



# LUND UNIVERSITY

## Lymphocytes in atherosclerosis

Wigren, Maria; Nilsson, Jan; Kolbus, Daniel

*Published in:*  
Clinica Chimica Acta

*DOI:*  
[10.1016/j.cca.2012.04.031](https://doi.org/10.1016/j.cca.2012.04.031)

2012

[Link to publication](#)

*Citation for published version (APA):*  
Wigren, M., Nilsson, J., & Kolbus, D. (2012). Lymphocytes in atherosclerosis. *Clinica Chimica Acta*, 413(19-20), 1562-1568. <https://doi.org/10.1016/j.cca.2012.04.031>

*Total number of authors:*  
3

### General rights

Unless other specific re-use rights are stated the following general rights apply:  
Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

Read more about Creative commons licenses: <https://creativecommons.org/licenses/>

### Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

LUND UNIVERSITY

PO Box 117  
221 00 Lund  
+46 46-222 00 00

## **Lymphocytes in atherosclerosis**

Maria Wigren, Jan Nilsson and Daniel Kolbus

Department of Clinical Sciences Malmö, Lund University, Sweden

Correspondence to:

Jan Nilsson, Department of Clinical Sciences, Entrance 72;91:12, Malmö University Hospital,  
20502 Malmö, Sweden, Phone: +4640391230, Fax: +4640391212, Email:

[jan.nilsson@med.lu.se](mailto:jan.nilsson@med.lu.se)

## **Abstract**

It is well established that atherosclerosis is caused by an inflammatory process in the arterial intima. However, it is only **in** recent years that it has become clear that this inflammation is modulated by immune responses against plaque antigens. These antigens are primarily believed to be modified self-antigens such as oxidized LDL. The immune system **is challenged** to determine whether these antigens should be regarded self and tolerated or non-self and eliminated. The latter will result in plaque development while the first will be protective. T cells are key effectors of both types of responses. An activation of regulatory T cells inhibits auto-reactive T effector cells and is anti-inflammatory. In contrast, if Th1 cells become activated in the plaque this is associated with increased inflammation and disease progression. The role of B cells in atherosclerosis remains to be clarified but some species of athero-protective antibodies have been identified. The elucidation of role of immune system in atherosclerosis has revealed new targets for intervention and both vaccines and antibody-based therapies are presently in or due to enter clinical testing.

## **Highlights**

◆ Atherosclerosis involves an inflammatory process in the arterial intima ◆ Immune responses against plaque antigens modulate plaque inflammation ◆ Regulatory T cells inhibits auto reactive T effector cells and inflammation ◆ Th1 cells are pro-inflammatory and promote plaque progression ◆ Novel therapies aim to mimic or selectively activate athero-protective immune responses

## **1. Introduction**

**The role of inflammation in the development of atherosclerosis has been a well-established fact for more than a century.** Originally this inflammation was solely regarded as a result of accumulation of toxic lipids, but recent research has revealed that it is regulated by a complex pattern of immune responses. These immune responses may drive disease progression by activating pro-inflammatory Th1 and NKT cells against self-antigens in the plaque but can also be protective by inducing anti-inflammatory regulatory T cells. It is likely that the latter dominate in pre-stages of the disease and help to clear away damaged cells and lipoproteins from the vascular wall before causing further tissue injury. As the disease advances the immune responses (presumably against the same self-antigens) shift towards Th1 and exuberates inflammation. According to this view the interactions between antigen presenting cells (APC) and T cells that determine if immune responses against self-antigens in the plaque will be tolerogenic or pro-inflammatory play a key role in the disease process. Consequently, there has been an increasing focus on the role of T cells in atherosclerosis. Increased lymphocyte number is well documented as an independent risk factor for cardiovascular disease (1, 2). Interestingly, decreased lymphocyte numbers also confer increased risk (3), further underlining their complex role of the immune system in atherosclerosis. This review will summarize our current understanding of the role of different lymphocytes in atherosclerosis and briefly discuss the possibility of developing novel treatments for cardiovascular disease by targeting these cells. The role of the immune system is summarized in the figure.

## **2. CD4<sup>+</sup> T cells**

CD4<sup>+</sup> T cells, also named T helper (Th) cells, are believed to play an important role in atherosclerosis. Initial evidence of their atherogenic potential came from studies with

hypercholesterolemic severe immune-deficient (SCID) mice that were injected with CD4<sup>+</sup> T cells resulting in increased atherosclerosis (4). Moreover, Zhou and co-workers reported that CD4<sup>+</sup> T cell-deficient *ApoE*<sup>-/-</sup> mice have reduced lesions in the aortic root (5). In contrast, Elhage and co-workers, found no difference in aortic root lesions but increased atherosclerosis in *en face* preparations of the total aorta in the same type mice (6) indicating that CD4<sup>+</sup> T cells may have site-specific actions. CD4<sup>+</sup> T cells are a heterogeneous group of cells that include both pro- and anti-inflammatory cells. IL-12 and IL-18 from antigen presenting cells promote naïve CD4<sup>+</sup> T cells to express the Th1-specific transcription factor T-box expressed in T cells (Tbet) along with the signal transducer and activator of transcription 4 (STAT4) leading to Th1 cell differentiation. T-bet deficiency reduces atherosclerosis in *Lldr*<sup>-/-</sup> mice indicating that Th1 cells are pro-atherogenic (7). Th1 cells produce pro-inflammatory cytokines such as IFN- $\gamma$  and TNF- $\alpha$ . Hypercholesterolemic mice lacking IFN- $\gamma$  or TNF- $\alpha$  are characterized by reduced atherosclerosis providing further support of a pro-atherogenic role of Th1 cells (8, 9). Th2 cells represent a lineage of CD4<sup>+</sup> T that primarily interact with B cells. The role of Th2 cells in atherosclerosis appears to be more complex than that of Th1 cells and both pro-and anti-atherogenic actions have been reported. Th2 cell differentiation is initiated by the Th2 cell signature cytokine IL-4. This cytokine prevents production of IFN- $\gamma$  leading to inhibition of Th1 cell differentiation. The role of IL-4 and Th2 cells in experimental atherosclerosis remains to be fully elucidated and hypercholesterolemic mice deficient in IL-4 have been reported to have reduced as well as unaltered atherosclerosis (10, 11). Activated Th2 cells typically also produce the cytokines IL-5, IL-9, IL-13 and IL-25 (12-14). IL-5 has a protective effect on atherosclerosis in hypercholesterolemic mice, a phenomena that has been attributed to the ability of IL-5 to stimulate the synthesis of so called natural antibodies from B cells of the B1 type (15, 16). These antibodies are IgM that bind to phospholipid-epitopes in oxLDL and prevent scavenger

receptor-mediated uptake of oxLDL in macrophages (15). The roles of IL-9, IL-13 and IL-25 in atherosclerosis development are poorly studied and remain to be fully elucidated. IL-33 is another cytokine with the potential to induce Th2 responses and IL-33 treatment has been shown to reduce atherosclerosis development in *ApoE*<sup>-/-</sup> mice (17).

