



LUND UNIVERSITY

Bactericidal activity of human eosinophilic granulocytes against *Escherichia coli*

Persson, Terese; Andersson, Pia; Bodelsson, Mikael; Laurell, Martin; Malm, Johan; Egesten, Arne

Published in:
Infection and Immunity

DOI:
[10.1128/IAI.69.6.3591-3596.2001](https://doi.org/10.1128/IAI.69.6.3591-3596.2001)

2001

[Link to publication](#)

Citation for published version (APA):

Persson, T., Andersson, P., Bodelsson, M., Laurell, M., Malm, J., & Egesten, A. (2001). Bactericidal activity of human eosinophilic granulocytes against *Escherichia coli*. *Infection and Immunity*, 69(6), 3591-3596. <https://doi.org/10.1128/IAI.69.6.3591-3596.2001>

Total number of authors:
6

General rights

Unless other specific re-use rights are stated the following general rights apply:

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

Read more about Creative commons licenses: <https://creativecommons.org/licenses/>

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

LUND UNIVERSITY

PO Box 117
221 00 Lund
+46 46-222 00 00

Bactericidal Activity of Human Eosinophilic Granulocytes against *Escherichia coli*

TERESE PERSSON,¹ PIA ANDERSSON,¹ MIKAEL BODELSSON,² MARTIN LAURELL,¹
JOHAN MALM,³ AND ARNE EGESTEN^{1*}

Sections for Medical Microbiology¹ and Clinical Chemistry,³ Department of Laboratory Medicine, Lund University,
Malmö University Hospital, Malmö, and Department of Anesthesia and Intensive Care,
Lund University Hospital, Lund,² Sweden

Received 13 November 2000/Returned for modification 8 January 2001/Accepted 12 March 2001

Eosinophils participate in allergic inflammation and may have roles in the body's defense against helminthic infestation. Even under noninflammatory conditions, eosinophils are present in the mucosa of the large intestine, where large numbers of gram-negative bacteria reside. Therefore, roles for eosinophils in host defenses against bacterial invasion are possible. In a system for bacterial viable counts, the bactericidal activity of eosinophils and the contribution of different cellular antibacterial systems against *Escherichia coli* were investigated. Eosinophils showed a rapid and efficient killing of *E. coli* under aerobic conditions, whereas under anaerobic conditions bacterial killing decreased dramatically. In addition, diphenylene iodonium chloride (DPI), an inhibitor of the NADPH oxidase and thereby of superoxide production, also significantly inhibited bacterial killing. The inhibitor of nitric oxide (NO) production L-N⁵-(1-iminoethyl)-ornithine dihydrochloride did not affect the killing efficiency, suggesting that NO or derivatives thereof are of minor importance under the experimental conditions used. To investigate the involvement of superoxide and eosinophil peroxidase (EPO) in bacterial killing, EPO was blocked by azide. The rate of *E. coli* killing decreased significantly in the presence of azide, whereas addition of DPI did not further decrease the killing, suggesting that superoxide acts in conjunction with EPO. Bactericidal activity was seen in eosinophil extracts containing granule proteins, indicating that oxygen-independent killing may be of importance as well. The findings suggest that eosinophils can participate in host defense against gram-negative bacterial invasion and that oxygen-dependent killing, i.e., superoxide acting in conjunction with EPO, may be the most important bactericidal effector function of these cells.

Eosinophilic granulocytes are believed to participate in the body's defense against infestation with parasites, especially helminths (such as nematodes and trematodes) in the larval stage (35). They are also typically present in the allergic inflammatory response seen in conditions such as allergy and asthma, where they may contribute to tissue damage and connective tissue remodeling (35). However, the roles of eosinophils in host defense functions are not settled. Even under normal, noninflammatory conditions, eosinophils reside in tissues close to mucosal membranes that are in close contact with a potentially hostile environment, such as the content of gram-negative bacteria of the large intestine (23). Furthermore, eosinophils possess the adhesion molecule $\alpha 4\beta 7$, which binds to MAdCAM-1, an adhesion molecule expressed by the endothelial cells of the vessels in the gastrointestinal tract, also suggesting roles for eosinophils after recruitment to these tissues (16). Their number increases dramatically during some conditions, such as ulcerative colitis, where the integrity of the mucosal barrier is disturbed and bacteria may invade the tissue (27).

Eosinophils have several properties that can contribute to bactericidal activity. They have a high content of cytotoxic proteins stored in cytoplasmic granules, they are potent pro-

ducers of reactive oxygen species, and they may also produce nitric oxide (NO) (35).

In the present study, we investigated the antibacterial activity of eosinophils against *Escherichia coli* and tried to define the contribution of different cellular antibacterial systems.

MATERIALS AND METHODS

L-N⁵-(1-iminoethyl)-ornithine dihydrochloride (L-NIO) and diphenylene iodonium chloride (DPI) were from Calbiochem, La Jolla, Calif. Recombinant human interleukin-5 (IL-5) was obtained from Pharmingen, San Diego, Calif. Griess reagent, L-arginine, cytochalasin B, phorbol 12-myristate 13-acetate (PMA), platelet-activating factor (PAF), and 2',7'-dichlorodihydrofluorescein diacetate (DCF) were purchased from Sigma, St. Louis, Mo. Lipopolysaccharide (LPS), from *E. coli* strain O55:B5, was obtained from Difco, Detroit, Mich. Dulbecco's modified Eagle's medium without glutamine and phenol red (DMEM) was purchased from ICN, Costa Mesa, Calif. Hanks' balanced salt solution with calcium and magnesium (HBSS) was from Gibco, Paisley, Scotland. Nitrate reductase and NADH were obtained from Cayman Chemical, Ann Arbor, Mich. Boron dipyrromethan (BODIPY)-labeled *E. coli* strain K-12 was from Molecular Probes Europe BV, Leiden, The Netherlands.

