Role of p38 Mitogen-Activated Protein Kinase in Middle Ear Mucosa Hyperplasia during Bacterial Otitis Media

Palacios, Sean D.; Pak, Kwang; Rivkin, Alexander Z; Kayali, Ayse G; Austen, Darrell; Aletsee, Christoph; Melhus, Asa; Webster, Nicholas J G; Ryan, Allen F.

Published in:
Infection and Immunity

DOI:
10.1128/IAI.72.8.4662-4667.2004

2004

Link to publication

Citation for published version (APA):

Total number of authors:
9
Role of p38 Mitogen-Activated Protein Kinase in Middle Ear Mucosa Hyperplasia during Bacterial Otitis Media

Sean D. Palacios,† Kwang Pak,‡ Alexander Z. Rivkin, Ayse G. Kayali, Darrell Austen, Christoph Aletsee, Åsa Melhus, Nicholas J. G. Webster, and Allen F. Ryan

Departments of Surgery/Otolaryngology and Medicine, University of California, San Diego, and VA Medical Center, La Jolla, California; Department of Otolaryngology, University of Wuerzburg, Wuerzburg, Germany; and Department of Microbiology, University of Lund, Lund, Sweden

Received 20 February 2004/Returned for modification 30 March 2004/Accepted 22 April 2004

Hyperplasia of the middle ear mucosa contributes to the sequelae of acute otitis media. Understanding the signal transduction pathways that mediate hyperplasia could lead to the development of new therapeutic interventions for this disease and its sequelae. Endotoxin derived from bacteria involved in middle ear infection can contribute to the hyperplastic response. The p38 mitogen-activated protein kinase (MAPK) is known to be activated by endotoxin as well as cytokines and other inflammatory mediators that have been documented in otitis media. We assessed the activation of p38 in the middle ear mucosa of an in vivo rat bacterial otitis media model. Strong activity of p38 was observed 1 to 6 h after bacterial inoculation. Activity continued at a lower level for at least 7 days. The effects of p38 activation were assessed using an in vitro model of rat middle ear mucosal hyperplasia in which mucosal growth is stimulated by nontypeable Haemophilus influenzae during acute otitis media. Hyperplastic mucosal explants treated with the p38 inhibitor SB203580 demonstrated significant inhibition of otitis media-stimulated mucosal growth. The results of this study suggest that intracellular signaling via p38 MAPK influences the hyperplastic response of the middle ear mucosa during bacterial otitis media.

Pathologic conditions related to otitis media include middle ear effusion, adhesive otitis, tympanic membrane perforation, tympanosclerosis, scarring of the middle ear mucosa, cholesteatoma, and atelectasis. These sequelae can lead to permanent damage of the middle ear cavity, resulting in hearing loss or impairment. A contributing factor to such morbidity during otitis media is the transformation of the inflamed middle ear mucosa. Normally, the middle ear mucosa consists of a simple squamous epithelium overlying a thin lamina propria that adheres to the underlying peristomeum. This minimal mucosa ranges from 15 to 20 μm thick. During otitis media the middle ear mucosa has the unique capacity to grow and proliferate to many times its original thickness, into a highly structured, pseudostratified, columnar epithelial complex. Hyperplasia of the mucosa is associated with many of the negative sequelae of middle ear infection. Production of mucus and other elements of middle ear effusions is closely linked to the production of additional mucosal cells and their transformation into a secretory phenotype. Expansion of the mucosa vasculature provides increased opportunity for leukocytic infiltration and tissue edema, while increases in stromal cells with their subsequent activation can lead to the generation of matrix components and fibrosis. The hyperplastic response of the middle ear mucosa occurs within a few days following any number of otitis media-related stimuli, including tubal obstruction, bacterial infection, endotoxin exposure, immunologic challenge, and exposure to inflammatory mediators such as cytokines. These stimuli are presumably linked to the hyperplastic response via intracellular signal transduction pathways.