### **3. CD4<sup>+</sup> regulatory T cells**

Regulatory T cells (Tregs) are a heterogeneous group of immune-inhibitory cells that suppress pathogenic immune responses primarily to self-antigens, but also against foreign antigens such as allergens and dietary antigens (18). Regulatory T cells are found in very low numbers in atherosclerotic plaques as compared to other chronically inflamed tissues (19, 20). This suggests that local tolerance is impaired in the plaques which could contribute to increased arterial inflammation. A protective role of regulatory T cells have been demonstrated in hypercholesterolemic mice deficient in co-stimulatory molecules important for regulatory T cell development as well as in mice which have been depleted of regulatory T cells through treatment with anti-CD25 antibody (22, 23). The athero-protective role of regulatory T cells is further substantiated by experiments in which regulatory T cells were eliminated in hypercholesterolemic mice through immunization with Foxp3-transfected dendritic cells, leading to regulatory T cell apoptosis and increased atherosclerosis (24). The anti-inflammatory cytokines IL-10, TGF- $\beta$  and the more recently discovered IL-35 are mediators of regulatory T cell suppression. IL-10 and TGF- $\beta$  have been shown to have anti-atherogenic effects in experimental atherosclerosis (25) (26) (27) (28). The translation of these experimental findings into clinical studies has been hampered by the difference in phenotypic definition of regulatory T cells in mice and humans. Whereas CD4<sup>+</sup>CD25<sup>+</sup> T cells in mice almost exclusively are functional suppressor cells (29) this phenotype is not always associated with suppressor function in humans (30) complicating characterization of human Tregs.

Recently, Schuler *et al.* reported that the ATP converting enzyme CD39 was present on CD4<sup>+</sup>CD25<sup>+</sup>FoxP3<sup>+</sup> T cells with suppressor function distinguishing these from the CD4<sup>+</sup>CD39<sup>+</sup>CD25<sup>-</sup>FoxP3<sup>-</sup> cells with no suppressor function (31). Since nearly all CD39<sup>+</sup>CD25<sup>+</sup> T cells were FoxP3<sup>+</sup>, this constitutes a promising marker of functional suppressor cells in humans. Studies evaluating the clinical significance of Tregs in atherosclerosis are few and report conflicting results. Decreased levels of circulating Tregs have been reported in patients with acute coronary syndrome (32). However, a study by Ammirati *et al* showed that circulating Tregs were not correlated with carotid intima media thickness (33). Taken together these data show that more studies on human Tregs and atherosclerosis needs to be performed before it can be concluded whether Tregs can be used as a clinical marker of cardiovascular disease severity and/or risk.

#### **4. CD8<sup>+</sup> T cells**

The impact of CD8<sup>+</sup> T cells on atherosclerosis development is less well characterized than that of CD4<sup>+</sup> T cells. Whereas MHC class I deficient C57Bl/6 mice on high-fat diet develop increased atherosclerosis (34), CD8<sup>+</sup> T cell-deficient *ApoE*<sup>-/-</sup> mice have similar lesion size as CD8-competent *ApoE*<sup>-/-</sup> mice (6). A prerequisite for specific involvement in the disease process is that the CD8<sup>+</sup> T cells are present in atherosclerotic lesions. Indeed, CD8<sup>+</sup> T cells are found together with CD4<sup>+</sup> T cells in lesions of both mice (35) and humans (36). In advanced human lesions they even appear to be the predominating T cell type (36). A possible pathological mechanism of CD8<sup>+</sup> T cells is CTL killing of vascular cells which in turn would exacerbates inflammation. Evidence for involvement of this mechanism was reported by Ludwig *et al.* in a mouse model expressing beta-galactosidase specifically in vascular smooth muscle cells. Upon injection of beta-galactosidase specific DCs the mice developed arteritis and atherosclerosis mediated by CD8<sup>+</sup> T cells (37). More recently Escalante *et al.*

showed that vascular cells can attenuate CD8<sup>+</sup> effector functions (38). Human vascular ECs were shown to up-regulate CD155 in response to IFN- $\gamma$  stimulation which in turn inhibited cytotoxic effects of activated CD8<sup>+</sup> T cells. However, such a regulatory mechanism may not withstand a strong cytotoxic response directed towards plaque specific antigens. We recently reported activation of CD8<sup>+</sup> T cells in response to diet-induced hypercholesterolemia in *ApoE*<sup>-/-</sup> mice and that in lymph nodes draining atherosclerotic lesions this activation precedes that of CD4<sup>+</sup> T cells (39). Accordingly, CD8<sup>+</sup> T cells that specifically recognize lesion antigens could be an important mediator of tissue destruction and chronic inflammation. Candidate antigens for CD8<sup>+</sup> T cell recognition are mimicry proteins of viral or bacterial origin. Jonasson *et al.* reported that patients with coronary artery disease had an expansion of CD8<sup>+</sup> T cells associated to cytomegalovirus infection (40). This notion is supported by observations that antibodies against cytomegalovirus cross-react with Heat shock protein 60 (HSP-60) and associate with atherosclerosis (41). Moreover, both CD8<sup>+</sup> and CD4<sup>+</sup> T cells isolated from human atherosclerotic plaque react with HSP60 and/or *Chlamydia pneumoniae* (42, 43) (44), implicating immune responses against these microorganisms in atherosclerosis.

## **5. CD8<sup>+</sup> regulatory T cells**

Several cell markers have been proposed to define CD8<sup>+</sup> suppressor subsets. CD8<sup>+</sup> Treg markers largely corresponds to those for CD4<sup>+</sup> T cells and include FoxP3, CTLA-4, IL-10, GITR, IL-16, CD11c, TGF-beta, CD103, CD122 and PD-1 (45). One mechanism of T effector cell suppression by regulatory T cells is capture of IL-2 by a high expression of IL-2 receptors. Accordingly, IL-2R beta (CD25) has been proposed to characterize suppressor function also of CD8<sup>+</sup> T cells in mice (46) and humans (47). Interestingly, also the IL-2R alpha (CD122) is representative of a regulatory CD8<sup>+</sup> T cells. These cells have recently being reported to suppress colitis in mice (48) and to have a human counterpart in CD8<sup>+</sup>CXCR3<sup>+</sup> T



cells (49). However, CD8<sup>+</sup>CD122<sup>+</sup> T cells contain both regulatory and memory T cell populations. Dai *et al.* (50) proposed that programmed death-1 (PD-1) defines the suppressor function of CD8<sup>+</sup>CD122<sup>+</sup> T cells, acting via IL-10 production. Interestingly, Gotsman *et al.* (51) found that PD-L1/L2/LDLR-deficient mice develop increased atherosclerosis characterized by an enhanced lesion infiltration of CD4<sup>+</sup> and CD8<sup>+</sup> T cells. Since the PD-ligands on APCs interact with PD-1, regulatory CD8<sup>+</sup> T cells may have a protective role in atherosclerosis. Evidence supporting this possibility was recently reported by Zhao *et al.* who showed that adoptively transferred CD8<sup>+</sup> T cells mediated the athero-protective effects of peptide immunization in *ApoE*<sup>-/-</sup> mice (52).