Bacteria. The *E. coli* strain ATCC 25922 4Q was obtained from the American Tissue Type Collection, Rockville, Md., and cultured on blood agar plates. Single colonies were picked and grown overnight at 35°C in tryptic soy broth. The bacteria were then centrifuged at 1,000 × *g* for 10 min, transferred to new nutrient broth, and allowed to grow for 2 h to be in log phase before use in experiments.

Isolation of eosinophils. Citrated blood was obtained from healthy volunteers after informed consent had been given, and eosinophils were isolated essentially as described previously (20). In short, after isolation of granulocytes on Ficoll-Paque (Pharmacia, Uppsala, Sweden), immunomagnetic beads coated with antibodies to CD16 (Miltenyi, Gladbach, Germany) were used to retrieve the neutrophils in a magnetic column, allowing the isolation of highly purified eosinophils. The purity of eosinophils was more than 96%; contaminating cells were neutrophils and lymphocytes, as judged by routine May-Grünwald-Giemsa staining. The cell viability was more than 98%, as judged by trypan blue exclusion.

* Corresponding author. Mailing address: Department of Medical Microbiology, Entrance 78, Malmö University Hospital, SE-205 02 Malmö, Sweden. Phone: 46 40 33 13 50. Fax: 46 40 33 62 34. E-mail: Arne.Egesten@medforsk.mas.lu.se.

Acid extraction of proteins from eosinophils. Pellets of frozen eosinophils were resuspended in ice-cold water and subjected to intermittent sonication in a water bath in the presence of ice for 10 s at 40 W until no cells could be seen settling to the bottom of the tube. The lysate was extracted with 0.16 N H₂SO₄ and kept on ice for 30 min with intermittent vortexing. After centrifugation at 23,000 × g for 20 min to sediment insoluble material, the recovered supernatant was dialyzed against 1 mM Tris-HCl (pH 7.4) until the pH of the dialysate was ≥7.0. Thereafter, the extracted protein was dialyzed against 1 mM Tris-HCl-150 mM NaCl at pH 7.4 in dialysis cassettes (Slide-A-Lyzer; Pierce, Rockford, Ill.) and stored at 4°C until use. The protein content was determined using a colorimetric assay based on bicinchoninic acid (BCA protein assay reagent; Pierce).

Incubation of bacteria with granulocytes. Bacteria were opsonized for 20 min at 37°C with 5% human pooled serum from 10 donors. This concentration of serum was not bactericidal. The bacteria were then centrifuged, washed, and resuspended in phosphate-buffered saline (PBS) (10⁹/ml), kept on ice and finally added to the experimental samples. Granulocytes (10⁷ in 0.1 ml of HBSS) and bacteria (10⁷ in 10 μl of PBS) were mixed with HBSS to a final volume of 0.5 ml. The cell suspensions were incubated at 37°C, with end-over-end turning. In some experiments, cells were preincubated with cytochalasin B (5 μg/ml) to disrupt the cytoskeleton, thereby inhibiting phagocytosis of bacteria.

Antibacterial assay. The bactericidal activity of eosinophils was determined by measuring the effect on bacterial colony-forming activity as described previously (28). From bacterial suspensions incubated with or without cells, aliquots were taken at the indicated time intervals and serially diluted in sterile ice-cold PBS. A 30-μl aliquot of the diluted sample was transferred to 5 ml of 1.3% (wt/vol) molten agar maintained at 48°C with 0.8% (wt/vol) nutrient broth and 0.5% (wt/vol) NaCl and poured into a petri dish. The temperature of the agar causes disruption of the granulocytes and release of intracellular bacteria. The agar was allowed to solidify, and the number of CFU was determined on the plates after overnight incubation at 35°C.

In some experiments, the agar was supplemented with 1 mg of bovine serum albumin/ml to allow sublethally injured bacteria to grow (29).

Incubation of bacteria with eosinophils under aerobic and anaerobic conditions. Cell suspensions were preincubated for 30 min at 37°C in an anaerobe box containing 10% H₂, 10% CO₂, and 80% N₂. In parallel, cells were incubated with bacteria in a normal atmospheric environment at the same temperature.

To investigate the function of the NADPH oxidase, cells were incubated with DCF (5 μM) under anaerobic or aerobic conditions and also moved from anaerobic to aerobic conditions or incubated with DPI followed by stimulation with PMA (100 ng/ml). The cells were then put on ice and fixed with formaldehyde to a final concentration of 1% (wt/vol) before measurement of the mean fluorescence intensity by flow cytometry using the fluorescein isothiocyanate channel.

DCF is a cell-permeative nonfluorescent probe that is de-esterified intracellularly and converts into highly fluorescent DCF upon oxidation (36). In short, a stem solution of DCF (5 mM) was prepared fresh every day by dissolving the substance in 95% (vol/vol) ethanol. This solution was kept in the dark at room temperature. Eosinophils at 10⁶/ml in HBSS were incubated with DCF (5 μM) for 15 min at 37°C before use in the experiments.

NO production. L-NIO was used to inhibit NO synthase (37). L-NIO (10 mM in PBS; final concentration of 100 μM) was added to the cells suspended in HBSS supplemented with 0.6 mM L-arginine. Bacterial incubations and viable counts were performed under aerobic conditions at 37°C. L-NIO incubated with bacteria alone did not affect the viability of the bacteria.