In other tissue systems the mitogen-activated protein kinase (MAPK) intracellular signaling pathways, including Erk, Jnk, and p38, have been shown to be linked to a variety of cellular responses, including cellular proliferation. The characteristic of the MAPK family makes them candidates for regulation of mucosal hyperplasia during infection. The p38 pathway in particular is a stress-induced kinase activated by various extracellular stimuli associated with otitis media, including bacterial endotoxin, osmotic shock, and inflammatory cytokines such as tumor necrosis factor and interleukin 1. The presence of endotoxin, elevated levels of inflammatory cytokines, and effusion in the middle ear during bacterial otitis media provides the potential for the activation of this pathway. In middle ear epithelial cell lines, p38 activation has been linked to the expression of genes encoding cytokines and mucin.

The p38 MAPK intracellular signaling pathway is the most recently characterized and least understood MAPK pathway. The intermediate effector molecules between initial extracellular stimulation by endotoxin or cytokines and downstream p38 activity have yet to be completely identified. It is known that p38 activation is carried out by a dual phosphorylation on Thr-180 and Tyr-182 within the Thr-Gly-Tyr motif located in subdomain VIII (12, 40). It is also known that there are four...
p38 isoforms termed α (38,000 Da), β (39,000 Da), γ (39,000 Da), and δ (38,000 Da) (8, 18). Specificity of isoform phosphorylation is believed to be partially responsible for p38’s diverse functional influence upon cellular physiology (13, 29). The functional variability of p38 makes it a potential agent in a number of middle ear mucosal changes that occur during otitis media.

Whether the p38 intracellular signaling pathway is activated during otitis media and what role it may play in the hyperplastic response of the middle ear mucosa have not been investigated. For this reason, we explored the presence of activated p38 at different postinoculation time points in a rat in vivo otitis media model. We also developed an in vitro model of hyperplastic middle ear mucosa and explored the effects of the p38α and p38β inhibitor, SB203580, on bacterially induced mucosal growth. The combined data from this study support multiple roles for p38 MAPK signaling in the hyperplastic response of the middle ear mucosa during bacterial otitis media.

**MATERIALS AND METHODS**

All experiments were performed according to National Institutes of Health guidelines on the care and use of laboratory animals and were approved by the institutional committee for animal experimentation.

**Assessment of p38 MAPK phosphorylation.** Male Sprague-Dawley rats weighing between 300 and 350 g were anesthetized with a mixture of ketamine and xylazine hydrochloride (Rompun) at 100 mg/ml and acetylprozine at 10 mg/ml, injected intramuscularly at a dose of 0.4 ml/100 g of body weight. Upon sedation, the animals were placed in the supine position and their bodies stabilized with masking tape. A 3-cm, vertical, midline incision was made between the animal’s mandible and clavicles, and middle ear bullae exposure was obtained bilaterally to re-cover the bulla holes, and the incisions were stapled closed. Each animal served as a control being cultured with DMSO (1 mM). The incisions were immediately placed in a petri dish containing phosphate-buffered saline (PBS). The middle ear mucosae were immediately frozen at −70 °C. The homogenates were centrifuged at 14,000  g at 4 °C, the supernatants were collected, and the protein concentrations were determined using a bicinchoninic acid assay. Each middle ear mucosal sample was immediately placed in a separate 60-mm-diameter Falcon petri dish coated with a thin layer of Sylgard from Dow Corning (Midland, Mich.). Culture medium consisting of a mixture of Dulbecco’s modified Eagle’s medium and Ham’s F-12 medium (3:1) supplemented with fetal calf serum (5%), hydrocortisone (0.4 μg/ml), isoproterenol (10 μM), penicillin (100 U/ml), and streptomycin (100 μg/ml) was then added (52). The middle ear mucosae were divided into roughly 0.5-mm² tissue explants with a Fine Science Tools (Foster City, Calif.) diamond knife. The explants from each bulla were then individually transplanted, epithelium uppermost, into single wells of a 24-well Falcon cell culture plate containing 170 μl of the above culture medium. The explants were then placed in an incubator at 37°C with 5% CO2 for 24 h and allowed to adhere to the culture plate surface. On day one, all wells with healthy, attached explants were randomly divided into four groups. The p38 inhibitor, SB203580 (Calbiochem, San Diego, Calif.), was reconstituted in dimethyl sulfoxide (DMSO) and added to one of three groups at 1, 10, and 100 mM in 400 μl of culture medium. The last group served as a negative control, the media receiving no addition of SB203580 and serving as a control for the DMSO concentration. The data were entered into Statview 5.0 and an analysis of variance was run with a P value criterion of <0.05. The tissue explants were also immunohistochemically stained with an anti-BrdU primary antibody (Sigma-Aldrich, St. Louis, Mo.) alone, the same concentration of DMSO used for all concentrations of SB203580. Each day, the medium from each well was removed and 400 μl of fresh culture medium was added. The presence of the appropriate concentration of SB203580 and/or DMSO was confirmed by testing a sample of tissue explants from each group at day three. The explants from each group were also treated with 5-bromo-2-deoxyuridine (BrdU) (Sigma-Aldrich) every day for the entire duration of the experiment. The explants were kept in culture for ten days.