## **6. Th17 cells**

A population of IL-17 producing T cells (Th 17 cells) constituting a non-Th1/Th2 lineage has recently been described (53). Th17 cells are protective against fungal and bacterial infections, (54) but have also been implicated in atherosclerosis. However, their role in the disease process remains controversial because both pro-atherogenic (55-58) and protective effects (59) have been reported. Th17 cells have been shown to target collagen V exacerbating atherosclerosis (60) and Xie *et al.* suggested that an imbalance between Th17 and regulatory T cells may result in the progression of atherosclerosis (61). However, de Boer *et al.* did not observe Th17 cells in human plaques (62). Instead, the pro-inflammatory IL-17A/F was expressed by plaque neutrophils while SMCs, ECs and B cells produced the anti-inflammatory IL-17E. A correlation between IL-17 and neutrophils was also observed in patients with coronary artery disease (CAD) (63). The plasma level of IL-17 has been reported to correlate with that of IFN- $\gamma$  in CAD patients (64). The same cytokine expression pattern was found in coronary artery-infiltrating T cells, indicating that these cytokines act synergistically to exacerbate atherosclerosis (64). Comparing the Th17/Th1 cell subsets in

peripheral blood of patients with acute coronary syndrome (ACS) Zhao *et al* found that patients had an increase in IL-17<sup>+</sup>/IFN- $\gamma$ <sup>+</sup> T cells, but not IL-17<sup>+</sup>/IFN- $\gamma$ <sup>-</sup> T cells, indicating that a subset of pro-inflammatory Th17 cells are activated in ACS (65). A recent report by von Vietinghoff *et al.* demonstrated that the athero-protective effects of the immunosuppressant mycophenolate mofetil (MMF) in *ApoE*<sup>-/-</sup> mice is associated with a decrease in plasma and aortic IL-17, aortic T cell proliferation and IL-17A dependent accumulation of inflammatory peritoneal macrophages (66). Although the mechanism of Th17 suppression was not clarified, this study provides an interesting concept of specific IL-17 inhibition.

## **7. B cells**

Th2 cell responses are typically associated with B cell activation, plasma cell differentiation and production of antigen-specific antibodies. B cells are present in atherosclerotic lesions, although they are less frequent than T cells (67). Early studies using splenectomised mice suggested that B cells have a protective role in atherosclerosis. The spleen is a major B cell reservoir and splenectomy leads to increased atherosclerosis which can be reversed by B cell replacement (68). It has also been shown that hypercholesterolemic mice deficient in B cells (through depletion of the gene encoding the  $\mu$ -chain of the BCR,  $\mu$ MT) have more atherosclerosis (69). However, the role of B cells in atherosclerosis is now debated since recent data from two independent groups show that blocking B cells by treatment with an antibody against CD20 decreases atherosclerosis development in mice (70, 71). The concept of an athero-protective role of B cells is also based on the observations that some IgM and IgG species protect against atherosclerosis. Anti-oxLDL IgM antibodies have athero-protective effects (72), possibly through their capacity to bind oxLDL and thereby inhibit oxLDL uptake by macrophages and prevent foam cell formation (73). The role of oxLDL-specific IgG remains a matter of controversy because epidemiological studies have reported both positive and inverse associations with cardiovascular disease (74). One possible

explanation to these inconsistent results may be technical difficulties to standardize the antigens used in the ELISAs. In contrast, experimental studies more uniformly point to a protective role of antibodies. Intravenous administration of polyclonal IgG antibodies reduces plaque formation in hypercholesterolemic mice (75, 76), possibly through activation of inhibitory Fc $\gamma$ IIb receptors (77). Antibodies against an aldehyde-modified peptide sequence in apolipoprotein B-100 have been produced by recombinant technique and been shown to inhibit the development of atherosclerosis as well as to induce regression of existing lesions in mice (78, 79). The ability of these antibodies to reduce atherosclerosis also in humans is presently being studied in a randomized clinical trial. Further support for the notion that latter type of antibodies are protective have come from clinical studies demonstrating an association between high levels of IgG autoantibodies to certain peptide sequences in apolipoprotein B-100 peptides and less atherosclerosis as well as lower risk for development of acute myocardial infarction (80, 81).

## **8. Antigen presenting cells in atherosclerosis**

Dendritic cells (DCs) are antigen presenting cells (APCs) that present various types of antigens to T cells leading to initiation and maintenance of immune responses as well as inhibition of activation of T cells. Whether a DC will activate or inhibit T cells depends on its pattern of cytokine release and expression of cell surface co-stimulatory molecules. In general terms it can be said that activation of innate immune receptors, such as the Toll-like receptors (TLR), turns DCs into APCs that activate T effector cells, while antigen presentation that occurs in absence of TLR activation induces immunological tolerance. DCs are therefore a crucial link between innate and adaptive immune responses. Mouse DCs are characterized by expression of CD11c and are present in healthy mouse aortas, primarily in the adventitia (82). In atherosclerotic-prone parts of the aortic arch, mRNA expression of CD11c is elevated

compared to the atherosclerosis-resistant parts. In contrast to healthy vessels the majority of the DCs in atherosclerotic aortas are found in the intima (83). Advanced human plaques have increased number of DCs compared to early plaques. Experimental models of atherosclerosis show that monocyte-derived DCs have an impaired potential to emigrate from advanced plaques compared to plaques from early stages of the disease (84). It has been reported that dyslipidemia impairs the ability of DCs to migrate into lymph nodes (85). The functional role of DCs in atherosclerosis is not fully understood but they potentially play an important role as APCs in T cell activation presenting plaque-derived antigens (86). It has recently been shown that the DC-derived chemokine CCL17 is present in both human and mouse atherosclerotic plaques and that CCL17 deficiency in hypercholesterolemic mice reduces disease development possibly through increased activation of Tregs (87). Modulation of the plaque DC phenotype represents a potential therapeutic target to decrease vascular inflammation and atherosclerosis development.

### **9. Potential atherosclerosis antigens**

When present in atherosclerotic plaques, DCs take up and present plaque-specific antigens. Activated DCs will migrate to adjacent lymph nodes where they activate T cells specific for these antigens. They may also accumulate in the plaque and activate T cells inside the plaque. Modified LDL particles, including minimally modified (mm) LDL, oxidized (ox) LDL and peptides sequences derived from ApoB100 are considered to be important autoantigens in atherosclerosis. When DCs take up LDL-derived antigens in a normal artery wall this will occur in absence of TLR activation. As discussed above such DCs will become tolerogenic and do not induce the activation of antigen-specific Th1 cells. In contrast, if DCs take up such antigens in the inflamed environment of an atherosclerotic plaque increased expression of co-stimulatory factors will favor activation of antigen-specific Th1 cells. As many as 10% of all