Eosinophils were incubated in DMEM (2 × 10⁶ cells/ml) at 37°C for 24 h in the absence or presence of IL-5 (1 nM) to investigate their NO production. The supernatants were recovered, and proteins were precipitated with ZnSO₄ (30% [wt/vol]) before measurement of NO, as determined by the Griess reaction, described below. The viability of eosinophils after 24 h of incubation was determined by trypan blue exclusion and was >96% in medium alone and >98% in the presence of IL-5.

As a positive control of NO production and its inhibition by L-NIO, rat aorta was used. After institutional approval was obtained, male Sprague-Dawley rats (200 to 250 g of body weight) were anesthetized with halothane and exsanguinated. The descending thoracic aorta was removed and cut into circular segments weighing 2.5 to 4 g. The segments were each incubated in 1 ml of DMEM. LPS was added to a final concentration of 100 ng/ml. Two segments were incubated without LPS and served as controls. L-NIO (100 μM) was added to one of the segments incubated with LPS. The segments were incubated with gentle agitation for 24 h at 37°C in an atmosphere containing 8% carbon dioxide in air. After incubation, the segments were removed, blotted on filter paper, and weighed. The DMEM was transferred to clean vials and centrifuged for 5 min at 11,000 ×

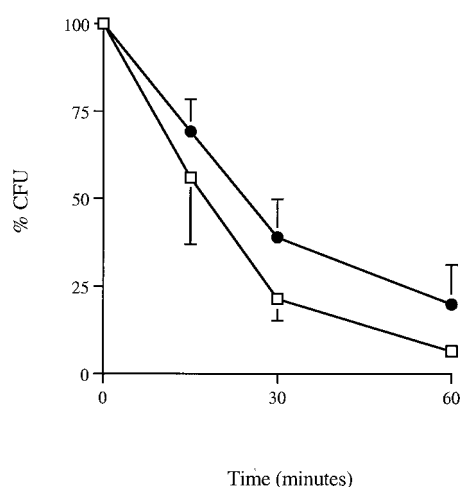


FIG. 1. Killing of *E. coli* by eosinophils under aerobic conditions at ratios of 1:1 (□) and 5:1 (●) as measured by viable counts. At the indicated time points, aliquots of the bacterial cell suspension were taken and assayed for viable bacteria. Bacterial viability is expressed as percent survival in the presence of eosinophils, where 100% represents the number of colonies formed at the indicated time points in the absence of eosinophils. Results are means ± SEMs of four independent experiments.

g at 4°C to remove cellular debris and stored at -20°C until assayed for nitrate and nitrite.

Nitrate and nitrite levels, reflecting NO production, were determined by the Griess reaction. In short, 100 μl of supernatant from duplicate incubations was transferred to the wells of a transparent microplate. Nitrate reductase and NADH were added, and the plate was incubated at room temperature for 2 h. Then, 100 μl of Griess reagent (40 mg/ml) was added, and absorbance was measured at 562 nm and compared with standards prepared by diluting sodium nitrate in DMEM. The detection limit for this assay of nitrite in DMEM was 750 nM.

Determination of phagocytosis. Eosinophils (10⁶/ml) were incubated with serum-opsonized BODIPY-labeled *E. coli* at a ratio of 1:100. The serum opsonization was performed as described above. Incubations were performed under both aerobic and anaerobic conditions at 37°C, in medium alone or after addition of IL-5 (1 nM) or PAF (1 μM). Aliquots were taken after 15, 30, and 60 min, and eosinophil binding of bacteria was determined using an Epics XL-MCL flow cytometry system (Beckman Coulter Inc., Fullerton, Calif.). Eosinophils were gated using their characteristics in side and forward scatter, excluding unbound bacteria. Fluorescence from extracellular bacteria was quenched by the addition of trypan blue. The percent intracellular bacteria was calculated by using the following formula: [mean fluorescence intensity after quenching/(initial mean fluorescence intensity - eosinophil autofluorescence)].

RESULTS

Killing of *E. coli*. The killing of *E. coli* by eosinophils was measured by viable count technique (Fig. 1). The killing of the bacteria reached a maximum after 1 h of incubation. Addition of the eosinophil-activating agents PAF (1 μM) or IL-5 (1 nM) (4, 26) did not further increase the killing efficiency (data not shown). The vast majority of bacteria were killed and not sublethally injured (data not shown).

Aerobic versus anaerobic conditions. Anaerobic conditions and pharmacological uncoupling of the NADPH oxidase under aerobic conditions reduced eosinophil killing of *E. coli* significantly compared with incubation of eosinophils and bacteria under aerobic conditions (Fig. 2a and b). These findings suggest that superoxide formed by NADPH oxidase is important for bacterial killing in this system.

