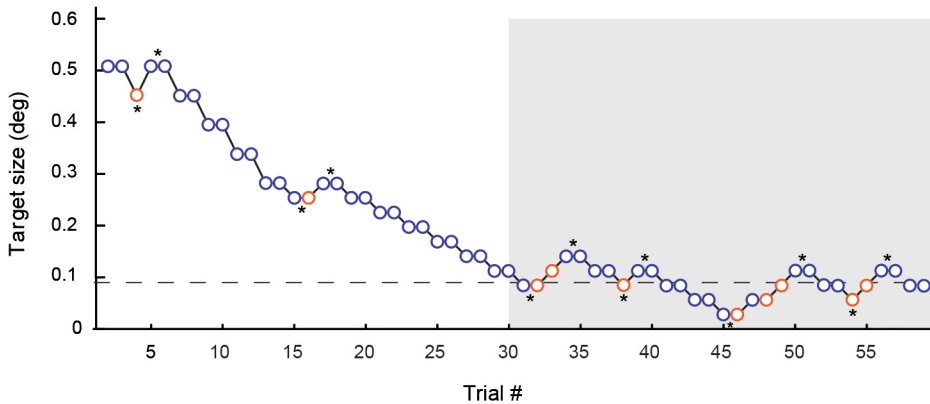


Figure 9. The 1-up/2-down staircase method. The stimulus intensity (in this example, target size) of the initial trials in a staircase session are well above the threshold. After two consecutive correct responses (blue circle) on the same intensity level, the stimulus intensity level decrease, while one incorrect response (red circle) is followed by an increase of stimulus intensity. Reversals (increase → decrease or decrease → increase) are indicated by asterisks. The threshold level (dashed line) is in this example calculated from the 8 last reversals (indicated by the grey background).

In the *method of adjustment*, which is one of Fechner's original methods, the test subject itself controls the increment or decrement of the stimulus intensity. The subject starts at a random intensity level and adjusts it gradually until the threshold is reached (Gescheider, 1997).

The *threshold tracking method* developed for testing hearing in humans (Békésy, 1947) resembles both the method of adjustment and the staircase method. It was later adapted by Blough (1955) to track visual sensitivity of pigeons. The pigeon was trained to peck at one key when a bright patch was present (S+) and to peck at another key when the bright patch was absent (S-), in a yes/no procedure. A peck on the first key would lower the luminance of the patch, while a peck on the second key would raise the intensity of the patch, causing it to fluctuate around the threshold intensity throughout the experiment. This adapted method has been used to track visual threshold curves during dark-adaptation in several species (Adler & Dalland, 1959; Blough, 1956; Hersloff et al., 1974; LaMotte & Brown, 1970; Wells et al., 1975).

Training animals to perform in psychophysical experiments is often time-consuming. Furthermore, sometimes the experimental subject continues to improve its visual performance even after a task has been learned (Blough & Blough, 2022; Blough, 1971; Chaib et al., 2019; Chaib et al., 2021; Ghim, 2003; Sandow & Hanke, 2024). This phenomenon is commonly referred to as the *learning effect* and may persist for a few trials or sometimes several months (Ghim, 2003; Gilbert, 1994). The learning effect is in some instances a consequence of actual improvement of sensory perception (i.e. *perceptual learning*; Tsushima & Watanabe, 2009).

We experienced a learning effect in all experiments included in this thesis. Despite successful training of the bird subjects for specific experimental tasks, we would notice a gradual improvement of the performance once we started running the staircase test sessions. The birds would reduce their threshold over several consecutive test session (in the experiments in paper I up to 9 tests sessions) before reaching a stable plateau of performance. We do not know if the birds increased their performance because of perceptual learning, or if they improved their ability to focus on the experimental tasks. A similar learning effect was noticed in single target acuity experiments with both the common sunfish (*Lepomis gibbosus*; Spratte et al., 2021) and harbour seal (*Phoca vitulina*; Sandow & Hanke, 2024).

Spatial vision in birds

The avian eye

For most species of birds, vision is the primary sense, and as a group they rely more on visual information than any other vertebrate class (Martin, 2017a). The high importance of vision is reflected in the anatomy of the avian visual system in several ways. For example, birds typically have large eyes that occupy a considerable portion of the cranial volume (Shimizu & Watanabe, 2012), and the part of their brain devoted to processing of visual information is greater than in other animals (Martin, 2017a). In addition, the avian retina is among the most complex of all and it expresses a large variation between species (Hart, 2001a; Meyer, 1977; Walls, 1942).

Photoreceptors

In common with most other vertebrates, the retina of birds has two major classes of photoreceptors, *cones* and *rods*. Both rods and cones are elongated cells which can be divided into an inner segment, containing the nucleus, organelles and as synaptic terminal, and an outer segment, which houses the visual pigments. Cones mediate vision at daylight, while they lose their function at night. Rods, on the other hand, are about 25-100 times more sensitive than cones (Martin, 2017a). This means that they work at significantly lower light intensities but also that they are saturated in daylight.

Most birds have four spectrally distinctive types of single cones which enable them to have tetrachromatic colour vision (Kelber, 2019). The various single cones are mainly characterized by their different pigments which makes them sensitive to light at different wavelengths; the V-cone have a λ_{\max} (peak absorbance) at 355-424 nm, the S-cone at 427-463 nm, the M-cone at 497-514 nm, and the L-cone at 505-630 nm (Hart & Hunt, 2007). The spectral sensitivity of bird cones is further affected by the oil droplet, a spherical organelle located at the distal end of the inner segment, through which incoming light is filtered before reaching the outer segment (Toomey & Corbo, 2017). V-cones have transparent oil droplets that are thought to increase the light catch of the outer segment (Wilby & Roberts, 2017). The other three types of single cones (S, M, and L) have carotenoid-containing oil droplets that act as

optical long-pass filters, improving colour contrast at the expense of overall sensitivity (Toomey & Corbo, 2017; Wilby & Roberts, 2017).

In addition to the four types of single cones birds also have one type of *double cone*. Double cones are present in most vertebrate groups where they show a great diversity in their pigment content and morphology (Bowmaker, 2012). In birds, double cones consist of one larger principal member and one smaller accessory member, which are thought to be optically and electrically coupled (Hart & Hunt, 2007). Both members express the same pigment as the L single cone (LWS opsin). However, while the pigmented oil droplet of the L single cone shifts its λ_{\max} to longer wavelengths, the principal member of the double cone has a clear, or almost clear, oil droplet which likely transmits a larger fraction of the incoming light (Wilby & Roberts, 2017). The accessory member most often lacks an oil droplet completely (Hart, 2001b).

Although the double cone is the most abundant photoreceptor in the retina of diurnal birds their function is not fully understood. Likely, they serve a function in luminance-mediated vision but not in spectral discrimination (Kelber, 2019; Goldsmith & Butler, 2005). Behavioural experiments indicate an involvement in the perception of luminance contrast and fine texture (Jones & Osorio, 2004; Lind & Kelber, 2011), although high-resolution vision likely also involve input from single cones (Lind & Kelber, 2011; Mitkus et al., 2017; Seifert et al., 2023). Motion perception, which is likely driven by luminance cues, is another suggested function of double cones (Bhagavatula et al., 2009; Campenhausen & Kirschfeld, 1998; Seifert et al., 2023; but see: Sun & Frost, 1997).

One of the difficulties with studying the function of double cones is that their λ_{\max} lies between that of M and L single cones. This makes it difficult to distinguish double cone stimulation from a weighted sum of M and L single cone stimulation (Osorio et al., 1999). In any case, double cones form multiple retinal networks, both with rods and single cones, indicating that they play a role in multiple visual channels (Günther et al., 2021; Seifert et al., 2020).

Retinal topography

In common with most vertebrates, the retinas of birds are not functionally homogeneous. Ganglion cell density, photoreceptor composition, and retinal wiring vary with spatial location (Hart, 2001a). Different parts of the retina sample light coming from different directions in the visual field. As the light from these directions usually differs in terms of for example spectral composition, contrast, mean luminance (Nilsson et al., 2022), as well as temporal aspects (Martin, 2017b), different parts of the retina need to fulfil different requirements.

Retinal ganglion cell topography

Retinal ganglion cell (RGC) density, which is associated with spatial acuity (see “Retinal factors affecting spatial acuity”), often varies substantially across different retinal locations (Martin, 2017a; Meyer, 1977). Although the topographic variation of RGC density follows a general pattern, it also differs between species and has been shown to correlate with factors such as habitat structure, foraging technique and vulnerability to predators.

Birds typically have one or two retinal regions with elevated ganglion cell density, referred to as *areae* (Meyer, 1977). The *area centralis*, which as the name suggests is located in the central retina, is the most prevalent. Since the eyes of most birds are located on the side of the head, the *areae centralis* of the two eyes are oriented laterally (with a horizontal angle slightly less than 90° to the midline of the beak) and thus view separate parts of the world.

Many species have an additional *area*, the position of which can vary but which is usually directed to the frontal visual field. This type of *area* typically has a temporal or dorso-temporal placement in the retina (*area temporalis* or *area dorso-temporalis*), and is associated with hunting of live prey, or a need for fine-tuned bill control (Coimbra et al., 2014; Coimbra et al., 2009; Lisney, Iwaniuk, Bandet, et al., 2012; Lisney et al., 2015; Potier, Mitkus, et al., 2020; Tyrrell & Fernandez-Juricic, 2017).

Increased RGC density is also often seen as an elongated horizontal *area* across the retina and is then referred to as a *visual streak*. According to the “terrain theory”, proposed by Hughes (1977), the visual streak is an adaptation in animals that occupy open habitats and provides them with a panoramic view of the free horizon. Studies of the retinas of birds have, on the other hand, provided inconclusive support for the terrain hypothesis (Lisney, Iwaniuk, Kolominsky, et al., 2012).

Budgerigars forage on the ground in a predominantly open habitat and should therefore, according to the terrain hypothesis, possess visual streaks. Topographical mapping of their RGC density nevertheless revealed only a weak visual streak in one of five retinas (Mitkus et al., 2014). The same study also found no visual streak in the retina of the closely related Bourke’s parakeet (*Neopsephotus bourkii*) which occupies the same habitat type. Both species possessed an *area centralis* while the budgerigars also had an *area nasalis* that projected slightly backwards in the visual field (fig. 10). Since budgerigars use their beak when climbing the *area nasalis* is suggested to be used in visual scanning for predators as the head mobility is constrained during this activity (Mitkus et al., 2014). Although budgerigars often forage on grass seeds that have fallen to the ground, they frequently climb up directly on sturdier grass plants (Higgins, 1999).

Cockatoos are, like budgerigars, seed eating psittacines that live in Australia. However, unlike budgerigars, cockatoos have been found to possess visual streaks

(Coimbra et al., 2014). Five of six species that have been studied also had a dorso-temporal *areae*, assumed to project into the frontal visual field. As these same species are known to use their feet to grasp and manipulate food items during foraging, the enhanced acuity in the frontal visual field is thought to serve a purpose in such activities (Coimbra et al., 2014). Only the cockatiel (*Nymphicus hollandicus*), which like budgerigars and Bourke's parrots, only use their beak in food manipulation, were found to lack a dorso-temporal *area*.

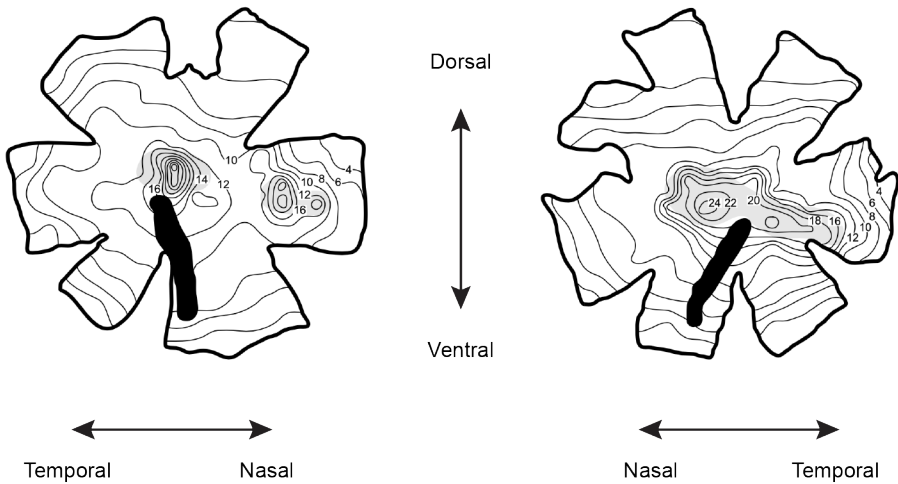


Figure 10. Retinal ganglion cell (RGC) topography in the budgerigar. Numbers represent $\times 1000$ cells mm^{-2} , grey shading indicates regions with RGCs stacked in layers, and black bars the position of the pecten oculi. Adapted from Mitkus et al. (2014).

The rock pigeon is another seed-eating species that, like cockatoos, has a retinal *area* that is directed into the frontal visual field (Querubin et al., 2009). Although rock pigeons are ground foragers, temporal or dorso-temporal *areae* are generally absent in species with this foraging practice (i.e. European starling [*Sturnus vulgaris*], brown-headed cowbird [*Molothrus ater*], house sparrow [*Passer domesticus*], house finch [*Carpodacus mexicanus*] and mourning dove [*Zenaida macroura*]: Dolan & Fernández-Juricic, 2010; tree sparrow [*Passer montanus*]: Rahman et al., 2006; seven phasianid species: Lisney, Iwaniuk, Kolominsky, et al., 2012); peafowl [*Pavo cristatus*]: Hart, 2002; red-winged blackbird [*Agelaius phoeniceus*]: Fernández-Juricic et al., 2019).

The RGC density distribution in the budgerigar, as well as Bourke's parrot, expresses a high inter-individual variation (Mitkus et al., 2014). Possibly, this is a consequence of domestication and human-controlled breeding, as the individuals examined in the cited study were not wild-caught. Phenotypic traits that exert a high

selection pressure on a species in its natural habitat (e.g. traits associated with predator avoidance or foraging) have been observed to exhibit a substantial variability in domesticated individuals (Mignon-Grasteau et al., 2005). Furthermore, Lisney et al. (2011) found evidence that domestication might have affected the temporal visual resolution of the chicken negatively. On the other hand, the chicken has not only been domesticated for a much longer time (~3500 years ago) than the budgie (less than 200 years ago) but has also been subjected to a higher artificial selection pressure, as it is used as livestock. Although the visual capacity of wild and domestic budgerigars has never been compared, their acoustic sense has been shown to be unaffected by domestication (Farabaugh et al., 1998).

Photoreceptors

As with RGCs, the distribution of photoreceptors also shows a high variation between bird species as well as across the same retina. The overall density of photoreceptors roughly follows the topographical pattern of RGC, although the photoreceptor to RGC convergence ratio is comparatively low in high density regions (Querubin et al., 2009).

The photoreceptor population of nocturnal owls consists mainly of rods (Fite, 1973; Lisney, Iwaniuk, Bandet, et al., 2012), while diurnal birds typically have a cone-dominated retina (Hart, 2001b). The *area* and *fovea* of many diurnal species have even been shown to lack rods entirely, presumably to accommodate a greater number of the smaller cones (Coimbra et al., 2015; Querubin et al., 2009).

The most prevalent cone type in diurnal birds is the double cone, which may constitute over half of the cone population in some species (Hart, 2001b). Many ground-foraging species (including the budgerigar) have the highest proportion of double cones in the ventral region of the retina, while arboreal species have the most double cones in the dorsal region (Hart, 2001a). This difference has been suggested to be an adaptation for detecting predators: while ground-foraging birds are vulnerable to airborne threats, arboreal birds often face attacks from below (Hart, 2001a). Despite their prevalence, double cones are absent in the central the *fovea* of some raptor species (Mitkus et al., 2017).

Foveae

In many species, the *area* or visual streak is accompanied by a *fovea*, which is an invagination in the inner layers of the retina (Bringmann, 2019; Meyer, 1977; Walls, 1942). As *foveae* overlies densely packed photoreceptors, with low RGC convergence ratio, they are commonly assumed to be involved in mediating high acuity vision. However, the complete function of the *fovea* is debated, some suggestions being the reduction of light scattering, image magnification, movement detection, and “focus indication” (reviewed in: Bringmann, 2019; Moore et al., 2017). *Foveae* located in the centre of the retina are widely distributed taxonomically and have been documented in species belonging to most lineages, such as raptors, psittacines,

passerines, fulmars, and Columbiformes (Bringmann, 2019). Some birds, such as the sacred kingfisher (*Halcyon sancta*), the laughing kookaburra (*Dacelo novaeguineae*), the least tern (*Sternula antillarum*), as well as most actively foraging raptors (Accipitriformes and Falconiformes), have both a central and a temporal *fovea* (Bringmann, 2019; Mitkus et al., 2017; Moroney & Pettigrew, 1987; Potier et al., 2016; Potier, Mitkus, et al., 2020). Having a temporal *fovea* without having a central *fovea* is comparatively rare but occurs in the common swift (*Apus apus*) and most species of owl (Strigiformes) (Bringmann, 2019; Fite, 1973; Lisney, Iwaniuk, Bandet, et al., 2012).

The visual field and eye movements in birds

Birds have their eyes positioned on the sides of their head. As a result, their visual field extends laterally around the head, typically leaving only a small “blind angle” at the back. Usually, the visual fields of the individual eyes have a small binocular overlap at the front, while most of the visual field is seen monocularly. The visual field variation seen in various species is suggested to be a product of primarily foraging method but also of the need for predator detection (Martin, 2017b).

When a bird spots a target of interest it will typically move it into either one of its central *areae/foveae* (*lateral fixation*) or the frontal visual field (*frontal fixation*). In general, lateral fixation is used for targets that are further away, while frontal fixation is used for targets nearby (Bloch & Martinoya, 1982; Kano et al., 2022; Martin & Katzir, 1999; Martinoya et al., 1983; Rounsley & McFadden, 2005). Many species have a refractive state that varies across the field of view: while the frontal visual field is myopic, the lateral visual field is emmetropic (Fitzke et al., 1985; Hodos & Erichsen, 1990). This means that they do not have to accommodate when switching between frontal fixation and lateral fixation, but also that they can forage on the ground while on the same time scan their surroundings for predators.

The frontal and lateral visual fields of birds differ not only in terms of optimal viewing distance but also in functionality. For example, moving targets are preferentially fixed by the lateral visual field, which is thought to be better adapted for predator detection (Evans et al., 1993; Maldonado et al., 1988). Indeed, information from frontal and lateral visual fields are associated with different processing pathways that are thought to handle separate aspects of visual information (Clark & Colombo, 2022; Güntürkün & Hahmann, 1999). Intraocular transfer is likely also restricted (Jimenez Ortega et al., 2008; Remy & Emmerton, 1991; Roberts et al., 1996), and birds often alternate between different parts of the visual field when inspecting unknown objects (Kano et al., 2022; Stamp Dawkins, 2002).

When a bird fixates a target in their frontal visual field, both eyes make a converging movement (Bloch et al., 1984; Martinoya et al., 1984). Frontal fixation is often made in association with pecking or lunging at a target, suggesting that the frontal visual

field is used for visual control of the beak and feet (Kano et al., 2022; Martin & Katzir, 1999; Martinoya et al., 1983). A larger degree of binocular overlap has been observed in species with a need for precise control of the bill or feet, primarily in foraging, but also in provisioning of young, and in nest construction (Martin, 2017b). Passerines generally have the widest binocular overlap, and the tool-using New Caledonian crow (*Corvus moneduloides*) has a maximum overlap of a full 61° (Troscianko et al., 2012).

It has been proposed that the function of the binocular overlap in birds is to provide depth perception by stereopsis. Although stereopsis has been demonstrated in the barn owl (*Tyto alba*; van der Willigen, 2011; van der Willigen et al., 1998) it is likely not widespread among birds. The binocular overlap may also increase visual sensitivity, which is important in nocturnal species that often have frontally oriented eyes (Read, 2021). Still, the primary purpose of frontal fixation is not necessarily the binocular overlap per se. The visual field of symmetrically converging eyes enables an expanding optic flow field in the direction of travel, which for example can be used for guidance of the beak (Martin, 2009). The binocular overlap has also been proposed to be a secondary consequence of minimizing the anterior blind area; viewing items in, or close to, the beak indirectly requires a wide binocular overlap (Tyrrell & Fernández-Juricic, 2017).

Species that are not at the top of the food chain must balance the need for binocular vision with having a wide cyclopean visual field (binocular + monocular visual fields) for predator detection (Martin, 2017b). A wider cyclopean field is often found in species which primarily rely on tactile senses when foraging (e.g. filter-feeding, or dabbling ducks, and shorebirds) and thus do not need to have precision control of the bill (Cantlay et al., 2023; Martin, 2017b). The Eurasian woodcock (*Scolopax rusticola*) and the mallard (*Anas platyrhynchos*), for example, both have total panoramic visual fields, and comprehensive visual coverage of the hemisphere (Martin, 1986, 1994).

Psittacines are extractive foragers, and although they use vision to locate food (mostly seeds, nuts and fruits), the beak, tongue, and in some species the feet, are used to extract the embedded eatable parts. At the tip of the upper mandible of parrots there are touch receptors (the “bill-tip organ”), which are used in food handling and object exploration (Martin & Martin, 2022). The visual field has so far only been measured in one psittacine, the Senegal parrot (*Poicephalus senegalus*), but the configuration is likely similar in closely related species (Martin & Martin, 2022). The Senegal parrot has a comparatively a wide frontal binocular overlap, but also a near total panoramic view above the head (Demery et al., 2011). The beak is located at the edge of the frontal binocular field, meaning they cannot see things held in it. Likely, the bill-tip organ of parrots compensates for a more comprehensive visual field around the beak, which instead extend above the head. Nevertheless, the Senegal parrot has a rather broad binocular overlap above the beak which allows visual inspection of objects up close (Demery et al., 2011).

Contrast sensitivity

Although birds have remarkably good vision in many ways, their contrast sensitivity is relatively low. Measurements of the maximum contrast sensitivity in different species range between 4.6 and 31 Michelson contrast⁻¹ (Blary et al., 2024; Ghim & Hodos, 2006; Haller et al., 2014; Harmening et al., 2009; Hirsch, 1982; Hodos et al., 2002; Jarvis et al., 2009; Lind et al., 2013; Lind et al., 2012; Potier et al., 2018; Raymond & Wolfe, 1981). In comparison, fishes have contrast sensitivities between 33 and 125 (Bilotta & Powers, 1991; Northmore & Dvorak, 1979; Northmore et al., 2007; Santon et al., 2019), primates between 90 and 200 (De Valois et al., 1974; Jacobs, 1977), and the domestic cat (*Felis silvestris*) 116 Michelson contrast⁻¹ (Bisti & Maffei, 1974). Among birds, the highest contrast sensitivities are found in raptors, notably Falconiformes species, but also in the raven (*Corvus corax*) (Blary et al., 2024; Hirsch, 1982). Why birds have such a low contrast sensitivity in general is not known but has been suggested to be a trade-off for other visual capacities such as UV-sensitivity (Blary et al., 2024; Ghim & Hodos, 2006).

The contrast sensitivity function (CSF) in birds can show some variation depending on the method applied. CSFs obtained from pattern electroretinogram (PERG) show a lower (by $\sqrt{2}$) peak sensitivity compared to behavioural experiments with operant conditioning (Hodos et al. 2002). Furthermore, studies that have used the optocollic reflex generally describe CSFs that are tuned to lower spatial frequencies than those that have used operant conditioning (Blary et al. 2024).

The CSF of budgerigars for grating stimuli has been measured in two different studies, both using operant conditioning and a two-alternative forced choice (2AFC) procedure. The maximum contrast sensitivity was estimated to be 10.2 Michelson contrast⁻¹ at 1.4 cycles degree⁻¹ (Lind & Kelber, 2011), and 13.3 Michelson contrast⁻¹ at 1.7 cycles degree⁻¹ (Haller et al., 2014) respectively. A similar contrast sensitivity was found in a brightness discrimination experiment with spatially separated homogeneously grey stimuli (11 Michelson contrast⁻¹: Lind et al., 2013). The contrast sensitivity of budgerigars is thus in the same range as for other granivorous species (Blary et al., 2024; Ghim & Hodos, 2006; Hodos et al., 2002; Jarvis et al., 2009; Lind et al., 2012).

In Paper I we tested the budgerigar detection threshold for single (non-periodic) targets with different contrast to the background. The targets all had a negative contrast to the background varying between >-99 and -41 in Weber contrast (>99 and 25 in Michelson contrast) and had a luminance profile of a single period of a sine wave. We found a similar spatial frequency-dependent contrast sensitivity for single targets as had previously been measured for gratings (fig.11). However, since we only included a limited range of contrasts in our experiments, it is not possible to draw any conclusions on peak contrast sensitivity or contrast sensitivity for low spatial frequencies.

Spatial acuity

Some birds have the sharpest visual acuity of all animals. The wedge-tailed eagle (*Aquila audax*) has an acuity of 138 cycles degree⁻¹ (Reymond, 1985) and the Egyptian vulture (*Neophron percnopterus*) and Indian vulture (*Gyps indicus*) of 135 cycles degree⁻¹ (Fischer, 1969). In comparison, humans have a spatial acuity of around 60 cycles degree⁻¹ (Campbell & Green, 1965). Human spatial acuity is still very impressive compared to most animal species; birds included. In fact, the exceptionally sharp vision of some raptors is not common in birds, whose visual acuity shows a great variation with 84% of all species having an acuity below 30 cycles degree⁻¹ (Caves et al., 2018).

One of the main drivers of avian visual capacity, including spatial acuity, is thought to be foraging (Martin, 2017a). Species which need to detect single food-items at a distance, primarily those feeding on vertebrates or scavenged prey (e.g. diurnal raptors), generally have the highest spatial acuity (Caves et al., 2024). In contrast, ground foraging species which feed on seeds or invertebrates (e.g. many small passerines, parrots, and pigeons) tend to have low spatial acuity (Coimbra et al., 2014; Dolan & Fernández-Juricic, 2010; Donner, 1951; Moore et al., 2015).

Budgerigars, which feed primarily on grass seeds, have a similar comparatively low spatial acuity as other small ground foraging birds. Their spatial acuity has been assessed with both behavioural experiments of grating acuity and anatomical measurements based on RGC density, methods which have yielded similar results. In the behavioural experiments, spatial acuity was estimated by extrapolating the cut-off point from the behaviourally measures CSF. Studies by Lind and Kelber (2011) and by Haller et al. (2014) reported spatial acuities of 10 cycles degree⁻¹ and 7.7 cycles degree⁻¹, respectively. Spatial acuity based on anatomical measurements was estimated to 7.9 cycles degree⁻¹ by Mitkus et al. (2014).

Single target acuity in birds

In addition to foraging, predator detection is believed to have a major impact on shaping vision in birds (Martin, 2017b). Ground-foraging birds that live in open habitats are visually exposed to aerial predators. High spatial acuity is thought to benefit these species because it allows them to detect predators at greater distances (Caves et al., 2024; Tisdale & Fernández-Juricic, 2009). The distance from which a bird can detect a predator, is often assumed to be deductible from their grating acuity. However, as discussed in “Feature detection below the theoretical resolution limit”, several non-avian animals have shown a higher acuity for single targets compared to gratings (sand lizard (*Lacerta agilis*), Ehrenhardt, 1937; human, Hecht et al., 1947; dragonflies (Odonata) and flies (Diptera), O’Carroll & Wiederman, 2014; harbour seal: Sandow & Hanke, 2024; carpenter bee (*Xylocopa tenuiscapa*),

Somanathan et al., 2017; honey bee (*Apis mellifera*), Vallet & Coles, 1993). This made us wonder how well budgerigars, which inhabit mostly open landscapes, can detect single targets against a plain background.

In Paper I we assessed the target acuity of budgerigars using three different circular targets, two of which had a negative contrast to the background (lower luminance than the background), and one which was isoluminant with the background (see “Aperiodic visual stimuli – target stimuli”). The first target (which we tested at five different contrast levels, see “Spatial vision in birds - Contrast sensitivity”) had a radial luminance profile shaped like a single period of a sine wave function (Fig. 11a), which facilitated direct comparison with budgerigar acuity measured with sinusoidal gratings.

The target size was measured as the *full width at half maximum* (fig. 11a-c), which equals half a period of a sine wave in a grating with the same resolution. From our data we estimated that budgerigars can detect a “sinusoidal target” subtending 0.065 degrees of their visual field, a measure which corresponds to a sinusoidal grating of 7.7 cycles degree⁻¹. This is very similar to the previous estimates of grating acuity in budgies, both from behavioural and anatomical studies (Haller et al., 2014; Lind & Kelber, 2011; Lind et al., 2012; Mitkus et al., 2014), suggesting that budgerigars are as good at detecting gratings as they are at detecting single targets with a sinusoidal luminance profile (fig. 11d).

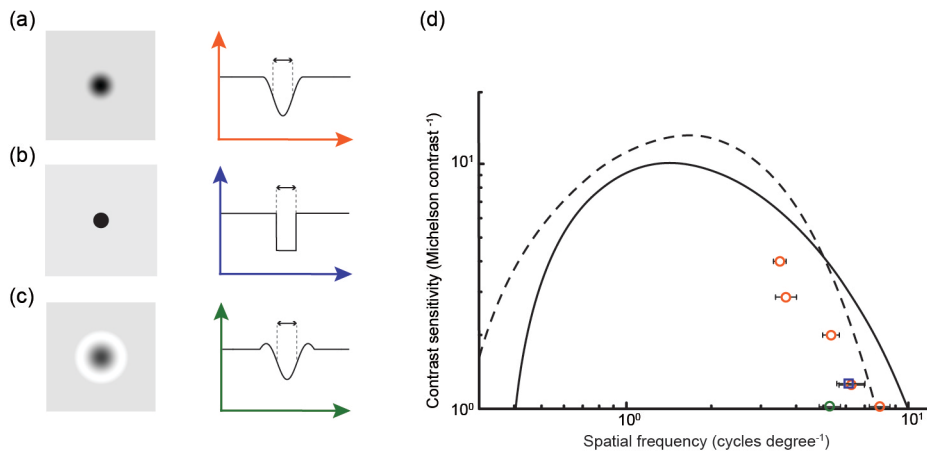


Figure 11. Single target acuity of budgerigars. (a-c) Single target designs and target luminance profiles from the experiments in Paper I: (a) sinusoidal target, (b) square-wave target and (c) isoluminant target. The small black arrows indicate the full width at half maximum. (d) The single target thresholds (color codes from luminance profiles a-c) plotted together with the contrast sensitivity function for budgerigars by Lind and Kelber (2011) and Haller et al. (2014), with a solid line and dashed line respectively.

The second target in our study had a radial luminance profile with the shape of a single period of a square wave (a “dot”; fig. 11b). Interestingly, this target had a significantly higher detection threshold (0.098 degrees, or 5.1 cycles degree⁻¹) than the sinusoidal target with the same contrast.

Previous research has established that the luminance profile of grating stimuli (sine or square wave) does not affect the CSF in budgerigars (Lind et al., 2012). This does not apply to humans which have a contrast sensitivity that is 1.27 ($4/\pi$) times higher for square gratings than for sinusoidal gratings (Campbell & Robson, 1968). This can be explained by the Fourier transform: while a sinusoidal grating contains a single spatial frequency, a square wave grating contains additional higher frequency components. The higher sensitivity for the square wave grating is believed to reflect the summed response at all frequencies. However, the cut-off frequency for both gratings will be the same, as the highest spatial frequencies are filtered out by the optics. human spatial acuity is thus not affected by the luminance profile of the grating (Campbell & Robson, 1968).

We could rule out that the higher detection threshold for square wave targets, compared to sinewave targets, was a consequence of its higher harmonics as this would have produced the opposite result. A different result would also have been expected if the target sizes at threshold were below the size of Ricco’s area (see “Feature detection below the theoretical resolution limit”); a summation of the luminance over the square wave target would have resulted in a higher contrast compared to the sinewave target (Supplementary material A, Paper I). Instead, we found that the two measures of target acuity agreed if we considered only the portion of the target that has at least 10% Michelson contrast (equivalent to the contrast detection threshold in budgies) against the background.

Although the sinewave target acuity in budgerigars correspond well with the previously measured grating acuity, their square wave target acuity is lower. Given that real targets typically have “sharp edges”, rather than gradual transitions to the background, square wave target acuity is likely to provide more realistic estimates of detection distance to such.

The detection threshold for the third, isoluminant, target was very similar to the detection threshold for the sinewave target with the same contrast to the background. The two targets were basically the same, except the isoluminant target was extended with a light “annulus” that made it overall isoluminant with the background (fig. 11c). Thus, the target would appear “invisible” at sizes below Ricco’s area.

Altogether our results indicate that the target acuity in budgerigars is limited by their retinal sampling frequency. Probably their low contrast sensitivity does not allow for detection of targets smaller than Ricco’s area. A lower detection threshold for (square wave) single targets compared to gratings was also found in the lagoon triggerfish (*Rhinecanthus aculeatus*: Champ et al., 2014), which like the budgerigar has a relatively low luminance contrast sensitivity (van den Berg et al., 2020).

A single target usually has a much smaller spatial extent compared to a grating. The "effective" stimulus size therefore differs, which can affect its visibility in several ways. In humans, the contrast sensitivity for gratings increase with the angular size of the stimulus, but also with the number grating cycles (Chen et al., 2019; King-Smith & Kulikowski, 1975; Robson & Graham, 1981). This effect is thought to depend on spatial summation of responses (King-Smith & Kulikowski, 1975), or probability summation (Meese & Summers, 2012; Robson & Graham, 1981). The existence of a similar mechanism in birds would likely affect the visibility of target and grating stimuli to different extents.

Furthermore, the sampling frequency is not homogenous throughout the avian retina, but the highest acuity is restricted to specific locations (i.e. *areae*). The detection of a single target, in particular if it approaches the threshold, thus relies on the image of the target being projected onto this particular retinal location. The same problem does not arise for a grating stimulus because of its greater spatial extent.

The targets in the experiments in Paper I were presented in the centre of a circular stimulus windows, in an attempt improve target localization by the birds. "Spatial cueing" has shown to increase the speed and accuracy of target localization in chickens (Sridharan et al., 2014). However, whether spatial cueing had any effect on target detection in our study is difficult to evaluate.

Motion vision in birds

Birds participate in numerous activities which require fast motion vision; a lot of them make rapid flight manoeuvres in dense vegetation, others catch evasive prey in mid-air or fly in large acrobatic murmurations. Birds in general have good motion vision and the highest critical flicker fusion frequency (CFF) of all vertebrates, surpassed only by insects across the animal kingdom (Inger et al., 2014; Lafitte et al., 2022). Both birds and insects are known for their ability to fly, and flight control is a behaviour that is believed to require fast visual perception (Lafitte et al., 2022).

Although the temporal acuity of bird vision is generally high, it also shows appreciable variation between species. The main driver of high temporal acuity is thought to be foraging strategy, and the highest CFFs have been measured in species hunting fast-flying prey (collared flycatcher [*Ficedula albicollis*]: 128.1 Hz, pied flycatcher [*F. hypoleuca*]: 138.2 Hz, bluetit [*Cyanistes caeruleus*]: 130.3 Hz, Boström et al., 2016; peregrine falcon [*Falco peregrinus*]: 124.5 Hz, Potier, Lieuvain, et al., 2020). In contrast, species that eat static food (e.g. seeds or nectar) have lower temporal acuity (budgerigar: 84.2 Hz, Boström et al., 2017; Anna's hummingbird [*Calypte anna*]: 70-80 Hz, Goller et al., 2019; chicken: 87 Hz, Lisney et al., 2011).