For each individual concentration of SB203580, six to nine explants were evaluated. Each explant was photographed each day with an RT SPOT color digital camera and their surface areas calculated using SPOT computer software calibrated to the appropriate magnification. The data were entered into Statview 5.0 and an analysis of variance was run with a P value criterion of <0.05. The tissue explants were also immunohistochemically stained with an anti-BrdU primary antibody (Sigma-Aldrich) followed by a biotinylated secondary antibody (Santa Cruz Biotechnologies, Santa Cruz, Calif.). After the appropriate biotin and avidin blocks, the cultures were treated with a fluorescein isothiocyanate-labeled strepavidin (DAKO, Carpinteria, Calif.). Each well was then photographed again under various magnifications with a GFP fluorescent filter.

**Inhibition of noninfected mucosae with SB203580.** The middle ear mucosae of six male Sprague-Dawley rats that had not been infected with H. influenzae were dissected, divided, and cultured in the manner described above. The explants were divided into three groups. The first group was cultured with 10 nM SB203580, the second group was cultured with 100 nM SB203580, and the last group served as a control being cultured with DMSO (1 μM). The fluorescein-stained, DAPI-counterstained explants and tissue sections were examined on an Olympus BX50 microscope. Representative photomicrographs taken on days 1, 3, 7, and 10. Analysis of variance was run with a P criterion of <0.05.

**Inhibition of infected mucosae with SB203580.** The bullae of six male Sprague-Dawley rats weighing between 300 and 350 g were injected with nontypeable H. influenzae (NTHI) at 105 cells per ml in the manner described above. After 48 hours, the animals were anesthetized and sacrificed by decapitation. After each animal was sacrificed, the middle ear bullae were surgically extracted and the middle ear mucosae, bilaterally, were dissected as described above. Each middle ear mucosal sample was immediately placed in a separate 60-mm-diameter Falcon petri dish coated with a thin layer of Sylgard from Dow Corning (Midland, Mich.). Culture medium consisting of a mixture of Dulbecco’s modified Eagle’s medium and Ham’s F-12 medium (3:1) supplemented with fetal calf serum (5%), hydrocortisone (0.4 μg/ml), isoproterenol (10 μM), penicillin (100 U/ml), and streptomycin (100 μg/ml) was then added (52). The middle ear mucosae were divided into roughly 0.5-mm² tissue explants with a Fine Science Tools (Foster City, Calif.) diamond knife. The explants from each bulla were then individually transplanted, epithelium uppermost, into single wells of a 24-well Falcon cell culture plate containing 170 μl of the above culture medium. The explants were then placed in an incubator at 37°C with 5% CO2 for 24 h and allowed to adhere to the culture plate surface. On day one, all wells with healthy, attached explants were randomly divided into four groups. The p38 inhibitor, SB203580 (Calbiochem, San Diego, Calif.), was reconstituted in dimethyl sulfoxide (DMSO) and added to one of three groups at 1, 10, and 100 mM in 400 μl of culture medium. The last group served as a negative control, the media receiving no addition of SB203580 and serving as a control for the DMSO concentration. The data were entered into Statview 5.0 and an analysis of variance was run with a P value criterion of <0.05. The tissue explants were also immunohistochemically stained with an anti-BrdU primary antibody (Sigma-Aldrich) followed by a biotinylated secondary antibody (Santa Cruz Biotechnologies, Santa Cruz, Calif.). After the appropriate biotin and avidin blocks, the cultures were treated with a fluorescein isothiocyanate-labeled strepavidin (DAKO, Carpinteria, Calif.). Each well was then photographed again under various magnifications with a GFP fluorescent filter.