T cells cloned from human atherosclerotic plaques react by secreting IFN- $\gamma$  when stimulated with oxLDL indicating that oxLDL-specific T cells induce a Th1 response in the plaque (88). Taken together, these observations suggest that atherosclerosis is aggravated by an autoimmune Th1 response against oxidized LDL. Interestingly, the first studies to test this hypothesis by immunizing of hypercholesterolemic animals with oxLDL or ApoB100 derived peptide fragments unexpectedly resulted in reduced development of atherosclerosis demonstrating that protective immune responses also exists (89-91). This apparent contradiction has subsequently been resolved and it is now clear that Th1 responses against oxLDL indeed are pro-atherogenic and that immunization can shift these immune responses towards tolerance and generation of athero-protective antibodies (92, 93). An atherosclerosis vaccine based on ApoB100 derived peptide has been developed for human use and is expected to enter in to clinical studies relatively soon (93). Another potential approach for immune-modulation to prevent and treat atherosclerosis is so called DC vaccination. This means that autologous DCs are isolated and exposed to a specific antigen *ex vivo* under conditions that either favors subsequent Th1/ Th2 or Treg activation. Accordingly, vaccination of hypercholesterolemic mice with DCs that have been treated with IL-10 (to become tolerogenic) and pulsed with ApoB100 have been shown to decrease atherosclerosis development and increases Treg generation (94). In contrast, non-tolerogenic DCs pulsed with malondialdehyde modified LDL increased atherosclerosis development in *ApoE*<sup>-/-</sup> mice (95) further demonstrating that similar antigens can give rise to both protective and atherogenic immune responses depending on how they are presented. Heat shock proteins (HSPs) represent another family of autoantigens that have been implicated in atherosclerosis. HSPs are highly conserved proteins produced by cells in response to stress factors. Serum levels of HSP60 have been shown to be elevated in subjects with atherosclerosis and to correlate with carotid intima media thickness (96). Immunization of hypercholesterolemic

mice with HSP65 have yielded inconsistent results as early atherosclerosis was increased in one report (97) and decreased in another one (98) . A possible explanation to the divergent results may be difference in the adjuvants used. Infectious agents such as *Chlamydia pneumoniae* have also been suggested to play a role in atherosclerosis. *C. pneumoniae* is a small, intracellular bacterium causing pneumonia that has been shown to infiltrate arteries through peripheral mononuclear cells infecting endothelial cells, vascular smooth muscle cells or monocytes/macrophages. Viable bacteria (99) and non-replicating aberrant bodies (100) has been identified in human atherosclerotic plaques. As T cells isolated from human atheroma respond to Chlamydia antigens (101) and pneumococcal vaccination decreases murine plaque formation (102) a molecular mimicry association between bacterial antigens and self-antigens such as ox LDL and HSPs has been discussed as a possible disease mechanism in atherosclerosis However, the specific impact of *C. pneumoniae* infections on atherosclerosis progression in the long-term perspective still remains to be thoroughly characterized.

## **10. Challenges in transferring mouse data to humans**

As has been outlined above there is convincing experimental evidence that the immune system plays a key role in modulating the atherosclerotic disease process. However, it should be kept in mind that the support for this concept rests almost entirely on studies performed in different mouse models. Also these experiments have been carried out on animals kept in sterile environment in which the general immunological challenge is unnaturally low as compared to humans. Although the immune systems of mice and humans share homology there are also some important differences. Mouse blood has a comparably high lymphocyte fraction whereas human blood is neutrophil-rich. In addition, humans have a more pronounced CD4<sup>+</sup>/CD8<sup>+</sup> T cell ratio, while the Th1-Th2 differentiation is less clear (103) .

Moreover, gender differences in lymphocyte numbers and sampling site should be thoroughly controlled to avoid introduction of misleading bias (104) . As characterization of lymphocyte populations differ, cell surface markers coupled to a functional mechanism apparent in mice may not be functional in humans. Thus, translation of data from mice to man has to be performed with cautiousness in order to minimize misinterpretation.

### **11. Lymphocytes in development and quantification of atherosclerotic plaques – disparities in mice and humans**

Experimental mouse models of atherosclerosis research are commonly based on gene defects and diets which generate several-fold higher plasma cholesterol level than in humans. This difference may cause a more rapid cholesterol accumulation and lesion development which is different from the slow build-up of human lesions that proceeds during decades in man (105). Differences in lipid metabolism may also affect immune responses differently in mouse and humans. Potentially, cell populations that respond to an immediate imbalance may dominate in mouse models of atherosclerosis, while the gradual development of human lesions involves different immune response. To overcome these problems mouse diets that only moderately increase cholesterol could be used but a likely downside of this approach is that only minimal atherosclerosis will develop. Additionally, genetically modified humanized mouse models could be used to better mimic the human immune response.

The location of plaque development represents another possible important limitation in the use of mouse models for atherosclerosis. Mouse lesions are usually studied at the aortic root where they first develop in mice whereas lesions rarely develop at this location in humans. Accordingly, lesion specific response to an intervention in mice may not be representative the human situation.

## **12. Concluding remarks and perspectives**

The identification and characterization of the important role of the immune system in atherosclerosis has significantly increased our understanding of the disease process and also helped to identify novel targets for prevention and treatment of cardiovascular disease. There is large clinical need for such treatments because it appears difficult to achieve risk-reductions greater than 30-40 % using the present risk factor intervention strategy. Novel treatments need to directly target the actual disease in the arterial wall and it is likely that modulation of plaque-specific immune responses represent a particularly promising approach to achieve this. Experimental animal studies have demonstrated the presence of both protective and atherogenic immune responses and evaluated candidate therapeutic strategies to specifically activate or mimic the protective immune responses. It has been clearly shown that the T cells are important effectors of both types of immune responses. The challenge for the future is to translate the knowledge obtained in experimental models into the clinic. The difficulties facing this process should not be underestimated. However, it should be kept in mind that the concept of atherosclerosis as a disease with partial autoimmune etiology is relatively new as compared to diseases such as type 1 diabetes, rheumatoid arthritis, systemic lupus erythematosus and multiple sclerosis and that there is much to be learned from attempts to develop immune-based therapies for these diseases.

### **Disclosures**

Jan Nilsson is signed as co-inventor on several patents on immune-modulatory therapies for atherosclerosis.



## Figure legend

**Role of T cells in atherosclerosis.** T cells entering into an atherosclerotic plaque will be exposed to antigens presented by a dendritic cell (DC) or by a macrophage. If the antigen presentation is done by an activated DC CD4<sup>+</sup> T cells will differentiate into Th1 cell secreting pro-inflammatory cytokines such as INF- $\gamma$  and TNF- $\alpha$ , while presentation by a tolerogenic DC will result in formation of a suppressive regulatory T cell (Treg) secreting anti-inflammatory cytokines such as IL-10 and TGF- $\beta$ . Activated CD8<sup>+</sup> and Th17 T cells are also present in atherosclerotic plaques but their functional roles remains to be fully elucidated. Monocytes exposed to pro-inflammatory cytokines in the plaque will become activated and contribute to both plaque progression and plaque de-stabilization. Monocytes also differentiate into macrophages that take up oxidized (ox) LDL and develop into foam cells. Lipids and cellular debris released from dead foam cells contribute to the development of a necrotic core.