Self-induced motion

Birds rely on self-induced retinal image motion, optic flow, to guide their behaviour in several different contexts. For example, translational optic flow provides flying birds with information about their own speed and distance to external objects. Budgerigars have been shown to balance the speed of the optic flow on both their eyes when they pass through narrow passages, a behaviour that enables a centred route between obstacles thus preventing collisions (Bhagavatula et al., 2011). Interestingly, not all species use the same strategy to maintain safe flight: Anna’s hummingbird instead uses the retinal image expansion rate for flight control (Dakin et al., 2016).

Translational optic flow can also be used to monitor flight speed (Schiffner & Srinivasan, 2015, 2016). Flying through corridors with dense foliage requires a lot of motion control and the ability to quickly fine-tune the course, something which is difficult at high speeds. Budgerigars achieve a safe flight by altering their speed in response to the magnitude of translational optic flow: they will fly slow if they experience a strong optic flow, fly fast if they experience weak optic flow (Schiffner & Srinivasan, 2015). This relationship between optic flow and flight speed is not linear, but budgerigars switch between two distinct flight speeds that likely correspond to local flight speed optima (Altshuler & Srinivasan, 2018; Hedenström & Ålerstam, 1995; Schiffner & Srinivasan, 2016).

Birds are thought to use the expanding optic flow field in front of them to estimate the *time-to-contact*⁵ with approaching objects (Lee & Kalmus, 1980). A similar strategy might also be utilized to time foot extension before landing, and to “streamline” before plummeting in gannets (Davies & Green, 1990; Lee et al., 1993; Lee & Kalmus, 1980; Lee & Reddish, 1981). Information derived from the expanding optic flow field is also likely used for controlling the bill, for example when eating or feeding chicks.

While translational and expanding optic flow provides knowledge about the position or distance to external objects, rotational optic flow only informs the bird about its own rotation. To separate the optic flow generated by rotation from that of translation, birds have been demonstrated to make stabilizing eye and head movements when they change the direction of flight (Eckmeier et al., 2008; Kress et al., 2015; Ros & Biewener, 2017). While the body change direction gradually during a turn, the head makes several fast saccadic movements interspersed with short periods of constant gaze orientation (Eckmeier et al., 2008; Kress et al., 2015; Ros & Biewener, 2017). During manoeuvring flights and obstacle avoidance, birds may also fixate salient edges in their frontal visual field to stabilize their gaze and facilitate

⁵ The *time-to-contact* is derived from the optical parameter τ . τ is defined as the angular distance between a point r and the focus of expansion (the radius, for circular objects), divided by the expansion velocity v at a given time t ($\tau(t) = r(t) / v(t)$)

extraction of information from the optic flow field (Eckmeier et al., 2008; Kress et al., 2015; Miñano et al., 2023; Raudies et al., 2012).

Object motion

Object motion is needed for the detection of predators, prey, or conspecifics, and is thus of great importance to most birds. Moving targets have been found to catch attention and induce lateral visual fixation in birds (Evans et al., 1993; Maldonado et al., 1988). For example, naïve chickens are predisposed to be attracted to moving objects, especially if exhibiting sudden changes in speed or direction (Rosa-Salva et al., 2016), a motion pattern that is believed to signal animacy (Abrams & Christ, 2003; Pratt et al., 2010; Tremoulet & Feldman, 2000).

Object motion that is induced by the presence of a predator is thought to be of particular importance to many birds, and they express strong innate reactions to targets mimicking the movement of a predator (e.g. Dessborn et al., 2012). Chickens react defensively also to simple visual targets moving in the dorsal visual field, presumably of the same reason (Evans et al., 1993; Hébert et al., 2019). The defence response of adult chickens is stronger if the moving target is large or fast, although a variety of moving targets induce visual fixation (Evans et al., 1993).

In birds, object motion is thought to be analysed by the *optic tectum*. The optic tectum is responsible for processing information about the location and relevance of visual targets and is also involved in attentional orientation behaviour (Knudsen & Schwarz, 2017). Tectal neurons in birds are highly responsive to targets that loom or drift in a contrasting direction relative its surrounding, which indicates that these stimuli induce attentional “pop-out” (Huang et al., 2022; Niu et al., 2020; Zahar et al., 2012).

Birds are capable of extracting valuable information simply from the motion of simple targets. They can categorize moving targets based on their speed or direction, suggesting that dynamic properties might contribute to the recognition of other animals or objects in their environment (Herbranson et al., 2002). This could allow birds to identify other individuals as predators or kin by their specific motion pattern at distances too great to resolve relevant spatial details.

The effect of motion on contrast sensitivity

Motion does not only catch the attention of birds but can also affect their visual threshold. Haller et al. (2014) demonstrated that the contrast sensitivity in budgerigars for “small-field” (6.7°) gratings stimuli increase with horizontal drift. Contrast sensitivity increased for all spatial frequencies included in the study, although the greatest changes were observed for very high ($6.5 \text{ cycles deg}^{-2}$) and very low ($0.48 \text{ cycles deg}^{-2}$) frequencies. The contrast sensitivity maximum occurred at the same

spatial frequency whether the grating was moving or not but increased from 14 (Michelson contrast⁻¹) for the static stimulus to 17.4 for the grating moving at the highest velocity (6 degrees s⁻¹). In comparison, motion does not affect the maximum contrast sensitivity in humans but shifts its position to lower spatial frequencies. In chickens, it was found that the contrast sensitivity of the optokinetic reflex increases with grating drift velocity (Shi & Stell, 2013). However, only low spatial frequencies (0.1-0.5 cycles deg⁻²) were tested in this study.

Other studies of the effect of movement on contrast sensitivity in birds have produced somewhat inconsistent results. Hodos et al. (2003) found that counter-phase sinusoidal modulation (1-32 Hz) of gratings, reduces the contrast sensitivity in pigeons (operant conditioning). In contrast, the contrast sensitivity of an American kestrel, increased with counter-phase modulation (abrupt changes at 0.25 Hz; Hirsch, 1982).

Detection of moving single targets

In Paper II we aimed to find out the effect of motion on single target acuity in budgerigars. We knew from Paper I that their spatial acuity assessed from sharp-edged (square wave) targets is higher than their grating acuity. Because motion has the potential to both increase the attentional capture of a visual stimulus and increase its perceived contrast, we wanted to find out whether adding motion would increase the visibility of a single target stimulus.

The training of the budgerigars for the experiments in Paper I was tedious, and the static targets were surprisingly difficult to condition. Martinoya et al. (1983) suggest that motion might facilitate the conditioning of visual stimuli when they are viewed through the lateral visual field. The reason for using moving target stimuli was thus twofold: 1) find out the effect of motion on the detection threshold for single targets, and 2) more time efficient training of the test subjects.

The experiment in Paper II included a single circular black target that moved semi-randomly within an “invisible” square. The target had a speed of 1.69 degrees s⁻¹, which was similar to the drift velocity that produced the greatest contrast sensitivity for high frequency gratings (1.4 degrees s⁻¹) in Haller et al. (2014). The detection threshold we found for the moving target stimulus was, however, very similar to the threshold for the static square wave target in Paper I. Although motion did not improve target acuity in our study, it is difficult to say whether we would get the same results with a different type of motion, such as lateral drifting. Furthermore, Haller et al. (2014) found the highest contrast sensitivity for the fastest driving gratings in their study. It is thus possible that a higher speed would also have improved the target acuity of the budgerigars in our experiment.

Random target movements did (to our knowledge) not capture the attention or facilitate the training process of the budgerigars. We had previously attempted to

condition both budgerigars and zebra finches (*Taeniopygia guttata*) to a moving target presented on a horizontally placed monitor (method based on Lind, 2016) but were unsuccessful. An attempt by Simon Potier (personal communication) to condition Harris's hawks to the moving target from Paper II did not succeed either.

The initial disinterest in the moving target on the part of the budgerigars is consistent with observations reported from studies using head-restrained birds. Small simple moving targets presented in the lateral visual field do not capture the attention of either pigeons (small light-emitting diode: Bloch et al., 1984) or starlings (4° black dot: Tyrrell et al., 2014). Instead, fast and unexpected movements by larger objects are much more effective for pigeons, while starlings rather fixate images of real moving mealworms or raptors.

Visual exposure to close range small targets in freely moving pigeons and chickens, on the other hand, tend to elicit pecking (e.g. Bird, Goodwin & Hess, 1969; Blough, 1977; Goodale, 1983; Osorio et al., 1999; Wilkie & Saksida, 1994). Pecking behaviour is associated with foraging and exploration, among other things, and is controlled by vision in the frontal visual field (Goodale, 1983). Given the functional difference, it is possible that fixation in the frontal and lateral visual field is induced by different types of stimuli.

The experimental arena described in Paper II forced the budgerigars to view the stimuli from a distance of 0.73 m. Overhead video recordings confirmed that the birds used their lateral visual field when viewing the stimuli during the experiment. Future studies of single target acuity in birds should therefore preferably be done with more ecologically relevant target shapes, for example a predator silhouette, to improve the visual attention. Presenting the stimuli overhead could also improve target relevance for the birds.

Vision in different light intensities

Luminance sensitivity

Dark adaptation of the visual system of birds behaves in a similar way to that of humans. The adaptation curve (lowest detectable luminance threshold as a function of elapsed time) for birds usually shows two distinct segments that reflect the different timescales with which cones and rods recover their light sensitivity. The *cone segment* of the curve begins with a comparatively rapid drop, followed by a progressively slower decline. After up to 30 minutes, the threshold begins to drop more quickly again, at the so-called "rod-cone brake", when the recovered sensitivity of the rods becomes noticeable. The *rod segment* continues with a gradually slower threshold decrease. Full adaptation can take up to about 60 minutes to reach, for

long or intense pre-exposure to light. The dark adaptation curve of pigeons reveal that their cone segment contribute to a proportionally larger part of the total threshold drop compared to for humans (Blough, 1956). This difference likely reflects that the avian retina is numerically dominated by cones, whereas the human retina is dominated by rods. Except for the pigeon, dark adaptation curves have been measured in only a few bird species, including the European starling (Adler & Dalland, 1959), the ring-billed gull (*Larus delawarensis*), the gray gull (*L. modestus*: Emond et al., 2006) the black-bellied tree duck (*Dendrocygna autumnalis*: Hersloff et al., 1974) and the mallard (Wells et al., 1975).

In addition to light intensity, the switch between cone- and rod-dominated vision is also likely controlled by the time of day. The rod activity of the chicken and the Japanese quail appears to be blocked during the day, while it is active during the night, regardless of the light level (Manglapus et al., 1998; Schaeffel et al., 1991). Spectral sensitivity measurements of the photoreceptors in budgerigars suggest a possible presence of a similar mechanism, as no rod activity could be observed at light intensities as low as 0.02 cd m^{-2} (Lind et al., 2014).

Effect of light intensity on spatial and temporal acuity

The spatial acuity in birds typically increases with the light intensity of the stimulus up to a maximum, whereby it plateaus or decreases slightly (Donner, 1951; Fite, 1973; Gover et al., 2009; Hodos & Leibowitz, 1977; Hodos et al., 1976; Lind et al., 2012; Martin & Gordon, 1974; Raymond, 1985; Raymond, 1987). Spatial acuity peaks at different light levels in different species, which has been suggested to relate to the natural light range within which a species is active (Donner, 1951).

The temporal acuity of bird vision is affected by luminance in a similar way as spatial acuity. The integration time of visual signals is shortened, resulting in the flicker-fusion frequency (FFF) increasing logarithmically with light intensity, due to a shortened integration time of the visual signal (Boström et al., 2017; Boström et al., 2016; Lisney et al., 2011; Potier, Lieuvain, et al., 2020).

Best visual acuity is obtained when a bird is fully adapted to the luminance of the stimulus, even for higher light intensities. The spatial acuity of pigeons measured with a 1 cd m^{-2} grating is significantly lower when a bird has been preadapted to scotopic light intensity compared to photopic light intensity (Hodos and Leibowitz, 1977).

Fast luminance adaptation in birds

There is not much knowledge about how birds cope with fast changes in light intensity. Yet flying birds are likely to be subject to even more rapid light changes than most terrestrial vertebrates, as they move quickly between sky and protective

vegetation. Adaptation mechanisms involving functional reconfigurations of retinal circuits take time and are therefore not particularly useful at such fast luminance transitions (Schwartz & Levine, 2021).

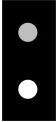
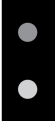





The pupillary light response is thought to smooth out fast changes in luminance by rapidly constricting the pupils in response to increasing light levels and dilating them as light levels drop (Douglas, 2018). The irises of birds are partly innervated by striated muscles (instead of only smooth muscle fibres as in mammals, fish, and amphibia), which enables comparatively fast (100-150 msec) constriction of the pupil (Douglas, 2018). The pupillary light response is likely to have only a limited effect on luminance adaptation in most birds, as the pupil can typically only change its area by a factor of 3-4 (less than 2, in budgerigars; Douglas, 2018; Lind & Kelber, 2009). An exception is the king penguin (*Aptenodytes patagonicus*) which has been suggested to use pupillary constriction to prevent scotopic light adaptation before foraging at deep waters (Martin, 1999). The pupils of king penguins have a rather extreme dynamic range and are capable of a 300-fold change in area.

Although the pupil of birds partly controls the light flux to the retina, this is likely not its only function. Pupillary constriction is thought to prevent blurring of the image in bright light by limiting the effect of spherical and chromatic aberration as light passes through the lens (Douglas, 2018; Kröger et al., 1999; Lind et al., 2008). Furthermore, rapid constrictions and dilations of the pupil, so-called "eye-pinning", occur in psittacines in contexts related to arousal or ambivalence (Brockway, 1964b; Gregory & Hopkins, 1974). Although seemingly well known among bird breeders and pet bird keepers, research on eye-pinning is, to my knowledge, scarce.

Visual sensitivity following a fast luminance drop

A situation that exposes birds to both rapid and large shifts in light intensity is the feeding of nestlings in tree cavities. Although cavity nests offer a safe place, the shielded design blocks out light, limiting vision (Wesołowski, 2007). Only a few percent of the incoming light reaches down to the tree cavity nests of passerines, where the median illuminance is 0.1-0.2 lux or lower (Maziarz & Wesolowski, 2014; Wesolowski & Maziarz, 2012). Still, individual feeding bouts only take a few seconds (Podkowa et al., 2019), a time span too short for complete dark-adaptation (Blough, 1956). Despite this, birds seem to use their vision in several behaviours in the nest-cavity. For example, visual cues are likely important for egg localisation during incubation (Avilés et al., 2006), but also for discovering the eggs laid by a nest-parasite (Di Giovanni et al., 2023; Yang et al., 2022); the visual saliency of nestlings have been shown to improve food transfer from parent to nestling (Dugas, 2015; Heeb et al., 2003; Podkowa et al., 2019; Wiebe & Slagsvold, 2009, 2012), but also affect allocation of food between siblings (Bize et al., 2006; Jourdie et al., 2004).

Table 1 Luminance contrast and brightness thresholds in budgerigars

Michelson contrast	Weber contrast	Resolution degrees (cycles deg ⁻¹)	Target or stimulus luminance cd m ⁻² S+(S-	Background luminance cd m ⁻²	Viewing distance mm	Stimulus or target size degrees	Stimulus design	Reference
0.29	0.45	9.6 (0.52)	200/110	0.225	12	9.6		Paper III
0.27	0.42	9.6 (0.52)	10.4/6.03	0.018	12	9.6		Paper III
0.37	0.54	9.6 (0.52)	0.68/0.31	0.0008	12	9.6		Paper III
0.26	0.41	9.6 (0.52)	0.190/0.11	0.00024	12	9.6		Paper III
0.09	0.18	–	47*	8.5	1268	3.6		Lind et al. 2013
0.10	–	(1.4)	50/50**	10-11	1268	3.6		Lind et al. 2011
0.07	–	(1.9)	63/63**	8-10	1268	6.7		Haller et al. 2014

**Mean luminance of the S+ and S-

*Mean luminance of the stripes in the grating

A lot of research on the role of vision in brood care has been conducted through experimental manipulation of nest lighting conditions or visual characteristics of nestlings. In many studies, factors such as increase in nestling mass (Bize et al., 2006; Heeb et al., 2003; Jourdie et al., 2004) or parental feeding behaviour (Border et al., 2023; Dugas, 2015; Podkowa et al., 2019) are often used as a proxy for visual discrimination or detection. The visual capacity of birds during rapid decreases in light intensity has never been directly tested.

The experiments in Paper III were designed to find out how well the vision of a cavity-nesting bird, the budgerigar, copes with rapid drops in light intensity, equivalent to what they encountered when entering a nest-cavity. We trained budgerigars to enter a small, dimly lit, chamber, the “decision box”, in which they were presented with visual stimuli in a 2AFC trial (see “Stimulus presentation”). The subject left the decision box between each trial to readapt to the higher light level outside.

The stimuli, bright circular targets (9.6° in diameter) on a dark background, were presented under four different light levels (ranging between 0.47 and 469 lux). We tested the birds’ ability to detect a single bright target from the background (the *absolute threshold* for luminance) as well as their ability to distinguish between two different bright targets (the *difference threshold* for luminance).

Interestingly, the birds more or less always responded already about 1 second after entering the decision box instead of waiting longer for vision to adapt. In passerines, low nest light levels result in reduced feeding efficiency and more time-consuming feeding (Dugas, 2015; Podkowa et al., 2019), likely because of the reduced visual sensitivity prior to full adaptation. We had expected the budgies to take longer to respond to the lowest light levels in our experiment. Although we were unable to show any effect of light level on response times, this does not rule out the presence of such in different contexts. In a 2AFC setting, long decision times are costly, since they result in fewer choices per time unit. Instead, a strategy of making quick, but not always correct, choices can be more cost-effective, especially when it comes to difficult decisions (Drugowitsch et al., 2012). In a feeding context, on the other hand, the more cost-effective strategy is probably to spend a few extra seconds in the nest to secure a safe delivery of food.

In the absolute threshold experiment, we found a similar threshold ($\sim 0.11\text{--}0.14\text{ cd m}^{-2}$) for the three lowest light levels, while it was significantly higher (0.83 cd m^{-2}) for the brightest light level. This confirms that the ambient illuminance has only an indirect role in the light sensitivity of the visual system in budgerigars, which instead adapt to the background luminance of visual stimuli. Although stimulus background luminance differed for all four light levels, the backgrounds at the three lowest levels were likely too dark to affect the birds’ luminance sensitivity. As opposed to this, the background at the highest light level reached above their luminance threshold, lowering their light sensitivity compared to the other three levels.

The experiment testing the difference threshold revealed that the budgerigars were close to equally good at detecting luminance contrast (0.41-0.54 Weber contrast) at all four light levels. This result is consistent with Weber's law, suggesting that birds partially adapt to the prevailing luminance already within a second.

The luminance difference thresholds we found in this experiment is notably higher than brightness or luminance contrast thresholds for budgerigars from other studies (Table 1). Fully adapted, and tested in bright light, budgerigars are able to detect static gratings with a 7.1-9.8% Michelson contrast and discriminate between homogenous grey fields with 11% Michelson contrast, the latter being equivalent to a Weber contrast of 0.18 (Haller et al., 2014; Lind et al., 2013; Lind & Kelber, 2011). It is difficult to say to what extent the relatively high luminance difference threshold in our experiments is due to insufficient adaptation and how much is due to other factors. If the spatial stimulus structure had a large impact, the result would likely be more similar to that of our previous study of luminance discrimination in budgerigars, which, like this one, used two homogeneous, spatially separated, grey fields as stimuli (Lind et al., 2013). Background luminance may also have affected the result. The targets in Paper III were significantly brighter than the background (Table 1), which may have impaired visual performance as contrast sensitivity is generally highest when target and background luminance match (Whittle, 1992). A more comprehensive understanding of the role of the adaptation state for the result would have required us to also test the birds under unchanged light conditions. However, this was not possible due to technical difficulties in providing the same high light level throughout the experimental setup.

Differences between budgerigars and cavity nesting passerines

Although with the study in Paper III we had the ambition to study the vision of cavity-nesting birds in general, the results must be interpreted specifically for budgerigars. In fact, there are several differences between the nesting behaviour of budgerigars compared to cavity-nesting passerines, which could reflect the relative importance of different sensory cues. Passerine nestlings vocalize and open their mouths widely in the direction of the parent when they beg for food. Nestlings in species with dark nests have rictal flanges that are larger, brighter, and have a higher contrast to the gape and surrounding, facilitating parental targeting of the mouth when feeding (Aviles et al., 2008; Hunt et al., 2003; Kilner & Davies, 1998). Budgerigar nestlings, on the other hand, do not expand their mouths when begging, nor do they have visually conspicuous rictal flanges. When a budgerigar feed its offspring it grasps its beak, at right angles, and regurgitates seeds directly into the crop.

Due to extreme hatching asynchrony in budgerigars (>2 days between hatchings), nestlings commonly vary greatly in size and development (fig. 12; Stamps et al., 1985). While all nestlings are able to vocalize, the begging behaviour in older nestlings also involves head bobbing, wing flapping and attempts of beak-grasping. Smaller chicks are less mobile and cannot even lift their head the first week after

hatching. The female parent still selectively feeds the smallest chick first, often without prior begging, by placing it on its back and grasping its beak (Stamps et al., 1985).

Beak grasping behaviour during feeding is typical of psittacines and suggests that touch plays a prominent role in this context. The touch receptor organ in the upper mandible of psittacines is used to manipulate and explore objects by tactile cues, compensating for the limited vision in the frontal visual field (Martin & Martin, 2020). It is therefore likely that budgerigars use touch more than sight to transfer food to their offspring.



Figure 12. Budgerigar nestlings. Budgerigars hatch asynchronously, and the average age difference between nestlings is about two days. The picture shows three nestlings of different ages, as well as two eggs, belonging to the same clutch.

Distinguishing between the different chicks in the dark nest surely involves multiple sensory modalities. The vigorous movements made by older nestlings could be a visual signal. But the targeting of the smallest, often passive, nestlings likely involve of additional senses. Unlike most passerines, budgerigar embryos communicate vocally with their parents even before hatching (Berlin & Clark, 1998). Another possible sensory modality is olfaction, which has been reported to be part of the social communication between adult budgerigars (Zhang et al., 2010).

Lastly, budgerigars are probably not guided as much by light conditions as passerines when choosing nestsite. The depth of budgerigar nests has been reported to vary between 26 cm and several meters below the entrance hole (Schrader, 1975; Wyndham, 1981). Passerines build their nests closer to the entrance with a considerably smaller variation (collared flycatcher: 2-38 cm: Maziarz & Wesolowski, 2014; marsh tits *Poecile palustris*: 8-14 cm, great tits 7-29 cm: Wesolowski & Maziarz, 2012).

Conclusions

In the studies included in this thesis I have explored the abilities of birds, with the budgerigar as my model, to detect and differentiate between single target stimuli. Me and my coauthors found that behavioural measurements of spatial acuity and contrast sensitivity that are based on grating stimuli do not necessarily translate to thresholds for single target stimuli. Although most animals tested are better at resolving single targets compared to gratings, budgerigars appear to be just as good or slightly better at resolving gratings (Paper I and II). The same single target acuity was measured for both for static (Paper I) and semi-randomly moving targets (Paper II). A major contributor to the low target acuity in budgerigars is likely their poor capacity to perceive luminance contrast. Low contrast sensitivity is a general trait in birds, so it is likely that other species also have relatively low target acuity. It would be interesting to find out if single target acuity is equally low in a species that hunt individual prey on the wing (e.g. flycatchers, hobbies, swallows).

Although we did not find any difference in the single target acuity for static and moving targets, an effect of motion in target detection cannot be excluded. Since we only tested one target speed and semi-random movement, we can only draw conclusions regarding these. To more comprehensively investigate the effect of motion on target acuity, different target speeds and types of motion, for example drifting and acceleration, would need to be tested. Furthermore, even if motion does not affect single target acuity, it may well affect the visibility of larger, low contrast, targets. It would also be interesting to find out how motion of a target affects the contrast sensitivity in birds in low light levels. For example, do budgerigar nestlings' head-

bobbing and wing flapping also enhance their contrast against the background, besides attracting attention?

In Paper III, we found that budgerigars have about the same contrast sensitivity after a small as after a large decrease in light intensity, indicating that some of their luminance adaptation occurs very quickly. The time course of rapid (milliseconds–minutes) luminance adaptation would be an intriguing study. Such a study would also benefit from including how colour vision is affected by rapid changes in light level. Furthermore, species from separate bird lineages build their nests in cavities and they may well have developed different strategies to cope with rapid light changes. Thus, it would be interesting to explore the visual performance of, for example, a cavity-nesting passerine species in an experiment similar to ours.

Quantification of various aspects of the visual environment of birds in relevant contexts would enable the design of behavioural experiments that can answer ecologically relevant questions. In particular, I believe that more research on potential interactions between spatial and temporal properties of visual stimuli would contribute to a deepened understanding of visual perception in birds.

This thesis has hopefully contributed with a small piece to the puzzle of visual perception and ecology of birds. Although me and my coauthors succeeded in finding answers to some of our original research questions, the experimental results together with various unexpected observations throughout the experimental process, have generated many new questions. I am excited to find out what future studies in this field of research will reveal.

Acknowledgements

My research, and this doctoral thesis, would not have been possible without the support and contributions from family, friends and colleagues. It has been a pleasure to be part of Lund Vision Group, thank you all for a fun and interesting time. I especially wish to thank my supervisors and co-authors. Almut, thank you for giving me this fantastic opportunity, you have been such an inspiration to me. Olle, thank you, especially for being such a great support. Eric, the field work in Panama and Australia gave me memories for life but also the confidence I needed to start my PhD candidacy; for this I am very grateful. Thank you also for proving that it is possible to be glamorous and stylish even during field work. Dan, thank you for everything, especially for all the support and patience. Jan-Åke, I am grateful for your help with the nest box cameras. Although they never produced any papers they contributed to important insights. Mikael, thank you for helping me when I was stuck in mathematical calculations and programming, and of course for all the conversations about life, the universe and everything.

My entire family has been very important to me throughout this journey, and I dedicate this thesis to my main inspiration in life: my two daughters Flora and Lilja. Thank you both for everything, especially for your curiosity and love of nature, and for always asking such clever questions. And last but not least, Josef, thank you for always being there for me, for always getting me back on track when I doubt myself, and for making me believe that I can write a doctoral thesis.

References

- Abrams, R. A., & Christ, S. E. (2003). Motion onset captures attention. *Psychological Science*, 14(5), 427. <https://doi.org/10.1111/1467-9280.01458>
- Adler, H. E., & Dalland, J. I. (1959). Spectral thresholds in the starling (*Sturnus vulgaris*). *Journal of Comparative and Physiological Psychology*, 52(4), 438-445. <https://doi.org/10.1037/h0039935>
- Adrian, W. (1989). Visibility of targets: model for calculation. *Lighting Research & Technology*, 21(4), 181-188. <https://doi.org/10.1177/096032718902100404>
- Ahissar, E., & Arieli, A. (2001). Figuring space by time. *Neuron*, 32(2), 185-201. [https://doi.org/10.1016/S0896-6273\(01\)00466-4](https://doi.org/10.1016/S0896-6273(01)00466-4)
- Aho, A. C., Donner, K., Helenius, S., Larsen, L. O., & Reuter, T. (1993). Visual performance of the toad (*Bufo bufo*) at low light levels: retinal ganglion cell responses and prey-catching accuracy. *Journal of Comparative Physiology A*, 172(6), 671-682. <https://doi.org/10.1007/bf00195393>
- Aho, A. C., Donner, K., & Reuter, T. (1993). Retinal origins of the temperature effect on absolute visual sensitivity in frogs. *The Journal of Physiology*, 463(1), 501-521. <https://doi.org/10.1113/jphysiol.1993.sp019608>
- Akre, K. L., & Johnsen, S. (2014). Psychophysics and the evolution of behaviour. *Trends in Ecology and Evolution*, 29(5), 291-300. <https://doi.org/10.1016/j.tree.2014.03.007>
- Altshuler, D. L., & Srinivasan, M. V. (2018). Comparison of visually guided flight in insects and birds. *Frontiers in Neuroscience*, 12. <https://doi.org/10.3389/fnins.2018.00157>
- Aviles, J. M., Perez-Contreras, T., Navarro, C., & Soler, J. J. (2008). Dark nests and conspicuousness in color patterns of nestlings of altricial birds. *American Naturalist*, 171(3), 327-338. <https://doi.org/10.1086/527493>
- Avilés, J. M., Soler, J. J., & Pérez-Contreras, T. (2006). Dark nests and egg colour in birds: A possible functional role of ultraviolet reflectance in egg detectability. *Proceedings: Biological Sciences*, 273(1603), 2821-2829. <https://doi.org/10.1098/rspb.2006.3674>
- Baccus, S. A., & Meister, M. (2002). Fast and slow contrast adaptation in retinal circuitry. *Neuron*, 36(5), 909-919. [https://doi.org/10.1016/S0896-6273\(02\)01050-4](https://doi.org/10.1016/S0896-6273(02)01050-4)
- Baden, T., Euler, T., & Berens, P. (2020). Understanding the retinal basis of vision across species. *Nature Reviews Neuroscience*, 21(1), 5. <https://doi.org/10.1038/s41583-019-0242-1>

- Baird, E., Srinivasan, M. V., Zhang, S., & Cowling, A. (2005). Visual control of flight speed in honeybees. *Journal of Experimental Biology*, 208(20), 3895-3905. <https://doi.org/10.1242/jeb.01818>
- Barbur, J. L., & Stockman, A. (2010). Photopic, mesopic and scotopic vision and changes in visual performance. In D. A. Dartt (Ed.), *Encyclopedia of the Eye* (pp. 323-331). Academic Press. <https://doi.org/10.1016/B978-0-12-374203-2.00233-5>
- Barlow, H. B. (1953). Summation and inhibition in the frog's retina. *Journal of Physiology*, 119(1), 69-88. <https://doi.org/10.1113/jphysiol.1953.sp004829>
- Barlow, H. B. (1957). Increment thresholds at low intensities considered as signal-noise discriminations. *Journal of Physiology*, 136(3), 469-488. <https://doi.org/10.1113/jphysiol.1957.sp005774>
- Barlow, H. B. (1958). Temporal and spatial summation in human vision at different background intensities. *Journal of Physiology*, 141(2), 337-350. <https://doi.org/10.1113/jphysiol.1958.sp005978>
- Barlow, H. B., Fitzhugh, R., & Kuffler, S. W. (1957). Change of organization in the receptive fields of the cat's retina during dark adaptation. *Journal of Physiology*, 137(3), 338-354. <https://doi.org/10.1113/jphysiol.1957.sp005817>
- Barten, P. G. J. (1999). *Contrast Sensitivity of the Human Eye and its Effects on Image Quality*. SPIE International Society for Optical Engineering. <http://ebooks.spiedigitallibrary.org/>
- Békésy, G. v. (1947). A new audiometer. *Acta Oto-Laryngologica*, 35(5-6), 411-422. <https://doi.org/10.3109/00016484709123756>
- Berlin, K. E., & Clark, A. B. (1998). Embryonic calls as care-soliciting signals in budgerigars, *Melopsittacus undulatus* [Article]. *Ethology*, 104(6), 531-544. <Go to ISI>://WOS:000074391200006
- Berson, D. M. (2020). The sensory organ: eye, receptors, retina. In B. Fritzsche (Ed.), *The Senses: A Comprehensive Reference* (2nd ed., Vol. 1, pp. 31-35). Elsevier Inc. <https://doi.org/10.1016/B978-0-12-805408-6.00008-7>
- Bhagavatula, P., Claudianos, C., Ibbotson, M., & Srinivasan, M. (2009). Edge detection in landing budgerigars (*Melopsittacus undulatus*). *Plos One*, 4(10), Article e7301. <https://doi.org/10.1371/journal.pone.0007301>
- Bhagavatula, P. S., Ibbotson, M. R., Srinivasan, M. V., & Claudianos, C. (2011). Optic flow cues guide flight in birds. *Current Biology*, 21(21), 1794-1799. <https://doi.org/10.1016/j.cub.2011.09.009>
- Bianco, I. H., Kampff, A. R., & Engert, F. (2011). Prey capture behavior evoked by simple visual stimuli in larval zebrafish. *Frontiers in Systems Neuroscience*, 5, 101. <https://doi.org/10.3389/fnsys.2011.00101>
- Bilotta, J., & Powers, M. K. (1991). Spatial contrast sensitivity of goldfish: mean luminance, temporal frequency and a new psychophysical technique. *Vision Research*, 31(3), 577-585. [https://doi.org/10.1016/0042-6989\(91\)90108-H](https://doi.org/10.1016/0042-6989(91)90108-H)
- BIPM. (2019). Le Système international d'unités / The International System of Units ('The SI Brochure'). In (9th ed.): Bureau International des Poids et Mesures.