**Inhibition of noninfected mucosae with SB203580.** The middle ear mucosae of six male Sprague-Dawley rats that had not been infected with H. influenzae were dissected, divided, and cultured in the manner described above. The explants were divided into three groups. The first group was cultured with 10 nM SB203580, the second group was cultured with 100 nM SB203580, and the last group served as a control being cultured with DMSO (1 μM). The fluorescein-stained, DAPI-counterstained explants and tissue sections were examined on an Olympus BX50 microscope. Representative photomicrographs taken on days 1, 3, 7, and 10. Analysis of variance was run with a P criterion of <0.05.
Evaluation of p38 in vitro. The presence and activation of p38 and the effects of SB203580 were determined in our in vitro culture system. Middle ear mucosal explants were prepared from healthy rat middle ears and from middle ears inoculated with NTHI 48 h previously. Explants were cultured as above for 72 h, with or without 100 nM SB203580. Explants were then harvested and assayed by Western blotting for total and phospho-p38 as described above.

RESULTS

Activation of p38 in the infected middle ear mucosa. Figure 1 illustrates the levels of total and activated p38 in the middle ear mucosa at various in vivo postinoculation time points. Total p38 protein (Fig. 1A and C) showed a gradual rise over the period evaluated. No phospho-p38 was detected in untreated, control mucosae (0 h in Fig. 1B). A high level of p38 activity was observed 1 and 6 h after inoculation with NTHI. Lower levels of phospho-p38 were seen at 24 and 48 h. While very little p38 activity was present at 72 h, moderate phosphorylation was again observed at 7 days. The ratio of phospho-p38 to total p38 (Fig. 1D) showed strong phosphorylation at 1 and 6 h and weaker phosphorylation at later periods. Similar patterns of p38 activation were observed in three additional series of mucosal samples (data not shown). Injection of saline into the middle ear had no effect on either total or phospho-p38 (data not shown).

p38 inhibition with SB203580 in infected middle ear mucosa. Figure 2 illustrates the effects of the p38 inhibitor, SB203580, on bacterial growth in the middle ear. Figure 2 shows that SB203580 inhibits bacterial growth in a dose-dependent manner. Significant inhibition of growth was observed at 10 nM (P < 0.0001) and 100 nM (P < 0.0001). Bars represent means, and vertical lines represent 1 SEM.
SB203580, on the outgrowth surface area of middle ear mucosal explants cultured 48 h postinoculation with NTHI. Explants from infected rats showed a high rate of growth throughout the 10-day culture period when not treated with SB203580. Such tissue explants, cultured in the presence of BrdU, demonstrated BrdU-positive cell nuclei throughout the entire outgrowth when immunohistochemically stained with a fluorescein isothiocyanate-labeled anti-BrdU antibody (data not shown). SB203580 at 1 nM (P = 0.8342) did not have a significant effect on this hyperplastic tissue outgrowth. SB203580 at 10 nM (P < 0.0001) and 100 nM (P < 0.0001) had a significant inhibitory effect compared with the negative controls. Outgrowths at SB203580 concentrations of 10 and 100 nM, however, were not significantly different compared to each other (P = 0.2497).