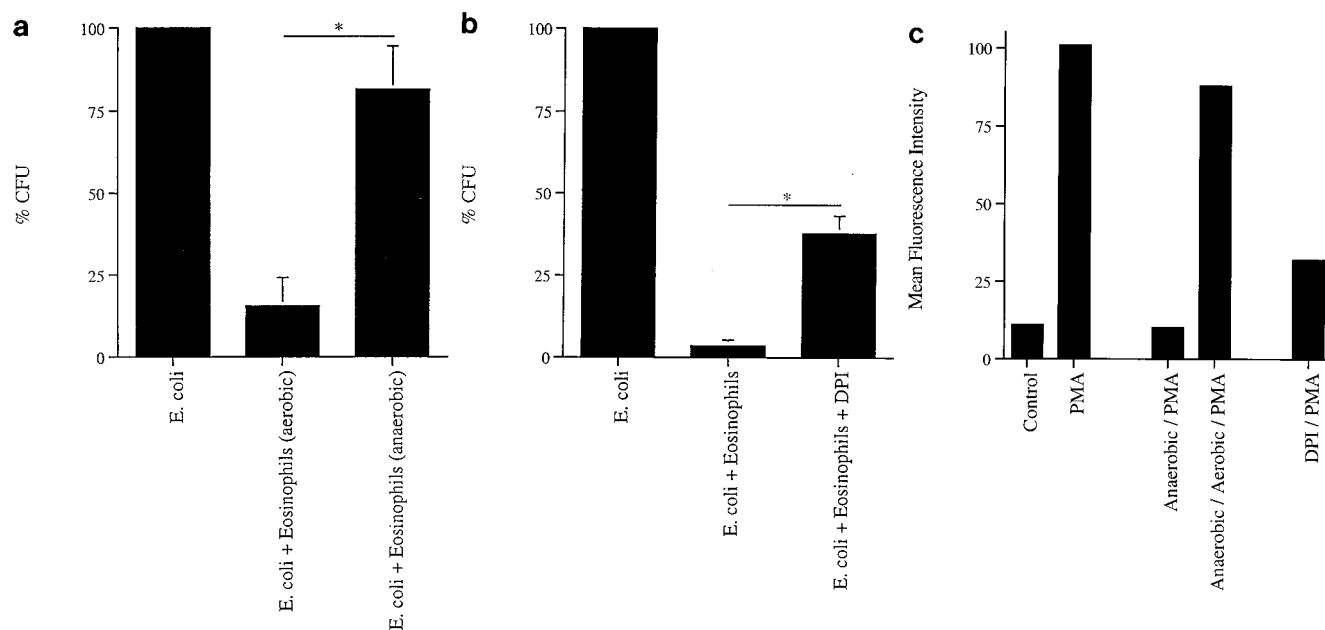


FIG. 2. Killing of *E. coli* by eosinophils is O_2 dependent. (a) Eosinophils were preincubated under aerobic and anaerobic conditions for 30 min before the addition of *E. coli*. After 60 min of incubation, viable bacteria were counted. Results are means \pm SEMs of four independent experiments. *, $P < 0.05$ (Student's *t* test for paired observations). (b) Pharmacological uncoupling of NADPH oxidase by DPI. Eosinophils were preincubated at 37°C for 10 min in the absence or presence of DPI (10 μ M) before incubation with *E. coli* in the continued absence or presence of DPI for 60 min. *, $P < 0.05$ (Student's *t* test for paired observations). Results are means plus SEMs of five independent experiments. (c) EPO activity as determined by DCF. Eosinophils were loaded with DCF and incubated under aerobic or anaerobic conditions for 30 min, under anaerobic followed by aerobic conditions for 30 min each, or in the presence of DPI (10 μ M) under aerobic conditions for 30 min; cells were subsequently stimulated with PMA (100 ng/ml) for 15 min. The resulting mean fluorescence intensity was determined by flow cytometry and reflects the oxidase activity in the cells. Results of one representative independent experiment out of three are shown.

To determine the efficiency of anaerobic conditions and DPI in inhibiting the oxidase activity, cells were loaded with DCF, and the oxidase-dependent fluorescence was measured (13) (Fig. 2c). PMA was used to induce a respiratory burst in the cells. Anaerobic conditions and DPI both decreased the PMA-induced respiratory burst. When the cells were first incubated for 30 min under anaerobic conditions and then subjected to aerobic conditions for 30 min before stimulation with PMA, a respiratory burst almost of the order of that in stimulated cells held under aerobic conditions was seen. This shows that decreased killing during anaerobic conditions was not attributable to poor viability of the cells.

Inhibition of NO synthesis. No significant reduction in eosinophil killing of *E. coli* was seen in the presence of the NO synthesis inhibitor L-NIO (Fig. 3a). This suggests that the generation of NO does not contribute to the killing of bacteria in this system. No NO production could be detected from resting or IL-5-stimulated eosinophils after 24 h of incubation, as determined by the Griess reaction (five separate experiments; data not shown). However, both constitutive and LPS-stimulated NO production from segments of rat aorta was inhibited by the same concentration of L-NIO (Fig. 3b).

Antibacterial activity of eosinophil protein extracts. Protein extracts from purified eosinophils killed bacteria in a dose-dependent manner (Fig. 4). The 50% effective dose was approximately 3 μ g/ml after 60 min of incubation with the bacteria.

Involvement of superoxide and eosinophil peroxidase (EPO) in bacterial killing. EPO was inhibited by the addition of azide

(5 mM) (Fig. 5). Azide at this concentration did not kill bacteria itself, nor did it affect the viability of eosinophils as judged by trypan blue exclusion. The presence of azide significantly decreased the killing of *E. coli*, whereas the subsequent addition of DPI did not further decrease the killing.

Phagocytosed versus bacteria bound to the cell surface. To determine whether the bacteria were phagocytosed or bound to the surface of eosinophils, *E. coli* cells labeled with the fluorescent dye BODIPY were used. After 60 min of incubation under aerobic conditions, 11.6% \pm 3.2% (mean \pm standard error of the mean [SEM] for three separate experiments) of the bacteria were phagocytosed, as determined by quenching of extracellular BODIPY by trypan blue. There was no difference between the numbers of phagocytosed bacteria during aerobic and anaerobic incubation.