## References

1. Dworacka M, Winiarska H, Borowska M, Abramczyk M, Bobkiewicz-Kozłowska T, Dworacki G. Pro-atherogenic alterations in T-lymphocyte subpopulations related to acute hyperglycaemia in type 2 diabetic patients. *Circ J*. 2007;71(6):962-7. Epub 2007/05/29.
2. Szodoray P, Timar O, Veres K, Der H, Szomjak E, Lakos G, et al. TH1/TH2 imbalance, measured by circulating and intracytoplasmic inflammatory cytokines--immunological alterations in acute coronary syndrome and stable coronary artery disease. *Scand J Immunol*. 2006;64(3):336-44. Epub 2006/08/22.
3. Nunez J, Minana G, Bodi V, Nunez E, Sanchis J, Husser O, et al. Low lymphocyte count and cardiovascular diseases. *Curr Med Chem*. 2011;18(21):3226-33. Epub 2011/06/16.
4. Zhou X, Nicoletti A, Elhage R, Hansson GK. Transfer of CD4(+) T cells aggravates atherosclerosis in immunodeficient apolipoprotein E knockout mice. *Circulation*. 2000;102(24):2919-22. Epub 2000/01/11.
5. Zhou X, Robertson AK, Rudling M, Parini P, Hansson GK. Lesion development and response to immunization reveal a complex role for CD4 in atherosclerosis. *Circ Res*. 2005;96(4):427-34. Epub 2005/01/22.
6. Elhage R, Gourdy P, Bouchet L, Jawien J, Fouque MJ, Fievet C, et al. Deleting TCR alpha beta+ or CD4+ T lymphocytes leads to opposite effects on site-specific atherosclerosis in female apolipoprotein E-deficient mice. *Am J Pathol*. 2004;165(6):2013-8. Epub 2004/12/08.
7. Buono C, Binder CJ, Stavrakis G, Witztum JL, Glimcher LH, Lichtman AH. T-bet deficiency reduces atherosclerosis and alters plaque antigen-specific immune responses. *Proc Natl Acad Sci U S A*. 2005;102(5):1596-601. Epub 2005/01/25.
8. Buono C, Come CE, Stavrakis G, Maguire GF, Connelly PW, Lichtman AH. Influence of interferon-gamma on the extent and phenotype of diet-induced atherosclerosis in the LDLR-deficient mouse. *Arterioscler Thromb Vasc Biol*. 2003;23(3):454-60. Epub 2003/03/05.
9. Branen L, Hovgaard L, Nitulescu M, Bengtsson E, Nilsson J, Jovinge S. Inhibition of tumor necrosis factor-alpha reduces atherosclerosis in apolipoprotein E knockout mice. *Arterioscler Thromb Vasc Biol*. 2004;24(11):2137-42. Epub 2004/09/04.
10. Davenport P, Tipping PG. The role of interleukin-4 and interleukin-12 in the progression of atherosclerosis in apolipoprotein E-deficient mice. *Am J Pathol*. 2003;163(3):1117-25. Epub 2003/08/26.
11. King VL, Cassis LA, Daugherty A. Interleukin-4 does not influence development of hypercholesterolemia or angiotensin II-induced atherosclerotic lesions in mice. *Am J Pathol*. 2007;171(6):2040-7. Epub 2007/12/07.
12. Klemann R, Zadelaar S, Kooistra T. Cytokines and atherosclerosis: a comprehensive review of studies in mice. *Cardiovasc Res*. 2008;79(3):360-76. Epub 2008/05/20.
13. Noelle RJ, Nowak EC. Cellular sources and immune functions of interleukin-9. *Nat Rev Immunol*. 2010;10(10):683-7. Epub 2010/09/18.
14. Caruso R, Stolfi C, De Nitto D, Pallone F, Monteleone G. The dual role of interleukin-25 in the control of immune-mediated pathologies. *Curr Mol Med*. 2011;11(1):26-30. Epub 2010/12/30.
15. Binder CJ, Hartvigsen K, Chang MK, Miller M, Broide D, Palinski W, et al. IL-5 links adaptive and natural immunity specific for epitopes of oxidized LDL and protects from atherosclerosis. *J Clin Invest*. 2004;114(3):427-37. Epub 2004/08/03.

16. Sampi M, Ukkola O, Paivansalo M, Kesaniemi YA, Binder CJ, Horkko S. Plasma interleukin-5 levels are related to antibodies binding to oxidized low-density lipoprotein and to decreased subclinical atherosclerosis. *J Am Coll Cardiol*. 2008;52(17):1370-8. Epub 2008/10/23.
17. Miller AM, Xu D, Asquith DL, Denby L, Li Y, Sattar N, et al. IL-33 reduces the development of atherosclerosis. *J Exp Med*. 2008;205(2):339-46. Epub 2008/02/13.
18. Corthay A. How do regulatory T cells work? *Scand J Immunol*. 2009;70(4):326-36. Epub 2009/09/16.
19. de Boer OJ, van der Meer JJ, Teeling P, van der Loos CM, van der Wal AC. Low numbers of FOXP3 positive regulatory T cells are present in all developmental stages of human atherosclerotic lesions. *PLoS One*. 2007;2(1):e779. Epub 2007/08/23.
20. Veillard NR, Steffens S, Burger F, Pelli G, Mach F. Differential expression patterns of proinflammatory and antiinflammatory mediators during atherogenesis in mice. *Arterioscler Thromb Vasc Biol*. 2004;24(12):2339-44. Epub 2004/10/02.
21. Wigren M, Bengtsson D, Duner P, Olofsson K, Bjorkbacka H, Bengtsson E, et al. Atheroprotective effects of Alum are associated with capture of oxidized LDL antigens and activation of regulatory T cells. *Circ Res*. 2009;104(12):e62-70. Epub 2009/05/30.
22. Gotsman I, Grabie N, Gupta R, Dacosta R, MacConmara M, Lederer J, et al. Impaired regulatory T-cell response and enhanced atherosclerosis in the absence of inducible costimulatory molecule. *Circulation*. 2006;114(19):2047-55. Epub 2006/10/25.
23. Ait-Oufella H, Salomon BL, Potteaux S, Robertson AK, Gourdy P, Zoll J, et al. Natural regulatory T cells control the development of atherosclerosis in mice. *Nature medicine*. 2006;12(2):178-80. Epub 2006/02/08.
24. van Es T, van Puijvelde GH, Foks AC, Habets KL, Bot I, Gilboa E, et al. Vaccination against Foxp3(+) regulatory T cells aggravates atherosclerosis. *Atherosclerosis*. 2010;209(1):74-80. Epub 2009/09/22.
25. Mallat Z, Besnard S, Duriez M, Deleuze V, Emmanuel F, Bureau MF, et al. Protective role of interleukin-10 in atherosclerosis. *Circ Res*. 1999;85(8):e17-24. Epub 1999/10/16.
26. Mallat Z, Gojova A, Marchiol-Fournigault C, Esposito B, Kamate C, Merval R, et al. Inhibition of transforming growth factor-beta signaling accelerates atherosclerosis and induces an unstable plaque phenotype in mice. *Circ Res*. 2001;89(10):930-4. Epub 2001/11/10.
27. Pinderski Oslund LJ, Hedrick CC, Olvera T, Hagenbaugh A, Territo M, Berliner JA, et al. Interleukin-10 blocks atherosclerotic events in vitro and in vivo. *Arterioscler Thromb Vasc Biol*. 1999;19(12):2847-53. Epub 1999/12/11.
28. Robertson AK, Rudling M, Zhou X, Gorelik L, Flavell RA, Hansson GK. Disruption of TGF-beta signaling in T cells accelerates atherosclerosis. *J Clin Invest*. 2003;112(9):1342-50. Epub 2003/10/22.
29. Sakaguchi S, Yamaguchi T, Nomura T, Ono M. Regulatory T cells and immune tolerance. *Cell*. 2008;133(5):775-87. Epub 2008/05/31.
30. Cvetanovich GL, Hafler DA. Human regulatory T cells in autoimmune diseases. *Curr Opin Immunol*. 2010;22(6):753-60. Epub 2010/09/28.
31. Schuler PJ, Harasymczuk M, Schilling B, Lang S, Whiteside TL. Separation of human CD4+CD39+ T cells by magnetic beads reveals two phenotypically and functionally different subsets. *J Immunol Methods*. 2011;369(1-2):59-68. Epub 2011/04/26.
32. Mor A, Luboshits G, Planer D, Keren G, George J. Altered status of CD4(+)CD25(+) regulatory T cells in patients with acute coronary syndromes. *European heart journal*. 2006;27(21):2530-7. Epub 2006/09/07.