- Bird Goodwin, E., & Hess, E. H. (1969). Innate visual form preferences in the pecking behavior of young chicks. *Behaviour*, 34(4), 223-237.
<http://www.jstor.org/stable/4533285>
- Bisti, S., & Maffei, L. (1974). Behavioural contrast sensitivity of the cat in various visual meridians. *The Journal of Physiology*, 241(1), 201-210.
<https://doi.org/https://doi.org/10.1113/jphysiol.1974.sp010649>
- Bize, P., Piault, R., Moureau, B., & Heeb, P. (2006). A UV signal of offspring condition mediates context-dependent parental favouritism. *Proceedings of the Royal Society B-Biological Sciences*, 273(1597), 2063-2068.
<https://doi.org/10.1098/rspb.2006.3546>
- Blackwell, H. R. (1946). Contrast thresholds of the human eye. *Journal of the Optical Society of America*, 36, 624-643. <https://doi.org/10.1364/JOSA.36.000624>
- Blake, R. (1998). The behavioural analysis of animal vision. In R. Carpenter & J. Robson (Eds.), *Vision Research: A Practical Guide to Laboratory Methods* (pp. 137-160). Oxford University Press.
<https://doi.org/10.1093/acprof:oso/9780198523192.003.0006>
- Blary, C. L. M., Duriez, O., Bonadonna, F., Caro, S. P., Mitkus, M., Besnard, A., & Potier, S. (2024). Low achromatic contrast sensitivity in birds: a common attribute shared by many phylogenetic orders [Article]. *The Journal of experimental biology*, 227(3). <https://doi.org/10.1242/jeb.246342>
- Bloch, S., & Martinoya, C. (1982). Comparing frontal and lateral viewing in the pigeon. I. Tachistoscopic visual acuity as a function of distance. *Behavioural Brain Research*, 5(3), 231-244. [https://doi.org/10.1016/0166-4328\(82\)90031-6](https://doi.org/10.1016/0166-4328(82)90031-6)
- Bloch, S., Rivaud, S., & Martinoya, C. (1984). Comparing frontal and lateral viewing in the pigeon. III. Different patterns of eye movements for binocular and monocular fixation. *Behavioural Brain Research*, 13(2), 173-182.
[https://doi.org/10.1016/0166-4328\(84\)90147-5](https://doi.org/10.1016/0166-4328(84)90147-5)
- Blough, D., & Blough, P. (2022). Animal psychophysics. In W. K. Honig & J. E. R. Staddon (Eds.), *Handbook of Operant Behavior* (pp. 514-539). Routledge.
<https://doi.org/10.4324/9781003256670>
- Blough, D. S. (1955). Method for tracing dark adaptation in the pigeon. *Science*, 121(3150), 703-704. <https://doi.org/10.1126/science.121.3150.703>
- Blough, D. S. (1956). Dark adaptation in the pigeon. *Journal of Comparative and Physiological Psychology*, 49(5), 425-430. <https://doi.org/10.1037/h0043257>
- Blough, D. S. (1957). Spectral sensitivity in the pigeon. *Journal of the Optical Society of America*, 47, 827-833. <https://doi.org/10.1364/JOSA.47.000827>
- Blough, D. S. (1977). Visual search in the pigeon: hunt and peck method. *Science*, 196(4293), 1013-1014. <https://www.jstor.org/stable/1744684>
- Blough, P. M. (1971). The visual acuity of the pigeon for distant targets. *Journal of the Experimental Analysis of Behavior*, 15(1), 57-67.
<https://doi.org/10.1901/jeab.1971.15-57>
- Border, S. E., Haas, L. E., Steines, M. E., & Dugas, M. B. (2023). Nestling mouth colors mediate parental favoritism but do not influence detectability. *Behavioral Ecology*, 34(4), 581-592. <https://doi.org/10.1093/beheco/arad026>

- Boström, J., Haller, N., Dimitrova, M., Odeen, A., & Kelber, A. (2017). The flicker fusion frequency of budgerigars (*Melopsittacus undulatus*) revisited. *Journal of Comparative Physiology A* 203, 15-22.
<https://doi.org/10.1007/s00359-016-1130-z>
- Boström, J. E., Dimitrova, M., Canton, C., Håstad, O., Qvarnström, A., & Ödeen, A. (2016). Ultra-rapid vision in birds. *Plos One*, 11(3), 7, Article e0151099.
<https://doi.org/10.1371/journal.pone.0151099>
- Bowmaker, J. K. (2012). Evolution of the vertebrate eye. In O. F. Lazareva, T. Shimizu, & E. A. Wasserman (Eds.), *How Animals See the World: Comparative Behavior, Biology, and Evolution of Vision*. (pp. 441-472). Oxford University Press.
<https://doi.org/10.1093/acprof:oso/9780195334654.003.0024>
- Bringmann, A. (2019). Structure and function of the bird fovea. *Anatomia, Histologia, Embryologia*, 48(3), 177-200. <https://doi.org/10.1111/ahc.12432>
- Brockway, B. F. (1964a). Ethological studies of the budgerigar (*Melopsittacus undulatus*): non-reproductive behavior. *Behaviour*, 22, 193-222.
<https://doi.org/10.1163/156853964x00021>
- Brockway, B. F. (1964b). Ethological studies of the budgerigar: reproductive behaviour. *Behaviour*, 23, 294-324. <https://doi.org/10.1163/156853964x00193>
- Brooke, M. D. (1989). Determination of the absolute visual threshold of a nocturnal seabird, the common diving petrel *Pelecanoides urinatrix*. *Ibis*, 131(2), 290-294.
<https://doi.org/10.1111/j.1474-919X.1989.tb02772.x>
- Burr, D. C. (1981). Temporal summation of moving images by the human visual system. *Proceedings of the Royal Society of London. Series B, Biological Sciences*, 211(1184), 321-339. <https://doi.org/10.1098/rspb.1981.0010>
- Burr, D. C., & Ross, J. (1982). Contrast sensitivity at high velocities. *Vision Research*, 22(4), 479-484. [https://doi.org/10.1016/0042-6989\(82\)90196-1](https://doi.org/10.1016/0042-6989(82)90196-1)
- Campbell, F. W., & Green, D. G. (1965). Optical and retinal factors affecting visual resolution [Article]. *The Journal of Physiology*, 181(3), 576-593-593.
<https://doi.org/10.1113/jphysiol.1965.sp007784>
- Campbell, F. W., & Robson, J. G. (1968). Application of fourier analysis to the visibility of gratings. *The Journal of Physiology*, 197(3), 551-566.
<https://doi.org/10.1113/jphysiol.1968.sp008574>
- Campanhausen, M., & Kirschfeld, K. (1998). Spectral sensitivity of the accessory optic system of the pigeon. *Journal of Comparative Physiology A*, 183(1), 1-6.
<https://doi.org/10.1007/s003590050229>
- Cantlay, J., Martin, G., McClelland, S., Potier, S., O'Brien, M., Fernández-Juricic, E., Bond, A., & Portugal, S. (2023). Binocular vision and foraging in ducks, geese and swans (Anatidae). *Proceedings of the Royal Society B*, 290.
<https://doi.org/10.1098/rspb.2023.1213>
- Carlile, P. A., Peters, R. A., & Evans, C. S. (2006). Detection of a looming stimulus by the Jacky dragon: selective sensitivity to characteristics of an aerial predator. *Animal Behaviour*, 72(3), 553-562. <https://doi.org/10.1016/j.anbehav.2005.10.027>

- Caves, E. M., Brandley, N. C., & Johnsen, S. (2018). Visual acuity and the evolution of signals. *Trends in Ecology & Evolution*, 33(5), 358-372. <https://doi.org/10.1016/j.tree.2018.03.001>
- Caves, E. M., Fernández-Juricic, E., & Kelley, L. A. (2024). Ecological and morphological correlates of visual acuity in birds. *Journal of Experimental Biology*, 227(2), Article jeb246063. <https://doi.org/10.1242/jeb.246063>
- Chaib, S., Ljungholm, M., Lind, O., & Kelber, A. (2019). Single target acuity is not higher than grating acuity in a bird, the budgerigar. *Vision Research*, 160, 37-42. <https://doi.org/10.1016/j.visres.2019.04.005>
- Chaib, S., Mussoi, J. G., Lind, O., & Kelber, A. (2021). Visual acuity of budgerigars for moving targets. *Biology Open*, 10(9), Article bio058796. <https://doi.org/10.1242/bio.058796>
- Champ, C., Wallis, G., Vorobyev, M., Siebeck, U., & Marshall, J. (2014). Visual acuity in a species of coral reef fish: *Rhinecanthus aculeatus*. *Brain, Behavior and Evolution*, 83(1), 31-42. <https://doi.org/10.1159/000356977>
- Chen, C.-C., Yeh, Y.-H. C., & Tyler, C. W. (2019). Length summation in noise. *Journal of Vision*, 19(9), 11. <https://doi.org/10.1167/19.9.11>
- Chen, E. Y., Marre, O., Fisher, C., Schwartz, G., Levy, J., da Silveira, R. A., & Berry, M. J., 2nd. (2013). Alert response to motion onset in the retina. *The Journal of Neuroscience*, 33(1), 120-132. <https://doi.org/10.1523/jneurosci.3749-12.2013>
- Christ, S. E., & Abrams, R. A. (2008). The attentional influence of new objects and new motion. *Journal of Vision*, 8(3), 27, 21-28. <https://doi.org/10.1167/8.3.27>
- Clark, W., & Colombo, M. (2022). Seeing the forest for the trees, and the ground below my beak: global and local processing in the pigeon's visual system. *Frontiers in Psychology*, 13, Article 888528. <https://doi.org/10.3389/fpsyg.2022.888528>
- Coimbra, J. P., Collin, S. P., & Hart, N. S. (2014). Topographic specializations in the retinal ganglion cell layer correlate with lateralized visual behavior, ecology, and evolution in cockatoos. *The Journal of comparative neurology*, 522(15), 3363-3385. <https://doi.org/10.1002/cne.23637>
- Coimbra, J. P., Collin, S. P., & Hart, N. S. (2015). Variations in retinal photoreceptor topography and the organization of the rod-free zone reflect behavioral diversity in Australian passerines. *The Journal of comparative neurology*, 523(7), 1073-1094. <https://doi.org/10.1002/cne.23718>
- Coimbra, J. P., Yamada, E. S., Trévia, N., Picanço-Diniz, C. W., Marceliano, M. L. V., & Andrade-Da-Costa, B. L. d. S. (2009). Number and distribution of neurons in the retinal ganglion cell layer in relation to foraging behaviors of tyrant flycatchers. *The Journal of comparative neurology*, 514(1), 66-73. <https://doi.org/10.1002/cne.21992>
- Cowie, A. (2014). *Experimental Studies of Social Foraging in Budgerigars, Melopsittacus undulatus* [doctoral thesis, University of St Andrews]. British Library EThoS. <http://hdl.handle.net/10023/6552>
- Cronin, T. W. (2020). Visual ecology. In B. Fritzsch (Ed.), *The Senses: A Comprehensive Reference* (2nd ed., pp. 66-95). Elsevier. <https://doi.org/10.1016/B978-0-12-809324-5.23902-3>

- Cronin, T. W., Johnsen, S., Marshall, N. J., & Warrant, E. J. (2014). *Visual Ecology*. Princeton University Press.
- Crossland, M. D. (2010). Acuity. In D. A. Dartt (Ed.), *Encyclopedia of the Eye*, (pp. 27-32). Academic Press. <https://doi.org/10.1016/B978-0-12-374203-2.00234-7>
- Crumey, A. (2014). Human contrast threshold and astronomical visibility. *Monthly Notices of the Royal Astronomical Society*, 442(3), 2600-2619. <https://doi.org/10.1093/mnras/stu992>
- Dakin, R., Fellows, T. K., & Altshuler, D. L. (2016). Visual guidance of forward flight in hummingbirds reveals control based on image features instead of pattern velocity. *Proceedings of the National Academy of Sciences*, 113(31), 8849-8854. <https://doi.org/10.1073/pnas.1603221113>
- David, C. T. (1982). Compensation for height in the control of groundspeed by *Drosophila* in a new, 'barber's pole' wind tunnel. *Journal of Comparative Physiology*, 147(4), 485-493. <https://doi.org/10.1007/BF00612014>
- Davies, M. N. O., & Green, P. R. (1990). Optic flow-field variables trigger landing in hawk but not in pigeons. *Naturwissenschaften*, 77(3), 142-144. <https://doi.org/10.1007/BF01134481>
- De Franceschi, G., Vivattanasam, T., Saleem, A. B., & Solomon, S. G. (2016). Vision guides selection of freeze or flight defense strategies in mice. *Current Biology*, 26(16), 2150-2154. <https://doi.org/10.1016/j.cub.2016.06.006>
- De Valois, K., K., & De Valois, R., L. (1991). *Spatial Vision*. Oxford University Press.
- De Valois, R. L., Morgan, H., & Snodderly, D. M. (1974). Psychophysical studies of monkey vision-III. Spatial luminance contrast sensitivity tests of macaque and human observers. *Vision Research*, 14(1), 75-81. [https://doi.org/10.1016/0042-6989\(74\)90118-7](https://doi.org/10.1016/0042-6989(74)90118-7)
- Demery, Z. P., Chappell, J., & Martin, G. R. (2011). Vision, touch and object manipulation in senegal parrots *Poicephalus senegalus*. *Proceedings of the Royal Society B*, 278(1725), 3687-3693. <https://doi.org/10.1098/rspb.2011.0374>
- Demirel, S., Anderson, R. S., Dakin, S. C., & Thibos, L. N. (2012). Detection and resolution of vanishing optotype letters in central and peripheral vision. *Vision Research*, 59, 9-16. <https://doi.org/10.1016/j.visres.2012.02.010>
- Dessborn, L., Englund, G., Elmberg, J., & Arzél, C. (2012). Innate responses of mallard ducklings towards aerial, aquatic and terrestrial predators. *Behaviour*, 149(13/14), 1299-1317. <https://doi.org/10.1163/1568539X-00003014>
- Di Giovanni, A. J., Villa, J., Stanback, M. T., Thompson, C. F., Sakaluk, S. K., Hauber, M. E., & Hanley, D. (2023). Decision rules for egg-color-based rejection by two cavity-nesting hosts of the brown-headed cowbird. *Journal of Experimental Biology*, 226(14). <https://doi.org/10.1242/jeb.245188>
- Dolan, T., & Fernández-Juricic, E. (2010). Retinal ganglion cell topography of five species of ground-foraging birds. *Brain, Behavior and Evolution*, 75(2), 111-121. <https://doi.org/10.1159/000305025>
- Donner, K. (1987). Adaptation-related changes in the spatial and temporal summation of frog retinal ganglion cells. *Acta Physiologica Scandinavica*, 131(4), 479-487. <https://doi.org/10.1111/j.1748-1716.1987.tb08267.x>

- Donner, K. (2021). Temporal vision: measures, mechanisms and meaning. *Journal of Experimental Biology*, 224(15). <https://doi.org/10.1242/jeb.222679>
- Donner, K. O. (1951). The visual acuity of some passerine birds. *Acta Zoologica Fennica*, 66.
- Douglas, R. H. (2018). The pupillary light responses of animals; a review of their distribution, dynamics, mechanisms and functions. *Progress in Retinal and Eye Research*, 66, 17-48. <https://doi.org/10.1016/j.preteyeres.2018.04.005>
- Douglas, R. H., & Cronin, T. W. (2016). Visual matched filtering in vertebrates. In G. von der Emde & E. Warrant (Eds.), *The Ecology of Animal Senses: Matched Filters for Economical Sensing* (pp. 169-203). Springer International Publishing. <https://doi.org/10.1007/978-3-319-25492-0>
- Drugowitsch, J., Moreno-Bote, R. N., Pouget, A., Churchland, A. K., & Shadlen, M. N. (2012). The cost of accumulating evidence in perceptual decision making [Article]. *Journal of Neuroscience*, 32(11), 3612-3628. <https://doi.org/10.1523/JNEUROSCI.4010-11.2012>
- Dugas, M. B. (2015). Detectability matters: conspicuous nestling mouth colours make prey transfer easier for parents in a cavity nesting bird. *Biology Letters*, 11, Article 20150771. <https://doi.org/10.1098/rsbl.2015.0771>
- Dunn, F. A., Lankheet, M. J., & Rieke, F. (2007). Light adaptation in cone vision involves switching between receptor and post-receptor sites. *Nature*, 449(7162), 603-606. <https://doi.org/10.1038/nature06150>
- Eckmeier, D., Geurten, B. R. H., Kress, D., Mertes, M., Kern, R., Egelhaaf, M., & Bischof, H.-J. (2008). Gaze strategy in the free flying zebra finch (*Taeniopygia guttata*). *Plos One*, 3(12), Article e3956. <https://doi.org/10.1371/journal.pone.0003956>
- Ehrenhardt, H. (1937). Formensehen und sehschärfebestimmungen bei eidechsen. *Zeitschrift für vergleichende Physiologie*, 24(2), 248-304. <https://doi.org/10.1007/BF00338282>
- Ehud, e., & Amos, e. (2012). Seeing via miniature eye movements: a dynamic hypothesis for vision. *Frontiers in Computational Neuroscience*, 6, Article 89. <https://doi.org/10.3389/fncom.2012.00089>
- Emond, M. P., McNeil, R., Cabana, T., Guerra, C. G., & Lachapelle, P. (2006). Comparing the retinal structures and functions in two species of gulls (*Larus delawarensis* and *Larus modestus*) with significant nocturnal behaviours. *Vision Research*, 46(18), 2914-2925. <https://doi.org/10.1016/j.visres.2006.02.023>
- Enroth-Cugell, C., & Robson, J. G. (1966). The contrast sensitivity of retinal ganglion cells of the cat. *The Journal of Physiology*, 187(3), 517-552. <https://doi.org/10.1113/jphysiol.1966.sp008107>
- Evans, C. S., Macedonia, J. M., & Marler, P. (1993). Effects of apparent size and speed on the response of chickens, *Gallus gallus*, to computer-generated simulations of aerial predators. *Animal Behaviour*, 46(1), 1-11. <https://doi.org/10.1006/anbe.1993.1156>
- Ewert, J. P., Buxbaum-Conradi, H., Dreisvagt, F., Glagow, M., Merkel-Harff, C., Röttgen, A., Schürg-Pfeiffer, E., & Schwippert, W. W. (2001). Neural modulation of visuomotor functions underlying prey-catching behaviour in anurans: perception,

- attention, motor performance, learning. *Comparative Biochemistry and Physiology Part A*, 128(3), 417-460. [https://doi.org/10.1016/S1095-6433\(00\)00333-0](https://doi.org/10.1016/S1095-6433(00)00333-0)
- Fain, G. L., Matthews, H. R., Cornwall, M. C., & Koutalos, Y. (2001). Adaptation in vertebrate photoreceptors. *Physiological Reviews*, 81(1), 117-151. <https://doi.org/10.1152/physrev.2001.81.1.117>
- Farabaugh, S. M., Dent, M. L., & Dooling, R. J. (1998). Hearing and vocalizations of wild-caught Australian budgerigars (*Melopsittacus undulatus*). *Journal of Comparative Psychology*, 112(1), 74-81. <https://doi.org/10.1037/0735-7036.112.1.74>
- Fernández-Juricic, E., Baumhardt, P. E., Tyrrell, L. P., Elmore, A., DeLiberto, S. T., & Werner, S. J. (2019). Vision in an abundant North American bird: The red-winged blackbird. *The Auk*, 136(3), Article ukz039. <https://doi.org/10.1093/auk/ukz039>
- Fischer, A. B. (1969). Laboruntersuchungen und freilandbeobachtungen zum sehvermögen und verhalten von altweltgeiern. *Zoologische Jahrbücher Abteilung für Systematik, Geographie und Biologie der Tiere*, 96, 81-132.
- Fite, K. V. (1973). Anatomical and behavioral correlates of visual acuity in the great horned owl. *Vision Research*, 13(2), 219-230. [https://doi.org/10.1016/0042-6989\(73\)90101-6](https://doi.org/10.1016/0042-6989(73)90101-6)
- Fitzke, F. W., Hayes, B. P., Hodos, W., Holden, A. L., & Low, J. C. (1985). Refractive sectors in the visual field of the pigeon eye. *The Journal of Physiology*, 369(1), 33-44. <https://doi.org/10.1113/jphysiol.1985.sp015886>
- Fox, R., Lehmkuhle, S. W., & Bush, R. C. (1977). Stereopsis in the falcon. *Science*, 197(4298), 79-81. <https://doi.org/10.1126/science.867054>
- Franconeri, S. L., & Simons, D. J. (2003). Moving and looming stimuli capture attention. *Perception & Psychophysics*, 65(7), 999-1010. <https://doi.org/10.3758/BF03194829>
- Frazor, R. A., & Geisler, W. S. (2006). Local luminance and contrast in natural images. *Vision Research*, 46(10), 1585-1598. <https://doi.org/10.1016/j.visres.2005.06.038>
- Frost, B. J. (2010). A taxonomy of different forms of visual motion detection and their underlying neural mechanisms. *Brain, Behavior & Evolution*, 75(3), 218-235. <https://doi.org/10.1159/000314284>
- Gaynes, J. A., Budoff, S. A., Grybko, M. J., Hunt, J. B., & Poleg-Polsky, A. (2022). Classical center-surround receptive fields facilitate novel object detection in retinal bipolar cells. *Nature Communications*, 13(1), Article 5575. <https://doi.org/10.1038/s41467-022-32761-8>
- Gescheider, G. A. (1997). *Psychophysics: The Fundamentals* (3rd ed.). Lawrence Erlbaum Associates.
- Ghim, M. M. (2003). *Spatial Contrast Sensitivity of Birds* [doctoral thesis, University of Maryland]. Digital Repository at the University of Maryland. <http://hdl.handle.net/1903/65>
- Ghim, M. M., & Hodos, W. (2006). Spatial contrast sensitivity of birds. *Journal of Comparative Physiology A*, 192(5), 523-534. <https://doi.org/10.1007/s00359-005-0090-5>

- Gibson, J. J. (2015). *The Ecological Approach to Visual Perception*. Psychology Press.
<https://doi.org/10.4324/9781315740218>
- Gilbert, C. D. (1994). Early perceptual learning. *Proceedings of the National Academy of Sciences*, 91(4), 1195-1197. <https://www.jstor.org/stable/2364065>
- Glezer, V. D. (1965). The receptive fields of the retina. *Vision Research*, 5(10), 497-525.
[https://doi.org/10.1016/0042-6989\(65\)90084-2](https://doi.org/10.1016/0042-6989(65)90084-2)
- Goldsmith, T. H., & Butler, B. K. (2003). The roles of receptor noise and cone oil droplets in the photopic spectral sensitivity of the budgerigar, *Melopsittacus undulatus*. *Journal of Comparative Physiology A*, 189(2), 135-142.
<https://doi.org/10.1007/s00359-002-0385-8>
- Goldsmith, T. H., & Butler, B. K. (2005). Color vision of the budgerigar (*Melopsittacus undulatus*): hue matches, tetrachromacy, and intensity discrimination. *Journal of Comparative Physiology A*, 191(10), 933-951. <https://doi.org/10.1007/s00359-005-0024-2>
- Goller, B., Fellows, T. K., Dakin, R., Tyrrell, L., Fernández-Juricic, E., & Altshuler, D. L. (2019). Spatial and temporal resolution of the visual system of the Anna's hummingbird (*Calypte anna*) relative to other birds. *Physiological and Biochemical Zoology*, 92(5), 481-495. <https://doi.org/10.1086/705124>
- Goodale, M. A. (1983). Visually guided pecking in the pigeon (*Columba livia*). *Brain Behavior and Evolution*, 22(1), 22-41. <https://doi.org/10.1159/000121504>
- Gover, N., Jarvis, J. R., Abeyesinghe, S. M., & Wathes, C. M. (2009). Stimulus luminance and the spatial acuity of domestic fowl (*Gallus g. domesticus*). *Vision Research*, 49(23), 2747-2753. <https://doi.org/10.1016/j.visres.2009.08.011>
- Gregory, R., & Hopkins, P. (1974). Pupils of a talking parrot. *Nature*, 252(5485), 637-638.
<https://doi.org/10.1038/252637a0>
- Günther, A., Dedek, K., Haverkamp, S., Irsen, S., Briggman, K. L., & Mouritsen, H. (2021). Double cones and the diverse connectivity of photoreceptors and bipolar cells in an avian retina. *The Journal of Neuroscience*, 41(23), 5015-5028.
<https://doi.org/10.1523/jneurosci.2495-20.2021>
- Güntürkün, O., & Hahmann, U. (1999). Functional subdivisions of the ascending visual pathways in the pigeon [Article]. *Behavioural Brain Research*, 98(2), 193-201.
[https://doi.org/10.1016/S0166-4328\(98\)00084-9](https://doi.org/10.1016/S0166-4328(98)00084-9)
- Hall, J. R., Cuthill, I. C., Baddeley, R., Shohet, A. J., & Scott-Samuel, N. E. (2013). Camouflage, detection and identification of moving targets. *Proceedings of the Royal Society B*, 280(1758), Article 20130064.
<https://doi.org/10.1098/rspb.2013.0064>
- Haller, N. K., Lind, O., Steinlechner, S., & Kelber, A. (2014). Stimulus motion improves spatial contrast sensitivity in budgerigars (*Melopsittacus undulatus*). *Vision Research*, 102, 19-25. <https://doi.org/10.1016/j.visres.2014.07.007>
- Harmening, W. M., Nikolay, P., Orłowski, J., & Wagner, H. (2009). Spatial contrast sensitivity and grating acuity of barn owls. *Journal of Vision*, 9(7), Article 13.
<https://doi.org/10.1167/9.7.13>

- Hart, N. S. (2001a). Variations in cone photoreceptor abundance and the visual ecology of birds. *Journal of Comparative Physiology A*, 187(9), 685-697. <https://doi.org/10.1007/s00359-001-0240-3>
- Hart, N. S. (2001b). The visual ecology of avian photoreceptors. *Progress in Retinal and Eye Research*, 20(5), 675-703. [https://doi.org/10.1016/s1350-9462\(01\)00009-x](https://doi.org/10.1016/s1350-9462(01)00009-x)
- Hart, N. S. (2002). Vision in the peafowl (Aves: *Pavo cristatus*). *Journal of Experimental Biology*, 205(24), 3925-3935. <https://doi.org/10.1242/jeb.205.24.3925>
- Hart, N. S., & Hunt, D. M. (2007). Avian visual pigments: characteristics, spectral tuning, and evolution. *The American Naturalist*, 169(S1), S7-S26. <https://doi.org/10.1086/510141>
- Haug, L. I., & Florsheim, A. (2010). Training of animals. In M. D. Breed & J. Moore (Eds.), *Encyclopedia of Animal Behavior* (pp. 439-445). Academic Press. <https://doi.org/10.1016/B978-0-08-045337-8.00164-9>
- Hébert, M., Versace, E., & Vallortigara, G. (2019). Inexperienced preys know when to flee or to freeze in front of a threat. *Proceedings of the National Academy of Sciences*, 116(46), 22918-22920. <https://doi.org/10.1073/pnas.1915504116>
- Hecht, S., Haig, C., & Chase, A. M. (1937). The influence of light adaptation on subsequent dark adaptation of the eye. *Journal of General Physiology*, 20(6), 831-850. <https://doi.org/10.1085/jgp.20.6.831>
- Hecht, S., Ross, S., & Mueller, C. G. (1947). The visibility of lines and squares at high brightness. *Journal of the Optical Society of America*, 37, 500-507. <https://doi.org/10.1364/JOSA.37.000500>
- Hedenström, A., & Ålerstam, T. (1995). Optimal flight speed of birds. *Philosophical Transactions of the Royal Society B*, 348(1326), 471-487. <https://doi.org/10.1098/rstb.1995.0082>
- Heeb, P., Schwander, T., & Faoro, S. (2003). Nestling detectability affects parental feeding preferences in a cavity-nesting bird. *Animal Behaviour*, 66(4), 637-642. <https://doi.org/10.1006/anbe.2003.2238>
- Henze, M. J., Ott, M., & Schaeffel, F. (2004). Variations in the off-axis refractive state in the eye of the Vietnamese leaf turtle (*Geoemyda spengleri*). *Journal of Comparative Physiology A*, 190(2), 131-137. <https://doi.org/10.1007/s00359-003-0480-5>
- Herbranson, W., Fremouw, T., & Shimp, C. (2002). Categorizing a moving target in terms of its speed, direction, or both. *Journal of the Experimental Analysis of Behavior*, 78(3), 249-270. <https://doi.org/10.1901/jeab.2002.78-249>
- Hersloff, L., Lehner, P. N., Bolen, E. G., & Rylander, M. K. (1974). Visual sensitivity in the black-bellied duck (*Dendrocygna autumnalis*), a crepuscular species. *Journal of Comparative and Physiological Psychology*, 86(3), 486-492. <https://doi.org/10.1037/h0036177>
- Heukamp, A. S., Warwick, R. A., & Rivlin-Etzion, M. (2020). Topographic variations in retinal encoding of visual space. *Annual Review of Vision Science*, 6, 237-259. <https://doi.org/10.1146/annurev-vision-121219-081831>
- Higgins, P. J. (1999). *Handbook of Australian, New Zealand & Antarctic Birds* (Vol. 4: Parrots to Dollarbirds). Oxford University Press.

- Hirsch, J. (1982). Falcon visual sensitivity to grating contrast. *Nature*, 300(5887), 57-58. <https://doi.org/10.1038/300057a0>
- Hodos, W. (2012). What birds see and what they don't: luminance, contrast, and spatial and temporal resolution. In O. F. Lazareva, T. Shimizu, & E. A. Wasserman (Eds.), *How Animals See the World: Comparative Behavior, Biology, and Evolution of Vision* (pp. 5-24). Oxford University Press.
- Hodos, W., & Erichsen, J. T. (1990). Lower-field myopia in birds: an adaptation that keeps the ground in focus. *Vision Research*, 30(5), 653-657. [https://doi.org/10.1016/0042-6989\(90\)90091-X](https://doi.org/10.1016/0042-6989(90)90091-X)
- Hodos, W., Ghim, M. M., Potocki, A., Fields, J. N., & Storm, T. (2002). Contrast sensitivity in pigeons: a comparison of behavioral and pattern ERG methods. *Documenta Ophthalmologica*, 104(1), 107-118. <https://doi.org/10.1023/a:1014427615636>
- Hodos, W., & Leibowitz, R. W. (1977). Near-field visual acuity of pigeons: effects of scotopic adaptation and wavelength. *Vision Research*, 17(3), 463-467. [https://doi.org/10.1016/0042-6989\(77\)90040-2](https://doi.org/10.1016/0042-6989(77)90040-2)
- Hodos, W., Leibowitz, R. W., & Bonbright, J. C. (1976). Near-field visual-acuity of pigeons: effect of head location and stimulus luminance. *Journal of the Experimental Analysis of Behavior*, 25(2), 129-141. <https://doi.org/10.1901/jeab.1976.25-129>
- Hodos, W., Potocki, A., Ghim, M. M., & Gaffney, M. (2003). Temporal modulation of spatial contrast vision in pigeons (*Columba livia*). *Vision Research*, 43(7), 761-767. [https://doi.org/10.1016/S0042-6989\(02\)00417-0](https://doi.org/10.1016/S0042-6989(02)00417-0)
- Huang, S., Niu, X., Wang, J., Wang, Z., Xu, H., & Shi, L. (2022). Visual responses to moving and flashed stimuli of neurons in domestic pigeon (*Columba livia domestica*) optic tectum. *Animals*, 12(14), Article 1798. <https://doi.org/10.3390/ani12141798>
- Hughes, A. (1977). The topography of vision in mammals of contrasting life style: comparative optics and retinal organisation. In F. Crescitelli (Ed.), *The Visual System in Vertebrates* (pp. 613-756). Springer.
- Hunt, S., Kilner, R. M., Langmore, N. E., & Bennett, A. T. D. (2003). Conspicuous, ultraviolet-rich mouth colours in begging chicks. *Proceedings of the Royal Society B*, 270(suppl_1), S25-S28. <https://doi.org/doi:10.1098/rsbl.2003.0009>
- Ibbotson, M. R., & Meffin, H. (2020). Visual information processing. In B. Fritzsche (Ed.), *The Senses: A Comprehensive Reference* (2nd ed., Vol. 1, pp. 36-53). Elsevier. <https://doi.org/10.1016/B978-0-12-809324-5.24225-9>
- Inger, R., Bennie, J., Davies, T. W., & Gaston, K. J. (2014). Potential biological and ecological effects of flickering artificial light. *Plos One*, 9(5), Article e98631. <https://doi.org/10.1371/journal.pone.0098631>
- Intoy, J., & Rucci, M. (2020). Finely tuned eye movements enhance visual acuity. *Nature Communications*, 11, Article 795. <https://doi.org/10.1038/s41467-020-14616-2>
- Isa, T., Marquez-Legorreta, E., Grillner, S., & Scott, E. K. (2021). The tectum/superior colliculus as the vertebrate solution for spatial sensory integration and action. *Current Biology*, 31(11), R741-R762. <https://doi.org/10.1016/j.cub.2021.04.001>

- Jacobs, G. H. (1977). Visual capacities of the owl monkey (*Aotus trivirgatus*)-II. Spatial contrast sensitivity. *Vision Research*, 17(7), 821-825.
[https://doi.org/10.1016/0042-6989\(77\)90125-0](https://doi.org/10.1016/0042-6989(77)90125-0)
- Jarvis, J. R., Abeyesinghe, S. M., McMahon, C. E., & Wathes, C. M. (2009). Measuring and modelling the spatial contrast sensitivity of the chicken (*Gallus g. domesticus*). *Vision Research*, 49(11), 1448-1454.
<https://doi.org/10.1016/j.visres.2009.02.019>
- Jékely, G. (2009). Evolution of phototaxis. *Philosophical Transactions: Biological Sciences*, 364(1531), 2795-2808. <https://www.jstor.org/stable/40486053>
- Jiang, X., & Mahroo, O. A. (2022). Human retinal dark adaptation tracked in vivo with the electroretinogram: insights into processes underlying recovery of cone- and rod-mediated vision. *Journal of Physiology*, 600(21), 4603-4621.
<https://doi.org/10.1113/JP283105>
- Jimenez Ortega, L., Stoppa, K., Guentuerkuen, O., & Troje, N. F. (2008). Limits of intraocular and interocular transfer in pigeons. *Behavioural Brain Research*, 193(1), 69-78. <https://doi.org/10.1016/j.bbr.2008.04.022>
- Johnsen, S. (2012). *The Optics of Life: A Biologist's Guide to Light in Nature*. Princeton University Press.
- Jones, C. D., & Osorio, D. (2004). Discrimination of oriented visual textures by poultry chicks. *Vision Research*, 44(1), 83-89.
<https://doi.org/10.1016/j.visres.2003.08.014>
- Jourdie, V., Moureau, B., Bennett, A. T. D., & Heeb, P. (2004). Ultraviolet reflectance by the skin of nestlings. *Nature*, 431(7006), 262-262.
<https://doi.org/10.1038/431262a>
- Kano, F., Naik, H., Keskin, G., Couzin, I. D., & Nagy, M. (2022). Head-tracking of freely-behaving pigeons in a motion-capture system reveals the selective use of visual field regions. *Scientific Reports*, 12, Article 19113.
<https://doi.org/10.1038/s41598-022-21931-9>
- Kaplan, E. (2020). Luminance sensitivity and contrast detection. In B. Fritzsche (Ed.), *The Senses: A Comprehensive Reference* (2nd ed., pp. 199-211). Elsevier.
<https://doi.org/10.1016/B978-0-12-805408-6.00294-3>
- Kelber, A. (2019). Bird colour vision - from cones to perception. *Current Opinion in Behavioral Sciences*, 30, 34-40. <https://doi.org/10.1016/j.cobeha.2019.05.003>
- Kilner, R., & Davies, N. B. (1998). Nestling mouth colour: ecological correlates of a begging signal. *Animal Behaviour*, 56, 705-712.
<https://doi.org/10.1006/anbe.1998.0785>
- King-Smith, P. E., & Kulikowski, J. J. (1975). The detection of gratings by independent activation of line detectors. *The Journal of Physiology*, 247(2), 237-271.
<https://doi.org/10.1113/jphysiol.1975.sp010930>
- Kirwan, J. D., Bok, M. J., Smolka, J., Foster, J. J., Hernández, J. C., & Nilsson, D.-E. (2018). The sea urchin *Diadema africanum* uses low resolution vision to find shelter and deter enemies. *Journal of Experimental Biology*, 221(14), Article jeb176271. <https://doi.org/10.1242/jeb.176271>

- Kirwan, J. D., Graf, J., Smolka, J., Mayer, G., Henze, M. J., & Nilsson, D.-E. (2018). Low-resolution vision in a velvet worm (Onychophora). *Journal of Experimental Biology*, 221(11), Article jeb175802. <https://doi.org/10.1242/jeb.175802>
- Kirwan, J. D., & Nilsson, D.-E. (2019). A millipede compound eye mediating low-resolution vision. *Vision Research*, 165, 36-44. <https://doi.org/10.1016/j.visres.2019.09.003>
- Knudsen, E. I., & Schwarz, J. S. (2017). The optic tectum: a structure evolved for stimulus selection. In J. H. Kaas (Ed.), *Evolution of Nervous Systems* (2nd ed., pp. 387-408). Academic Press. <https://doi.org/10.1016/B978-0-12-804042-3.00016-6>
- Kral, K. (2003). Behavioural-analytical studies of the role of head movements in depth perception in insects, birds and mammals. *Behavioural Processes*, 64(1), 1-12. [https://doi.org/10.1016/S0376-6357\(03\)00054-8](https://doi.org/10.1016/S0376-6357(03)00054-8)
- Kress, D., van Bokhorst, E., & Lentink, D. (2015). How lovebirds maneuver rapidly using super-fast head saccades and image feature stabilization. *Plos One*, 10(7), Article e0133341. <https://doi.org/10.1371/journal.pone.0129287>
- Kröger, R. H. H., Campbell, M. C. W., Fernald, R. D., & Wagner, H. J. (1999). Multifocal lenses compensate for chromatic defocus in vertebrate eyes. *Journal of Comparative Physiology A*, 184(4), 361-369. <https://doi.org/10.1007/s003590050335>
- Kuffler, S. W. (1953). Discharge patterns and functional organization of mammalian retina. *Journal of Neurophysiology*, 16(1), 37-68. <https://doi.org/10.1152/jn.1953.16.1.37>
- Kugler, K., Luksch, H., Peremans, H., Vanderelst, D., Wiegrebe, L., & Firzlaff, U. (2019). Optic and echo-acoustic flow interact in bats. *Journal of Experimental Biology*, 222(6), Article jeb195404. <https://doi.org/10.1242/jeb.195404>
- Lafitte, A., Sordello, R., Legrand, M., Nicolas, V., Obein, G., & Reyjol, Y. (2022). A flashing light may not be that flashy: a systematic review on critical fusion frequencies. *Plos One*, 17(12), Article e0279718. <https://doi.org/10.1371/journal.pone.0279718>
- Lamb, T. D., Collin, S. P., & Pugh, E. N., Jr. (2007). Evolution of the vertebrate eye: opsins, photoreceptors, retina and eye cup. *Nature Reviews Neuroscience*, 8(12), 960-975. <https://doi.org/10.1038/nrn2283>
- Lamb, T. D., & Pugh, E. N. (2004). Dark adaptation and the retinoid cycle of vision. *Progress in Retinal and Eye Research*, 23(3), 307-380. <https://doi.org/10.1016/j.preteyeres.2004.03.001>
- LaMotte, R. H., & Brown, J. L. (1970). Dark adaptation and spectral sensitivity in the cat. *Vision Research*, 10(8), 703-716. [https://doi.org/10.1016/0042-6989\(70\)90017-9](https://doi.org/10.1016/0042-6989(70)90017-9)
- Land, M. F. (1999). Motion and vision: why animals move their eyes. *Journal of Comparative Physiology A*, 185, 341-352. <https://doi.org/10.1007/s003590050393>
- Land, M. F. (2014). Eye movements of vertebrates and their relation to eye form and function. *Journal of Comparative Physiology A*, 201(2), 195-214. <https://doi.org/10.1007/s00359-014-0964-5>
- Land, M. F. (2019). Eye movements in man and other animals. *Vision Research*, 162, 1-7. <https://doi.org/10.1016/j.visres.2019.06.004>