Treatment of eosinophils with cytochalasin B, an agent that disrupts the cytoskeleton and thereby inhibits phagocytosis, did not affect the bacterial killing efficiency of eosinophils (data not shown).

Taken together, these data suggest that the bacterial killing occurs mainly on the surface of the cells.

DISCUSSION

In the present study we show that eosinophils can kill *E. coli* and that the killing, to a large degree, is oxygen dependent under the conditions used. We also show that there is oxygen-independent bactericidal activity in eosinophil protein extracts.

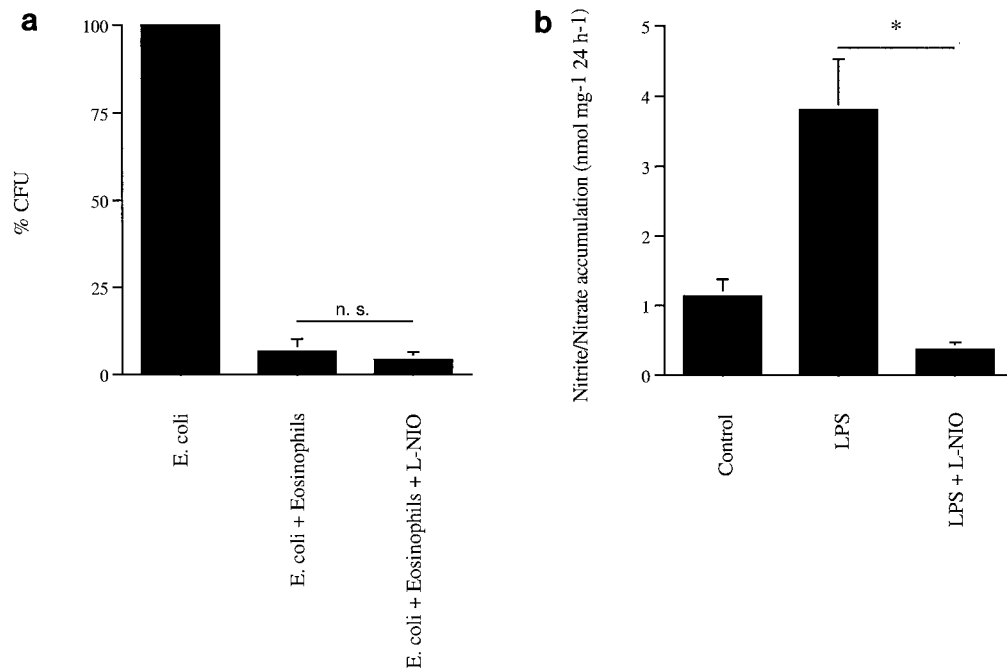


FIG. 3. Killing of *E. coli* by eosinophils during inhibited NO production. (a) Eosinophils were incubated with bacteria in the absence or presence of the NO synthase inhibitor L-NIO (100 μ M). Means \pm SEMs from three independent experiments are shown. n. s., not significant. (b) The efficiency of L-NIO (100 μ M) in inhibiting NO synthase and thereby NO production was determined by measurement of the NO metabolites NO_2^- and NO_3^- by the Griess reaction. Segments of rat thoracic aorta were incubated in the absence or presence of LPS (100 ng/ml) and in the presence of LPS plus L-NIO (100 μ M). Results are means plus SEMs from five independent experiments. *, $P < 0.05$ (Student's *t* test for paired observations).

Furthermore, our data suggest that the bactericidal activity of eosinophils is dependent on the interaction between superoxide generated by the NADPH oxidase and the granule-bound peroxidase EPO.

Several studies have shown that eosinophils are less capable of killing bacteria than neutrophils (2, 10, 32). However, many of these were performed at a time when it was difficult to

obtain cells of high purity. In addition, some data regarding the bactericidal capacities of eosinophils are conflicting. Some investigators have reported that eosinophils are as efficient phagocytes as are neutrophils (11), especially after stimulation by certain cytokines (3). More recently, it has been shown that

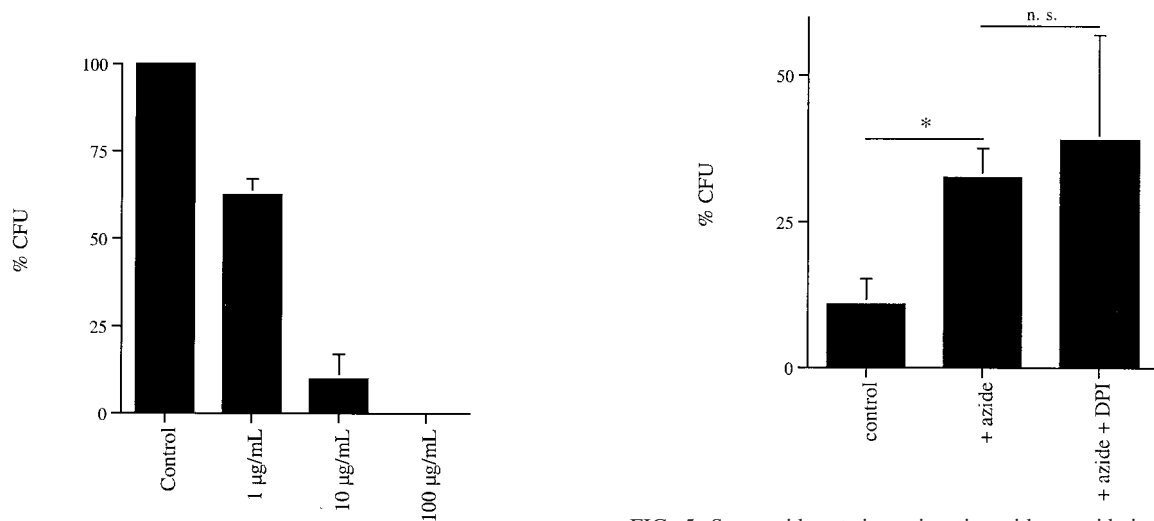


FIG. 4. Presence of bactericidal activity against *E. coli* in eosinophil crude protein extracts. Bacteria were incubated with extracts at the indicated concentrations for 60 min. Viable bacteria were assayed by viable counts.