33. Ammirati E, Cianflone D, Banfi M, Vecchio V, Palini A, De Metrio M, et al. Circulating CD4<sup>+</sup>CD25<sup>hi</sup>CD127<sup>lo</sup> regulatory T-Cell levels do not reflect the extent or severity of carotid and coronary atherosclerosis. *Arterioscler Thromb Vasc Biol.* 2010;30(9):1832-41. Epub 2010/06/12.
34. Fyfe AI, Qiao JH, Lusis AJ. Immune-deficient mice develop typical atherosclerotic fatty streaks when fed an atherogenic diet. *J Clin Invest.* 1994;94(6):2516-20. Epub 1994/12/01.
35. Roselaar SE, Kakkanathu PX, Daugherty A. Lymphocyte populations in atherosclerotic lesions of apoE <sup>-/-</sup> and LDL receptor <sup>-/-</sup> mice. Decreasing density with disease progression. *Arterioscler Thromb Vasc Biol.* 1996;16(8):1013-8. Epub 1996/08/01.
36. Gewaltig J, Kummer M, Koella C, Cathomas G, Biedermann BC. Requirements for CD8 T-cell migration into the human arterial wall. *Hum Pathol.* 2008;39(12):1756-62. Epub 2008/08/19.
37. Ludewig B, Freigang S, Jaggi M, Kurrer MO, Pei YC, Vlk L, et al. Linking immune-mediated arterial inflammation and cholesterol-induced atherosclerosis in a transgenic mouse model. *Proc Natl Acad Sci U S A.* 2000;97(23):12752-7. Epub 2000/10/26.
38. Escalante NK, von Rossum A, Lee M, Choy JC. CD155 on human vascular endothelial cells attenuates the acquisition of effector functions in CD8 T cells. *Arterioscler Thromb Vasc Biol.* 2011;31(5):1177-84. Epub 2011/02/19.
39. Kolbus D, Ramos OH, Berg KE, Persson J, Wigren M, Bjorkbacka H, et al. CD8<sup>+</sup> T cell activation predominate early immune responses to hypercholesterolemia in ApoE<sup>-/-</sup> mice. *BMC Immunol.* 2010;11(1):58. Epub 2010/12/04.
40. Jonasson L, Tompa A, Wikby A. Expansion of peripheral CD8<sup>+</sup> T cells in patients with coronary artery disease: relation to cytomegalovirus infection. *J Intern Med.* 2003;254(5):472-8. Epub 2003/10/11.
41. Bason C, Corrocher R, Lunardi C, Puccetti P, Olivieri O, Girelli D, et al. Interaction of antibodies against cytomegalovirus with heat-shock protein 60 in pathogenesis of atherosclerosis. *Lancet.* 2003;362(9400):1971-7. Epub 2003/12/20.
42. Rossmann A, Henderson B, Heidecker B, Seiler R, Fraedrich G, Singh M, et al. T-cells from advanced atherosclerotic lesions recognize hHSP60 and have a restricted T-cell receptor repertoire. *Exp Gerontol.* 2008;43(3):229-37. Epub 2008/01/30.
43. Benagiano M, D'Elis MM, Amedei A, Azzurri A, van der Zee R, Ciervo A, et al. Human 60-kDa heat shock protein is a target autoantigen of T cells derived from atherosclerotic plaques. *J Immunol.* 2005;174(10):6509-17. Epub 2005/05/10.
44. Nadareishvili ZG, Koziol DE, Szekely B, Ruetzler C, LaBiche R, McCarron R, et al. Increased CD8(+) T cells associated with Chlamydia pneumoniae in symptomatic carotid plaque. *Stroke.* 2001;32(9):1966-72. Epub 2001/09/08.
45. Suzuki M, Konya C, Goronzy JJ, Weyand CM. Inhibitory CD8<sup>+</sup> T cells in autoimmune disease. *Hum Immunol.* 2008;69(11):781-9. Epub 2008/09/25.
46. Bienvenu B, Martin B, Auffray C, Cordier C, Becourt C, Lucas B. Peripheral CD8<sup>+</sup>CD25<sup>+</sup> T lymphocytes from MHC class II-deficient mice exhibit regulatory activity. *J Immunol.* 2005;175(1):246-53. Epub 2005/06/24.
47. Cosmi L, Liotta F, Lazzeri E, Francalanci M, Angeli R, Mazinghi B, et al. Human CD8<sup>+</sup>CD25<sup>+</sup> thymocytes share phenotypic and functional features with CD4<sup>+</sup>CD25<sup>+</sup> regulatory thymocytes. *Blood.* 2003;102(12):4107-14. Epub 2003/08/02.
48. Endharti AT, Okuno Y, Shi Z, Misawa N, Toyokuni S, Ito M, et al. CD8<sup>+</sup>CD122<sup>+</sup> regulatory T cells (Tregs) and CD4<sup>+</sup> Tregs cooperatively prevent and cure CD4<sup>+</sup> cell-induced colitis. *J Immunol.* 2011;186(1):41-52. Epub 2010/11/26.