p38 inhibition with SB203580 in noninfected mucosa. Figure 3 illustrates the effects of the p38 inhibitor on the explant outgrowth surface area of cultured, noninfected middle ear mucosal explants. SB203580 at concentrations of 10 and 100 nM had no significant effect on mucosal growth compared to controls that were not treated with the inhibitor.

p38 activation in culture. After 72 h in culture, the level of total p38 was found to be higher in explants previously exposed to bacteria than in normal, noninfected explants. Similarly, phospho-p38 was elevated in bacterially exposed explants. Treatment with SB203580 had no effect on either total or phospho-p38 for either normal or bacterially exposed mucosal explants (Fig. 4).

**DISCUSSION**

The results of this study reveal evidence for a functional role of p38 signaling in middle ear mucosal hyperplasia during bacterial otitis media. Introduction of NTHI into the middle ear of rats produces otitis media that peaks in severity 2 to 5 days after inoculation, followed by recovery that is complete at around 2 weeks (35). We found that the mucosa of middle ears challenged in vivo with NTHI showed sharply elevated levels of activated p38 immediately after inoculation, declining to lower levels for the next few days. One week later, phospho-p38 was elevated, due partially to increasing levels of total p38 (Fig. 1). The early phospho-p38 response observed suggests that p38 is involved in the initial phases of otitis media. However, elevated levels of phospho-p38 at 7 days suggest that it may also play a role in recovery.

The early variation in activated p38 was produced by changes in phosphorylation, since total p38 showed no increase in the first hours after inoculation. This is in contrast to our prior data on the Erk MAPKs, in which variation in mucosal phospho-Erk largely paralleled changes in total Erk (39).

A role for p38 in the mucosal response to bacteria is also supported by our in vitro data. Both total and phospho-p38 were elevated in mucosal cultures that had previously been exposed to NTHI in vivo. Moreover, inhibition of p38 signaling produced a significant, dose-dependent suppression of bacterially stimulated growth in middle ear mucosal explant cultures (Fig. 2) but had no effect on noninfected middle ear explants (Fig. 3). Since all of the cells present in the middle ear cultures were labeled by BrdU, the observed growth represents cell proliferation rather than migration from the explant. Thus inhibition of p38 markedly reduced bacterially induced mucosal proliferation. The in vitro growth of noninfected mucosal samples presumably occurred via a p38-independent mecha-
nism, perhaps mediated by growth factors present in the media and/or produced by the samples themselves.

The function of an individual intracellular signaling pathway can be evaluated only if inhibitors that selectively suppress targeted enzymes in that pathway are available. SB203580, which operates through competition with ATP for the same docking site on the p38 kinase (55, 57), specifically inhibits members of the p38 MAPK subgroup, not the Erk or Jnk MAPKs (8). SB203580 inhibits only the α and β isoforms of p38 and has little or no effect on the γ or δ isoforms (17). This is caused by the presence of amino acids larger than threonine at position 106 of the latter isoforms, which prevent accommodation of the 4-fluorophenyl moiety of SB203580 (8). Since SB203580 does not inhibit the γ or δ isoforms, and individual isoform identity based upon molecular weight cannot be determined from Figure 1, it cannot be concluded from the data of our study whether or not the γ or δ isoforms are present or if they have significant functional involvement in mucosal hyperplasia during otitis media. However, since inhibition of the α and β isoforms via SB203580 eliminates the majority of the bacterially induced growth of middle ear mucosa in vitro, it can be argued that the γ and δ isoforms do not play a major role. Of course, our in vitro model may differ from the in vivo situation.