- Land, M. F., & Nilsson, D.-E. (2012). *Animal eyes* (2nd ed.). Oxford University Press.
- Lee, D. N., Davies, M. N. O., Green, P. R., & van der Weel, F. R. (1993). Visual control of velocity of approach by pigeons when landing. *Journal of Experimental Biology*, 180(1), 85-104. <https://doi.org/10.1242/jeb.180.1.85>
- Lee, D. N., & Kalmus, H. (1980). The optic flow field: The foundation of vision. *Philosophical Transactions of the Royal Society B*, 290(1038), 169-179. <http://www.jstor.org/stable/2395423>
- Lee, D. N., & Reddish, P. E. (1981). Plummeting gannets: A paradigm of ecological optics. *Nature*, 293(5830), 293-294. <https://doi.org/10.1038/293293a0>
- Legge, G. E. (1984). Binocular contrast summation—II. Quadratic summation. *Vision Research*, 24(4), 385-394. [https://doi.org/10.1016/0042-6989\(84\)90064-6](https://doi.org/10.1016/0042-6989(84)90064-6)
- Levitt, H. (1971). Transformed up-down methods in psychoacoustics. *The Journal of the Acoustical Society of America*, 49, 467-477. <https://doi.org/10.1121/1.1912375>
- Lewis, P., Rosen, R., Unsbo, P., & Gustafsson, J. (2011). Resolution of static and dynamic stimuli in the peripheral visual field. *Vision Research*, 51(16), 1829-1834. <https://doi.org/10.1016/j.visres.2011.06.011>
- Lie, I. (1980). Visual detection and resolution as a function of retinal locus. *Vision Research*, 20(11), 967-974. [https://doi.org/10.1016/0042-6989\(80\)90079-6](https://doi.org/10.1016/0042-6989(80)90079-6)
- Lind, O. (2016). Colour vision and background adaptation in a passerine bird, the zebra finch (*Taeniopygia guttata*). *Royal Society Open Science*, 3, Article 160383. <https://doi.org/10.1098/rsos.160383>
- Lind, O., Chavez, J., & Kelber, A. (2014). The contribution of single and double cones to spectral sensitivity in budgerigars during changing light conditions. *Journal of Comparative Physiology A*, 200(3), 197-207. <https://doi.org/10.1007/s00359-013-0878-7>
- Lind, O., Karlsson, S., & Kelber, A. (2013). Brightness discrimination in budgerigars (*Melopsittacus undulatus*). *Plos One*, 8(1), Article e54650. <https://doi.org/10.1371/journal.pone.0054650>
- Lind, O., & Kelber, A. (2009). The intensity threshold of colour vision in two species of parrot [Articl]. *Journal of Experimental Biology*, 212(22), 3693-3699. <https://doi.org/10.1242/jeb.035477>
- Lind, O., & Kelber, A. (2011). The spatial tuning of achromatic and chromatic vision in budgerigars. *Journal of Vision*, 11(7), Article 2. <https://doi.org/10.1167/11.7.2>
- Lind, O., Sunesson, T., Mitkus, M., & Kelber, A. (2012). Luminance-dependence of spatial vision in budgerigars (*Melopsittacus undulatus*) and Bourke's parrots (*Neopsephotus bourkii*). *Journal of Comparative Physiology A*, 198(1), 69-77. <https://doi.org/10.1007/s00359-011-0689-7>
- Lind, O. E., Kelber, A., & Kröger, R. H. H. (2008). Multifocal optical systems and pupil dynamics in birds. *Journal of Experimental Biology*, 211(17), 2752-2758. <https://doi.org/10.1242/jeb.018630>
- Lisney, T. J., Ekesten, B., Tauson, R., Håstad, O., & Ödeen, A. (2012). Using electroretinograms to assess flicker fusion frequency in domestic hens *Gallus*

- gallus domesticus*. *Vision Research*, 62, 125-133.
<https://doi.org/10.1016/j.visres.2012.04.002>
- Lisney, T. J., Iwaniuk, A. N., Bandet, M. V., & Wylie, D. R. (2012). Eye shape and retinal topography in owls (Aves: Strigiformes). *Brain, Behavior and Evolution*, 79(4), 218-236. <https://doi.org/10.1159/000337760>
- Lisney, T. J., Iwaniuk, A. N., Kolominsky, J., Bandet, M. V., Corfield, J. R., & Wylie, D. R. (2012). Interspecific variation in eye shape and retinal topography in seven species of galliform bird (Aves: Galliformes: Phasianidae). *Journal of Comparative Physiology A*, 198(10), 717-731. <https://doi.org/10.1007/s00359-012-0742-1>
- Lisney, T. J., Rubene, D., Rózsa, J., Løvlie, H., Håstad, O., & Ödeen, A. (2011). Behavioural assessment of flicker fusion frequency in chicken *Gallus gallus domesticus*. *Vision Research*, 51(12), 1324-1332.
<https://doi.org/10.1016/j.visres.2011.04.009>
- Lisney, T. J., Wylie, D. R., Kolominsky, J., & Iwaniuk, a. N. (2015). Eye morphology and retinal topography in hummingbirds (Trochilidae: Aves). *Brain, Behavior & Evolution*, 86(3-4), 176-190. <https://doi.org/10.1159/000441834>
- Lu, Z.-L., & Sperling, G. (2012). Black–white asymmetry in visual perception. *Journal of Vision*, 12(10), Article 8. <https://doi.org/10.1167/12.10.8>
- Maldonado, P. E., Maturana, H., & Varela, F. J. (1988). Frontal and lateral visual-system in birds - frontal and lateral gaze. *Brain Behavior and Evolution*, 32(1), 57-62.
<https://doi.org/10.1159/000116532>
- Malone, J. (2017). Animal psychophysics: The study of sensation in nonverbal organisms. In *APA handbook of comparative psychology: Perception, learning, and cognition*, Vol. 2. (pp. 3-24). American Psychological Association.
<https://doi.org/10.1037/0000012-001>
- Manglapus, M. K., Buelow, N. F., Barlow, R. B., & Uchiyama, H. (1998). Circadian rhythms of rod-cone dominance in the Japanese quail retina. *Journal of Neuroscience*, 18(12), 4775-4784. <https://doi.org/10.1523/jneurosci.18-12-04775.1998>
- Manglapus, M. K., Pierce, M. E., Barlow, R. B., Iuvone, P. M., & Underwood, H. (1999). Dopamine mediates circadian rhythms of rod-cone dominance in the Japanese quail retina. *Journal of Neuroscience*, 19(10), 4132-4141.
<https://doi.org/10.1523/jneurosci.19-10-04132.1999>
- Marquez-Legorreta, E., Piber, M., & Scott, E. K. (2020). Visual escape in larval zebrafish: stimuli, circuits, and behavior. In R. T. Gerlai (Ed.), *Behavioral and Neural Genetics of Zebrafish* (pp. 49-71). Academic Press. <https://doi.org/10.1016/B978-0-12-817528-6.00004-8>
- Martin, G. R. (1977). Absolute visual threshold and scotopic spectral sensitivity in the tawny owl *Strix aluco*. *Nature*, 268(5621), 636-638.
<https://doi.org/10.1038/268636a0>
- Martin, G. R. (1986). Total panoramic vision in the mallard duck, *Anas platyrhynchos*. *Vision Research*, 26(8), 1303-1305. [https://doi.org/10.1016/0042-6989\(86\)90112-4](https://doi.org/10.1016/0042-6989(86)90112-4)

- Martin, G. R. (1994). Visual fields in woodcocks *Scolopax rusticola* (Scolopacidae: Charadriiformes). *Journal of Comparative Physiology A*, 174(6), 787-793. <https://doi.org/10.1007/bf00192728>
- Martin, G. R. (1999). Eye structure and foraging in King Penguins *Aptenodytes patagonicus*. *Ibis*, 141(3), 444-450. <https://doi.org/10.1111/j.1474-919X.1999.tb04413.x>
- Martin, G. R. (2009). What is binocular vision for? A birds' eye view. *Journal of Vision*, 9(11), Article 14. <https://doi.org/10.1167/9.11.14>
- Martin, G. R. (2017a). *The Sensory Ecology of Birds*. Oxford University Press.
- Martin, G. R. (2017b). What drives bird vision? Bill control and predator detection overshadow flight. *Frontiers in Neuroscience*, 11, Article 619. <https://doi.org/10.3389/fnins.2017.00619>
- Martin, G. R., & Gordon, I. E. (1974). Visual acuity in the tawny owl (*Strix aluco*). *Vision Research*, 14(12), 1393-1397. [https://doi.org/10.1016/0042-6989\(74\)90014-5](https://doi.org/10.1016/0042-6989(74)90014-5)
- Martin, G. R., & Katzir, G. (1999). Visual fields in short-toed eagles, *Circus gallicus* (Accipitridae), and the function of binocularity in birds. *Brain, Behavior and Evolution*, 53(2), 55-66. <https://doi.org/10.1159/000006582>
- Martin, G. R., & Martin, R. O. (2020). Psittaciformes sensory systems. In J. Vonk & T. Shackelford (Eds.), *Encyclopedia of Animal Cognition and Behavior* (pp. 1-16). Springer International Publishing. https://doi.org/10.1007/978-3-319-47829-6_1704-1
- Martin, G. R., & Martin, R. O. (2022). Psittaciformes sensory systems. In J. Vonk & T. K. Shackelford (Eds.), *Encyclopedia of Animal Cognition and Behavior* (pp. 5767-5782). Springer International Publishing. https://doi.org/10.1007/978-3-319-55065-7_1704
- Martinez-Conde, S., & Macknik, S. L. (2008). Fixational eye movements across vertebrates: Comparative dynamics, physiology, and perception. *Journal of Vision*, 8(14), Article 28. <https://doi.org/10.1167/8.14.28>
- Martinoya, C., Le Houezec, J., & Bloch, S. (1984). Pigeon's eyes converge during feeding: Evidence for frontal binocular fixation in a lateral-eyed bird. *Neuroscience Letters*, 45(3), 335-339. [https://doi.org/10.1016/0304-3940\(84\)90248-9](https://doi.org/10.1016/0304-3940(84)90248-9)
- Martinoya, C., Rivaud, S., & Bloch, S. (1983). Comparing frontal and lateral viewing in the pigeon. II. Velocity thresholds for movement discrimination. *Behavioural Brain Research*, 8(3), 375-385. [https://doi.org/10.1016/0166-4328\(83\)90182-1](https://doi.org/10.1016/0166-4328(83)90182-1)
- Masland, R. H. (2001). The fundamental plan of the retina. *Nature Neuroscience*, 4(9), 877-886. <https://doi.org/10.1038/nn0901-877>
- Masland, Richard H. (2012). The neuronal organization of the retina. *Neuron*, 76(2), 266-280. <https://doi.org/10.1016/j.neuron.2012.10.002>
- Maziarz, M., & Wesolowski, T. (2014). Does darkness limit the use of tree cavities for nesting by birds? *Journal of Ornithology*, 155(3), 793-799. <https://doi.org/10.1007/s10336-014-1069-1>

- McFadden, S. A., & Wild, J. M. (1986). Binocular depth perception in the pigeon. *Journal of the Experimental Analysis of Behavior*, 45(2), 149-160. <https://doi.org/10.1901/jeab.1986.45-149>
- Meese, T. S., & Summers, R. J. (2012). Theory and data for area summation of contrast with and without uncertainty: Evidence for a noisy energy model. *Journal of Vision*, 12(11), Article 9. <https://doi.org/10.1167/12.11.9>
- Menkhorst, P., Rogers, D., Clarke, R., Davies, J., Marsack, P., & Franklin, K. (2017). *The Australian Bird Guide*. CSIRO Publishing.
- Meyer, D. B. (1977). The avian eye and its adaptations. In F. Crescitelli (Ed.), *The Visual System in Vertebrates* (Vol. 5, pp. 549-611). Springer Berlin Heidelberg. https://doi.org/10.1007/978-3-642-66468-7_10
- Mignon-Grasteau, S., Boissy, A., Bouix, J., Faure, J.-M., Fisher, A. D., Hinch, G. N., Jensen, P., Le Neindre, P., Mormède, P., Prunet, P., Vandeputte, M., & Beaumont, C. (2005). Genetics of adaptation and domestication in livestock. *Livestock Production Science*, 93(1), 3-14. <https://doi.org/10.1016/j.livprodsci.2004.11.001>
- Millodot, M., & Blough, P. (1971). The refractive state of the pigeon eye. *Vision Research*, 11(9), 1019-1022. [https://doi.org/10.1016/0042-6989\(71\)90223-9](https://doi.org/10.1016/0042-6989(71)90223-9)
- Miñano, S., Golodetz, S., Cavallari, T., & Taylor, G. K. (2023). Through hawks' eyes: Synthetically reconstructing the visual field of a bird in flight. *International Journal of Computer Vision*, 131(6), 1497-1531. <https://doi.org/10.1007/s11263-022-01733-2>
- Mitkus, M., Chaib, S., Lind, O., & Kelber, A. (2014). Retinal ganglion cell topography and spatial resolution of two parrot species: budgerigar (*Melopsittacus undulatus*) and Bourke's parrot (*Neopsephotus bourkii*). *Journal of Comparative Physiology A*, 200(5), 371-384. <https://doi.org/10.1007/s00359-014-0894-2>
- Mitkus, M., Olsson, P., Toomey, M. B., Corbo, J. C., & Kelber, A. (2017). Specialized photoreceptor composition in the raptor fovea. *Journal of Comparative Neurology*, 525(9), 2152-2163. <https://doi.org/10.1002/cne.24190>
- Moore, B. A., Pita, D., Tyrrell, L. P., & Fernández-Juricic, E. (2015). Vision in avian emberizid foragers: Maximizing both binocular vision and fronto-lateral visual acuity. *Journal of Experimental Biology*, 218(9), 1347-1358. <https://doi.org/10.1242/jeb.108613>
- Moore, B. A., Tyrrell, L. P., Kamilar, J. M., Collin, S. P., Dominy, N. J., Hall, M. I., Heesy, C. P., Lisney, T. J., Loew, E. R., Moritz, G. L., Nava, S. S., Warrant, E., Yopak, K. E., & Fernández-Juricic, E. (2017). Structure and function of regional specializations in the vertebrate retina. In J. H. Kaas (Ed.), *Evolution of Nervous Systems* (2nd ed., pp. 351-372). Academic Press. <https://doi.org/10.1016/B978-0-12-804042-3.00008-7>
- Mora, C. V., Davison, M., & Walker, M. M. (2009). Conditioning as a technique for studying the sensory systems involved in animal orientation, homing and navigation – a review. *The Journal of Navigation*, 62(4), 571-585. <https://doi.org/10.1017/S0373463309990063>

- Moreno, J., Morales, J., Lobato, E., Merino, S., Tomás, G., & Martínez-De La Puente, J. (2005). Evidence for the signaling function of egg color in the pied flycatcher *Ficedula hypoleuca*. *Behavioral Ecology*, 16(5), 931-937. <https://doi.org/10.1093/beheco/ari072>
- Moroney, M. K., & Pettigrew, J. D. (1987). Some observations on the visual optics of kingfishers (Aves, Coraciformes, Alcedinidae). *Journal of Comparative Physiology A*, 160(2), 137-149. <https://doi.org/10.1007/BF00609721>
- Murphy-Baum, B. L., Schwartz, G. W., & Awatramani, G. B. (2021). Direction selectivity. In G. W. Schwartz (Ed.), *Retinal Computation* (pp. 200-229). Academic Press. <https://doi.org/10.1016/B978-0-12-819896-4.00012-3>
- Nilsson, D.-E., Smolka, J., & Bok, M. (2022). The vertical light-gradient and its potential impact on animal distribution and behavior. *Frontiers in Ecology and Evolution*, 10, Article 951328. <https://doi.org/10.3389/fevo.2022.951328>
- Niu, X., Huang, S., Yang, S., Wang, Z., Li, Z., & Shi, L. (2020). Comparison of pop-out responses to luminance and motion contrasting stimuli of tectal neurons in pigeons. *Brain Research*, 1747, Article 147068. <https://doi.org/10.1016/j.brainres.2020.147068>
- Northmore, D. P. M., & Dvorak, C. A. (1979). Contrast sensitivity and acuity of the goldfish. *Vision Research*, 19(3), 255-261. [https://doi.org/10.1016/0042-6989\(79\)90171-8](https://doi.org/10.1016/0042-6989(79)90171-8)
- Northmore, D. P. M., Oh, D. J., & Celenza, M. A. (2007). Acuity and contrast sensitivity of the bluegill sunfish and how they change during optic nerve regeneration. *Visual Neuroscience*, 24(3), 319-331. <https://doi.org/10.1017/S0952523807070307>
- O'Carroll, D. C., & Wiederman, S. D. (2014). Contrast sensitivity and the detection of moving patterns and features. *Philosophical Transactions of the Royal Society B*, 369(1636), Article 20130043. <https://doi.org/10.1098/rstb.2013.0043>
- Olsson, P., Lind, O., & Kelber, A. (2018). Chromatic and achromatic vision: parameter choice and limitations for reliable model predictions. *Behavioral Ecology*, 29(2), 273-282. <https://doi.org/10.1093/beheco/arx133>
- Osorio, D., Mikló, A., & Gonda, Z. (1999). Visual ecology and perception of coloration patterns by domestic chicks. *Evolutionary Ecology*, 13, 673-689. <https://doi.org/10.1023/A:1011059715610>
- Ott, M. (2006). Visual accommodation in vertebrates: mechanisms, physiological response and stimuli. *Journal of Comparative Physiology A*, 192(2), 97-111. <https://doi.org/10.1007/s00359-005-0049-6>
- Perlman, I., & Normann, R. A. (1998). Light adaptation and sensitivity controlling mechanisms in vertebrate photoreceptors. *Progress in Retinal and Eye Research*, 17(4), 523-563. [https://doi.org/10.1016/s1350-9462\(98\)00005-6](https://doi.org/10.1016/s1350-9462(98)00005-6)
- Pettigrew, J. D., Dreher, B., Hopkins, C. S., McCall, M. J., & Brown, M. (1988). Peak density and distribution of ganglion cells in the retinae of microchiropteran bats: Implications for visual acuity. *Brain, Behavior and Evolution*, 32(1), 39-56. <https://doi.org/10.1159/000116531>

- Pettigrew, J. D., Wallman, J., & Wildsoet, C. F. (1990). Saccadic oscillations facilitate ocular perfusion from the avian pecten. *Nature*, 343(6256), 362-363. <https://doi.org/10.1038/343362a0>
- Podkowa, P., Malinowska, K., & Surmacki, A. (2019). Light affects parental provisioning behaviour in a cavity-nesting passerine. *Journal of Avian Biology*, 50(11), Article e02254. <https://doi.org/10.1111/jav.02254>
- Podkowa, P., & Surmacki, A. (2017). The importance of illumination in nest site choice and nest characteristics of cavity nesting birds. *Scientific Reports*, 7, Article 1329. <https://doi.org/10.1038/s41598-017-01430-y>
- Potier, S., Bonadonna, F., Kelber, A., Martin, G. R., Isard, P. F., Dulaurent, T., & Duriez, O. (2016). Visual abilities in two raptors with different ecology. *Journal of Experimental Biology*, 219(17), 2639-2649. <https://doi.org/10.1242/jeb.142083>
- Potier, S., Lieuvain, M., Pfaff, M., & Kelber, A. (2020). How fast can raptors see? *Journal of Experimental Biology*, 223(1), Article jeb209031. <https://doi.org/10.1242/jeb.209031>
- Potier, S., Mitkus, M., & Kelber, A. (2018). High resolution of colour vision, but low contrast sensitivity in a diurnal raptor. *Proceedings of the Royal Society B*, 285(1885), Article 20181036. <https://doi.org/doi:10.1098/rspb.2018.1036>
- Potier, S., Mitkus, M., & Kelber, A. (2020). Visual adaptations of diurnal and nocturnal raptors. *Seminars in Cell & Developmental Biology*, 106, 116-126. <https://doi.org/10.1016/j.semcdb.2020.05.004>
- Pratt, J., Radulescu, P. V., Guo, R. M., & Abrams, R. A. (2010). It's alive!: Animate motion captures visual attention. *Psychological Science*, 21(11), 1724-1730. <https://doi.org/10.1177/0956797610387440>
- Procacci, N. M., Allen, K. M., Robb, G. E., Ijekah, R., Lynam, H., & Hoy, J. L. (2020). Context-dependent modulation of natural approach behaviour in mice. *Proceedings of the Royal Society B*, 287(1934), Article 20201189. <https://doi.org/doi:10.1098/rspb.2020.1189>
- Querubin, A., Hie, R. L., Provis, J. M., & Bumsted O'Brien, K. M. (2009). Photoreceptor and ganglion cell topographies correlate with information convergence and high acuity regions in the adult pigeon (*Columba livia*) retina. *Journal of Comparative Neurology*, 517(5), 711-722. <https://doi.org/10.1002/cne.22178>
- Rahman, M. L., Sugita, S., Aoyama, M., & Sugita, S. (2006). Number, distribution and size of retinal ganglion cells in the jungle crow (*Corvus macrorhynchos*). *Anatomical Science International*, 81(4), 253-259. <https://doi.org/10.1111/j.1447-073x.2006.00151.x>
- Raudies, F., Mingolla, E., & Neumann, H. (2012). Active gaze control improves optic flow-based segmentation and steering. *Plos One*, 7(6), Article e38446. <https://doi.org/10.1371/journal.pone.0038446>
- Read, J. C. A. (2021). Binocular vision and stereopsis across the animal kingdom. *Annual Review of Vision Science*, 7, 389-415. <https://doi.org/10.1146/annurev-vision-093019-113212>

- Remy, M., & Emmerton, J. (1991). Directional dependence of intraocular transfer of stimulus detection in pigeons (*Columba livia*). *Behavioral Neuroscience*, 105(5), 647-652. <https://doi.org/10.1037/0735-7044.105.5.647>
- Reymond, L. (1985). Spatial visual acuity of the eagle *Aquila audax*: A behavioural, optical and anatomical investigation. *Vision Research*, 25(10), 1477-1491. [https://doi.org/10.1016/0042-6989\(85\)90226-3](https://doi.org/10.1016/0042-6989(85)90226-3)
- Reymond, L. (1987). Spatial visual acuity of the falcon, *Falco berigora*: A behavioural, optical and anatomical investigation. *Vision Research*, 27(10), 1859-1874. [https://doi.org/10.1016/0042-6989\(87\)90114-3](https://doi.org/10.1016/0042-6989(87)90114-3)
- Reymond, L., & Wolfe, J. (1981). Behavioural determination of the contrast sensitivity function of the eagle *Aquila audax*. *Vision Research*, 21(2), 263-271. [https://doi.org/10.1016/0042-6989\(81\)90120-6](https://doi.org/10.1016/0042-6989(81)90120-6)
- Reynolds, S. J., Martin, G. R., & Cassey, P. (2009). Is sexual selection blurring the functional significance of eggshell coloration hypotheses? *Animal Behaviour*, 78(1), 209-215. <https://doi.org/10.1016/j.anbehav.2009.03.003>
- Rider, A. T., Henning, G. B., & Stockman, A. (2019). Light adaptation controls visual sensitivity by adjusting the speed and gain of the response to light. *Plos One*, 14(8), e0220358. <https://doi.org/10.1371/journal.pone.0220358>
- Rider, A. T., Henning, G. B., & Stockman, A. (2022). A reinterpretation of critical flicker-frequency (CFF) data reveals key details about light adaptation and normal and abnormal visual processing. *Progress in Retinal and Eye Research*, 87, Article 101001. <https://doi.org/10.1016/j.preteyeres.2021.101001>
- Rieke, F., & Rudd, M. E. (2009). The challenges natural images pose for visual adaptation. *Neuron*, 64(5), 605-616. <https://doi.org/10.1016/j.neuron.2009.11.028>
- Riggs, L. A., Ratliff, F., Cornsweet, J. C., & Cornsweet, T. N. (1953). The disappearance of steadily fixated visual test objects. *Journal of the Optical Society of America*, 43(6), 495-501. <https://doi.org/10.1364/JOSA.43.000495>
- Roberts, W. A., Phelps, M. T., Macuda, T., Brodbeck, D. R., & Russ, T. (1996). Intraocular transfer and simultaneous processing of stimuli presented in different visual fields of the pigeon. *Behavioral Neuroscience*, 110(2), 290-299. <https://doi.org/10.1037/0735-7044.110.2.290>
- Robson, J. G. (1966). Spatial and temporal contrast-sensitivity functions of the visual system. *Journal of the Optical Society of America*, 56(8), 1141-1142.
- Robson, J. G., & Graham, N. (1981). Probability summation and regional variation in contrast sensitivity across the visual field. *Vision Research*, 21(3), 409-418. [https://doi.org/10.1016/0042-6989\(81\)90169-3](https://doi.org/10.1016/0042-6989(81)90169-3)
- Ros, I. G., & Biewener, A. A. (2017). Pigeons (*C. livia*) follow their head during turning light: Head stabilization underlies the visual control of flight. *Frontiers in Neuroscience*, 11, Article 655. <https://doi.org/10.3389/fnins.2017.00655>
- Rosa-Salva, O., Grassi, M., Lorenzi, E., Regolin, L., & Vallortigara, G. (2016). Spontaneous preference for visual cues of animacy in naïve domestic chicks: The case of speed changes. *Cognition*, 157, 49-60. <https://doi.org/10.1016/j.cognition.2016.08.014>

- Rossi, E. A., & Roorda, A. (2010). The relationship between visual resolution and cone spacing in the human fovea. *Nature Neuroscience*, 13(2), 156-157.
<https://doi.org/10.1038/nn.2465>
- Rounsley, K. J., & McFadden, S. A. (2005). Limits of visual acuity in the frontal field of the rock pigeon (*Columba livia*). *Perception*, 34(8), 983-993.
<https://doi.org/10.1068/p5205>
- Rucci, M., Ahissar, E., & Burr, D. (2018). Temporal coding of visual space. *Trends in Cognitive Sciences*, 22(10), 883-895. <https://doi.org/10.1016/j.tics.2018.07.009>
- Rucci, M., Iovin, R., Poletti, M., & Santini, F. (2007). Miniature eye movements enhance fine spatial detail. *Nature*, 447(7146), 852-855.
<https://doi.org/10.1038/nature05866>
- Rushton, S. K., Bradshaw, M. F., & Warren, P. A. (2007). The pop out of scene-relative object movement against retinal motion due to self-movement. *Cognition*, 105(1), 237-245. <https://doi.org/10.1016/j.cognition.2006.09.004>
- Santon, M., Münch, T. A., & Michiels, N. K. (2019). The contrast sensitivity function of a small cryptobenthic marine fish. *Journal of Vision*, 19(2), Article 1.
<https://doi.org/10.1167/19.2.1>
- Sandow, L. -M., & Hanke, F. D. (2024). Aerial single target acuity of harbor seals (*Phoca vitulina*) for stationary and moving targets of varying contrast. *Vision Research*, 218, Article 108389. <https://doi.org/10.1016/j.visres.2024.108389>
- Schaeffel, F., Hagel, G., Eikermann, J., & Collett, T. (1994). Lower-field myopia and astigmatism in amphibians and chickens. *Journal of the Optical Society of America A*, 11(2), 487-495. <https://doi.org/10.1364/JOSAA.11.000487>
- Schaeffel, F., Rohrer, B., Lemmer, T., & Zrenner, E. (1991). Diurnal control of rod function in the chicken. *Visual Neuroscience*, 6(6), 641-653.
<https://doi.org/10.1017/s0952523800002637>
- Schiffner, I., & Srinivasan, M. V. (2015). Direct evidence for vision-based control of flight speed in budgerigars. *Scientific Reports*, 5, Article 10992.
<https://doi.org/10.1038/srep10992>
- Schiffner, I., & Srinivasan, M. V. (2016). Budgerigar flight in a varying environment: Flight at distinct speeds? *Biology Letters*, 12(6), Article 20160221.
<https://doi.org/10.1098/rsbl.2016.0221>
- Scholtyssek, C., Dacke, M., Kröger, R., & Baird, E. (2014). Control of self-motion in dynamic fluids: fish do it differently from bees. *Biology Letters*, 10(5), 20140279.
<https://doi.org/10.1098/rsbl.2014.0279>
- Schrader, N. (1975). The breeding of budgerigars in Western N. S. W. *Australian Bird Watcher*, 6, 118-122.
- Schwartz, G. W., & Levine, J. (2021). Luminance adaptation. In G. W. Schwartz (Ed.), *Retinal Computation* (pp. 26-46). Academic Press. <https://doi.org/10.1016/B978-0-12-819896-4.00003-2>
- Schwartz, G. W., & Swygart, D. (2020). Circuits for feature selectivity in the inner retina. In B. Fritsch (Ed.), *The Senses: A Comprehensive Reference* (2:nd ed., pp. 275-292). Elsevier. <https://doi.org/10.1016/B978-0-12-809324-5.24186-2>

- Schwartz, G. W., & Swygart, D. (2021). Object motion sensitivity. In G. W. Schwartz (Ed.), *Retinal Computation* (pp. 230-244). Academic Press.
<https://doi.org/10.1016/B978-0-12-819896-4.00013-5>
- Seifert, M., Baden, T., & Osorio, D. (2020). The retinal basis of vision in chicken. *Seminars in Cell & Developmental Biology*, 106, 106-115.
<https://doi.org/10.1016/j.semcdb.2020.03.011>
- Seifert, M., Roberts, P. A., Kafetzis, G., Osorio, D., & Baden, T. (2023). Birds multiplex spectral and temporal visual information via retinal On- and Off-channels. *Nature Communications*, 14(1), 5308. <https://doi.org/10.1038/s41467-023-41032-z>
- Shapley, R., & Enroth-Cugell, C. (1984). Visual adaptation and retinal gain controls. *Progress in Retinal Research*, 3, 263-346. [https://doi.org/10.1016/0278-4327\(84\)90011-7](https://doi.org/10.1016/0278-4327(84)90011-7)
- Shi, Q., & Stell, W. K. (2013). Die fledermaus: Regarding optokinetic contrast sensitivity and light-adaptation, chicks are mice with wings. *Plos One*, 8(9), Article e75375. <https://doi.org/10.1371/journal.pone.0075375>
- Shi, Q., Teves, M. M., Lillywhite, A., Pagtalunan, E. B., & Stell, W. K. (2020). Light adaptation in the chick retina: Dopamine, nitric oxide, and gap-junction coupling modulate spatiotemporal contrast sensitivity. *Experimental Eye Research*, 195, Article 108026. <https://doi.org/10.1016/j.exer.2020.108026>
- Shimizu, T., & Watanabe, S. (2012). The avian visual system: Overview. In O. F. Lazareva, T. Shimizu, & E. A. Wasserman (Eds.), *How Animals See the World: Comparative Behavior, Biology, and Evolution of Vision*. (pp. 473-482). Oxford University Press.
- Siddiqi, A., Cronin, T. W., Loew, E. R., Vorobyev, M., & Summers, K. (2004). Interspecific and intraspecific views of color signals in the strawberry poison frog *Dendrobates pumilio*. *Journal of Experimental Biology*, 207(14), 2471-2485.
<https://doi.org/10.1242/jeb.01047>
- Sivak, J. G. (1976). The accommodative significance of the "ramp" retina of the eye of the stingray. *Vision Research*, 16(9), 945-950, IN942-IN943.
[https://doi.org/10.1016/0042-6989\(76\)90225-X](https://doi.org/10.1016/0042-6989(76)90225-X)
- Skinner, B. F. (1957). The experimental analysis of behavior. *American Scientist*, 45(4), 343-371. <http://www.jstor.org.ludwig.lub.lu.se/stable/27826953>
- Snyder, A. W., & Miller, W. H. (1977). Photoreceptor diameter and spacing for highest resolving power. *Journal of the Optical Society of America*, 67(5), 696-698.
<https://doi.org/10.1364/JOSA.67.000696>
- Solomon, S. G., Janbon, H., Bimson, A., & Wheatcroft, T. (2023). Visual spatial location influences selection of instinctive behaviours in mouse. *Royal Society Open Science*, 10(4), Article 230034. <https://doi.org/10.1098/rsos.230034>
- Somanathan, H., Borges, R. M., Warrant, E. J., & Kelber, A. (2017). Visual adaptations for mate detection in the male Carpenter bee *Xylocopa tenuiscapa*. *Plos One*, 12(1), Article e0168452. <https://doi.org/10.1371/journal.pone.0168452>
- Souza, G. d. S., Gomes, B. D., & Silveira, L. C. d. L. (2011). Comparative neurophysiology of spatial luminance contrast sensitivity. *Psychology and Neuroscience*, 4(1), 29-48. <https://doi.org/10.3922/j.psns.2011.1.005>