49. Shi Z, Okuno Y, Rifa'i M, Endharti AT, Akane K, Isobe K, et al. Human CD8+CXCR3+ T cells have the same function as murine CD8+CD122+ Treg. *Eur J Immunol.* 2009;39(8):2106-19. Epub 2009/07/18.
50. Dai H, Wan N, Zhang S, Moore Y, Wan F, Dai Z. Cutting edge: programmed death-1 defines CD8+CD122+ T cells as regulatory versus memory T cells. *J Immunol.* 2010;185(2):803-7. Epub 2010/06/16.
51. Gotsman I, Grabie N, Dacosta R, Sukhova G, Sharpe A, Lichtman AH. Proatherogenic immune responses are regulated by the PD-1/PD-L pathway in mice. *J Clin Invest.* 2007;117(10):2974-82. Epub 2007/09/15.
52. Zhao Xiaoning C-Y, Dimayuga Paul C, Zhou Jianchang, Yano Juliana , Lio Wei Man, Chan Lai Fan , Li Xiaojun, Kirzner Jonathan , Trinidad Portia , Cercek Bojan, and Shah Prediman K Abstract 18244: Adoptive Cell Transfer Demonstrates that CD8+ T cells Mediate the Anti-atherogenic Effects of the ApoB-100 Peptide P210 Immunization in ApoE<sup>-/-</sup> Mice *Circulation.* 2010;122:A18244.
53. Miossec P. IL-17 and Th17 cells in human inflammatory diseases. *Microbes Infect.* 2009;11(5):625-30. Epub 2009/04/18.
54. Milner JD, Sandler NG, Douek DC. Th17 cells, Job's syndrome and HIV: opportunities for bacterial and fungal infections. *Curr Opin HIV AIDS.* 2010;5(2):179-83. Epub 2010/06/15.
55. van Es T, van Puijvelde GH, Ramos OH, Segers FM, Joosten LA, van den Berg WB, et al. Attenuated atherosclerosis upon IL-17R signaling disruption in LDLr deficient mice. *Biochem Biophys Res Commun.* 2009;388(2):261-5. Epub 2009/08/08.
56. Smith E, Prasad KM, Butcher M, Dobrian A, Kolls JK, Ley K, et al. Blockade of interleukin-17A results in reduced atherosclerosis in apolipoprotein E-deficient mice. *Circulation.* 2010;121(15):1746-55. Epub 2010/04/07.
57. Gao Q, Jiang Y, Ma T, Zhu F, Gao F, Zhang P, et al. A critical function of Th17 proinflammatory cells in the development of atherosclerotic plaque in mice. *J Immunol.* 2010;185(10):5820-7. Epub 2010/10/19.
58. Madhur MS, Funt SA, Li L, Vinh A, Chen W, Lob HE, et al. Role of interleukin 17 in inflammation, atherosclerosis, and vascular function in apolipoprotein e-deficient mice. *Arterioscler Thromb Vasc Biol.* 2011;31(7):1565-72. Epub 2011/04/09.
59. Taleb S, Romain M, Ramkhelawon B, Uyttenhove C, Pasterkamp G, Herbin O, et al. Loss of SOCS3 expression in T cells reveals a regulatory role for interleukin-17 in atherosclerosis. *J Exp Med.* 2009;206(10):2067-77. Epub 2009/09/10.
60. Dart ML, Jankowska-Gan E, Huang G, Roenneburg DA, Keller MR, Torrealba JR, et al. Interleukin-17-dependent autoimmunity to collagen type V in atherosclerosis. *Circ Res.* 2010;107(9):1106-16. Epub 2010/09/04.
61. Xie JJ, Wang J, Tang TT, Chen J, Gao XL, Yuan J, et al. The Th17/Treg functional imbalance during atherogenesis in ApoE<sup>(-/-)</sup> mice. *Cytokine.* 2010;49(2):185-93. Epub 2009/10/20.
62. de Boer OJ, van der Meer JJ, Teeling P, van der Loos CM, Idu MM, van Maldegem F, et al. Differential expression of interleukin-17 family cytokines in intact and complicated human atherosclerotic plaques. *J Pathol.* 2010;220(4):499-508. Epub 2009/12/19.
63. Wang Z, Lee J, Zhang Y, Wang H, Liu X, Shang F, et al. Increased Th17 cells in coronary artery disease are associated with neutrophilic inflammation. *Scand Cardiovasc J.* 2011;45(1):54-61. Epub 2011/01/14.
64. Eid RE, Rao DA, Zhou J, Lo SF, Ranjbaran H, Gallo A, et al. Interleukin-17 and interferon-gamma are produced concomitantly by human coronary artery-infiltrating T cells and act synergistically on vascular smooth muscle cells. *Circulation.* 2009;119(10):1424-32. Epub 2009/03/04.

65. Zhao Z, Wu Y, Cheng M, Ji Y, Yang X, Liu P, et al. Activation of Th17/Th1 and Th1, but not Th17, is associated with the acute cardiac event in patients with acute coronary syndrome. *Atherosclerosis*. 2011;217(2):518-24. Epub 2011/04/29.
66. von Vietinghoff S, Koltsova EK, Mestas J, Diehl CJ, Witztum JL, Ley K. Mycophenolate mofetil decreases atherosclerotic lesion size by depression of aortic T-lymphocyte and interleukin-17-mediated macrophage accumulation. *J Am Coll Cardiol*. 2011;57(21):2194-204. Epub 2011/05/21.
67. Zhou X, Hansson GK. Detection of B cells and proinflammatory cytokines in atherosclerotic plaques of hypercholesterolaemic apolipoprotein E knockout mice. *Scand J Immunol*. 1999;50(1):25-30. Epub 1999/07/15.
68. Caligiuri G, Nicoletti A, Poirier B, Hansson GK. Protective immunity against atherosclerosis carried by B cells of hypercholesterolemic mice. *J Clin Invest*. 2002;109(6):745-53. Epub 2002/03/20.
69. Major AS, Fazio S, Linton MF. B-lymphocyte deficiency increases atherosclerosis in LDL receptor-null mice. *Arterioscler Thromb Vasc Biol*. 2002;22(11):1892-8. Epub 2002/11/12.
70. Ait-Oufella H, Herbin O, Bouaziz JD, Binder CJ, Uyttenhove C, Laurant L, et al. B cell depletion reduces the development of atherosclerosis in mice. *J Exp Med*. 2010;207(8):1579-87. Epub 2010/07/07.
71. Kyaw T, Tay C, Khan A, Dumouchel V, Cao A, To K, et al. Conventional B2 B cell depletion ameliorates whereas its adoptive transfer aggravates atherosclerosis. *J Immunol*. 2010;185(7):4410-9. Epub 2010/09/08.
72. Tsimikas S, Brilakis ES, Lennon RJ, Miller ER, Witztum JL, McConnell JP, et al. Relationship of IgG and IgM autoantibodies to oxidized low density lipoprotein with coronary artery disease and cardiovascular events. *J Lipid Res*. 2007;48(2):425-33. Epub 2006/11/10.
73. Nilsson J, Nordin Fredrikson G, Schioppa A, Shah PK, Jansson B, Carlsson R. Oxidized LDL antibodies in treatment and risk assessment of atherosclerosis and associated cardiovascular disease. *Curr Pharm Des*. 2007;13(10):1021-30. Epub 2007/04/14.
74. Nilsson J, Kovanen PT. Will autoantibodies help to determine severity and progression of atherosclerosis? *Curr Opin Lipidol*. 2004;15(5):499-503. Epub 2004/09/14.
75. Persson L, Boren J, Nicoletti A, Hansson GK, Pekna M. Immunoglobulin treatment reduces atherosclerosis in apolipoprotein E<sup>-/-</sup> low-density lipoprotein receptor<sup>-/-</sup> mice via the complement system. *Clinical and experimental immunology*. 2005;142(3):441-5. Epub 2005/11/22.
76. Nicoletti A, Kaveri S, Caligiuri G, Bariety J, Hansson GK. Immunoglobulin treatment reduces atherosclerosis in apo E knockout mice. *J Clin Invest*. 1998;102(5):910-8. Epub 1998/09/03.
77. Zhao M, Wigren M, Duner P, Kolbus D, Olofsson KE, Bjorkbacka H, et al. FcγRIIB inhibits the development of atherosclerosis in low-density lipoprotein receptor-deficient mice. *J Immunol*. 2010;184(5):2253-60. Epub 2010/01/26.
78. Schioppa A, Bengtsson J, Soderberg I, Janciauskiene S, Lindgren S, Ares MP, et al. Recombinant human antibodies against aldehyde-modified apolipoprotein B-100 peptide sequences inhibit atherosclerosis. *Circulation*. 2004;110(14):2047-52. Epub 2004/09/29.
79. Schioppa A, Frendeus B, Jansson B, Soderberg I, Ljungcrantz I, Araya Z, et al. Recombinant antibodies to an oxidized low-density lipoprotein epitope induce rapid regression of atherosclerosis in apobec-1<sup>(-/-)</sup>/low-density lipoprotein receptor<sup>(-/-)</sup> mice. *J Am Coll Cardiol*. 2007;50(24):2313-8. Epub 2007/12/11.
80. Fredrikson GN, Schioppa A, Berglund G, Alm R, Shah PK, Nilsson J. Autoantibody against the amino acid sequence 661-680 in apo B-100 is associated with