The sensitivity of the middle ear mucosa to SB203580 also needs to be taken into consideration. There was a potent inhibitory effect on mucosal growth at SB203580 concentrations of 10 to 100 nM, with no significant difference in inhibition between these two concentrations. Studies in most other tissue systems (6, 7) require higher concentrations of SB203580 (1 to 10 μM) to produce an inhibitory effect. Some cell types, however, such as platelets and THP-1 monocytes, display significant inhibition of p38 kinase activity at comparable concentrations of SB203580 (1 to 100 nM) (7), with bacterially exposed middle ear mucosa displaying a sensitivity similar to that of platelets, characterized by inhibition saturating at 10 nM. It should be noted that this potent inhibitory response may be related to an increased intracellular level of SB203580 due to exposure over the ten-day culture period. Long exposure time has been proposed to explain the low threshold of response to SB203580 in THP-1 monocytes (7). Of course, as with all inhibitors, it is also possible that SB203580 inhibits protein kinases or enzymes other than p38 that have not been evaluated or identified and which may relate to mucosal cell proliferation. It should also be noted that hydrocortisone was present in the culture media for both control and experimental mucosal samples. This could have influenced the growth of the mucosa, but would have affected both untreated and inhibitor-treated mucosa equally.

Total p38 and phospho-p38 were both elevated in in vitro mucosal explants previously exposed to bacteria, compared to noninfected in vitro explants. This agrees with the in vivo results and also is consistent with the effects of SB203580 on mucosal growth in bacterially exposed mucosal explants. SB203580 had no effect on total p38 levels or on p38 phosphorylation, consistent with its action downstream of p38 activation.

The p38 pathway is involved in a broad range of cellular responses in other tissue systems, including proliferation, migration, differentiation, and apoptosis (11, 22, 28, 36, 56). The divergent physiological functions of p38 may be explained in part by the existence of four different p38 isoforms. It is therefore possible that phosphorylation of different p38 isoforms at distinct times might contribute to the differential behavior of the middle ear mucosa during the time course of otitis media.

The data of this study suggest a significant role played by p38 MAPK signaling in middle ear mucosal hyperplasia. Our results complement the work of others that implicates p38 activation in the production of inflammatory cytokines and mucin (32, 49, 53, 54), which contribute to otitis media pathogenesis. These data may be clinically relevant to the treatment of bacterial otitis media. It is clear that both the middle ear mucosal hyperplastic responses and the p38 MAPK signaling pathway are highly complex and coordinated biological processes. Introducing an inhibitor in vivo might disrupt this process, blocking growth and/or proliferation of the hyperplastic middle ear mucosa and reducing the number of recurrent episodes and the severity of the sequelae of acute otitis media. However, our results suggest that timing and inhibitor specificity could play an important role if p38 inhibition is to be clinically effective. Introducing an inhibitor during the early stages of infection may cause a significant reduction in mucosal hyperplasia by inhibiting cellular proliferation and enhancing cellular apoptosis as has been demonstrated in Jurkat cells (48). However, if an inhibitor of p38 were introduced later during infection, it might possibly inhibit mucosal recovery by delaying apoptosis. Inhibition of p38 has been shown to block apoptosis in other epithelial cells (31). This could increase the duration of the mucosal hyperplasia. Additionally, even at saturated levels of SB203580, some mucosal growth still takes place in vitro. This suggests involvement of alternative pathways independent of p38 in middle ear mucosal hyperplasia, possibly indicating the need for multiple inhibitors to completely suppress the hyperplastic response. We have shown that inhibition of the Erk/ MAPKs also decreases middle ear mucosal proliferation (39). A more complete understanding of which intracellular pathways are involved and the specifics of which p38 isoforms are most active during otitis media will better enable the rational design of therapeutic interventions for this disease.

ACKNOWLEDGEMENTS

This work was supported by grant DC00129 from the NIH/ NIDCD and by the Research Service of the United States Department of Veteran Affairs.

REFERENCES