- Spratte, C., Sandow, L.-M., Schnermann, J. S., & Hanke, F. D. (2021). Single target acuity in the common sunfish (*Lepomis gibbosus*). *Journal of Experimental Biology*, 224(20), Article jeb243068. <https://doi.org/10.1242/jeb.243068>
- Sridharan, D., Ramamurthy, D. L., Schwarz, J. S., & Knudsen, E. I. (2014). Visuospatial selective attention in chickens. *Proceedings of the National Academy of Sciences*, 111(19), E2056-E2065. <https://doi.org/doi:10.1073/pnas.1316824111>
- Stamp Dawkins, M. (2002). What are birds looking at? Head movements and eye use in chickens. *Animal Behaviour*, 63(5), 991-998. <https://doi.org/10.1006/anbe.2002.1999>
- Stamps, J., Clark, A., Arrowood, P., & Kus, B. (1985). Parent-offspring conflict in budgerigars. *Behaviour*, 94(1/2), 1-40. <https://doi.org/10.1163/156853985X00253>
- Stamps, J., Clark, A., Arrowood, P., & Kus, B. (1989). Begging behavior in budgerigars. *Ethology*, 81, 177-192. <https://doi.org/10.1111/j.1439-0310.1989.tb00765.x>
- Stamps, J., Clark, A., Kus, B., & Arrowood, P. (1987). The effects of parent and offspring gender on food allocation in budgerigars. *Behaviour*, 101, 177-199. <https://doi.org/10.1163/156853987x00422>
- Storchi, R., Rodgers, J., Gracey, M., Martial, F. P., Wynne, J., Ryan, S., Twining, C. J., Cootes, T. F., Killick, R., & Lucas, R. J. (2019). Measuring vision using innate behaviours in mice with intact and impaired retina function. *Scientific Reports*, 9(1), Article 10396. <https://doi.org/10.1038/s41598-019-46836-y>
- Sumner-Rooney, L., Kirwan, J. D., Lowe, E., & Ullrich-Lüter, E. (2020). Extraocular vision in a brittle star Is mediated by chromatophore movement in response to ambient light. *Current Biology*, 30(2), 319-327. <https://doi.org/10.1016/j.cub.2019.11.042>
- Sun, H. J., & Frost, B. J. (1997). Motion processing in pigeon tectum: Equiluminant chromatic mechanisms. *Experimental Brain Research*, 116(3), 434-444. <https://doi.org/10.1007/PL00005771>
- Temple, S. E. (2011). Why different regions of the retina have different spectral sensitivities: A review of mechanisms and functional significance of intraretinal variability in spectral sensitivity in vertebrates. *Visual Neuroscience*, 28(4), 281-293. <https://doi.org/10.1017/S0952523811000113>
- Thibos, L. N., Bradley, A., Xu, R., & Lopez-Gil, N. (2019). Ricco's law and absolute threshold for foveal detection of black holes. *Journal of the Optical Society of America A*, 36(4), B35-B43. <https://doi.org/10.1364/JOSAA.36.000B35>
- Tinbergen, N., & Perdeck, A. C. (1950). On the stimulus situation releasing the begging response in the newly hatched herring gull chick (*Larus argentatus argentatus* pont.). *Behaviour*, 3(1), 1-39. <https://www.jstor.org/stable/4532715>
- Tisdale, V., & Fernández-Juricic, E. (2009). Vigilance and predator detection vary between avian species with different visual acuity and coverage. *Behavioral Ecology*, 20(5), 936-945. <https://doi.org/10.1093/beheco/arp080>
- Toomey, M. B., & Corbo, J. C. (2017). Evolution, development and function of vertebrate cone oil droplets. *Frontiers in Neural Circuits*, 11, Article 97. <https://doi.org/10.3389/fncir.2017.00097>

- Tremoulet, P. D., & Feldman, J. (2000). Perception of animacy from the motion of a single object. *Perception*, 29(8), 943-951. <https://doi.org/10.1068/p3101>
- Troscianko, J., Chappell, J., Martin, G. R., Von Bayern, A. M. P., & Rutz, C. (2012). Extreme binocular vision and a straight bill facilitate tool use in New Caledonian crows. *Nature Communications*, 3, Article 1110. <https://doi.org/10.1038/ncomms2111>
- Tsushima, Y., & Watanabe, T. (2009). Roles of attention in perceptual learning from perspectives of psychophysics and animal learning. *Learning and Behavior*, 37(2), 126-132. <https://doi.org/10.3758/lb.37.2.126>
- Tuten, W. S., Cooper, R. F., Tiruveedhula, P., Dubra, A., Roorda, A., Cottaris, N. P., Brainard, D. H., & Morgan, J. I. W. (2018). Spatial summation in the human fovea: Do normal optical aberrations and fixational eye movements have an effect? *Journal of Vision*, 18(8), Article 6. <https://doi.org/10.1167/18.8.6>
- Tyrrell, L. P., Butler, S. R., Yorzinski, J. L., & Fernández-Juricic, E. (2014). A novel system for bi-ocular eye-tracking in vertebrates with laterally placed eyes. *Methods in Ecology and Evolution*, 5(10), 1070-1077. <https://doi.org/10.1111/2041-210X.12249>
- Tyrrell, L. P., & Fernandez-Juricic, E. (2017). The hawk-eyed songbird: Retinal morphology, eye shape, and visual fields of an aerial insectivore. *American Naturalist*, 189(6), 709-717. <https://doi.org/10.1086/691404>
- Tyrrell, L. P., & Fernández-Juricic, E. (2017). Avian binocular vision: It's not just about what birds can see, it's also about what they can't. *Plos One*, 12(3), e0173235. <https://doi.org/10.1371/journal.pone.0173235>
- Vallet, A. M., & Coles, J. A. (1993). The perception of small objects by the drone honeybee. *Journal of Comparative Physiology A*, 172(2), 183-188. <https://doi.org/10.1007/BF00189395>
- van den Berg, C. P., Hollenkamp, M., Mitchell, L. J., Watson, E. J., Green, N. F., Marshall, N. J., & Cheney, K. L. (2020). More than noise: Context-dependent luminance contrast discrimination in a coral reef fish (*Rhinecanthus aculeatus*). *Journal of Experimental Biology*, 223(21), Article jeb232090. <https://doi.org/10.1242/jeb.232090>
- van der Willigen, R. F. (2011). Owls see in stereo much like humans do. *Journal of Vision*, 11(7), Article 10. <https://doi.org/10.1167/11.7.10>
- van der Willigen, R. F., Frost, B. J., & Wagner, H. (1998). Stereoscopic depth perception in the owl. *NeuroReport*, 9(6), 1233-1237. <https://doi.org/10.1097/00001756-199804200-00050>
- Volbrecht, V. J., Shrago, E. E., Scheffrin, B. E., & Warner, J. S. (2000). Spatial summation in human cone mechanisms from 0° to 20° in the superior retina. *Journal of the Optical Society of America A* 17(3), 641-650. <https://doi.org/10.1364/josaa.17.000641>
- Vorobyev, M., Brandt, R., Peitsch, D., Laughlin, S. B., & Menzel, R. (2001). Colour thresholds and receptor noise: Behaviour and physiology compared. *Vision Research*, 41(5), 639-653. [https://doi.org/10.1016/S0042-6989\(00\)00288-1](https://doi.org/10.1016/S0042-6989(00)00288-1)

- Vorobyev, M., & Osorio, D. (1998). Receptor noise as a determinant of colour thresholds. *Proceedings of the Royal Society B*, 265(1394), 351-358. <https://doi.org/10.1098/rspb.1998.0302>
- Voss, J., & Bischof, H. J. (2009). Eye movements of laterally eyed birds are not independent. *Journal of Experimental Biology*, 212(10), 1568-1575. <https://doi.org/10.1242/jeb.024950>
- Wagner, H., Pappé, I., & Nalbach, H. O. (2022). Optocollic responses in adult barn owls (*Tyto furcata*). *Journal of Comparative Physiology A*, 208(2), 239-251. <https://doi.org/10.1007/s00359-021-01524-z>
- Walls, G. L. (1942). *The Vertebrate Eye and its Adaptive Radiation*. Cranbrook Institute of Science.
- Warrant, E. J. (1999). Seeing better at night: Life style, eye design and the optimum strategy of spatial and temporal summation. *Vision Research*, 39(9), 1611-1630. [https://doi.org/10.1016/S0042-6989\(98\)00262-4](https://doi.org/10.1016/S0042-6989(98)00262-4)
- Warren Jr, W. H., Morris, M. W., & Kalish, M. (1988). Perception of translational heading from optical flow. *Journal of Experimental Psychology: Human Perception and Performance*, 14(4), 646-660. <https://doi.org/10.1037/0096-1523.14.4.646>
- Warwick, R. A., Heukamp, A. S., Riccitelli, S., & Rivlin-Etzion, M. (2023). Dopamine differentially affects retinal circuits to shape the retinal code. *The Journal of Physiology*, 601(7), 1265-1286. <https://doi.org/10.1113/JP284215>
- Wells, M. C., Lehner, P. N., Bolen, E. G., & Rylander, M. K. (1975). Comparison of scotopic sensitivity in diurnal (*Anas platyrhynchos*) and crepuscular (*Dendrocygna autumnalis*) ducks *Journal of Comparative Physiological Psychology*, 88(2), 940-944. <https://doi.org/10.1037/h0076444>
- Wesołowski, T. (2007). Lessons from long-term hole-nester studies in a primeval temperate forest. *Journal of Ornithology*, 148(2), 395-405. <https://doi.org/10.1007/s10336-007-0198-1>
- Wesolowski, T., & Maziarz, M. (2012). Dark tree cavities - a challenge for hole nesting birds? *Journal of Avian Biology*, 43(5), 454-460. <https://doi.org/10.1111/j.1600-048X.2012.05704.x>
- Westheimer, G. (1975). Visual acuity and hyperacuity. *Investigative Ophthalmology*, 14(8), 570-572.
- Westheimer, G. (2009a). Hyperacuity. In L. R. Squire (Ed.), *Encyclopedia of Neuroscience* (pp. 45-50). Oxford: Academic Press.
- Westheimer, G. (2009b). Visual acuity: Information theory, retinal image structure and resolution thresholds. *Progress in Retinal and Eye Research*, 28(3), 178-186. <https://doi.org/10.1016/j.preteyeres.2009.04.001>
- Westheimer, G., & McKee, S. P. (1977). Spatial configurations for visual hyperacuity. *Vision Research*, 17(8), 941-947. [https://doi.org/10.1016/0042-6989\(77\)90069-4](https://doi.org/10.1016/0042-6989(77)90069-4)
- Whittle, P. (1992). Brightness, discriminability and the crispening effect. *Vision Research*, 32(8), 1493-1507. [https://doi.org/10.1016/0042-6989\(92\)90205-w](https://doi.org/10.1016/0042-6989(92)90205-w)

- Wiebe, K. L., & Slagsvold, T. (2009). Mouth coloration in nestling birds: Increasing detection or signalling quality? *Animal Behaviour*, 78(6), 1413-1420. <https://doi.org/10.1016/j.anbehav.2009.09.013>
- Wiebe, K. L., & Slagsvold, T. (2012). Parents take both size and conspicuousness into account when feeding nestlings in dark cavity nests. *Animal Behaviour*, 84(6), 1307-1312. <https://doi.org/10.1016/j.anbehav.2012.09.014>
- Wilby, D., & Roberts, N. W. (2017). Optical influence of oil droplets on cone photoreceptor sensitivity. *Journal of Experimental Biology*, 220(11), 1997-2004. <https://doi.org/10.1242/jeb.152918>
- Wilkie, D. M., & Saksida, L. M. (1994). The spatial distribution of pigeons' target detection. *Behavioural Brain Research*, 60(2), 137-140. [https://doi.org/10.1016/0166-4328\(94\)90140-6](https://doi.org/10.1016/0166-4328(94)90140-6)
- Williams, D. R., & Coletta, N. J. (1987). Cone spacing and the visual resolution limit. *Journal of the Optical Society of America A*, 4(8), 1514-1523. <https://doi.org/10.1364/JOSAA.4.001514>
- Williams, W. A. (2022). Innate Behaviors. In J. Vonk & T. K. Shackelford (Eds.), *Encyclopedia of Animal Cognition and Behavior* (pp. 3445-3454). Cham: Springer.
- Wurtz, R. H. (1998). Optic flow: A brain region devoted to optic flow analysis? *Current Biology*, 8(16), R554-R556. [https://doi.org/10.1016/s0960-9822\(07\)00359-4](https://doi.org/10.1016/s0960-9822(07)00359-4)
- Wylie, D. (2013). Processing of visual signals related to self-motion in the cerebellum of pigeons. *Frontiers in Behavioral Neuroscience*, 7. <https://doi.org/10.3389/fnbeh.2013.00004>
- Wyndham, E. (1980a). Diurnal cycle, behavior and social organization of the budgerigar *Melopsittacus undulatus*. *Emu*, 80, 25-33. <https://doi.org/10.1071/mu9800025>
- Wyndham, E. (1980b). Environment and food of the budgerigar *Melopsittacus undulatus*. *Australian Journal of Ecology*, 5(1), 47-61. <https://doi.org/10.1111/j.1442-9993.1980.tb01231.x>
- Wyndham, E. (1981). Breeding and Mortality of Budgerigars *Melopsittacus Undulatus*. *Emu*, 81(4), 240-243. <https://doi.org/10.1071/MU9810240>
- Yang, C., Møller, A. P., & Liang, W. (2022). Light matters: Nest illumination alters egg rejection behavior in a cavity-nesting bird. *Avian Research*, 13, Article 100016. <https://doi.org/10.1016/j.avrs.2022.100016>
- Yovanovich, C. A. M., Koskela, S. M., Nevala, N., Kondrashev, S. L., Kelber, A., & Donner, K. (2017). The dual rod system of amphibians supports colour discrimination at the absolute visual threshold. *Philosophical Transactions of the Royal Society B*, 372(1717), Article 20160066. <https://doi.org/10.1098/rstb.2016.0066>
- Zahar, Y., Wagner, H., & Gutfreund, Y. (2012). Responses of tectal neurons to contrasting stimuli: An electrophysiological study in the barn owl. *Plos One*, 7(6), Article e39559. <https://doi.org/10.1371/journal.pone.0039559>
- Zhang, J. X., Wei, W., Zhang, J. H., & Yang, W. H. (2010). Uropygial gland-secreted alkanols contribute to olfactory sex signals in budgerigars. *Chemical Senses*, 35(5), 375-382. <https://doi.org/10.1093/chemse/bjq025>

Ölveczky, B. P., Baccus, S. A., & Meister, M. (2003). Segregation of object and background motion in the retina. *Nature*, 423(6938), 401-408.
<https://doi.org/10.1038/nature01652>

Paper I





Single target acuity is not higher than grating acuity in a bird, the budgerigar

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ARTICLE INFO

Keywords:

Target detection
Target acuity
Spatial resolution
Bird vision
Contrast sensitivity
Visual acuity

ABSTRACT

We examined the capacity of budgerigars (*Melopsittacus undulatus*) to visually detect dark single targets against a brighter background and established their spatial resolution limit for such targets. While the sampling density of the retina limits the resolution of gratings, target detection is theoretically limited by contrast sensitivity. This allows many animals to detect single targets smaller than their visual resolution limit, but this is not the case for budgerigars. The budgerigars were able to detect a high contrast circular target with a luminance profile of a single period of a sine wave subtending 0.065 degrees of their visual field, corresponding to a spatial acuity of 7.7 cycles degree⁻¹, a measurement in line with the previously measured grating acuity of budgerigars (7.7 and 10 cycles degree⁻¹). This result is different from findings on the spatial acuity of humans, who can detect single targets much smaller than predicted by their acuity for gratings. The low contrast sensitivity of budgerigar vision might be one of the reasons why the single target acuity is not higher than grating acuity. Adding a bright surround to the target did not influence detection threshold significantly. However, the threshold was slightly higher for a target with a square-wave luminance profile than for a target with a sinusoidal luminance profile.

1. Introduction

Spatial resolution of vision is traditionally measured as the capacity of the eye to resolve a grating of dark and light bars (de Valois & de Valois, 1990). In order to resolve a grating, the images of adjacent dark and light bars must fall on the receptive fields of separate sampling units (photoreceptors or, in many cases, retinal ganglion cells) in the retina (Land & Nilsson, 2012). If the grating is any finer, each unit will sample both light and dark areas, which will reduce the perceived contrast, and ultimately make the image appear uniformly grey. Grating acuity thus is a measure of how fine detail in a visual scene the eye is able to resolve.

It is often assumed that it is possible to deduce, from grating acuity, the size of the smallest single target that an animal can detect. However, different physical mechanisms determine how fine gratings and how small objects an eye can resolve. While the retinal sampling density sets the limit to grating acuity, single target acuity is limited by contrast sensitivity. Thus, a single target smaller than the receptive field of a retinal sampling unit can still be detected if it has high contrast to the background (O'Carroll & Wiederman, 2014).

While gratings have been used extensively when investigating the visual acuity of vertebrates, determining single target acuity has been a

common approach in work with insects (for examples see Giurfa & Vorobyev, 1998, Somanathan, Borges, Warrant, & Kelber, 2017, Spaethe & Chittka, 2003, Vallet & Coles, 1993). Behavioural tests have revealed target detection below the resolution limit for gratings in several insect species. Drone honey bees (*Apis mellifera*) are able to detect a dummy queen bee only subtending an angle of 0.41 degree in their visual field (Vallet & Coles, 1993) and a recent study describes male carpenter bees (*Xylocopa tenuiscapa*) reacting to a flying female covering less than 0.1 degree of their visual field (Somanathan et al., 2017), although the interommatidial angles in both species, 0.5 degree and 0.7 degree respectively, predict much lower grating acuity.

Humans are able to detect a black square against the sky subtending 1/5 of the width of a single line in the finest grating they can resolve, while a black single line is visible even at about 1/100 of the width of a single black stripe in such a grating (Hecht, Ross, & Mueller, 1947). Few such studies have been performed on other vertebrates, but Ehrenhardt (1937) reported that sand lizards (*Lacerta agilis*) were able to detect a single black line against a bright background when it had 1/10 of the width of one stripe in the finest grating that the animals could resolve.

The choice of the most suitable stimulus for investigation of visual acuity thus depends on the question to be asked. Grating acuity is a good measure of how small details in a cluttered environment an

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animal can detect while target acuity determines in which distance a small single object can be detected on an even background such as the sky (Land, 1997). The visual capabilities of birds have been widely studied but to our knowledge no one has investigated how small single targets or objects they can detect. Budgerigars (*Melopsittacus undulatus*) have been used extensively as model for the avian visual system, and several aspects of their spatial vision have been investigated. The spatial resolution threshold of a budgerigar for a stationary grating is 7.7 to 10 cycles degree⁻¹ (Haller, Lind, Steinlechner, & Kelber, 2014, Lind & Kelber, 2011, Lind, Sunesson, Mitkus, & Kelber, 2012). The maximum contrast sensitivity for stationary achromatic gratings is 10.2 to 14 (corresponding to 9.8–7.1 Michelson contrast, for spatial frequencies around 1–2 cycles degree⁻¹; Haller et al., 2014, Lind & Kelber, 2011, Lind et al., 2012). Budgerigars need a similar achromatic contrast (9 Michelson contrast) to discriminate two spatially separated homogenous fields (Lind, Karlsson, & Kelber, 2013), while they can detect drifting gratings at only 5.8 Michelson contrast (Haller et al., 2014). Birds in general have low contrast sensitivity, ranging between 7 and 30, compared to humans, which have a contrast sensitivity of about 175 (Lind et al., 2012).

In this study we first determined the detection threshold for dark targets with different achromatic contrasts to the background, in short, the target acuity of budgerigars. To make the results comparable to previous, work, we used targets with sinusoidal luminance profiles. Second, we ask whether the luminance profile of the target affects its detectability. Does a target with a square-wave luminance profile have the same detection threshold as the target with sinusoidal luminance profile?

Previous studies on budgerigar grating acuity revealed no difference between visual acuity of sinusoidal patterns and square-wave patterns (Lind & Kelber, 2011, Lind et al., 2012), but in humans a higher detectability for square-wave patterns has been observed (Campbell & Robson, 1968). If the contrast sensitivity was high enough, these two targets could theoretically be detected beneath the resolution limit of the retina. This should not be possible for a target with the same overall luminance as the background. Therefore, we also tested the ability of budgerigars to detect a target with a sinusoidal luminance profile similar to the first stimulus but with a bright surround and thus, the same overall luminance as the background.

2. Methods

2.1. Animals

We used four budgerigars (one female and three males) in our behavioural experiments. The birds were fed a parakeet seed mix as well as fruits and vegetables. One day prior to the weekly training/testing period the seed mix was removed from the cage but the birds always had access to fruits or vegetables. During the training and test sessions, usually on four days/week, the seed mix was used as reward. All experiments were performed following Swedish legislation, under the permit M111-14 from the local authority for animal ethics.

2.2. Experimental apparatus

The experiments took place in a flight cage (length: 1580 mm, width: 860 and height: 670 mm) constructed of metal net except for the floor and one of the short end walls that were made of matte-grey plastic board (Fig. 1). The plastic wall had two circular openings of 90 mm diameter placed 330 mm apart, which allowed the presentation of stimuli on an LCD-screen behind the wall. Beneath each window a feeder with removable lid and a perch was positioned. A vertical grey plastic board divided the cage into two equally sized compartments starting between the stimulus windows and 1160 mm into the cage.

A starting perch was positioned opposite the stimulus windows. Centred on this perch, a bird had a good view of both stimulus

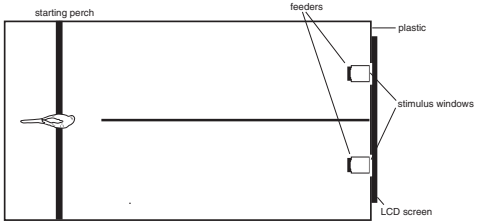


Fig. 1. Sketch of the experimental cage viewed from above with the bird sitting on the starting perch.

windows. The experimenter was always situated behind the grey wall, out of sight for the bird, and monitored the behaviour of the bird on an external screen showing the live feed of a camera attached to the experimental cage.

2.3. Stimuli

Each stimulus pair consisted of one homogeneously grey field (137 candela/m²) and one identical grey field with a circular target in its centre. The stimuli were created in Matlab (v. 8.5.0.197613, The MathWorks Inc.) as PNG-images and presented in Microsoft PowerPoint (v. 14.7.2.170228).

2.3.1. Experiment 1: Sinusoidal targets of different contrasts

In Experiment 1, we determined the detection threshold for a single dark circular target with a gradual transition between the darker centre and the brighter background. The luminance profile of this target resembled a single cycle of a sinusoidal wave, which allowed for direct comparison to previous behavioural tests of grating acuity in the same species (Haller et al., 2014, Lind & Kelber, 2011, Lind et al., 2012). This target was presented with 5 different contrasts to the background (C1, C2, C3, C4, and C5). All stimuli had the same background luminance while the target luminance differed. We give contrast levels as Weber contrast, which is commonly used for contrasts between a single target and its background (O'Carroll & Wiederman, 2014, Rigosi, Wiederman, & O'Carroll, 2017). The equivalent Michelson contrasts are given in Table 1 for easier comparison with previous studies. Target size is given as full width at half amplitude (Fig. 2a), which allows for direct comparison to sine wave gratings.

2.3.2. Experiments 2 and 3: Targets with different luminance profiles

In experiments 2 and 3 we investigated whether the luminance profile of the target affected the detection threshold. The aim of experiment 2 was to compare the detection thresholds for targets with square wave luminance profile to those with a sine wave profile. The stimulus used in experiment 2 had a dark circular target with a sharp edge and a square luminance profile (Fig. 2b). We used the same background luminance as in experiment 1, and the highest contrast (C1) to the background.

A square wave grating and a sine wave grating with the same fundamental frequency and contrast have the same overall luminance.

Table 1
Contrast levels used in the experiments expressed as Weber contrast and Michelson contrast.

	C1	C2	C3	C4	C5
Weber contrast $C_W = \frac{I_0 - I_b}{I_b}$	> 99	89	68	53	41
Michelson contrast $C_M = \frac{I_{max} - I_{min}}{I_{max} + I_{min}}$	> 99	80	50	35	25

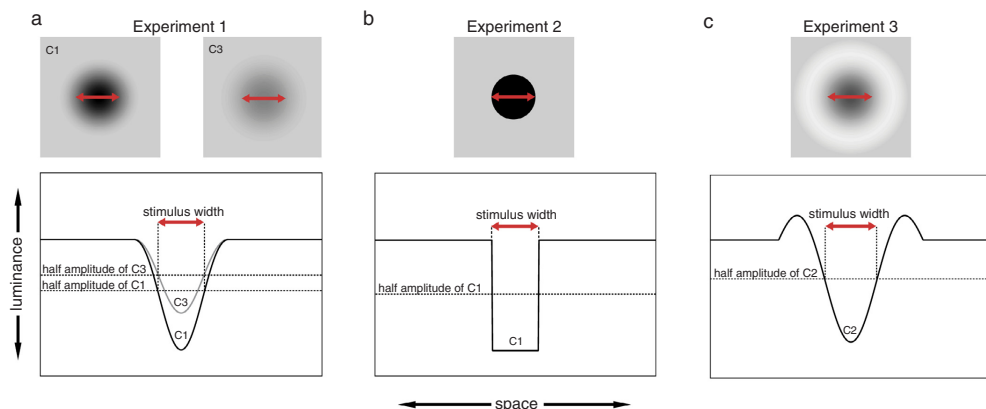


Fig. 2. Luminance profiles of the three types of stimuli. a) The sinusoidal target (Experiment 1), b) the square wave target (Experiment 2) and c) the target with a bright surround (Experiment 3).

However, a circular target with a sine wave luminance profile (the C1 target in experiment 1) decreases the luminance of the stimulus 19% more than a square wave target (the target in experiment 2; see [Supplementary Material A](#) for the calculations).

As we define the size of the sinusoidal target as full width at half maximum, the area of the target with contrast below half amplitude (or 50 Weber contrast to background) will extend outside the defined “target area”. However, budgerigars are able to detect a Michelson contrast of about 10 ([Lind et al., 2013](#), [Lind & Kelber, 2011](#), [Lind et al., 2012](#)). If we measure the size of the targets as the area with at least 10 Michelson contrast to the background, the diameter of the C1 sine wave target will increase with about 59% compared to full width at half maximum, while the diameter of the square wave target will not change.

In experiment 3, we used a target with the same overall luminance as the background. This way, the bird should be unable to discriminate the stimuli with and without targets, if the target was smaller than the acceptance angle of one sampling unit of the eye. This target was similar to the target used in experiment 1, but had a bright surround ([Fig. 2c](#)). The size of the target was measured as the full width at half amplitude between the darkest and brightest part. It was not possible to display this target with the highest contrast (C1) on our monitor without creating sharp transitions between dark and bright areas. In order to avoid such unwanted artefacts we therefore chose to present this target with contrast C2 (89 Weber contrast between target centre and background and 92 Weber contrast between target centre and bright surround)

2.4. Training and testing

We trained each bird individually to associate the stimulus without target, the positive stimulus, with a food reward, and the stimulus with target, the negative stimulus, with absence of a reward. This way, the bird will experience two positive stimuli, if it is unable to resolve the target, and we avoid a scenario in which the bird stops making choices because of the lack of a rewarding stimulus, which would be impossible to separate from the lack of motivation of a different cause.

During the training and test sessions a bird was sitting in the middle of the starting perch facing the two stimulus windows (see [Fig. 1](#)). Each trial started with the screen displaying one stimulus pair – the grey background with the circular dark target as negative stimulus and the background alone as positive stimulus. If the bird flew to the perch in

front of the positive stimulus the feeder was opened and the bird was allowed to eat for around four seconds. If the bird was flying to the negative stimulus, the screen turned black and the feeder remained closed. The bird had unlimited viewing time of the stimuli before making its choice. A new trial began once the bird had returned to and centred on the starting perch. The positive and negative stimuli were presented in the right or left window in a pseudo-random order ([Gellermann, 1933](#)).

In the initial training sessions with a bird, we used a large target (subtending 1 degree of the visual field at the decision point). A bird was considered to have learned to associate the positive stimulus with the food reward when it performed $\geq 80\%$ correct choices in two consecutive training sessions of 20 choices each. Once a bird had fulfilled this criterion we decreased the size of the stimulus stepwise using an adaptive 2-down/1-up staircase procedure ([Levitt, 1971](#)) in each session of 20 trials. This frequently used method implies that, following two correct choices by the subject the experimenter reduces the size of the target presented in the next trial, while after a single incorrect choice stimulus size is increased. Such a process of reversals results in fluctuation around a target size, for which the probability of making two correct choices in a row equals the probability of making one incorrect choice. This corresponds to the point on a psychometric curve, in which the probability of a correct choice is 70.7% ([Levitt, 1971](#), see [Fig. 3](#) for an example). We chose this probability level to allow for easier comparison of our results with previous data sets on spatial resolution and contrast sensitivity of budgerigars, in which stimuli were presented a set number of times in random order. In those experiments, following binomial statistics, a level of 72.5% correct was significantly different from chance ([Lind & Kelber, 2011](#)).

In order to keep the birds motivated, we started each session using a fairly large target and decreased target size in small steps. Over several staircase training sessions, all birds improved in motivation and performance, and we increased the number of trials per session to between 40 and 60. A bird was considered to have reached its detection threshold, when the performance did not improve over three consecutive sessions.

2.5. Analysis

The detection threshold was calculated as the mean stimulus size at the reversals (two correct choices following an incorrect choice or an incorrect choice following two correct choices) during the last 20 trials

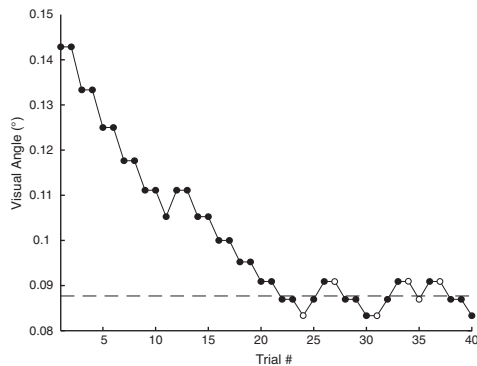


Fig. 3. An example test session with a bird (Pippi) in experiment 1. The detection threshold (indicated by the dashed line) was estimated by calculating the mean of the stimulus sizes in the reversals (indicated by open circles) of the 20 last trials.

in a test session (dashed line in Fig. 3). To avoid estimation bias we used an even number of reversal points per session excluding the first reversal point if needed. The results for each contrast level and experiment include three test sessions per bird.

To examine the effect of contrast level on detection threshold we fitted a linear mixed model (LMM) with random intercepts to our data from experiment 1 using the `{lmerTest}` package (Kuznetsova, Brockhoff, & Christensen, 2017) in RStudio (v. 1.1.463; RStudio Team, 2016). The model had contrast level as fixed effect and bird identity as random effect. This model was compared to a reduced model, excluding the fixed effect of contrast, with a log likelihood ratio test (Quinn & Keough, 2002). We performed a Tukey's HSD test for post-hoc analysis using the `{multcomp}` package (Hothorn, Bretz, & Westfall, 2008) in RStudio.

To test whether detection thresholds differed between targets with different luminance profiles, we compared the results from experiment 2 (targets with C1 contrast and a sharp edge) and the results from experiment 3 (sinusoidal target with contrast C2 and bright surround) to those obtained with the same contrast level in experiment 1. We used the `{lmerTest}` package in R to fit LMMs with random intercepts and luminance profile as fixed effects and bird identity as random effect. These models were analysed by comparing them to reduced models, excluding the fixed effects of luminance profiles, with a log likelihood ratio test.

3. Results

3.1. Experiment 1: Detection threshold for sinusoidal targets

All four birds learned to discriminate the stimulus without the target from the stimulus with the target. Over four to eight staircase training sessions, their performance improved before it reached a constant level (see Supplementary Material Fig. B1). In the test sessions, the birds were able to detect the target with the highest contrast to the background (C1) when it had a diameter of 0.065 ± 0.008 degrees. This corresponds to resolution of a sinusoidal grating with a spatial frequency of $7.7 \text{ cycles degree}^{-1}$ (95% confidence interval: $6.8 - 9.0 \text{ cycles degree}^{-1}$). The smallest target detected by any bird was 0.056 degrees in diameter (Pippi, C1; Fig. 4a). The model including contrast level as fixed effect provided a significantly better fit to the data than a reduced model ($\chi^2 = 110.8$, $df = 4$, $p < 0.001$) and had a lower Akaike Information Criterion (AIC) value (-321.8 versus -219.9),

indicating that detection threshold varied with contrast level. The detection thresholds differed significantly between all contrast levels except between C2 and C3 and between C4 and C5 (for statistics see Supplementary Material Table C1).

3.2. Experiments 2 and 3: Targets with different luminance profiles

In Experiment 2, using a square wave target, the birds reached a detection threshold of 0.098 ± 0.008 degrees (Fig. 5), corresponding to the resolution of a sinusoidal grating of $5.1 \text{ cycles degree}^{-1}$. When the data from Experiment 2 was analysed together with C1 data from Experiment 1, a model including luminance profile as fixed factor had a significantly better fit to our data than a reduced model ($\chi^2 = 37.2$, $df = 1$, $p < 0.001$) and had a lower AIC value (144.8 versus -109.6) This indicates that luminance profile had an effect on the detection thresholds in this experiment.

To test whether this difference resulted from the difference in overall luminance between these two targets, we compared the two targets on a unit-less scale that describes how much each target reduces the overall luminance of the stimulus ("change in luminance"). On this scale, the target in experiment 1 changes the luminance by 19% more than the target in experiment 2, given the same diameter (see Supplementary Material A). Thus, at detection threshold - since the relative change in luminance is a function of the area of the target - the larger square wave target changed overall luminance by almost 90% more than the sinusoidal target. We repeated the statistical analysis using "change in luminance" as the dependent variable and still found the luminance profile to have an effect on detection threshold ($\chi^2 = 24.3$, $df = 1$, $p < 0.001$, AIC: -229.9 versus -207.6).

We also determined which portion of the sinusoidal target had at least 10 Michelson contrast to the background, taking into account that budgerigars are able to detect brightness contrasts of ≈ 10 Michelson contrast (Lind et al., 2013). Measuring the size of the targets in this way, the mean detection threshold for the sine-wave target was 0.094 degrees. We repeated the statistical analysis, and for this comparison, found no significant effect of luminance profile on detection threshold ($\chi^2 = 2.08$, $df = 1$, $p = 0.15$, AIC: -145.2 versus -145.1).

Finally, in experiment 3, we tested the birds using a target with a bright surround. The birds were able to detect a target with a diameter of 0.085 ± 0.008 degrees (Fig. 5). The data from Experiment 3 was analysed together with the C2 data from Experiment 1 and a model including luminance profile as fixed effect did not show a better fit than a reduced model ($\chi^2 = 0.85$, $df = 1$, $p = 0.36$, AIC: -144.1 versus 145.2), indicating no effect of luminance profile on detection threshold in this experiment.

4. Discussion

We tested how small circular targets budgerigars are able to detect, depending on their luminance profiles and contrasts to the background. The targets used in Experiment 1 had a luminance profile similar to one cycle of a sine function allowing us to directly compare the detection thresholds with previously measured spatial resolution and contrast sensitivity of budgerigars for sinusoidal gratings (Haller et al., 2014, Lind & Kelber, 2011). With the highest contrast to the background (C1), the birds could detect a target subtending 0.065 ± 0.008 degrees of the visual field, equivalent to a resolution threshold of $7.7 \text{ cycles degree}^{-1}$. This acuity measurement is similar to the grating acuity obtained by Haller et al. (2014; $7.7 \text{ cycles degree}^{-1}$; Fig. 4b) but the limit obtained by Lind and Kelber (2011) and Lind et al. (2012; $10 \text{ cycles degree}^{-1}$) lies outside the 95% confidence interval of our measurement (see Results). Mitkus, Chaib, Lind, and Kelber (2014) estimated spatial acuity for budgerigars based on maximal ganglion cell density in the retina to be $6.9 \text{ cycles degree}^{-1}$, which is inside the 95% confidence interval of our threshold.