- decreased carotid stenosis and cardiovascular events. *Atherosclerosis*. 2007;194(2):e188-92. Epub 2007/01/12.
81. Sjogren P, Fredrikson GN, Samnegard A, Ericsson CG, Ohrvik J, Fisher RM, et al. High plasma concentrations of autoantibodies against native peptide 210 of apoB-100 are related to less coronary atherosclerosis and lower risk of myocardial infarction. *European heart journal*. 2008;29(18):2218-26. Epub 2008/07/31.
  82. Moos MP, John N, Grabner R, Nossmann S, Gunther B, Vollandt R, et al. The lamina adventitia is the major site of immune cell accumulation in standard chow-fed apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol*. 2005;25(11):2386-91. Epub 2005/09/24.
  83. Jongstra-Bilen J, Haidari M, Zhu SN, Chen M, Guha D, Cybulsky MI. Low-grade chronic inflammation in regions of the normal mouse arterial intima predisposed to atherosclerosis. *J Exp Med*. 2006;203(9):2073-83. Epub 2006/08/09.
  84. Llodra J, Angeli V, Liu J, Trogan E, Fisher EA, Randolph GJ. Emigration of monocyte-derived cells from atherosclerotic lesions characterizes regressive, but not progressive, plaques. *Proc Natl Acad Sci U S A*. 2004;101(32):11779-84. Epub 2004/07/29.
  85. Angeli V, Llodra J, Rong JX, Satoh K, Ishii S, Shimizu T, et al. Dyslipidemia associated with atherosclerotic disease systemically alters dendritic cell mobilization. *Immunity*. 2004;21(4):561-74. Epub 2004/10/16.
  86. Koltsova EK, Ley K. How dendritic cells shape atherosclerosis. *Trends Immunol*. 2011. Epub 2011/08/13.
  87. Weber C, Meiler S, Doring Y, Koch M, Drechsler M, Megens RT, et al. CCL17-expressing dendritic cells drive atherosclerosis by restraining regulatory T cell homeostasis in mice. *J Clin Invest*. 2011;121(7). Epub 2011/06/03.
  88. Stemme S, Faber B, Holm J, Wiklund O, Witztum JL, Hansson GK. T lymphocytes from human atherosclerotic plaques recognize oxidized low density lipoprotein. *Proc Natl Acad Sci U S A*. 1995;92(9):3893-7. Epub 1995/04/25.
  89. Ameli S, Hultgardh-Nilsson A, Regnstrom J, Calara F, Yano J, Cercek B, et al. Effect of immunization with homologous LDL and oxidized LDL on early atherosclerosis in hypercholesterolemic rabbits. *Arterioscler Thromb Vasc Biol*. 1996;16(8):1074-9. Epub 1996/08/01.
  90. Yuan Z, Kishimoto C, Sano H, Shioji K, Xu Y, Yokode M. Immunoglobulin treatment suppresses atherosclerosis in apolipoprotein E-deficient mice via the Fc portion. *Am J Physiol Heart Circ Physiol*. 2003;285(2):H899-906. Epub 2003/07/16.
  91. Fredrikson GN, Soderberg I, Lindholm M, Dimayuga P, Chyu KY, Shah PK, et al. Inhibition of atherosclerosis in apoE-null mice by immunization with apoB-100 peptide sequences. *Arterioscler Thromb Vasc Biol*. 2003;23(5):879-84. Epub 2003/03/22.
  92. Nilsson J, Hansson GK. Autoimmunity in atherosclerosis: a protective response losing control? *J Intern Med*. 2008;263(5):464-78. Epub 2008/04/16.
  93. Nilsson J, Wigren M, Shah PK. Regulatory T cells and the control of modified lipoprotein autoimmunity-driven atherosclerosis. *Trends Cardiovasc Med*. 2009;19(8):272-6. Epub 2010/05/08.
  94. Hermansson A, Johansson DK, Ketelhuth DF, Andersson J, Zhou X, Hansson GK. Immunotherapy with tolerogenic apolipoprotein B-100-loaded dendritic cells attenuates atherosclerosis in hypercholesterolemic mice. *Circulation*. 2011;123(10):1083-91. Epub 2011/03/02.
  95. Hjerpe C, Johansson D, Hermansson A, Hansson GK, Zhou X. Dendritic cells pulsed with malondialdehyde modified low density lipoprotein aggravate atherosclerosis in Apoe(-/-) mice. *Atherosclerosis*. 2010;209(2):436-41. Epub 2009/11/10.

96. Xu Q, Schett G, Perschinka H, Mayr M, Egger G, Oberhollenzer F, et al. Serum soluble heat shock protein 60 is elevated in subjects with atherosclerosis in a general population. *Circulation*. 2000;102(1):14-20. Epub 2000/07/06.
97. Afek A, George J, Gilburd B, Rauova L, Goldberg I, Kopolovic J, et al. Immunization of low-density lipoprotein receptor deficient (LDL-RD) mice with heat shock protein 65 (HSP-65) promotes early atherosclerosis. *J Autoimmun*. 2000;14(2):115-21. Epub 2000/03/24.
98. Klingenberg R, Ketelhuth DF, Strodthoff D, Gregori S, Hansson GK. Subcutaneous immunization with heat shock protein-65 reduces atherosclerosis in Apoe(-/-) mice. *Immunobiology*. 2011. Epub 2011/07/30.
99. Nystrom-Rosander C, Edvinsson M, Thelin S, Hjelm E, Friman G. Chlamydia pneumoniae: Specific mRNA in aorta ascendens in patients undergoing coronary artery by-pass grafting. *Scandinavian journal of infectious diseases*. 2006;38(9):758-63. Epub 2006/08/30.
100. Borel N, Summersgill JT, Mukhopadhyay S, Miller RD, Ramirez JA, Pospischil A. Evidence for persistent Chlamydia pneumoniae infection of human coronary atheromas. *Atherosclerosis*. 2008;199(1):154-61. Epub 2007/11/22.
101. Curry AJ, Portig I, Goodall JC, Kirkpatrick PJ, Gaston JS. T lymphocyte lines isolated from atheromatous plaque contain cells capable of responding to Chlamydia antigens. *Clinical and experimental immunology*. 2000;121(2):261-9. Epub 2000/08/10.
102. Binder CJ, Horkko S, Dewan A, Chang MK, Kieu EP, Goodyear CS, et al. Pneumococcal vaccination decreases atherosclerotic lesion formation: molecular mimicry between Streptococcus pneumoniae and oxidized LDL. *Nature medicine*. 2003;9(6):736-43. Epub 2003/05/13.
103. Mestas J, Hughes CC. Of mice and not men: differences between mouse and human immunology. *J Immunol*. 2004;172(5):2731-8. Epub 2004/02/24.
104. Doeing DC, Borowicz JL, Crockett ET. Gender dimorphism in differential peripheral blood leukocyte counts in mice using cardiac, tail, foot, and saphenous vein puncture methods. *BMC Clin Pathol*. 2003;3(1):3. Epub 2003/09/16.
105. Goncalves I, Stenstrom K, Skog G, Mattsson S, Nitulescu M, Nilsson J. Short communication: Dating components of human atherosclerotic plaques. *Circ Res*. 2010;106(6):1174-7. Epub 2010/02/20.