FIG. 5. Superoxide acts in conjunction with peroxide in eosinophil killing of *E. coli*. Eosinophils were incubated with *E. coli* in medium alone (control), after addition of azide (5 mM), and in the presence of the combination of azide (5 mM) and DPI (5 μ M). *, $P < 0.05$; n. s., not significant (Student's *t* test for paired observations).

they can, at least under certain circumstances, show bactericidal activities comparable to those of neutrophils (17, 38).

The microbicidal systems of granulocytes can be divided into oxygen-dependent and -independent systems (15, 24). The production of superoxide by the membrane-bound NADPH oxidase, of eosinophils is three to six times greater than that of neutrophils (12). The product of the oxidase system, superoxide, seems to have limited cytotoxicity, and the hydrogen peroxide that is formed by dismutation of superoxide is bactericidal only at high concentrations (19). Eosinophils are exceptionally rich in peroxidase, e.g., EPO, which is present in the matrix of eosinophil granules at 15 $\mu\text{g}/10^6$ eosinophils (9, 35). In systems using purified EPO, superoxide, and hydrogen peroxide, the generation of potent cytotoxic species such as the hydroxyl radical in eosinophils has been observed (31). When combined with H_2O_2 and halides (for example, chloride, iodide, or bromide), EPO catalyzes the generation of highly cytotoxic hypochlorous acids (22). Recently, thiocyanate was suggested to be an important substrate in this reaction, generating hypothiocyanite and cyanate (1). Our findings obtained by using azide to block the heme group of EPO and DPI to block the NADPH oxidase system suggest that the oxidase system acting in conjunction with EPO is important for the bactericidal activity of eosinophils. This finding is in agreement with the finding that myeloperoxidase and NADPH oxidase act in conjunction in neutrophils (18), but it is in contrast to an earlier report showing that EPO did not contribute to the bactericidal activity of intact eosinophils (7).

The bactericidal activity of EPO is pH dependent (22). During incubation, the pH of the medium decreased, but there was no difference between the pHs of the media under aerobic and anaerobic conditions (data not shown), arguing against the possibility that such an effect contributed to the difference in bacterial killing.

It has been suggested that eosinophils can produce NO through a constitutive expression of inducible NO synthase (14). NO can react with superoxide and form peroxynitrate, which possesses potent cytotoxic properties against, for example, *E. coli* (6). Therefore, eosinophil NO production together with superoxide production by the oxidase system may present a powerful source of cytotoxicity (33). In our experiments, however, we were not able to detect NO production by the eosinophils, nor did we see any effects of the inducible NO synthase inhibitor L-NIO on bacterial killing by eosinophils. The reason for this could be the source of cells or the means of detection used. These methodological problems have been reported in experiments with neutrophils (37).

We found potent bactericidal activity in protein extracts from eosinophils. The oxygen-independent antibacterial effects of eosinophils may be attributable to several granule-bound proteins that have, in purified forms, been shown to possess bactericidal effects (25). The characteristic proteins making up the bulk content of eosinophil granules are the highly cationic proteins eosinophil cationic protein (ECP), EPO, eosinophil-derived neurotoxin (EDN), and major basic protein. ECP and eosinophil-derived neurotoxin possess RNase activity, but the bactericidal activity of at least ECP seems to be independent of its RNase activity (34). The effect of ECP on bacteria may instead come from its pore formation (40). Interestingly, the cytotoxicity of ECP can be potentiated by the presence of a

functioning oxidase system (39), but the mechanism behind this has not been characterized.

Other granule proteins possessing antibacterial activity have been reported to be present in eosinophil granules. Among these are the 14-kDa secretory phospholipase A_2 (5), the bactericidal/permeability-increasing protein (8), and the bacterial cell wall-degrading enzyme *N*-acetylmuramyl-L-alanine amidase (21). The contribution of each of these is difficult to determine, but together they could represent a powerful and, to a large degree, oxygen-independent bactericidal system. They may also be important for bactericidal activities after release to the extracellular environment.

Since IL-5 is important for eosinophil differentiation, increased susceptibility to infection in IL-5 knockout mice could be expected. Surprisingly, IL-5-deficient mice still produce basal levels of eosinophils that appear to be morphologically and functionally normal (30). In the case of host defense against parasitic infestation, the basal levels of eosinophils has been shown to compromise defense against several helminths. In addition, IL-5 deficient mice also appear to have functional deficiencies in B-1 B lymphocytes and in immunoglobulin A production (30). To our knowledge, the susceptibility to bacterial infection of IL-5 knockout mice has not been investigated. Furthermore, the regulation of the immune systems in mice is different at several levels from that in humans, and therefore firm conclusions with regard to the roles of eosinophils in mice and humans may be difficult to make.

Our study suggests that eosinophils can be efficient in host defense against invasion by gram-negative bacteria and that oxygen-dependent killing, i.e., superoxide acting in conjunction with EPO, may be the most important bactericidal effector function of these cells. This can be important at mucosal interfaces and in the mucosa of, for example, the large intestine, where the conditions are aerobic.