In contrast to humans and other animals, whose detection threshold

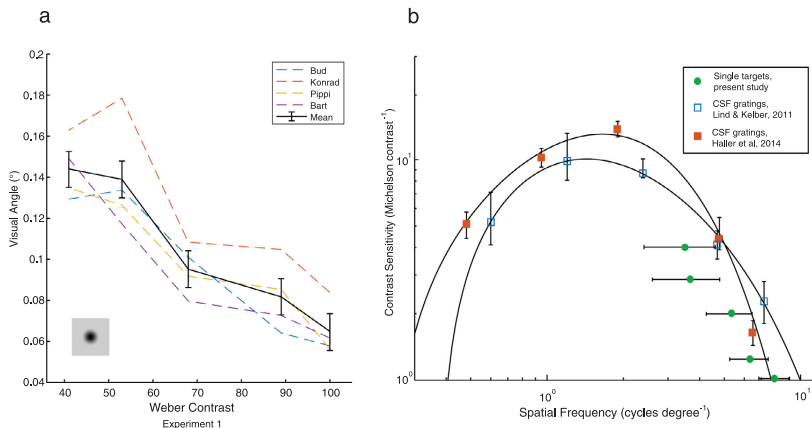


Fig. 4. Detection thresholds of four budgerigars for single targets with different contrasts to the background (Experiment 1). a) Detection thresholds (in degrees) as function of Weber contrast. Solid line: Mean values of all birds (\pm standard errors), dashed lines represent detection thresholds of each bird. b) Comparison between single target and grating acuity and contrast sensitivity (using Michelson contrast). Green circles: detection thresholds from Experiment 1, calculated as cycles degree⁻¹ (\pm 95% confidence interval). Solid lines: Contrast sensitivity functions of budgerigars with achromatic gratings from Lind and Kelber (2011), with open blue squares (\pm 95% confidence interval), and Haller et al. (2014), with filled red squares (\pm standard error).

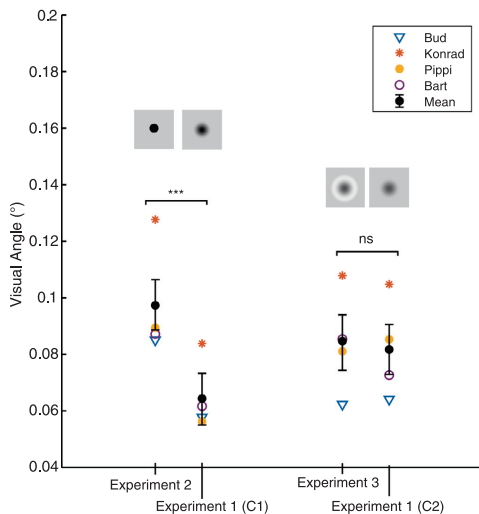


Fig. 5. Detection thresholds of budgerigars for single targets. Left: Targets with square luminance profiles (Experiment 2) compared to targets with sinusoidal luminance profiles (C1 Experiment 1), right: targets with a bright surround (Experiment 3) compared to sinusoidal luminance profiles (C2 Experiment 1). Black filled circles: mean values of all birds. Error bars give standard error. Other symbols: individual birds (see inset).

for single targets has been investigated (Hecht et al., 1947, O'Carroll & Wiederman, 2014), the budgerigars in our study were not better at detecting single targets compared to gratings. A possible reason for this could be their low contrast sensitivity of ≈ 10 , given as inverted Michelson contrast (Lind et al., 2013, Lind & Kelber, 2011). Birds generally have low contrast sensitivity (between 7 and 30; see Lind et al.,

2012, Potier, Mitkus, & Kelber, 2018), compared to humans or cats, which have a contrast sensitivity of 175 and 116, respectively (Lind et al., 2012). Detection of single targets smaller than the resolution threshold is limited by contrast sensitivity since such targets are seen as a small change in luminance in one sampling unit in the retina, and thus appear as having lower contrast to the background (Land & Nilsson, 2012, O'Carroll & Wiederman, 2014). Overall luminance of the stimuli used in Experiment 1 decreases proportionally to the square of target diameter (Eq. (A.3) in supplementary methods), thus the contrast between target and background decreases rapidly with target size.

Fig. 4b shows the detection thresholds for the stimuli in Experiment 1 as function of contrast sensitivity, using the Michelson contrast of the stimuli (see Table 1). Our results are in line with contrast sensitivity functions for budgerigars measured using gratings (Fig. 4b; Haller et al., 2014, Lind & Kelber, 2011, Lind et al., 2012). However, we never tested the budgerigars with low spatial frequencies so we do not know whether the contrast sensitivity function for single targets will have the same band-pass shape as the function for gratings (Lind et al., 2012).

Budgerigars could detect smaller targets with the highest contrast (C1) target in Experiment 1 than in Experiment 2 when we measured the size of the sinusoidal target as full width at half maximum. This difference was even larger when the target was measured as the overall decrease in stimulus luminance, compared to the stimulus without target (see Eq. (A.5) in Supplementary Methods). Interestingly, the contrast sensitivity of humans is $4/\pi$ (1.27) times higher for a square wave grating than for a sinusoidal grating with the same period (Campbell & Robson, 1968). However, this has not been demonstrated in budgerigars (Lind et al., 2012), and it has not been measured for single targets in humans.

For a sinusoidal target diameter adjusted for budgerigar contrast sensitivity (area with contrast ≥ 10 Michelson contrast included), acuity was similar to that of a square-wave target (see Results). This shows that comparisons between single targets require careful considerations of the observer's visual system.

Humans are better at detecting a single straight line than a single square of the same width (Hecht et al., 1947). A line covers a greater portion of the visual field and the receptive fields of more retinal sampling units than a square. It is thus possible that budgerigars can

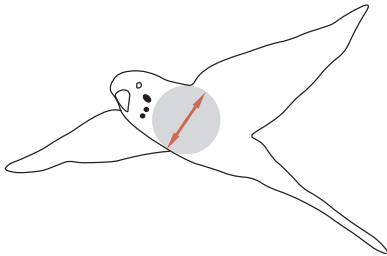


Fig. 6. Illustration of a flying budgerigar. The red arrow marks the core transect diameter.

also detect single lines that are finer than predicted from their target acuity.

The detection threshold for the target with bright surround (Experiment 3) was not different from the detection threshold for the sine-wave target with the same contrast but without surround (Experiment 1, C2; Fig. 5). The luminance profile of the target used in Experiment 3 was designed to have the same overall luminance as the background, which should theoretically make detection impossible when it is smaller than the receptive field of a sampling unit in the retina. Interestingly, “vanishing optotypes” have been used to measure spatial acuity of human subjects using letters. Like single targets or lines, simple black letters can be detected even when they are smaller than the resolution threshold would allow, simply because of the change in luminance. A “vanishing optotype”, with a bright surround similar to the bright surround that we used in Experiment 3, the letter “vanishes” at the level of resolution, as its overall luminance is equal to that of the background (Demirel, Anderson, Dakin, & Thibos, 2012, Howland, Ginsburg, & Campbell, 1978). In a similar way, because the detection limit for the sinusoidal target with and without a bright surround did not differ, we can conclude that the thresholds indeed are limited by the spatial acuity of the birds, and not by contrast sensitivity.

A budgerigar is able to detect a high contrast sharp-edged target subtending ≈ 0.1 degrees of its visual field. If we assume that a flying bird occupies at least an area the size of the core of its body (Fig. 6), we can estimate detection distance. A budgerigar would be able to spot a conspecific against the sky from about 25 m distance and a typical predator on small birds, the Brown falcon (*Falco berigora*), from the safe distance of 85 m.

A bird in flight, however, has a more complex shape than a circle, making this a rather conservative approximation. As a flying bird normally moves across the visual field of the observer (although see Kane & Zamani, 2014) it is possible that image motion also affects its visibility. Budgerigars have higher contrast sensitivity for moving than for stationary gratings (Haller et al., 2014). Therefore, we plan to investigate the influence of motion on the detectability of single targets in our next study.

Acknowledgements

We are grateful to James Foster and Lars Råberg for expert advises on statistics, to David Wilby for inspiring discussions on stimulus design and to Simon Potier for helpful comments on the manuscript. This study was financially supported by the Swedish Research Council (#2016-03298.3) and the K & A Wallenberg Foundation (Ultimate Vision), Stockholm.

Declarations of interest

None.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.visres.2019.04.005>.

References

- Campbell, F. W., & Robson, J. G. (1968). Application of Fourier analysis to the visibility of gratings. *The Journal of Physiology*, 197(3), 551–566. <https://doi.org/10.1113/jphysiol.1968.sp008574>.
- de Valois, R. L., & de Valois, K. K. (1990). *Spatial vision*. New York: Oxford University Press.
- Demirel, S., Anderson, R. S., Dakin, S. C., & Thibos, L. N. (2012). Detection and resolution of vanishing optotype letters in central and peripheral vision. *Vision Research*, 59, 9–16. <https://doi.org/10.1016/j.visres.2012.02.010>.
- Ehrenhardt, H. (1937). Formensehen und Sehschärfestimmungen bei Eidechsen. *Zeitschrift für vergleichende Physiologie*, 24(2), 248–304. <https://doi.org/10.1007/BF00338282>.
- Gellermann, L. W. (1933). Chance orders of alternating stimuli in visual discrimination experiments. *Pedagogical Seminary and Journal of Genetic Psychology*, 42(1), 206–208. <https://doi.org/10.1080/08856559.1933.10534237>.
- Giurfa, M., & Vorobyev, M. (1998). The angular range of achromatic target detection by honey bees. *Journal of Comparative Physiology A*, 183(1), 101–110. <https://doi.org/10.1007/s003590050238>.
- Haller, N. K., Lind, O., Steinlechner, S., & Kelber, A. (2014). Stimulus motion improves spatial contrast sensitivity in budgerigars (*Melospittacus undulatus*). *Vision Research*, 102, 19–25. <https://doi.org/10.1016/j.visres.2014.07.007>.
- Hecht, S., Ross, S., & Mueller, C. G. (1947). The visibility of lines and squares at high brightness. *Journal of the Optical Society of America*, 37, 500–507. <https://doi.org/10.1364/JOSA.37.000500>.
- Hothorn, T., Bretz, F., & Westfall, P. (2008). Simultaneous inference in general parametric models. *Biometrical Journal*, 50(3), 346–363. <https://doi.org/10.1002/bimj.200810425>.
- Howland, B., Ginsburg, A., & Campbell, F. (1978). High-pass spatial-frequency letters as clinical optotypes. *Vision Research*, 18(8), 1063–1066. [https://doi.org/10.1016/0042-6989\(78\)90036-6](https://doi.org/10.1016/0042-6989(78)90036-6).
- Kane, S. A., & Zamani, M. (2014). Falcons pursue prey using visual motion cues: New perspectives from animal-borne cameras. *Journal of Experimental Biology*, 217, 225–234. <https://doi.org/10.1242/jeb.092403>.
- Kuznetsova, A., Brockhoff, P. B., & Christensen, R. H. B. (2017). lmerTestPackage: Tests in linear mixed effects models. *Journal of Statistical Software*, 82(13), 1–26. <https://doi.org/10.18637/jss.v082.i13>.
- Land, M. F. (1997). Visual acuity in insects. *Annual Review of Entomology*, 42(1), 147–177. <https://doi.org/10.1146/annurev.ento.42.1.147>.
- Land, M. F., & Nilsson, D.-E. (2012). *Animal Eyes* (second ed.). Oxford: Oxford University Press.
- Levitt, H. (1971). Transformed up-down methods in psychoacoustics. *Journal of the Acoustical Society of America*, 49(2), 467–477. <https://doi.org/10.1121/1.1912375>.
- Lind, O., Karlsson, S., & Kelber, A. (2013). Brightness discrimination in budgerigars (*Melospittacus undulatus*). *PLoS One*, 8(1), e54650. <https://doi.org/10.1371/journal.pone.0054650>.
- Lind, O., & Kelber, A. (2011). The spatial tuning of achromatic and chromatic vision in budgerigars. *Journal of Vision*, 11(7), 2. <https://doi.org/10.1167/11.7.2>.
- Lind, O., Sunesson, T., Mitkus, M., & Kelber, A. (2012). Luminance-dependence of spatial vision in budgerigars (*Melospittacus undulatus*) and Bourke's parrots (*Neopsephos bourkii*). *Journal of Comparative Physiology A*, 198(1), 69–77. <https://doi.org/10.1007/s00359-011-0689-7>.
- Mitkus, M., Chaib, S., Lind, O., & Kelber, A. (2014). Retinal ganglion cell topography and spatial resolution of two parrot species: budgerigar (*Melospittacus undulatus*) and Bourke's parrot (*Neopsephos bourkii*). *Journal of Comparative Physiology A*, 200(5), 371–384. <https://doi.org/10.1007/s00359-014-0894-2>.
- O'Carroll, D. C., & Wiederman, S. D. (2014). Contrast sensitivity and the detection of moving patterns and features. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 369(1636), 20130043. <https://doi.org/10.1098/rstb.2013.0043>.
- Potier, S., Mitkus, M., & Kelber, A. (2018). High resolution of colour vision, but low contrast sensitivity in a diurnal raptor. *Proceedings of the Royal Society B: Biological Sciences*, 285(1885), 20181036. <https://doi.org/10.1098/rspb.2018.1036>.
- Quinn, G. P., & Keough, M. J. (2002). *Experimental design and data analysis for biologists*. Cambridge: Cambridge University Press.
- Rigosi, E., Wiederman, S. D., & O'Carroll, D. C. (2017). Visual acuity of the honey bee retina and the limits for feature detection. *Scientific Reports*, 7, 45972. <https://doi.org/10.1038/srep45972>.
- RStudio Team (2016). *RStudio: Integrated development for R*. Boston, MA: RStudio Inc. <http://www.rstudio.com>.
- Somanathan, H., Borges, R. M., Warrant, E. J., & Kelber, A. (2017). Visual adaptations for mate detection in the male carpenter bee *Xylocopa tenuiscapa*. *PLoS One*, 12(1), e0168452. <https://doi.org/10.1371/journal.pone.0168452>.
- Spaethe, J., & Chittka, L. (2003). Interindividual variation of eye optics and single object resolution in bumblebees. *Journal of Experimental Biology*, 206(19), 3447–3453. <https://doi.org/10.1242/jeb.00570>.
- Vallet, A. M., & Coles, J. A. (1993). The perception of small objects by the drone honeybee. *Journal of Comparative Physiology A: Sensory, Neural and Behavioral Physiology*, 172(2), 183–188. <https://doi.org/10.1007/BF00189395>.

Supplementary material

A. Supplementary methods – Calculation of overall change in luminance by targets in experiments 1 and 2

Since the detection of a single target smaller than the grating resolution threshold theoretically is limited by contrast it is of importance to know how much each target will decrease the overall luminance in the different stimulus types.

Given the same length and period, a single stripe from a sinusoidal grating and a stripe from a square wave grating will cause the same decrease in luminance. We can approximate this by integrating the area A_{cos} contained by one period of a cosine wave,

$$A_{cos} = - \int_0^P \frac{\cos\left(\frac{2\pi x}{P}\right) + 1}{2} dx = - \frac{P}{2} \quad \text{Eq. (A.1)}$$

where P is the period, and compare this to the area contained by one period of a square wave A_{sq}

$$A_{sq} = - \int_0^P y_{sq} dx = - \frac{P}{2} \quad y_{sq} \begin{cases} 1 & \text{when } 0 \leq x \leq \frac{P}{2} \\ 0 & \text{when } x > \frac{P}{2} \end{cases} \quad \text{Eq. (A.2)}$$

A single circular target, on the other hand, is two-dimensional and the decrease in luminance for a radially sinusoidal single wave can be approximated by the solid of revolution for half a period of a cosine function

$$V_{cos} = -\pi \int_0^1 \left(\frac{P}{2\pi}\right)^2 (\cos^{-1}(2y - 1))^2 dy \approx -\frac{P^2 2.93}{4\pi} \quad \text{Eq. (A.3)}$$

The equivalent approximation for a square wave V_{sq} is

$$V_{sq} = -\pi \int_0^1 \left(\frac{P_{sq}}{4}\right)^2 dy = -\frac{P^2 \pi}{16} \quad \text{Eq. (A.4)}$$

The quotient of V_{cos} and V_{sq} is

$$\frac{V_{cos}}{V_{sq}} \approx 1.19 \quad \text{Eq. (A.5)}$$

The C1 target in experiment 1 thus decreases the luminance 19% more than the target in experiment 2 given the same diameter.

B. Supplementary figure

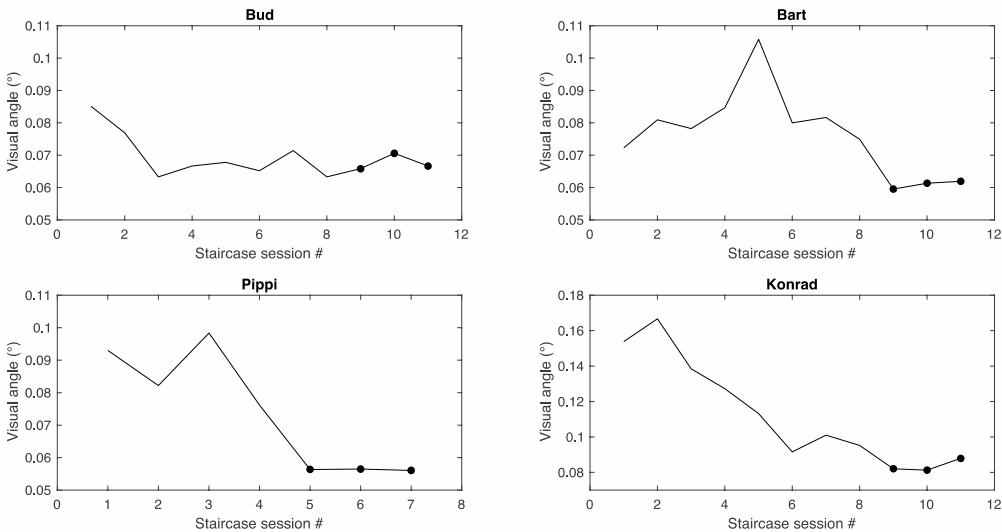


Figure B1. Learning curves of the birds in experiment 1 with stimulus contrast C1. Each data point shows the threshold (calculated as described in the methods, from reversal points) that the specific bird reached in staircase training sessions and the three test sessions (indicated by filled circles). Note that different birds needed different numbers (between four and 8) of training sessions before reaching a constant threshold, and that two birds (named Bart and Pippi) had motivation problems during the course of the training process. The fact that all birds showed a constant performance in the three test sessions was taken as proof that these represent their true thresholds.

C. Supplementary statistic analysis

Table C1. Multiple comparisons of means in Experiment 1: Tukey Contrasts.

	Estimate	Std. Error	z value	Pr(> z)
C2 - C1 == 0	0.017105	0.005718	2.992	0.0232 *
C3 - C1 == 0	0.030774	0.005718	5.382	<0.001 ***
C4 - C1 == 0	0.076184	0.005718	13.324	<0.001 ***
C5 - C1 == 0	0.082257	0.005718	14.386	<0.001 ***
C3 - C2 == 0	0.013668	0.005718	2.390	0.1178
C4 - C2 == 0	0.059078	0.005718	10.332	<0.001 ***
C5 - C2 == 0	0.065152	0.005718	11.394	<0.001 ***
C4 - C3 == 0	0.045410	0.005718	7.942	<0.001 ***
C5 - C3 == 0	0.051483	0.005718	9.004	<0.001 ***
C5 - C4 == 0	0.006074	0.005718	1.062	0.8259

D. Supplementary data tables

Table D1. Mean detection threshold of Experiment 1 from three test sessions averaged for all 4 birds. The overall mean (Mean± standard error) as estimated by the model.

Experiment 1	C1	C2	C3	C4	C5
Bud	0.058°	0.064°	0.102°	0.136°	0.132°
Konrad	0.084°	0.106°	0.109°	0.182°	0.164°
Pippi	0.056°	0.086°	0.093°	0.129°	0.141°
Bart	0.062°	0.073°	0.080°	0.119°	0.152°
Mean ± standard error	0.065 ± 0.008°	0.082 ± 0.008°	0.096 ± 0.008°	0.141 ± 0.008°	0.147 ± 0.008°

Table D2. Mean detection threshold of Experiments 2 and 3, average values from three test sessions with each of the four birds. The overall mean (Mean± standard error) as estimated by the model.

	Experiment 2	Experiment 3
Bud	0.085°	0.062°
Konrad	0.128°	0.108°
Pippi	0.089°	0.081°
Bart	0.087°	0.085°
Mean ± standard error	0.097 ± 0.008°	0.085 ± 0.009°

Paper II



RESEARCH ARTICLE

Visual acuity of budgerigars for moving targets

Sandra Chaib[‡], Juliane Gaviraghi Mussoi^{*}, Olle Lind and Almut Kelber

ABSTRACT

For a bird, it is often vital to visually detect food items, predators, or individuals from the same flock, i.e. moving stimuli of various shapes. Yet, behavioural tests of visual spatial acuity traditionally use stationary gratings as stimuli. We have behaviourally tested the ability of budgerigars (*Melopsittacus undulatus*) to detect a black circular target, moving semi-randomly at $1.69 \text{ degrees s}^{-1}$ against a brighter background. We found a detection threshold of 0.107 ± 0.007 degrees of the visual field for a target size corresponding to a resolution of a grating with a spatial frequency of 4.68 cycles degree⁻¹. This detection threshold is lower than the resolution limit for gratings but similar to the threshold for stationary single objects of the same shape. We conclude that the target acuity of budgerigars for moving single targets, just as for stationary single targets, is lower than their acuity for gratings.

KEY WORDS: Bird vision, Visual resolution, Dynamic acuity, Target detection, Object detection

INTRODUCTION

Vision is undoubtedly one of the primary senses of birds (Martin, 2017). The excellent colour vision (Kelber, 2019), as well as high spatial (Fischer, 1969; Reymond, 1985) and temporal resolution (Bostrom et al., 2016; Potier et al., 2020) of some species, are among the best in the animal kingdom.

In psychophysics, the common way to measure visual spatial acuity is determining the sinusoidal or square-wave grating of the highest spatial frequency that the eye can resolve. Assuming that the retinal mosaic limits spatial resolution of vision, the acuity limit is reached when adjacent black and white bars in a square-wave grating fall on the receptive fields of neighbouring sampling units (e.g. photoreceptors, or ganglion cells) in the retina (Land and Nilsson, 2012). This is a useful method to get a standardized comparative measurement of the resolving power of the eye (Barten, 1999; De Valois and De Valois, 1991). However, gratings rarely exist in nature, and thus, to understand how spatial resolution influences visually guided behaviour in an ecological context, other measures may be more interesting. For example, when asking at what distance a passerine can detect a conspecific, or a raptor can spot its prey, target acuity – which we define as the detection threshold, or the minimal resolvable angle, for small or distant

single objects, might give a more relevant answer (Chaib et al., 2019). In order to compare grating and target resolution, we assume that the size of the target (in degrees [deg] of visual field) equals half a cycle (one black or one white stripe) of a square wave grating.

Many animals, including humans, have been shown to possess higher acuity for single targets than for gratings (Ehrenhardt, 1937; Hecht et al., 1947; Vallet and Coles, 1993). Humans can resolve gratings with a spatial frequency of around 60 cycles deg⁻¹, meaning that a single black or white stripe in the grating is 0.0083 deg wide. However, we can detect a single black line on a uniformly bright background, for instance a rope in front of the sky, even when it is only 0.00012 deg wide (Hecht et al., 1947), thus about 70 times narrower. Thus, target acuity is theoretically limited by contrast sensitivity, while grating acuity is limited by the resolving power of the retina (O'Carroll and Wiederman, 2014). In a recent study, we showed that this was not the case for budgerigars (*Melopsittacus undulatus*, Shaw 1805) that have similar acuity for single targets and gratings (Chaib et al., 2019). Budgerigars can resolve gratings with 7.7 to 10 cycles deg⁻¹, in which one black or white stripe subtends 0.05 to 0.065 deg of their visual field (Haller et al., 2014; Lind and Kelber, 2011; Lind et al., 2012), while they can just detect single targets of between 0.065 and 0.098 deg size, depending on the luminance profile of the target (Chaib et al., 2019). The main reason for this difference between humans and birds is presumably the birds' lower sensitivity for achromatic contrast (Ghim and Hodós, 2006; Haller et al., 2014; Harmening et al., 2009; Hirsch, 1982; Hodós et al., 2002; Lind et al., 2013, 2012; Orłowski et al., 2012; Potier et al., 2018; Reymond and Wolfe, 1981). Birds require around 10% Michelson contrast to discern gratings, while humans need less than 1% (De Valois et al., 1974). A high contrast target smaller than the resolution limit determined for gratings, will be perceived as having lower and lower contrast to the background, with decreasing size. For a bird the detection threshold will be reached for a larger target compared to for a human.

Many natural targets that are vital for a bird, such as a soaring falcon or a flying prey animal, are not stationary but rather dynamic. Moving visual objects are not necessarily perceived in the same way as stationary objects. The movement of an object relative to the background can break camouflage (Hall et al., 2013) or catch the viewer's attention (Richard and Shawn, 2003; Rushton et al., 2007), thereby making the object more salient and potentially lower the detection threshold. In humans, visual acuity is mostly impaired as a function of movement (Brown, 1972; Lewis et al., 2011), but under some circumstances it can also be improved. For example, peripheral visual acuity is slightly improved by slow target motion (Brown, 1972).

To our knowledge, the effect of motion on acuity and contrast sensitivity of birds has only been investigated with gratings. The contrast sensitivity of budgerigars is higher for horizontally drifting than for stationary achromatic gratings (Haller et al., 2014). For high spatial frequency (6.5 cycles deg⁻¹) gratings, a velocity of 1.4 deg s⁻¹ almost doubles contrast sensitivity for budgerigars. While Tyrrell et al. (2014) found that starlings (*Sturnus vulgaris*)

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Received 6 May 2021; Accepted 29 July 2021

were not very likely to visually fixate a stationary or a moving black dot, the effect of movement on the detection threshold of single targets has not previously been investigated.

During our previous experiment (Chaib et al., 2019) it was surprisingly difficult to train budgerigars to the task of detecting stationary single targets. If the unexpectedly low visual acuity for stationary targets was influenced by the lack of motivation from the birds, this could potentially be overcome by movement of the target (Pratt et al., 2010; Richard and Shawn, 2003). As a result of this assumption, our expectation was that budgerigars could detect smaller moving targets than stationary targets.

RESULTS AND DISCUSSION

Five of seven budgerigars learned to associate the presence of the target with a reward. They were able to detect a moving black target with a diameter subtending 0.107 ± 0.007 deg (mean \pm sd) of their visual field (Fig. 1), corresponding to a black-and-white grating with a spatial frequency of 4.68 ± 0.32 cycles deg^{-1} (in which a black and a white stripe subtend 0.214 deg). The bird with the highest acuity could detect a target subtending 0.091 deg of the visual field (5.48 cycles deg^{-1}), while the bird with the lowest acuity could detect a target subtending 0.124 deg (4.04 cycles deg^{-1}). Just as for stationary targets, the detection threshold for single black targets was lower than expected on the basis of grating acuity (7.7 to 10 cycles deg^{-1} ; Chaib et al., 2019; Haller et al., 2014; Lind and Kelber, 2011).

We knew from our earlier study that budgerigars have a detection threshold of 0.098 ± 0.008 deg, corresponding to 5.1 ± 0.45 cycles deg^{-1} , for a stationary target of the same shape and contrast to the background as the moving target (Chaib et al., 2019). Three birds participated in both experiments. To find out whether these two detection thresholds differ significantly, we fitted a linear mixed-effects model with random intercepts to the combined data with experiment type (moving target versus stationary target) as a fixed effect and individual birds as a random effect. We compared this model to a reduced model excluding the experiment type (fixed effect) and did not

find a significant effect of experiment type on the detection threshold ($\chi^2=0.74$, d.f.=1, $P=0.39$, AIC full model: -114.9 , AIC reduced model: -116.2). This indicates that, contrary to our expectation, the detection threshold for moving targets is not significantly different from the detection threshold for stationary targets.

We have previously calculated that a budgerigar, with a target acuity of around 0.1 deg of the visual field, would be able to spot a soaring brown falcon *Falco berigora* from a distance of 85 m (Chaib et al., 2019). We conclude from the present study that for a falcon moving at 1.69 deg s^{-1} the distance would be roughly the same. However, a speed of 1.69 deg s^{-1} would correspond to a groundspeed of 2.5 m s^{-1} from this distance, which is considerably lower than the soaring flight speed measured in other falcons (Cochran and Applegate, 1986; Rosén and Hedenström, 2002). In this study, we chose a retinal speed of the target which had previously shown to increase contrast sensitivity in budgerigars (Haller et al., 2014). This does not rule out that an ecologically more relevant speed, similar to that of a flying raptor, might yield a different result.

We hoped that a moving target would make it easier and more intuitive for the birds to detect the target and thus to learn the task, but this was not the case. This may be related to the findings by Tyrrell et al. (2014) that starlings were not more likely to visually fixate a randomly moving black dot compared to a stationary black dot. Interestingly the starlings trained by Tyrrell et al. (2014) only fixated the dot in 25% of the trials. Visually relevant stimuli, like a moving mealworm or a Harris's hawk (moving or stationary) were more likely to be fixated by the birds than the dot (Tyrrell et al., 2014). The fact that a starling is more likely to fixate on a stationary image of a hawk compared to a moving dot suggests that stimulus shape might be a greater indicator of importance than movement. However, besides differing in shape, these stimuli also differed in size, contrast, and movement type making it difficult to separate shape as an exclusive factor (Tyrrell et al., 2014). Moreover, in studying visual acuity, using elaborate targets like raptor silhouettes provides difficulties in quantifying the size of the target and thus comparing to other measures of spatial vision.

Another relevant factor may be the stimulus position. Chickens react to a black round target moving in a straight line above their head by predator avoidance response, including visual fixation (Hébert et al., 2019). The position of the stimulus, as well as the pattern of movement, likely have an impact on the relevance of the stimulus for the bird. Birds which are naturally exposed to aerial predators, like budgerigars, starlings and chickens, might be prone to fixate a dorsal straight moving target. A randomly moving target in the horizontal field of view, on the other hand, might be of less importance to ground foraging birds, although starlings occasionally catch insects in the air (Tinbergen, 1981). It is possible that birds of prey, or birds specialized in hawking, are more prone to pay attention to small unidentifiable moving targets. However, Harris's hawks also have proved difficult to condition to small moving targets (Simon Potier, Lund University, personal communication).

Experiments in optimal foraging suggest that birds will spend more time foraging by walking (a low-cost way of travel) with a low yield compared to foraging by flying (a high-cost way of travel) with a high yield (Bautista et al., 2001). With this in mind, we had trained the birds to walk instead of fly in a smaller experimental arena. Our expectation was that the birds would be able to do more trials per session for a smaller food reward. However, we did not experience a great difference in the birds' willingness to participate in the experiment compared to in previous experiment when the birds were flying.

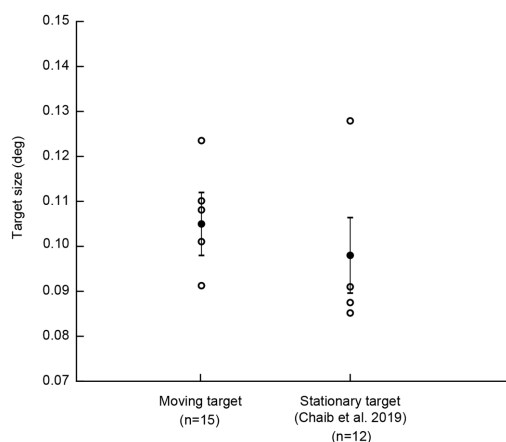


Fig. 1. Detection thresholds for moving, as well as stationary, single targets. Empty circles represent the thresholds for individual birds and filled circles represent means for all birds in the experiment. Error bars represent s.d.

To conclude, the target acuity of budgerigars is not better for moving targets than for stationary targets. Budgerigars do not instinctively visually fixate randomly moving black targets in the frontal or lateral visual field. It is possible that the position of the target might be of relevance and that a budgerigar might react differently to a dorsally presented target. An interesting future direction would be to investigate the moving target acuity in birds foraging on flying prey, like insects or small birds.

MATERIALS AND METHODS

Animals

Three female and four male budgerigars participated in the experiment. Three of these birds had also participated in a previous experiment with stationary targets (Chaib et al., 2019). The birds were fed a millet-based seed mix adapted for parakeets, vegetables and fruit except for experimental days when they received the seed mix only as a reward in the experiment, complemented by vegetables and fruit in the home cage. The birds participated in the experiments four consecutive days a week and rested for three days. All experiments were performed following Swedish legislation, under the permit M111-14 from the local authority for animal ethics.

Experimental setup

The experiments were performed in a y-maze with a removable top constructed of opaque polyacrylic sheets. A 15 cm wide, 20 cm long and 20 cm high compartment, the *start box*, would hold the bird at the start of each trial (Fig. 2A). The start box was open to two 73 cm long, 15 cm wide and 20 cm high corridors leading to two stimulus windows, each 15 cm high and 7 cm wide corresponding to 11.6×5.5 deg of visual angle as seen from the *decision line* (the boundary between the start box and the corridors; Fig. 1A). A monitor (32WL30MS, LG, Seoul, South Korea) positioned behind the stimulus windows displayed the stimuli (Fig. 1A). A feeder was positioned at the end of each corridor. Each feeder was connected to

a food dispenser (Lind, 2016) by a plastic tube (not shown in the figure).

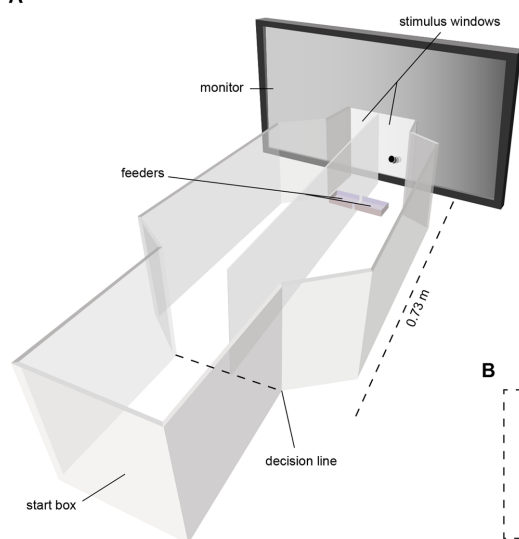
Stimuli

The rewarding stimulus consisted of a black dot (0.23 cd/m^2), the *target*, moving in a semi-random manner on a bright grey background (140 cd/m^2 ; $>99\%$ contrast). The direction in which the target moved for every new frame was normally distributed around the previous direction of travel. This way, the target had an erratic movement, although with smooth turns. The trajectory of the target centre never moved outside an area subtending 0.72×0.72 cm in the stimulus window and 0.56×0.56 deg of visual angle, as seen from the decision line. When the target reached the invisible boundary, the direction was reversed (Fig. 2B). It has been shown that budgerigars have higher contrast sensitivity for drifting than for stationary gratings (Haller et al., 2014). Thus, to obtain the most favourable conditions in the experiment, we set the target speed to 1.69 deg s^{-1} as seen from the decision point of the bird. This is close to the speed at which maximal contrast sensitivity was measured in the study by Haller et al. (2014). The unrewarding stimulus consisted of the same bright grey background as the rewarding stimulus but lacked the target. The stimuli were created in Matlab using the Psychophysics Toolbox extensions (Brainard, 1997; Pelli, 1997; Kleiner et al., 2007).

Experimental procedure

After an auditory start signal (three short consecutive tones), the rewarding stimulus, with the moving target, appeared in one of the stimulus windows – either the left or the right. The bird was trained to enter the corridor leading to the stimulus window presenting the target. When the bird made a correct choice, entering the corridor leading to the rewarding stimulus, a high pitch signal would sound, and a few seeds would be delivered into the feeder in that corridor. When the bird made an incorrect choice, and entered the corridor where no target was present, a low pitch signal would sound, and the

A



B

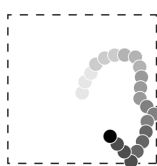


Fig. 2. The experimental setup and stimulus.

(A) The experimental setup. At the start of each trial, the bird was positioned in the start box viewing the monitor. When the target was displayed in one of the stimuli windows, the bird would make its choice by entering one of the corridors. The part of the monitor not visible in the stimuli windows was dark throughout the trials. The experimental arena was covered by a lid of transparent polyacrylic and a black fabric surrounded the sides of the setup (not seen in the figure). (B) An example of a target trajectory. The dashed line represents the invisible boundary in which the target centre moved.

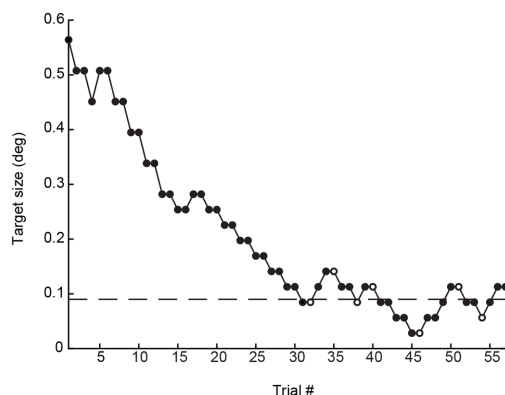


Fig. 3. An example of an adaptive staircase. Empty circles represent reversal points and the dashed line is the detection threshold of the session, calculated as the mean of the reversals in the last 25–30 trials. The example is taken from ‘female 2’ (Fig. S1D). All test sequences included in the analysis of the experiment can be found in the Supplementary Material (Fig. S1A–E).

stimuli would disappear from the stimuli windows. In both cases, a new trial started after the bird had returned to the start box and faced the corridors.

In the initial training sessions only the largest targets, with diameters of 1.44 and 0.71 cm or 1.13 and 0.56 deg of the visual field, were used. Once a bird had learned to choose the correct side in the y-maze, we started the staircase sessions, in which the size of the target was changed following an adaptive 1-up/2-down staircase procedure (Levitt, 1971; Fig. 3). In the staircase sessions, the initial target size was 0.56 deg of the visual field, which is well above the detection threshold of the birds (Chaib et al., 2019). Target size would decrease after two consecutive correct choices, but increase again after one incorrect choice, until target size fluctuated around the level at which the probability of a decrease of target size equals the probability of an increase of target size. This level corresponds to the point on a psychometric function where the probability of making a correct choice is 70.7% (Levitt, 1971). The staircase step sizes were ± 0.056 deg (of the diameter of the target) above a target size of 0.282 deg and ± 0.028 deg below this size. Each test session consisted of 45–60 trials depending on the motivation of the bird. Consistent with our experience from previous experiments using stationary targets, the birds improved their detection threshold during the first three to five training sessions until they reached a plateau (Chaib et al., 2019). If a bird did not improve over three sessions in a row, we concluded that this represented its maximal performance and ended the experiment.

Analysis

We analysed the data from the last three sessions for each bird, which are the sessions after the bird reached the performance plateau. The thresholds were estimated by averaging the reversal points, the values in the staircase where the curve slope changes direction (Fig. 3), of the last 25–30 trials (depending on the length of the session) in each of the three sessions. We used an even number of reversal points for each test session to avoid any estimation bias (Levitt, 1971). The individual thresholds for each test session obtained this way were compared to the detection thresholds for stationary targets that had been

determined in a previous experiment. A linear mixed-effects model with random intercept was fitted to the pooled data from both experiments, including birds participating in both experiments as well as birds only participating in one of the experiment. The model included with experiment type (stationary target or moving target) as a fixed effect and bird identity as a random effect, using the lmerTest package (Kuznetsova et al., 2017) in RStudio (v. 1.1.463; R Studio Team, 2016). This model was compared to a reduced model, excluding the fixed effect of experiment type, with a log-likelihood ratio test. Additionally, the two models were compared by their Akaike information criterion (AIC) values.

Acknowledgements

The authors wish to thank Robin Johnsson for helping us to build the experimental setup and for his contribution to training the birds, Mikael Ljungholm for helping us create the target, and two anonymous reviewers for constructive comments on the manuscript.

Competing interests

The authors declare no competing or financial interests.

Author contributions

Conceptualization: S.C., O.L., A.K.; Methodology: S.C., O.L., A.K.; Software: S.C., O.L.; Validation: S.C.; Formal analysis: S.C.; Investigation: S.C., J.G.M.; Resources: O.L., A.K.; Data curation: S.C.; Writing - original draft: S.C.; Writing - review & editing: S.C., J.G.M., O.L., A.K.; Visualization: S.C.; Supervision: O.L., A.K.; Project administration: A.K.; Funding acquisition: A.K.

Funding

This study was funded by Swedish Research Council (2016-03298_3).

References

- Barten, P. G. J. (1999). *Contrast sensitivity of the human eye and its effects on image quality*. Bellingham, WA: SPIE – International Society for Optical Engineering.
- Bautista, L. M., Tinbergen, J. and Kacelnik, A. (2001). To walk or to fly? How birds choose among foraging modes. *Proc. Natl. Acad. Sci. USA* **98**, 1089–1094.
- Boström, J. E., Dimitrova, M., Canton, C., Hästad, O., Qvarnström, A. and Ödeen, A. (2016). Ultra-rapid vision in birds. *PLoS ONE* **11**, 7. doi:10.1371/journal.pone.0151099
- Brainard, D. H. (1997). The psychophysics toolbox. *Spat. Vis.* **10**, 433–436. doi:10.1163/156856897X00357
- Brown, B. (1972). Resolution thresholds for moving targets at the fovea and in the peripheral retina. *Vis. Res.* **12**, 293–304. doi:10.1016/0042-6989(72)90119-8
- Chaib, S., Ljungholm, M., Lind, O. and Kelber, A. (2019). Single target acuity is not higher than grating acuity in a bird, the budgerigar. *Vis. Res.* **160**, 37–42. doi:10.1016/j.visres.2019.04.005
- Cochran, W. W. and Applegate, R. D. (1986). Speed of flapping flight of merlins and peregrine falcons. *Condor* **88**, 397–398. doi:10.2307/1368897
- De Valois, K. K. and DeValois, R. L. (1991). *Spatial Vision*. Oxford, UK: Oxford University Press.
- De Valois, R. L., Morgan, H. and Snodderly, D. M. (1974). Psychophysical studies of monkey vision-III. Spatial luminance contrast sensitivity tests of macaque and human observers. *Vis. Res.* **14**, 75–81. doi:10.1016/0042-6989(74)90118-7
- Ehrenhardt, H. (1937). Formensehen und Sehschärfbestimmungen bei Eidechsen. *Z. vergl. Physiol.* **24**, 248–304.
- Fischer, A. B. (1969). Laboruntersuchungen und Freilandbeobachtungen zum Sehvermögen und Verhalten von Altweltgeiern. *Zool. Jahrb. Syst. Ökol. Geogr. Tiere* **96**, 81–132.
- Ghim, M. M. and Hodos, W. (2006). Spatial contrast sensitivity of birds. *J. Comp. Physiol. A* **192**, 523–534. doi:10.1007/s00359-005-0090-5
- Hall, J. R., Cuthill, I. C., Baddeley, R., Shohet, A. J. and Scott-Samuel, N. E. (2013). Camouflage, detection and identification of moving targets. *Proc. Royal Soc. B* **280**, 20130064. doi:10.1098/rspb.2013.0064
- Haller, N. K., Lind, O., Steinelechner, S. and Kelber, A. (2014). Stimulus motion improves spatial contrast sensitivity in budgerigars (*Melopsittacus undulatus*). *Vis. Res.* **102**, 19–25. doi:10.1016/j.visres.2014.07.007
- Harmening, W. M., Nikolay, P., Orłowski, J. and Wagner, H. (2009). Spatial contrast sensitivity and grating acuity of barn owls. *J. Vis.* **9**, 13. doi:10.1167/9.7.13
- Hébert, M., Versace, E. and Vallortigara, G. (2019). Inexperienced preys know when to flee or to freeze in front of a threat. *Proc. Natl. Acad. Sci. USA* **116**, 22918–22920. doi:10.1073/pnas.1915504116

- Hecht, S., Ross, S. and Mueller, C. G. (1947). The visibility of lines and squares at high brightness. *J. Opt. Soc. Am. B* **37**, 500-507. doi:10.1364/JOSA.37.000500
- Hirsch, J. (1982). Falcon visual sensitivity to grating contrast. *Nature* **300**, 57-58. doi:10.1038/300057a0
- Hodos, W., Ghim, M. M., Potocki, A., Fields, J. N. and Storm, T. (2002). Contrast sensitivity in pigeons: a comparison of behavioral and pattern ERG methods. *Doc. Ophthalmol.* **104**, 107-118. doi:10.1023/A:1014427615636
- Kelber, A. (2019). Bird colour vision - from cones to perception. *Curr. Opin. Behav. Sci.* **30**, 34-40. doi:10.1016/j.cobeha.2019.05.003
- Kleiner, M., Brainard, D., Pelli, D., Ingling, A., Murray, R. and Broussard, C. (2007). What's new in Psychtoolbox-3? *Perception* **36**, 1-16.
- Kuznetsova, A., Brockhoff, P. B. and Christensen, R. H. B. (2017). lmerTest package: tests in linear mixed effects models. *J. Stat. Softw.* **82**, 1-26. doi:10.18637/jss.v082.i13
- Land, M. F. and Nilsson, D.-E. (2012). *Animal Eyes*. Oxford, UK: Oxford University Press.
- Levitt, H. (1971). Transformed up-down methods in psychoacoustics. *J. Acoust. Soc. Am.* **49**, 467-477. doi:10.1121/1.1912375
- Lewis, P., Rosen, R., Unsbo, P. and Gustafsson, J. (2011). Resolution of static and dynamic stimuli in the peripheral visual field. *Vis. Res.* **51**, 1829-1834. doi:10.1016/j.visres.2011.06.011
- Lind, O. (2016). Colour vision and background adaptation in a passerine bird, the zebra finch (*Taeniopygia guttata*). *Royal Soc. Open Sci.* **3**, 160383. doi:10.1098/rsos.160383
- Lind, O. and Kelber, A. (2011). The spatial tuning of achromatic and chromatic vision in budgerigars. *J. Vis.* **11**, 2. doi:10.1167/11.7.2
- Lind, O., Karlsson, S. and Kelber, A. (2013). Brightness discrimination in budgerigars (*Melopsittacus undulatus*). *PLoS ONE* **8**, e54650. doi:10.1371/journal.pone.0054650
- Lind, O., Sunesson, T., Mitkus, M. and Kelber, A. (2012). Luminance-dependence of spatial vision in budgerigars (*Melopsittacus undulatus*) and Bourke's parrots (*Neopsephotus bourkii*). *J. Comp. Physiol. A* **198**, 69-77. doi:10.1007/s00359-011-0689-7
- Martin, G. (2017). *The Sensory Ecology of Birds*. Oxford, UK: Oxford University Press.
- O'Carroll, D. C. and Wiederman, S. D. (2014). Contrast sensitivity and the detection of moving patterns and features. *Philos. Trans. R. Soc. Lond. B* **369**, 20130043. doi:10.1098/rstb.2013.0043
- Orlowski, J., Harmening, W. and Wagner, H. (2012). Night vision in barn owls: Visual acuity and contrast sensitivity under dark adaptation. *J. Vis.* **12**, 4. doi:10.1167/12.13.4
- Pelli, D. G. (1997). The VideoToolbox software for visual psychophysics: Transforming numbers into movies. *Spat. Vis.* **10**, 437-442. doi:10.1163/156856897X00366
- Potier, S., Mitkus, M. and Kelber, A. (2018). High resolution of colour vision, but low contrast sensitivity in a diurnal raptor. *Proc. Royal Soc. B* **285**, 20181036. doi:10.1098/rspb.2018.1036
- Potier, S., Lieuvain, M., Pfaff, M. and Kelber, A. (2020). How fast can raptors see? *J. Exp. Biol.* **223**, jeb209031. doi:10.1242/jeb.209031
- Pratt, J., Radulescu, P. V., Guo, R. M. and Abrams, R. A. (2010). It's alive!: animate motion captures visual attention. *Psychol. Sci.* **21**, 1724-1730. doi:10.1177/0956797610387440
- Reymond, L. (1985). Spatial visual acuity of the eagle *Aquila audax*: a behavioural, optical and anatomical investigation. *Vis. Res.* **25**, 1477-1491. doi:10.1016/0042-6989(85)90226-3
- Reymond, L. and Wolfe, J. (1981). Behavioural determination of the contrast sensitivity function of the eagle *Aquila audax*. *Vis. Res.* **21**, 263-271. doi:10.1016/0042-6989(81)90120-6
- Richard, A. A. and Shawn, E. C. (2003). Motion onset captures attention. *Psychol. Sci.* **14**, 427-432. doi:10.1111/1467-9280.01458
- Rosén, M. and Hedenström, A. (2002). Soaring flight in the Eleonora's falcon (*Falco eleonorae*). *Auk* **119**, 835-840. doi:10.1093/auk/119.3.835
- R Studio Team (2016). *RStudio: Integrated Development for R*. Boston, MA: RStudio, Inc.
- Rushton, S. K., Bradshaw, M. F. and Warren, P. A. (2007). The pop out of scene-relative object movement against retinal motion due to self-movement. *Cognition* **105**, 237-245. doi:10.1016/j.cognition.2006.09.004
- Tinbergen, J. M. (1981). Foraging decisions in starlings (*Sturnus vulgaris* L.). *Ardea* **69**, 1-67.
- Tyrrell, L. P., Butler, S. R., Yorzinski, J. L. and Fernández-Juricic, E. (2014). A novel system for bi-ocular eye-tracking in vertebrates with laterally placed eyes. *Methods Ecol. Evol.* **5**, 1070-1077. doi:10.1111/2041-210X.12249
- Vallet, A. M. and Coles, J. A. (1993). The perception of small objects by the drone honeybee. *J. Comp. Physiol. A Neuroethol. Sens. Neural. Behav. Physiol.* **172**, 183-188.

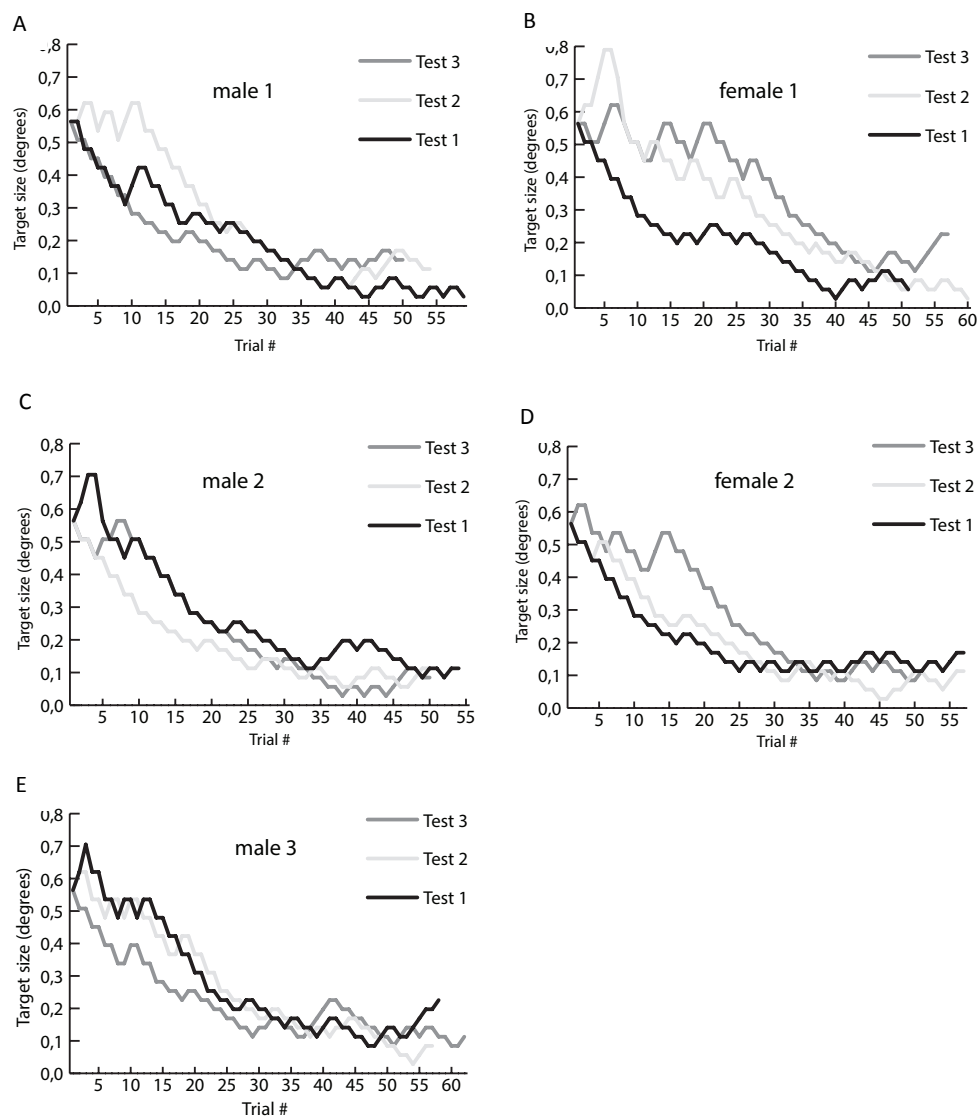


Fig. S1. Results from the moving target acuity test following an adaptive 2-down/1-up procedure. Fig A-E show the three test sessions from each of the five birds included in the analysis of the experiment.

Paper III



Research



Cite this article: Chaib S, Lind O, Kelber A. 2023 Fast visual adaptation to dim light in a cavity-nesting bird. *Proc. R. Soc. B* **290**: 20230596.

<https://doi.org/10.1098/rspb.2023.0596>

Received: 11 March 2023

Accepted: 17 April 2023

Subject Category:

Neuroscience and cognition

Subject Areas:

neuroscience, ecology, behaviour

Keywords:

cavity-nesting birds, budgerigar, vision, dark-adaptation, contrast vision, visual adaptation

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Electronic supplementary material is available online at <https://doi.org/10.6084/m9.figshare.c.6619772>.

Fast visual adaptation to dim light in a cavity-nesting bird

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Many birds move fast into dark nest cavities forcing the visual system to adapt to low light intensities. Their visual system takes between 15 and 60 min for complete dark adaptation, but little is known about the visual performance of birds during the first seconds in low light intensities. In a forced two-choice behavioural experiment we studied how well budgerigars can discriminate stimuli of different luminance directly after entering a darker environment. The birds made their choices within about 1 s and did not wait to adapt their visual system to the low light intensities. When moving from a bright facility into an environment with 0.5 log unit lower illuminance, the budgerigars detected targets with a luminance of 0.825 cd m⁻² on a black background. When moving into an environment with 1.7 or 3.5 log units lower illuminance, they detected targets with luminances between 0.106 and 0.136 cd m⁻². In tests with two simultaneously displayed targets, the birds discriminated similar luminance differences between the targets (Weber fraction of 0.41–0.54) in all light levels. Our results support the notion that partial adaptation of bird eyes to the lower illumination occurring within 1 s allows them to safely detect and feed their chicks.

1. Introduction

Birds use vision in a wide range of different light regimes, from bright daylight (approx. 10⁵ lux) to dim starlight under the canopy of trees (approx. 10⁻³ lux [1]). Although some species spend their awake time in dim light conditions, most birds are active in bright daylight, where their visual system allows them to scan the environment rapidly and in great detail [2]. Still, even nocturnal birds can use vision in daylight, and diurnal birds can to some extent see in dim light, which is important, as light levels in natural habitats are highly dynamic. During a day, skylight levels change by a factor of 1000, and a bird flying from an open field into the woods can experience a light intensity decrease by a factor of 100 or more [1]. Thus, bird eyes need continuous adaptation to match the present light conditions.

Vertebrate eyes employ several strategies to adapt to changing light conditions. One strategy is the pupillary light response which controls the amount of light reaching the retina [3]. In most land-living vertebrates, pupil dilation and constriction work within seconds [2–4]. Even though its dynamic range varies between different species, the pupillary light response accounts for a small amount of adaptation only (in budgerigars (*Melopsittacus undulatus*), for instance, changing the retinal illumination by less than a factor of 2 [4]), and its primary purpose has been attributed to other functions such as preventing a decrease in visual acuity in bright light owing to optical aberrations [2–4].

A second adaptation to vision in a broad range of light intensities is the ‘duplex retina’ of vertebrates, with two sets of photoreceptors, rods—active in dim (scotopic) light conditions, and cones—active during bright (photopic) light conditions [5,6]. At intermediate (mesopic) light levels both cones and rods contribute to vision. Birds have an even more complex retina; in addition

to rods and cones they have double cones, which can operate under somewhat lower light conditions than single cones [7].

Although the different receptor types allow the visual system to work in a wide intensity range, additional retinal mechanisms, at different light levels and with different time courses are involved in luminance adaptation [8–11].

Prolonged exposure to bright light ‘bleaches’ a part of the photoreceptor pigment which has to be regenerated before it can absorb photons again [12,13]. The recovery of full sensitivity, known as dark-adaptation, can take more than 40 min in humans [13,14] and has a similar time-course in birds [15–17]. Cone recovery is faster and occurs within just a few minutes [13].

In addition, fast mechanisms allow the vertebrate retina to cope with the smaller luminance changes associated with the change of gaze within a visual scene. Response gain is adjusted to the background luminance, enhancing the visual signal at dim backgrounds and preventing saturation in bright light [18]. These mechanisms act, within less than a second, at photoreceptor level and later stages in the retinal pathway [8,18–20].

As a result of luminance adaptation, the visual system can extract detailed information and keep contrast sensitivity largely constant in a wide range of light intensities [9]. The sensitivity of the adapted eye to luminance differences is proportional to the average light intensity, a relation referred to as Weber’s law ($\Delta I/I_B = k$ [9,10]). This linear relationship between background luminance and contrast sensitivity only breaks down at very high light levels owing to saturation of the photoreceptors and at low light levels owing to receptor noise [9].

The visual sensitivity and spatial resolution of birds have been studied after adaption to various light levels, and full dark adaptation has a similar time-course in birds as in humans [14,15]. By contrast, very little is known about the visual capacities of birds after fast changes of luminance. When provisioning their brood, cavity nesting birds may move frequently back and forth between sunlit foraging grounds and the tree cavities or nest-boxes, where they may face light intensities 1000-fold dimmer than outside [21,22]. Visits to the nest often take only a few seconds, probably too short for dark adaptation by slow mechanisms such as photopigment regeneration [15].

Several studies indicate that cavity nesting birds use visual cues when feeding their chicks: after experimental manipulation of the colour of the gape flanges of cavity-breeding passerine nestlings, brighter and more conspicuously coloured individuals gain more weight than their duller siblings [23–25]. Lower luminance contrast between chicks and their background or lower illumination in the nest make food transfer between parent and nestlings more difficult [26,27]. Breeding great tits (*Parus major*) choose brighter nest-boxes over darker ones [28], and in darker nest-boxes, both great tits and marsh tits (*Poecile palustris*) build nest cups closer to the entrance, probably to compensate for the dim illumination [29,30]. Some cavity nesting passerines also use vision to discriminate their eggs from the eggs of brood parasites and assess the fitness of the female (i.e. [31,32]).

Here we use the budgerigar, a species that nests in tree holes in the interior of Australia [33] as a model species to study luminance discrimination of birds that move fast from a bright to a dark environment. Pupil dynamics [4], contrast sensitivity [34–37], spatial acuity [34,36–39] and colour vision

[7,34,40–42] of the species have previously been investigated under different light levels. We performed two behavioural experiments to investigate (i) the detection threshold for bright targets on dark background, and (ii) the discrimination threshold for two targets of different luminance.

2. Methods

(a) Animals

Two female and two male budgerigars participated in the experiment. The budgerigars were held in the animal facility at the Department of Biology at Lund University and fed a parakeet seed mix as well as fresh fruit and vegetables. On days of training and testing (3–5 days per week), the seed mix was restricted and only used as a reward. However, the birds had always access to fresh fruit and vegetables in the housing cage. All experiments followed Swedish legislation, under the permit dn. 5.8.18-17189/2018 granted by the responsible authority (Malmö – Lunds djurförsöksetiska nämnd).

(b) Experimental set-up

The experimental set-up comprised two compartments, the large flight cage, and the smaller decision box (figure 1a). The flight cage was a net cage, 133 cm long, 65 cm high and 84 cm wide, with a grey plastic floor. A waiting perch was positioned halfway between and parallel to the short walls, and 20 cm above the floor, for the experimental bird. A camera (GoPro Hero, GoPro Inc., San Mateo, CA 94402, USA) on one short wall of the flight cage allowed us to monitor the behaviour of the bird without being seen. The opposite short wall was made of wood and separated the flight cage from the decision box. A bird reached the decision box by flying or by climbing a wooden ramp and entering a corridor (10 cm high and wide and 12 cm long), which protruded into the flight cage from an opening in the wooden wall 25 cm above the floor. This corridor prohibited light from the flight cage from entering the decision box through the opening.

The decision box had solid side walls and was 22 cm long, 20 cm wide and 36 cm high. The wall opposite the entrance was open to a monitor that displayed visual stimuli in two stimulus windows, one to the right and one to the left (figure 1b). Decision distance was controlled by a divider that protruded orthogonally from between the stimulus windows 12 cm into the box. A bird entering the decision chamber would view two different stimuli, one on the right and one on the left side of the wall and make a choice by entering the compartment on either side. The floor beneath each stimulus window had a sliding door which was controlled manually by the experimenter from the outside and could be opened to provide a reward of seeds.

Four white LEDs (LZC-00NW40, LED Engin Inc., San Jose, USA) and two fluorescent tubes (Biolux L18W/965; Osram, flicker frequency 9 500 Hz) illuminated the decision box and the flight cage indirectly from above by reflection from wrinkled aluminium foil to ensure an even illumination. This provided an illuminance of 1400 lux at the centre of the waiting perch (measured with a Hagner Luxmeter, Hagner AB, Solna, Sweden, pointing upwards). The illuminance in the smaller decision box was 469 lux (measured the same way as above, with the luxmeter placed between the entrance and the end of the dividing wall, 5 cm above the floor) at the brightest illuminance condition (light level 1). By placing neutral density filters (Lee filters, Andover, Hampshire, UK) on top of the transparent plastic roof of the decision box the light level was dimmed to obtain three additional illuminance conditions: 28 lux (light level 2), 1.83 lux (light level 3) and 0.47 lux (light level 4). The illuminance at light level 4 was extrapolated from the other

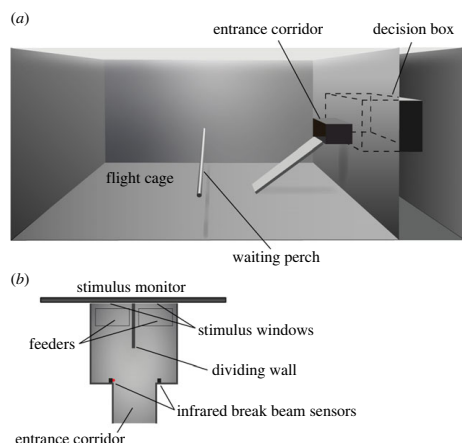


Figure 1. Experimental set-up. (a) Side view of the setup. (b) The decision box viewed from above.

values, as the instrument was not sensitive enough to give reliable readings at this light level. Luminance of the test targets was measured with a Hagner photometer (Hagner AB, Solna, Sweden) by placing the sensor close to the stimulus monitor at the position of the target. During tests at a specific light level, a neutral density filter corresponding to the filter used on top of the decision box was placed in front of the monitor to match the decrease in illuminance. A table of light conditions at the four different light levels can be found in the electronic supplementary material, table S1.

Just inside the entrance to the decision box, next to the corridor, a set of infrared break beam sensors (Adafruit Industries, New York, USA) monitored the time when the bird entered. The behaviour of the bird in the decision box was monitored using a Pi NoIR camera module. The break beam sensor and camera module were controlled by single-board computers (Raspberry Pi 2, model B, Raspberry Pi, Pencoed, Wales).

(c) Experiment 1: detection threshold

(i) Experimental procedures

We performed two experiments. Experiment 1 was designed to measure the detection threshold of the birds for a bright target on a dark background. At the start of each trial the bird was sitting on the waiting perch in the flight cage. In response to an auditory start signal, the bird flew or climbed up to the corridor and entered the decision box. As soon as the bird passed the break beam sensors, two stimuli were presented in the stimulus windows, one positive (rewarding) stimulus and one negative (unrewarding) stimulus. The negative stimulus was plain black while the positive stimulus was black with a bright circular target (figure 2a) of 2.2 cm diameter, and thus, extending 9.6° of the visual field of the bird when viewed from the decision point at the end of the dividing wall. The positive and negative stimuli were presented semi-randomly to the right and left according to Fellows [43]. The bird made a choice by approaching one of the stimuli. If the bird chose the positive stimulus, the feeder in front of it was opened and the bird was allowed to eat for a few seconds. If the bird chose the negative stimulus the feeder was not opened, and the target disappeared.

To initiate a new trial, the bird had to return to the waiting perch. The choice made by the bird was recorded manually and opening and closing of the feeder as well as the start of a

new trial were controlled by the experimenter who watched the bird via the two cameras. The minimum luminance detection threshold was determined using a one-up/two-down staircase procedure [44]. If the bird made two consecutive correct choices on the same level of difficulty, this was considered as a successful trial unit, and the luminance of the positive stimulus was decreased in the next trial. By contrast, when the bird made one incorrect choice, or, alternatively, one correct choice followed by one incorrect choice, on the same difficulty level, this was considered an unsuccessful trial unit and the luminance was increased for the next trial (figure 2a,c). To increase the motivation of the birds in the beginning of a test session each staircase started at a degree well above the threshold of the birds. Step sizes were larger at the beginning of a staircase and were decreased at two occasions to be smallest near the threshold, for increased precision (electronic supplementary material, table S2). One staircase session included between 38 and 100 choices depending on how fast the bird reached a steady state performance, meaning that it did not improve its performance in further trials. Each bird participated in three staircase sessions in each of the four light levels.

To investigate whether the period of adaptation to the different light intensities in the experimental set-up had an influence on the test outcome we measured two different time intervals for each trial. We defined the response time as the period between the time points when a bird entered the decision box (registered by the infrared break beam sensor) and when it made a choice (manually registered by the experimenter). To estimate the time that a bird spent in the high light intensity in the flight cage we measured the inter-trial time as the period between the time when a bird left the decision box and the onset of a new trial.

(ii) Analysis of luminance detection threshold

The detection thresholds were analysed in absolute luminance values and not in Weber fractions, in relation to the background luminance, which would have been another possibility. The reason for this choice was that the background luminance at light levels 3 and 4 was below the absolute luminance threshold for budgerigars [34], why it would probably not influence the detection thresholds.

For every staircase session, the threshold was calculated as the average of the values at the reversals of the last 20 choices (figure 2c). To avoid estimation bias, we used an even number of reversals for each staircase session, excluding the first reversal if needed.

We used the r-package ‘lmerTest’ [45] in Rstudio (v. 1.4.1106 [46]) to fit linear mixed effect models (LMMs), by maximum likelihood, to the staircase detection thresholds (three samples for each bird and light level). The dependent variable (luminance detection threshold) was log-transformed to better fit the assumption of normality. To test whether the detection thresholds differed between the different light levels we compared a full model including light level as a fixed effect, to a reduced model excluding this fixed effect, with a likelihood ratio test. Individual bird was included in both models as a random intercept to avoid pseudo-replication. We used the ‘multcomp’ package [47] in RStudio to perform Tukey’s *post hoc* test on the detection thresholds for the different light levels.

(iii) Analysis of time intervals

The response time and inter-trial time data are based on the same trials as the threshold data. The only exception are trials in which a bird entered and exited the decision box several times before making a choice. In these cases, the response time data were excluded from the analysis.

Response time data were fitted to an LMM. The dependent variable (response time) was inverse-transformed to better

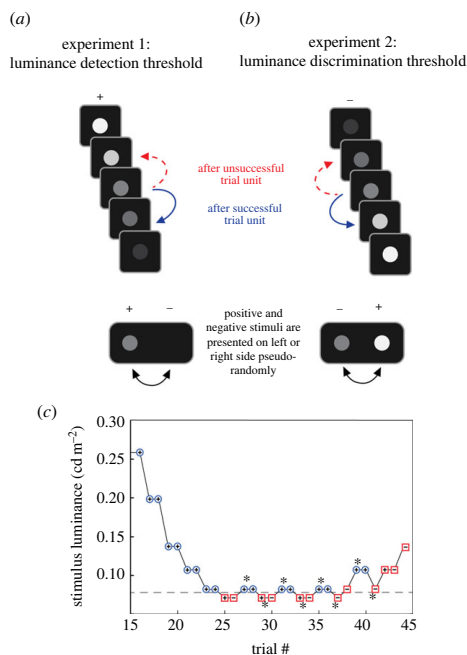


Figure 2. Example of stimuli from (a) experiment 1 and (b) experiment 2. The stimuli series (a,b) show stimuli of increasing difficulty from top to bottom. A successful trial unit (two consecutively correct trial choices including the same target luminance) resulted in an increase of difficulty (blue solid arrow), whereas an unsuccessful trial unit (either one incorrect trial choice or one correct choice followed by an incorrect trial choice including the same target luminance) resulted in a decrease in difficulty (red dashed arrow). (c) An example staircase from experiment 1. The plus and minus signs indicate trials with correct and incorrect choices, a blue coloured circle and red coloured squares indicate whether the choice belongs to a successful or unsuccessful trial unit. Asterisks indicate the reversals of the last 20 choices, which are included in the analysis of the threshold (dashed line). Note that the first reversal is removed from the analysis to give equal weight to successful and unsuccessful trial units.

meet the assumption of normality. The full model included the main effects of inter-trial time, light level, successful/unsuccessful trial unit, as well as the interactions between inter-trial time and successful/unsuccessful trial unit and light level and successful/unsuccessful trial. Individual bird was included in all models as a random intercept to account for pseudo-replication. We used Akaike information criterion (AIC) to find the most likely model and a likelihood ratio test to compare models.

(d) Experiment 2: discrimination threshold

(i) Experimental procedure

In experiment 2, we used the same test procedure as in experiment 1 to estimate the discrimination threshold for luminance differences between two simultaneously displayed targets. The positive stimulus consisted of a bright round target on a dark background with a constant luminance throughout the session (200, 10.4, 0.68 and 0.19 cd m^{-2} at levels 1 to 4, respectively). The negative stimulus was a target of lower luminance on the same dark background (figure 2b). Again, positive and negative stimuli were presented semi-randomly to the right and left

according to Fellows [43]. The targets had the same size as the target in experiment 1 and were separated by 30.7° (centre to centre). Using the same one-up/two-down staircase procedure, and gradually decreased step sizes (electronic supplementary material, table S3), we tested the birds until the choices fluctuated around the discrimination threshold (figure 2b). The birds made between 35 and 100 choices in each staircase session. Two of the birds, bird 1 and bird 3, participated in three test sessions for each light level, whereas bird 2 participated in three sessions only for level 1, 2 and 3, and bird 4 participated in three sessions for level 1 and 2 and one session for level 3. As in experiment 1, we included the values at the reversals of the last 20 choices and used an even number of reversals.

(ii) Analysis of luminance difference threshold

The luminance difference thresholds are expressed in Weber fractions. According to Weber's law, the minimum detectable difference of a new stimulus is proportional to the value of the reference stimulus. In our experiment we calculate the Weber fraction k as

$$k = \frac{\Delta I}{I_+}, \quad (2.1)$$

where ΔI is the luminance difference between the positive and the negative stimulus and I_+ is the luminance of the positive stimulus.

LMMs were fitted to the data and analysed in the same way as in experiment 1.