ACKNOWLEDGMENTS

This work was supported by grants from the Th C Bergh Foundation, the Magnus Bergvall Foundation, the Gorthon Foundation, the Greta & Johan Kock Foundations, the Malmö General Hospital Cancer Foundation, the Tore Nilsson Foundation, the Swedish Asthma and Allergy Association's Research Foundation, the Swedish Society of Medicine, the Zoéga Foundation, and the Alfred Österlund Foundation.

REFERENCES

1. Arlandson, M., T. Decker, V. A. Roongta, L. Bonilla, K. H. Mayo, J. C. MacPherson, S. L. Hazen, and A. Slungaard. 2001. Eosinophil peroxidase oxidation of thiocyanate: characterization of major reaction products and a potential sulfhydryl-targeted cytotoxicity system. *J. Biol. Chem.* **276**:215–224.
2. Baehner, R. L., and R. B. J. Johnston. 1974. Metabolic and bactericidal activities of human eosinophils. *Br. J. Haematol.* **20**:277–285.
3. Blom, M., A. T. Tool, P. T. Kok, L. Koenderman, D. Roos, and A. J. Verhoeven. 1994. Granulocyte-macrophage colony-stimulating factor, interleukin-3 (IL-3), and IL-5 greatly enhance the interaction of human eosinophils with opsonized particles by changing the affinity of complement receptor type 3. *Blood* **83**:2978–2984.
4. Blom, M., A. T. Tool, D. Roos, and A. J. Verhoeven. 1992. Priming of human eosinophils by platelet-activating factor enhances the number of cells able to bind and respond to opsonized particles. *J. Immunol.* **149**:3672–3677.
5. Blom, M., A. T. Tool, P. C. Wever, G. J. Wolbink, M. C. Brouwer, J. Calafat, A. Egesten, E. F. Knol, C. E. Hack, D. Roos, and A. J. Verhoeven. 1998. Human eosinophils express, relative to other circulating leukocytes, large amounts of secretory 14-kD phospholipase A_2 . *Blood* **91**:3037–3043.
6. Brunelli, L., J. P. Crow, and J. S. Beckman. 1995. The comparative toxicity of nitric oxide and peroxynitrite to *Escherichia coli*. *Arch. Biochem. Biophys.* **316**:327–334.
7. Bujak, J. S., and R. K. Root. 1974. The role of peroxidase in the bactericidal

- activity of human blood eosinophils. *Blood* **43**:727–736.
8. Calafat, J., H. Janssen, A. Tool, M. A. Dentener, E. F. Knol, H. F. Rosenberg, and A. Egesten . 1998. The bactericidal/permeability-increasing protein (BPI) is present in specific granules of human eosinophils. *Blood* **91**:4770–4775.
 9. Carlson, M., C. G. Peterson, and P. Venge. 1985. Human eosinophil peroxidase: purification and characterization. *J. Immunol.* **134**:1875–1879.
 10. Cline, M. J. 1972. Microbicidal activity of human eosinophils. *J. Reticuloendothel. Soc.* **12**:332–339.
 11. DeChatelet, L. R., R. A. Migler, P. S. Shirley, H. B. Muss, P. Szejda, and D. A. Bass. 1978. Comparison of intracellular bactericidal activities of human neutrophils and eosinophils. *Blood* **52**:609–617.
 12. DeChatelet, L. R., P. S. Shirley, L. C. McPhail, C. C. Huntley, H. B. Muss, and D. A. Bass. 1977. Oxidative metabolism of the human eosinophil. *Blood* **50**:525–535.
 13. DeLeo, F. R., L. A. Allen, M. Apicella, and W. M. Nauseef. 1999. NADPH oxidase activation and assembly during phagocytosis. *J. Immunol.* **163**:6732–6740.
 14. Del Poso, V., E. de Arruda-Chaves, B. de Andrés, B. Cárdbaba, A. López-Farré, S. Gallardo, I. Cortegano, L. Vidarte, A. Jurado, J. Sastre, P. Palomino, and C. Lahoz. 1997. Eosinophils transcribe and translate messenger RNA for inducible nitric oxide synthase. *J. Immunol.* **158**:859–864.
 15. Elsbach, P., J. Weiss, and O. Levy. 1999. Oxygen-independent antimicrobial systems of phagocytes, p. 801–817. *In* J. I. Gallin and R. Snyderman (ed.), *Inflammation: basic principles and clinical correlates*. Lippincott Williams & Wilkins Co., Baltimore, Md.
 16. Erle, D. J., M. J. Briskin, E. C. Butcher, A. Garcia-Pardo, A. I. Lazarovits, and M. Tidswell. 1994. Expression and function of the MAdCAM-1 receptor, integrin alpha 4 beta 7, on human leukocytes. *J. Immunol.* **153**:517–528.
 17. Fabian, I., Y. Kletter, S. Mor, C. Geller-Bernstein, M. Ben-Yaakov, B. Volovitz, and D. W. Golde. 1992. Activation of human eosinophil and neutrophil functions by haematopoietic growth factors: comparisons of IL-1, IL-3, IL-5 and GM-CSF. *Br. J. Haematol.* **80**:137–143.
 18. Hampton, B. M., A. J. Kettle, and C. C. Winterbourn. 1996. Involvement of superoxide and myeloperoxidase in oxygen-dependent killing of *Staphylococcus aureus* by neutrophils. *Infect. Immun.* **64**:3512–3517.
 19. Hampton, B. M., A. J. Kettle, and C. C. Winterbourn. 1998. Inside the neutrophil phagosome: oxidants, myeloperoxidase, and bacterial killing. *Blood* **92**:3007–3017.
 20. Hansel, T. T., I. J. M. De Vries, T. Iff, S. Rihs, M. Wandzilak, K. Blaser, and C. Walker. 1991. An improved immunomagnetic procedure for isolation of highly purified human blood eosinophils. *J. Immunol. Methods* **145**:105–110.
 21. Hoijer, A. M., M. J. Melief, J. Calafat, D. R. Roos, W. van den Beemd, J. J. van Dongen, and M. P. Hazenberg. 1997. Expression and intracellular localization of the human N-acetylmuramyl-L-alanine amidase, a bacterial cell wall-degrading enzyme. *Blood* **90**:1246–1254.
 22. Jong, E. C., W. R. Henderson, and S. J. Klebanoff. 1980. Bactericidal activity of eosinophil peroxidase. *J. Immunol.* **124**:1378–1382.
 23. Kato, M., G. M. Kephart, N. J. Talley, J. M. Wagner, M. G. Sarr, M. Bonno, T. W. McGovern, and G. J. Gleich. 1998. Eosinophil infiltration and degranulation in normal human tissue. *Anat. Rec.* **252**:418–425.
 24. Klebanoff, S. J. 1999. Oxygen metabolites from phagocytes, p. 721–768. *In* J. I. Gallin and R. Snyderman (ed.), *Inflammation: basic principles and clinical correlates*. Lippincott Williams & Wilkins Co., Baltimore, Md.
 25. Lehrer, R. I., D. Szklarek, A. Barton, T. Ganz, K. J. Hammann, and G. J. Gleich. 1989. Antibacterial properties of major basic protein and eosinophil cationic protein. *J. Immunol.* **142**:4428–4434.
 26. Lopez, A. F., C. J. Sanderson, J. R. Gamble, H. D. Campbell, I. G. Young, and M. A. Vadas. 1988. Recombinant human interleukin 5 is a selective activator of human eosinophil function. *J. Exp. Med.* **167**:219–224.
 27. Makiyama, K., S. Kanzaki, K. Yamasaki, W. Zee-Iriarte, and Y. Tsuji. 1995. Activation of eosinophils in the pathophysiology of ulcerative colitis. *J. Gastroenterol.* **30**:64–69.
 28. Mandic-Mulec, I., J. Weiss, and A. Zychlinsky. 1997. *Shigella flexneri* is trapped in polymorphonuclear leukocyte vacuoles and efficiently killed. *Infect. Immun.* **65**:110–115.
 29. Mannion, B. A., J. Weiss, and P. Elsbach. 1990. Separation of sublethal and lethal effects of polymorphonuclear leukocytes on *Escherichia coli*. *J. Clin. Investig.* **86**:631–641.
 30. Matthaei, K. I., P. Foster, and I. G. Young. 1997. The role of interleukin-5 (IL-5) in vivo: studies with IL-5 deficient mice. *Mem. Inst. Oswaldo Cruz* **92**:63–68.
 31. McCormick, M. L., T. L. Roeder, M. A. Railsback, and B. E. Britigan. 1994. Eosinophil peroxidase-dependent hydroxyl radical generation by human eosinophils. *J. Biol. Chem.* **269**:27914–27919.
 32. Mickenberg, I. D., R. K. Root, and S. M. Wolff. 1972. Bactericidal and metabolic properties of human eosinophils. *Blood* **39**:67–80.
 33. Oliveira, S. H., S. G. Fonseca, P. R. Romao, F. Figueiredo, S. H. Ferreira, and F. Q. Cunha. 1998. Microbicidal activity of eosinophils is associated with activation of the arginine-NO pathway. *Parasite Immunol.* **20**:405–412.
 34. Rosenberg, H. F. 1995. Recombinant human eosinophil cationic protein. Ribonuclease activity is not essential for cytotoxicity. *J. Biol. Chem.* **270**:7876–7881.
 35. Rosenberg, H. F. 1999. The eosinophil, p. 61–77. *In* J. I. Gallin and R. Snyderman (ed.), *Inflammation: basic principles and clinical correlates*. Lippincott Williams & Wilkins Co., Baltimore, Md.
 36. Rosenkranz, A. R., S. Schmaldienst, K. M. Stuhlmeier, W. Chen, W. G. Knapp, and J. Zlabinger. 1992. A microplate assay for the detection of oxidative products using 2',7'-dichlorofluorescein-diacetate. *J. Immunol. Methods* **156**:39–45.
 37. Wheeler, A. M., S. D. Smith, G. Garcia-Cardena, C. F. Nathan, R. M. Weiss, and W. C. Sessa. 1997. Bacterial infection induces nitric oxide synthase in human neutrophils. *J. Clin. Investig.* **99**:110–116.
 38. Yazdanbakhsh, M., C. M. Eckmann, A. A. Bot, and D. Roos. 1986. Bactericidal action of eosinophils from normal human blood. *Infect. Immun.* **53**:192–198.
 39. Yazdanbakhsh, M., P.-C. Tai, C. J. F. Spry, G. J. Gleich, and D. Roos. 1987. Synergism between eosinophil cationic protein and oxygen metabolites in killing of schistosomula of *Schistosoma mansoni*. *J. Immunol.* **138**:3443–3447.
 40. Young, J. D., C. G. Peterson, P. Venge, and Z. A. Cohn. 1986. Mechanism of membrane damage mediated by human eosinophil cationic protein. *Nature* **321**:613–616.

Editor: A. D. O'Brien