(iii) Analysis of time intervals

Response times and inter-trial times were measured and analysed in the same way as in experiment 1.

3. Results

(a) Experiment 1

(i) Luminance detection threshold

The results are based on a total of 48 staircase thresholds from four birds at four light levels. The birds were able to discriminate targets of similar luminance in the three darkest light levels (2–4), while the detection threshold on the brightest level (1) was considerably higher. Model comparison showed that a model including light level as a fixed effect and individual bird as a random effect had a significantly better fit to the data than a model only including individual bird as a random effect ($n = 48$, $\chi^2_3 = 126.7$, $p < 0.001$). We therefore concluded that light level had an effect on detection threshold. At light levels 2 and 4 the estimated detection thresholds were similar (0.110 cd m^{-2} , 95% confidence interval (CI) [0.092, 0.132], and 0.106 cd m^{-2} , 95% CI [0.091, 0.123]) (figure 3a), while the threshold at level 3 was significantly higher (0.136 cd m^{-2} , 95% CI [0.113, 0.163]) than at level 4 ($p < 0.04$). The threshold at level 1 (0.825 cd m^{-2} , 95% CI [0.685, 0.988]) was higher than the thresholds at all other light levels ($p < 0.001$).

(ii) Time intervals

The median response time in experiment 1 was 1.17 s ($n = 959$ choices by four birds, first to third quartile = $0.93\text{--}1.60 \text{ s}$). The model that best explains the distribution of response times included the effect of light level ($F = 3.27$, $p < 0.05$), successful/unsuccessful trial unit ($F = 24.14$, $p < 0.001$), as well as the interaction between these two ($F = 2.63$, $p < 0.05$), and this model had a significantly better fit to the data compared

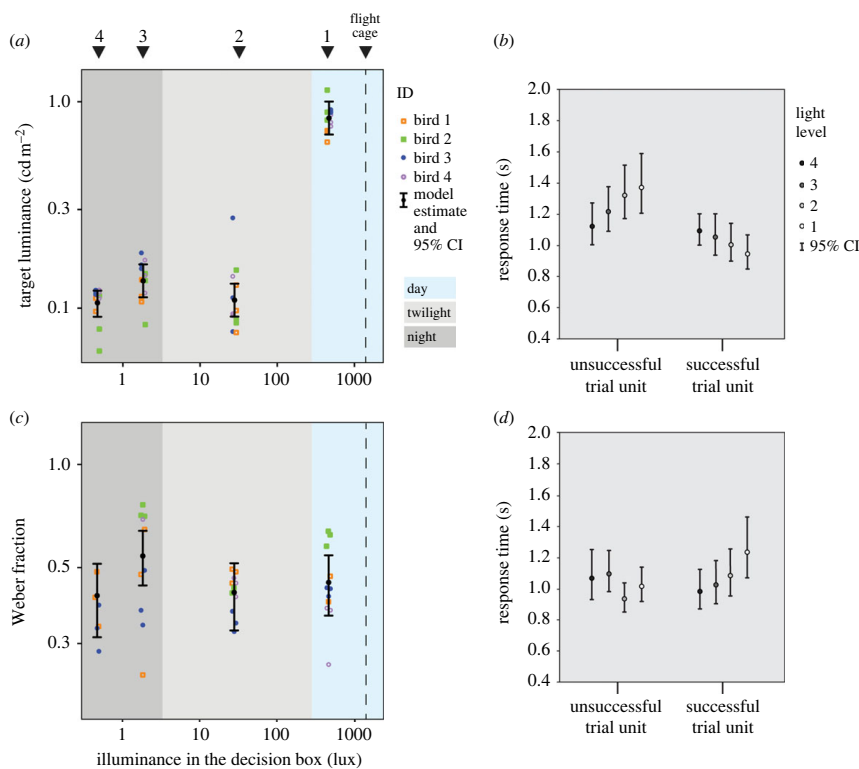


Figure 3. (a,b) Experiment 1: (a) detection threshold—the minimum luminance, at which birds could detect the target. Different colours and symbols represent results from individual birds, while black filled circles are the back-transformed model estimates of the thresholds. Numbers on top indicate illuminance in the decision box at the different light levels, dashed lines represent illuminance at the waiting perch in the flight cage. The definition of day, twilight and night illuminance follows Martin [1]. (b) Response times for trials in experiment 1 at each light level and separately for successful or unsuccessful trial units. Response times are the back-transformed estimates from the LMM. (c,d) Experiment 2: (c) discrimination threshold—the minimum luminance difference, expressed as Weber fraction (equation (2.1)), for which two bright targets could be discriminated. (d) Response times for experiment 2.

to the null model ($\chi^2_2 = 35.9$, $p < 0.001$). The birds had a shorter response time in successful trial units than in unsuccessful trial units. Median inter-trial time was 8.33 s (first to third quartile = 6.41–13.80 s).

(b) Experiment 2

(i) Luminance discrimination threshold

The results include a total of 40 staircase thresholds from four individuals tested at four light levels. Light level had a significant effect on luminance discrimination threshold ($n = 40$, $\chi^2_3 = 10.07$, $p < 0.05$). The smallest discriminable difference, a Weber fraction of 0.41 (95% CI [0.32, 0.53]) was found at level 4 (figure 3c). The threshold at level 1 was equivalent to a Weber fraction of 0.45 (95% CI [0.37, 0.55]), for level 2 it was 0.42 (95% CI [0.34, 0.53]), and for level 3 it was 0.54 (95% CI [0.46, 0.65]). The threshold at level 3 was significantly different to thresholds at level 2 ($p < 0.05$) and level 4 ($p < 0.05$).

(ii) Time intervals

The median response time in experiment 2 was 0.98 s ($n = 799$ choices by four birds, first to third quartile = 0.82–1.26 s). The most likely model included the fixed factors of inter-trial time ($F = 11.7$, $p < 0.001$), successful/unsuccessful trial unit ($F =$

32.5, $p < 0.001$) and light level ($F = 11.2$, $p < 0.001$), as well as the interaction between light level and successful/unsuccessful trial unit ($F = 2.76$, $p < 0.05$). This model had a significantly better fit to the data than the null model excluding all fixed effects ($\chi^2_3 = 95.0$, $p < 0.001$). Median inter-trial time for experiment 2 was 7.79 s (first to third quartile = 6.17–11.13 s).

4. Discussion

By training budgerigars to leave a bright (1400 lux) flight cage, enter a dark decision box and detect or discriminate stimuli of different luminance, our experiment simulates the sudden drop in light intensity that cavity nesting birds experience when entering a dark nest to care for their offspring. In experiment 1 we investigated the luminance threshold for the detection of a bright target on a dark background and found that it depended on the light level in the decision box. The detection threshold for the lower three light levels (0.47–28 lux) were in the same range, between 0.106 and 0.136 cd m^{-2} , while the threshold for the brightest level (469 lux) was considerably higher, 0.825 cd m^{-2} (figure 3a). We assume that the luminance of the background at the dimmest levels was too dim to be detected by the bird, hence the similar results for these

conditions. The lower luminance detection thresholds in the dimmer light levels suggest that the birds have a fast mechanism allowing them to adapt to these low levels.

In experiment 2, we determined the luminance discrimination thresholds for two bright targets on a dark background. The thresholds were in the same range at all four different light levels, with Weber fractions between 0.41 (level 4) and 0.54 (level 3; figure 3c). Without any adaptation we would have expected a gradual rise in Weber fraction with decreasing illuminance in the decision box [48]. In our data no such trend could be seen, indicating the existence of a fast luminance adaptation mechanism.

We cannot determine the mechanisms underlying the high sensitivity observed in the dim light conditions, but we can conclude that they function within about 1 s. As mentioned in the introduction, pupil dynamics probably play a minor role in this context [3,4]. Only few birds are capable of remarkable changes in pupil size. The king penguin (*Aptenodytes patagonicus*) constricts its pupils to tiny square-shaped pinholes in daylight allowing the retina to stay dark-adapted before diving to foraging grounds several hundred metres below the sea surface [49]. Dilation of the pupils to its maximum size under water increases retinal image illumination 300-fold. No such extreme pupil dynamics have been reported in any terrestrial bird species. Budgerigars are able to dilate their pupils from 2.3 mm to 3 mm, allowing their eyes to let in 1.7 times more light [4]. This can only account for a small fraction of the sensitivity increase seen between levels 1 and 2, in our experiments. Avian pupil constriction can happen within the tenth of a second, an ability attributed to the presence of striated muscle [3,50,51]. Pupil dilation, by contrast, requires several seconds to be completed [51,52]. The birds in our experiments took only around 1 s to locate the correct stimuli, clearly excluding pupil dynamics as the main mechanism of sensitivity increase.

In birds, fast visual adaptation has only been studied in the context of colour constancy. A study on chickens [53] indicates the presence of fast (or simultaneous) and slower (up to 5 min) adaptational mechanisms in the chromatic pathway. Primates have fast mechanisms which adjust retinal luminance sensitivity within less than a second [20,54], enabling retinal adaptation to the highly dynamic luminance variation experienced when actively exploring a visual scene [18,55]. Birds have a similar rate of gaze change as humans when scanning the environment [56] and thus probably need similarly rapid luminance adaptation.

We are not sure why, in both experiments, lower thresholds were found at light levels 2 and 4 than at the intermediate level 3 (figure 3a,c). Previous experiments have indicated that the single cones of budgerigars lose sensitivity at an illuminance between 2 and 10 lux, but double cones remain active at lower intensities ([7], and O. Lind 2013, unpublished data). This shift, happening between light levels 2 and 3 might explain at least part of the sensitivity drop seen at level 3.

In a previous study on brightness discrimination, budgerigars had a Weber fraction of 0.18 when tested with two spatially separated large achromatic fields [35]. In our measurements, we found considerably higher thresholds, a Weber fraction between 0.41 and 0.54, a difference probably resulting from the short adaptation period.

The median response time of the budgerigars was 1.17 s in experiment 1 and 0.98 s in experiment 2. Whether the choice was part of a successful or unsuccessful trial unit, had the largest effect on response time. Our expectation

had been that longer response times would reflect longer adaptation periods, and thus, correlate with successful trials. In both experiments, birds had a shorter response time in successful trials, but the differences were too small (within the range of 10^{-2} s) to allow conclusions about effects on adaptation. Shorter response times for correct than for incorrect choices have previously been found in studies of optimal decision making [57,58] and thus, are more likely a consequence of decision making than of adaptation. In primates the initial steep sensitivity rise (within 1 s) is followed by a slower adaptive change [9]. If the time course of early adaptation is similar in birds, then fast decision taking is probably an efficient behaviour, as feeding parents commute to the nest many times every day, and long decision times will come with a high cost [57]. The fast adaptation mechanisms are incomplete but sufficient to allow birds to see well enough in the nest.

Unlike many passerine chicks, budgerigar chicks do not beg for food with a wide-open gape with conspicuous flanges. Budgerigars are unable to lift the head until they are 6–8 days old and although they are able to vocalize, active begging behaviour (e.g. head-bobbing, moving towards the parent) are uncommon until they are around 11 days old [59,60]. During this period the parents initiate feeding events by beak-grasping [59]. Like other psittacine birds, budgerigars have specialized touch-receptors, referred to as the ‘bill-tip organ’, in the upper bill [61,62]. The bill-tip organ is used in object exploration and manipulation [61,62] and it is possible that budgerigars use tactile stimuli to a larger extent than visual stimuli in parent-offspring communication. Nevertheless, budgerigar chicks are individually targeted during feeding suggesting visual detection is involved [59]. Furthermore, the eggs and chicks of a domesticated budgerigar nest have a Weber contrast of about 0.4–0.6 to a nest background made of wood chips (electronic supplementary material, figure S1 and table S8). The typical substrate on which wild budgerigars lay their eggs consists of decaying wood and faeces [63] which probably provides an even higher contrast. Our study suggests that budgerigars adapt to the strong drop in light intensity, equivalent to that experienced when entering the nest, within less than 1 s. Their sensitivity then allows them to fast and efficiently feed the chicks using visual control.

Ethics. All experiments followed Swedish legislation, under the permit dnr. 5.8.18-17189/2018 granted by the responsible authority (Malmö – Lunds djurförsöksetiska nämnd).

Data accessibility. All data used in this manuscript are available from the Dryad Digital Repository: <https://doi.org/10.5061/dryad.qz612jmm8> [64].

The data are also provided in the electronic supplementary material [65].

Authors' contributions. S.C.: conceptualization, data curation, formal analysis, investigation, methodology, software, validation, visualization, writing—original draft, writing—review and editing; O.L.: conceptualization, methodology, supervision, writing—review and editing; A.K.: conceptualization, funding acquisition, methodology, project administration, resources, supervision, writing—review and editing.

All authors gave final approval for publication and agreed to be held accountable for the work performed therein.

Conflict of interest declaration. We declare we have no competing interests.

Funding. The study was funded by the Swedish Research Council (2016-03298_3).

Acknowledgements. We want to thank Camilla Aagaard at Fågelhuset in Kvärlöv for having us visit and photograph her birds. We are also grateful for the constructive feedback provided by two anonymous reviewers, which helped improve the manuscript.

References

- Martin GR. 2017 *The sensory ecology of birds*. 1st edn. Oxford, UK: Oxford University Press.
- Land MF, Nilsson D-E. 2012 *Animal eyes*, 2nd edn. Oxford, UK: Oxford University Press.
- Douglas RH. 2018 The pupillary light responses of animals; a review of their distribution, dynamics, mechanisms and functions. *Prog. Retin. Eye Res.* **66**, 17–48. (doi:10.1016/j.preteyeres.2018.04.005)
- Lind OE, Kelber A, Kröger RH. 2008 Multifocal optical systems and pupil dynamics in birds. *J. Exp. Biol.* **211**, 2752–2758. (doi:10.1242/jeb.018630)
- Walls GL. 1942 *The vertebrate eye and its adaptive radiation*. Bloomfield Hills, MI: Cranbrook Institute of Science.
- Hart NS. 2001 The visual ecology of avian photoreceptors. *Prog. Retin. Eye Res.* **20**, 675–703. (doi:10.1016/s1350-9462(01)00009-x)
- Lind O, Chavez J, Kelber A. 2014 The contribution of single and double cones to spectral sensitivity in budgerigars during changing light conditions. *J. Comp. Physiol. A* **200**, 197–207. (doi:10.1007/s00359-013-0878-7)
- Dunn FA, Lankheet MJ, Rieke F. 2007 Light adaptation in cone vision involves switching between receptor and post-receptor sites. *Nature* **449**, 603–606. (doi:10.1038/nature06150)
- Shapley R, Enroth-Cugell C. 1984 Visual adaptation and retinal gain controls. *Prog. Retin. Res.* **3**, 263–346. (doi:10.1016/0278-4327(84)90011-7)
- Pertman I, Normann RA. 1998 Light adaptation and sensitivity controlling mechanisms in vertebrate photoreceptors. *Prog. Retin. Eye Res.* **17**, 523–563. (doi:10.1016/s1350-9462(98)00005-6)
- Barlow HB, Fitzhugh R, Kuffler SW. 1957 Change of organization in the receptive fields of the cat's retina during dark adaptation. *J. Physiol.* **137**, 338–354. (doi:10.1113/jphysiol.1957.sp005817)
- Burkhardt D. 1994 Light adaptation and photopigment bleaching in cone photoreceptors *in situ* in the retina of the turtle. *J. Neurosci.* **14**, 1091–1105. (doi:10.1523/jneurosci.14-03-01091.1994)
- Lamb TD, Pugh EN. 2004 Dark adaptation and the retinoid cycle of vision. *Prog. Retin. Eye Res.* **23**, 307–380. (doi:10.1016/j.preteyeres.2004.03.001)
- Hecht S, Haig C, Chase AM. 1937 The influence of light adaptation on subsequent dark adaptation of the eye. *J. Gen. Physiol.* **20**, 831–850. (doi:10.1085/jgp.20.6.831)
- Blough DS. 1956 Dark adaptation in the pigeon. *J. Comp. Physiol. Psychol.* **49**, 425–430. (doi:10.1037/h0043257)
- Hersloff L, Lehner PN, Bolen EG, Rylander MK. 1974 Visual sensitivity in the black-bellied duck (*Dendrocygna autumnalis*), a crepuscular species. *J. Comp. Physiol. Psychol.* **86**, 486–492. (doi:10.1037/h0036177)
- Wells MC, Lehner PN, Bolen EG, Rylander MK. 1975 Comparison of scotopic sensitivity in diurnal (*Anas platyrhynchos*) and crepuscular (*Dendrocygna autumnalis*) ducks. *J. Comp. Physiol. Psychol.* **88**, 940–944. (doi:10.1037/h0076444)
- Rieke F, Rudd ME. 2009 The challenges natural images pose for visual adaptation. *Neuron* **64**, 605–616. (doi:10.1016/j.neuron.2009.11.028)
- Dunn FA, Rieke F. 2008 Single-photon absorptions evoke synaptic depression in the retina to extend the operational range of rod vision. *Neuron* **57**, 894–904. (doi:10.1016/j.neuron.2008.01.031)
- Hayhoe MM, Benimoff NI, Hood DC. 1987 The time-course of multiplicative and subtractive adaptation process. *Vision Res.* **27**, 1981–1996. (doi:10.1016/0042-6989(87)90062-9)
- Maziarz M, Wesolowski T. 2014 Does darkness limit the use of tree cavities for nesting by birds? *J. Ornithol.* **155**, 793–799. (doi:10.1007/s10336-014-1069-1)
- Reynolds SJ, Martin GR, Cassey P. 2009 Is sexual selection blurring the functional significance of eggshell coloration hypotheses? *Anim. Behav.* **78**, 209–215. (doi:10.1016/j.anbehav.2009.03.003)
- Heeb P, Schwander T, Faoro S. 2003 Nestling detectability affects parental feeding preferences in a cavity-nesting bird. *Anim. Behav.* **66**, 637–642. (doi:10.1006/anbe.2003.2238)
- Wiebe KL, Slagsvold T. 2009 Mouth coloration in nesting birds: increasing detection or signalling quality? *Anim. Behav.* **78**, 1413–1420. (doi:10.1016/j.anbehav.2009.09.013)
- Wiebe KL, Slagsvold T. 2012 Parents take both size and conspicuousness into account when feeding nestlings in dark cavity nests. *Anim. Behav.* **84**, 1307–1312. (doi:10.1016/j.anbehav.2012.09.014)
- Podkova P, Malinowska K, Surmacki A. 2019 Light affects parental provisioning behaviour in a cavity-nesting passerine. *J. Avian Biol.* **50**, e02254. (doi:10.1111/jav.02254)
- Dugas MB. 2015 Detectability matters: conspicuous nestling mouth colours make prey transfer easier for parents in a cavity nesting bird. *Biol. Lett.* **11**, 20150771. (doi:10.1098/rsbl.2015.0771)
- Podkova P, Surmacki A. 2017 The importance of illumination in nest site choice and nest characteristics of cavity nesting birds. *Sci. Rep.* **7**, 1329. (doi:10.1038/s41598-017-01430-y)
- Wesolowski T, Maziarz M. 2012 Dark tree cavities - a challenge for hole nesting birds? *J. Avian Biol.* **43**, 454–460. (doi:10.1111/j.1600-048X.2012.05704.x)
- Holveck MJ, Grégoire A, Doutrelant C, Lambrechts MM. 2019 Nest height is affected by lampost lighting proximity in addition to nestbox size in urban great tits. *J. Avian Biol.* **50**, e01798. (doi:10.1111/jav.01798)
- Manna TJ, Hanley D, Honza M, Capek M, Rutila J, Samaš P, Abolins-Abols M, Hauber ME. 2020 Fitting different visual models to behavioral patterns of parasitic egg rejection along a natural egg color gradient in a cavity-nesting host species. *Vis. Res.* **167**, 54–59. (doi:10.1016/j.visres.2019.12.007)
- Soler JJ, Navarro C, Contreras TP, Avilés JM, Cuervo JJ. 2008 Sexually selected egg coloration in spotless starlings. *Am. Nat.* **171**, 183–194. (doi:10.1086/524958)
- Wyndham E. 1980 Diurnal cycle, behavior and social organization of the budgerigar *Melopsittacus undulatus*. *Emu* **80**, 25–33. (doi:10.1071/mu9800025)
- Lind O, Kelber A. 2011 The spatial tuning of achromatic and chromatic vision in budgerigars. *J. Vision* **11**, 2. (doi:10.1167/11.7.2)
- Lind O, Karlsson S, Kelber A. 2013 Brightness discrimination in budgerigars (*Melopsittacus undulatus*). *PLoS ONE* **8**, e54650. (doi:10.1371/journal.pone.0054650)
- Haller NK, Lind O, Steinlechner S, Kelber A. 2014 Stimulus motion improves spatial contrast sensitivity in budgerigars (*Melopsittacus undulatus*). *Vision Res.* **102**, 19–25. (doi:10.1016/j.visres.2014.07.007)
- Chaib S, Ljungholm M, Lind O, Kelber A. 2019 Single target acuity is not higher than grating acuity in a bird, the budgerigar. *Vision Res.* **160**, 37–42. (doi:10.1016/j.visres.2019.04.005)
- Chaib S, Garivaghi Mussoi J, Lind O, Kelber A. 2021 Visual acuity of budgerigars for moving targets. *Biol. Open* **10**, bio058796. (doi:10.1242/bio.058796)
- Lind O, Sunesson T, Mitkus M, Kelber A. 2012 Luminance-dependence of spatial vision in budgerigars (*Melopsittacus undulatus*) and Bourke's parrots (*Neophaps bourkii*). *J. Comp. Physiol. A* **198**, 69–77. (doi:10.1007/s00359-011-0689-7)
- Lind O, Kelber A. 2009 The intensity threshold of colour vision in two species of parrot. *J. Exp. Biol.* **212**, 3693–3699. (doi:10.1242/jeb.035477)
- Chavez J, Kelber A, Vorobyev M, Lind O. 2014 Unexpectedly low UV-sensitivity in a bird, the budgerigar. *Biol. Lett.* **10**, 20140670. (doi:10.1098/rsbl.2014.0670)
- Goldsmith TH, Butler BK. 2005 Color vision of the budgerigar (*Melopsittacus undulatus*): hue matches, tetrachromacy, and intensity discrimination. *J. Comp. Physiol. A* **191**, 933–951. (doi:10.1007/s00359-005-0024-2)
- Fellows BJ. 1967 Chance stimulus sequences for discrimination tasks. *Psychol. Bull.* **67**, 87–92. (doi:10.1037/h0024098)
- Levitt H. 1971 Transformed up-down methods in psychoacoustics. *J. Acoust. Soc. Am.* **49**, 467–477. (doi:10.1121/1.1912375)
- Kuznetsova A, Brockhoff PB, Christensen RHB. 2017 lmerTest package: tests in linear mixed effects models. *J. Stat. Softw.* **82**, 1–26. (doi:10.18637/jss.v082.i13)
- R Core Team. 2021 *RStudio: integrated development for R*. Boston, MA: RStudio, Inc.
- Hothorn T, Bretz F, Westfall P. 2008 Simultaneous inference in general parametric models. *Biom. J.* **50**, 346–363. (doi:10.1002/bimj.200810425)
- Craik KJW. 1938 The effect of adaptation on differential brightness discrimination. *J. Physiol.* **92**, 406–421. (doi:10.1113/jphysiol.1938.sp003612)
- Martin GR. 1999 Eye structure and foraging in king penguins *Aptenodytes patagonicus*. *Ibis* **141**, 444–450. (doi:10.1111/j.1474-919X.1999.tb04413.x)
- Barbur JL, Prescott NB, Douglas RH, Jarvis JR, Wathes CM. 2002 A comparative study of stimulus-

- specific pupil responses in the domestic fowl (*Gallus gallus domesticus*) and the human. *Vision Res.* **42**, 249–255. (doi:10.1016/s0042-6989(01)00279-6)
51. Oliphant LW, Johnson MR, Murphy C, Howland H. 1983 The musculature and pupillary response of the great horned owl iris. *Exp. Eye Res.* **37**, 583–595. (doi:10.1016/0014-4835(83)90134-3)
 52. Moayed AA, Choh V, Hariri S, Liu C, Wong A, Bizheva K. 2012 Stimulus-specific pupil dynamics measured in birds (*Gallus gallus domesticus*) *in vivo* with ultrahigh resolution optical coherence tomography. *Investig. Ophthalmol. Vis. Sci.* **53**, 6863–6869. (doi:10.1167/iovs.12-10291)
 53. Olsson P, Wilby D, Kelber A. 2016 Quantitative studies of animal colour constancy: using the chicken as model. *Proc. R. Soc. B* **283**, 20160411. (doi:10.1098/rspb.2016.0411)
 54. Geisler WS. 1983 Mechanisms of visual sensitivity: backgrounds and early dark adaptation. *Vision Res.* **23**, 1423–1432. (doi:10.1016/0042-6989(83)90154-2)
 55. Frazor RA, Geisler WS. 2006 Local luminance and contrast in natural images. *Vision Res.* **46**, 1585–1598. (doi:10.1016/j.visres.2005.06.038)
 56. Land M. 2019 Eye movements in man and other animals. *Vision Res.* **162**, 1–7. (doi:10.1016/j.visres.2019.06.004)
 57. Drugowitsch J, Moreno-Bote RN, Pouget A, Churchland AK, Shadlen MN. 2012 The cost of accumulating evidence in perceptual decision making. *J. Neurosci.* **32**, 3612–3628. (doi:10.1523/JNEUROSCI.4010-11.2012)
 58. Palmer J, Huk AC, Shadlen MN. 2005 The effect of stimulus strength on the speed and accuracy of a perceptual decision. *J. Vision* **5**, 376–404. (doi:10.1167/5.5.1)
 59. Stamps J, Clark A, Arrowood P, Kus B. 1985 Parent-offspring conflict in budgerigars. *Behaviour* **94**, 1–40. (doi:10.1163/156853985X00253)
 60. Brittan-Powell EF, Dooling RJ, Farabaugh SM. 1997 Vocal development in budgerigars (*Melopsittacus undulatus*): contact calls. *J. Comp. Psychol.* **111**, 226–241. (doi:10.1037/0735-7036.111.3.226)
 61. Demery ZP, Chapelle J, Martin GR. 2011 Vision, touch and object manipulation in Senegal parrots *Poicephalus senegalus*. *Proc. R. Soc. B* **278**, 3687–3693. (doi:10.1098/rspb.2011.0374)
 62. Martin GR, Martin RO. 2021 Psittaciformes sensory systems. In *Encyclopedia of animal cognition and behavior* (eds J Vonk, T Shackelford), pp. 5767–5782. Cham, Switzerland: Springer.
 63. Wyndham E. 1981 Breeding and mortality of budgerigars *Melopsittacus undulatus*. *Emu* **81**, 240–243. (doi:10.1071/MU9810240)
 64. Chaib S, Olle L, Almut K. 2023 Data from: Fast visual adaptation to dim light in a cavity-nesting bird. Dryad Digital Repository. (doi:10.5061/dryad.qz612jmm8)
 65. Chaib S, Olle L, Almut K. 2023 Fast visual adaptation to dim light in a cavity-nesting bird. Figshare. (doi:10.6084/m9.figshare.c.6619772)

Supplementary material to:

Fast visual adaptation to dim light in a cavity-nesting bird

Chaib, S., Lind, O., and Kelber, A.
Proceedings of the Royal Society B
DOI: 10.1098/rspb.2023.0596

Experimental conditions

Table S1. Light conditions at the different light levels.

Level	ND filter ⁽¹⁾	Illuminance (lux)	Relative illuminance change ⁽²⁾	Background luminance (cd/m ²)	Luminance of positive stimulus in Exp. 2 (cd/m ²)
1	none	469	-0.48	2.25×10^{-1}	200
2	1.2	28.0	-1.68	1.8×10^{-2}	10.4
3	2.4	1.83	-2.88	8.0×10^{-4}	0.675
4	3	0.47	-3.48	2.37×10^{-4}	0.190

⁽¹⁾The optical density of the neutral-density (ND) filter used at the light level.
⁽²⁾The relative change in illuminance between the waiting perch (1400 lux) and the decision box, in log₁₀ units.

Table S2. Target luminance at the staircase steps used in Experiment 1. Units are given in cd/m².

Level 1	Level 2	Level 3	Level 4
63.5	7.98	0.850	0.248
48.7	6.35	0.675	0.219
38.5	4.60	0.526	0.190
28.0	3.27	0.415	0.170
18.8	2.64	0.300	0.150
13.5	2.00	0.257	0.144
7.10	1.49	0.214	0.138
3.62	0.973	0.198	0.132
2.61	0.677	0.191	0.129
1.77	0.380	0.183	0.126

1.26	0.319	0.175	0.123
1.16	0.259	0.167	0.120
1.05	0.229	0.159	0.117
0.950	0.198	0.152	0.114
0.846	0.168	0.144	0.111
0.743	0.138	0.136	0.108
0.639	0.107	0.129	0.105
0.536	0.077	0.123	0.102
0.432	0.071	0.116	0.099
	0.065	0.110	0.096
		0.103	0.093
			0.090
			0.088
			0.085
			0.083
			0.080
			0.078
			0.076
			0.073
			0.071
			0.068
			0.066

Table S3. Contrast between the targets of the positive and negative stimuli at the different staircase steps used in Experiment 2. Units are given in Weber contrast.

Level 1	Level 2	Level 3	Level 4
0.91	0.91	0.96	0.97
0.86	0.86	0.93	0.92
0.81	0.81	0.90	0.88
0.76	0.75	0.88	0.86
0.68	0.69	0.86	0.84
0.66	0.62	0.84	0.82
0.63	0.60	0.82	0.79
0.60	0.57	0.80	0.77
0.58	0.56	0.78	0.75
0.55	0.54	0.76	0.74
0.53	0.52	0.75	0.73
0.52	0.51	0.74	0.71
0.50	0.49	0.73	0.70

0.48	0.47	0.72	0.69
0.47	0.46	0.71	0.68
0.45	0.44	0.69	0.66
0.43	0.42	0.68	0.65
0.42	0.41	0.67	0.64
0.40	0.39	0.66	0.63
0.39	0.37	0.64	0.61
0.37	0.36	0.63	0.60
0.35		0.62	0.59
0.33		0.61	0.58
0.32		0.59	0.56
0.30		0.58	0.55
0.28		0.57	0.54
0.27		0.56	0.53
0.25		0.54	0.51
0.23		0.52	0.49
		0.50	0.48
		0.49	0.46
		0.47	0.45
		0.45	0.43
		0.44	0.42
		0.42	0.40
		0.40	0.38
		0.39	0.37
		0.37	0.35
		0.35	0.34
		0.34	0.32
		0.32	0.31
		0.30	0.29
		0.29	
		0.27	
		0.25	
		0.24	
		0.22	
		0.20	

Statistical analyses

Table S4. Fixed effects of luminance detection thresholds in Experiment 1. Thresholds were log-transformed before analysis.

Fixed effects	Estimate	Std. error	df	t-value	p
Level 4 (intercept)	-2.24533	0.07534	19.08516	-29.802	<0.001
Level 3	0.24775	0.09341	44.00000	2.652	<0.05
Level 2	0.03751	0.09341	44.00000	0.402	0.6899
Level 1	2.05039	0.09341	44.00000	21.950	<0.001

Table S5. Estimates of response time in Experiment 1. Response times were inversed before analysis.

	Fixed effects	Estimate	Std. error	df	t-value	p
Unsuccessful trial unit	Level 4 (intercept)	0.88971	0.05327	10.41653	16.703	<0.001
	Level 3	-0.06868	0.04830	955.07800	-1.422	0.15538
	Level 2	-0.13336	0.04896	955.32544	-2.724	<0.01
	Level 1	-0.16112	0.05070	955.24513	-3.178	<0.01
Successful trial unit	Level 4	0.02361	0.04224	955.18841	0.559	0.57633
	Level 3	0.05808	0.05957	955.10949	0.975	0.32985
	Level 2	0.10215	0.06000	955.48233	1.703	0.08898
	Level 1	0.16567	0.06112	955.31525	2.711	<0.01

Table S6. Fixed effects of luminance discrimination thresholds in Experiment 2.

Fixed effects	Estimate	Std. error	df	t-value	p
Level 4 (intercept)	0.41035	0.05117	14.20173	8.020	<0.001
Level 3	0.12690	0.04820	36.83606	2.633	<0.05
Level 2	0.00919	0.04743	37.32644	0.194	0.8474
Level 1	0.03615	0.04743	37.32644	0.762	0.4507

Table S7. Estimates of response time in Experiment 2. Response times were inversed before analysis.

	Fixed effects	Estimate	Std. error	df	t-value	p
Unsuccessful trial unit	Level 4 (intercept)	0.923	0.069	11.720	13.398	<0.001
	Level 3	-0.024	0.054	796.76	-0.452	0.651
	Level 2	0.130	0.053	796.830	2.465	<0.05
	Level 1	0.046	0.053	796.722	0.872	0.383
Successful trial unit	Level 4	0.080	0.051	795.225	1.561	0.119
	Level 3	0.039	0.065	795.242	0.598	0.550
	Level 2	-0.014	0.063	795.191	-0.215	0.829
	Level 1	0.125	0.063	795.287	1.990	<0.05
	Inter trial time	-0.0049	0.0014	798.935	-3.420	<0.001

Measurements of luminance contrasts from a domestic budgerigar nest



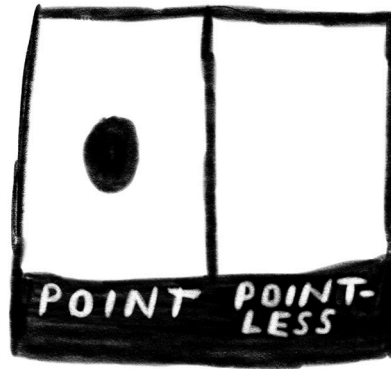
Figure S1. Domestic budgerigar nest. The chicks were all hatched at different occasions, and the smallest one (a) just a few days before the photograph was taken. We measured the mean pixel intensity at the cere of the two youngest chicks (a, b) and at an unhatched egg (c). The dashed line surrounds the area at which we measured the mean pixel intensity of the nest. The pixel intensity was measured as weighted RGB colours using ImageJ (Schneider et al. 2012). The photograph was taken in Olympus Raw Format (ORF) to ensure the contrasts in the image to be as accurate as possible. The image was transformed to a Tagged Image File Format (TIFF, no compression) to be able to import it to the ImageJ software.

Table S8. Luminance contrast in a domestic budgerigar nest.

	Mean pixel intensity	St Dev	Min	Max	Weber contrast to the nest background
Cere, small chick	47 072.620	4 303.325	22 779	57 604	0.53314
Cere, large chick	43 527.445	4 493.433	17 852	52 029	0.41767
Egg	48 395.001	6 221.896	21 740	57 934	0.57621
Nest	30 703.472	12 348.757	0	61 311	0

Reference

Schneider, C. A., Rasband, W. S., & Eliceiri, K. W. (2012) NIH Image to ImageJ: 25 years of image analysis. *Nat. Methods*, **9**, 671–675. (doi:10.1038/nmeth.2089)



List of papers

Chaib, S., Ljungholm, M., Lind, O., & Kelber, A. (2019). Single target acuity is not higher than grating acuity in a bird, the budgerigar. *Vision Research*, 160, 37–42. doi: 10.1016/j.visres.2019.04.005

Chaib, S., Mussoi, J. G., Lind, O., & Kelber, A. (2021). Visual acuity of budgerigars for moving targets. *Biology Open*, 10(9), Article bio058796. doi: 10.1242/bio.058796

Chaib, S., Lind, O., & Kelber, A. (2023) Fast visual adaptation to dim light in a cavity-nesting bird. *Proceedings of the Royal Society B*. 290(1998), Article 20230596. doi: 10.1098/rspb.2023.0596