



LUND UNIVERSITY

Improving the pathogenicity of a nematode-trapping fungus by genetic engineering of a subtilisin with nematotoxic activity.

Åhman, Johan; Johansson, Tomas; Olsson, Maja; Punt, P J; van den Hondel, C A M J J; Tunlid, Anders

Published in:
Applied and Environmental Microbiology

DOI:
[10.1128/AEM.68.7.3408-3415.2002](https://doi.org/10.1128/AEM.68.7.3408-3415.2002)

2002

[Link to publication](#)

Citation for published version (APA):

Åhman, J., Johansson, T., Olsson, M., Punt, P. J., van den Hondel, C. A. M. J. J., & Tunlid, A. (2002). Improving the pathogenicity of a nematode-trapping fungus by genetic engineering of a subtilisin with nematotoxic activity. *Applied and Environmental Microbiology*, 68(7), 3408-3415. <https://doi.org/10.1128/AEM.68.7.3408-3415.2002>

Total number of authors:
6

General rights

Unless other specific re-use rights are stated the following general rights apply:
Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal

Read more about Creative commons licenses: <https://creativecommons.org/licenses/>

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

LUND UNIVERSITY

PO Box 117
221 00 Lund
+46 46-222 00 00

Improving the Pathogenicity of a Nematode-Trapping Fungus by Genetic Engineering of a Subtilisin with Nematotoxic Activity

Johan Åhman,¹† Tomas Johansson,¹ Maja Olsson,¹ Peter J. Punt,²
Cees A. M. J. J. van den Hondel,² and Anders Tunlid^{1*}

Department of Microbial Ecology, Lund University, S-223 62 Lund, Sweden,¹ and Department of Applied Microbiology and Gene Technology, TNO Voeding, 3700 AJ Zeist, The Netherlands²

Received 26 December 2001/Accepted 10 April 2002

Nematophagous fungi are soil-living fungi that are used as biological control agents of plant and animal parasitic nematodes. Their potential could be improved by genetic engineering, but the lack of information about the molecular background of the infection has precluded this development. In this paper we report that a subtilisin-like extracellular serine protease designated *PII* is an important pathogenicity factor in the common nematode-trapping fungus *Arthrobotrys oligospora*. The transcript of *PII* was not detected during the early stages of infection (adhesion and penetration), but high levels were expressed concurrent with the killing and colonization of the nematode. Disruption of the *PII* gene by homologous recombination had a limited effect on the pathogenicity of the fungus. However, mutants containing additional copies of the *PII* gene developed a higher number of infection structures and had an increased speed of capturing and killing nematodes compared to the wild type. The paralyzing activity of *PII* was verified by demonstrating that a heterologous-produced *PII* (in *Aspergillus niger*) had a nematotoxic activity when added to free-living nematodes. The toxic activity of *PII* was significantly higher than that of other commercially available serine proteases. This is the first report showing that genetic engineering can be used to improve the pathogenicity of a nematode-trapping fungus. In the future it should be possible to express recombinant subtilisins with nematocidal activity in other organisms that are present in the habitat of parasitic nematodes (e.g., host plant).

Many nematodes are parasites on plants or animals and can cause serious crop losses and severe diseases. The development of new methods to control such nematodes is of major importance because the traditional methods that are based on the use of nematicides and antihelminthic drugs are associated with major environmental and health concerns. In addition, resistance development in target organisms requires new nematicides to be developed continuously. An alternative strategy is to control parasitic nematodes by using their natural enemies (23). Among them, the nematophagous fungi, which comprise a group of mainly soil-living fungi that are infectious to nematodes, have drawn the largest interest. A common strategy has been to mass-produce the fungi on solid substrates followed by application to the environment. Although there are several reports of successful biological control of both plant and animal parasitic nematodes by using nematophagous fungi (e.g., references 24 and 31), they are commonly perceived to perform poorly compared to traditional nematicides and antihelminthics.

One way to improve the control potential of nematophagous fungi would be to use genetic engineering to increase the pathogenicity and survival of the introduced fungus. Until recently, however, such a development has been precluded due to the lack of information about the molecular background of

fungal infections of nematodes (11). We have identified two putative pathogenicity factors in the common nematophagous fungus *Arthrobotrys oligospora*, a carbohydrate binding protein (lectin) and an extracellular serine protease designated *PII* (1, 20, 26). *PII* belongs to the subtilisin family of serine proteases (27). Extracellular subtilisins have also been isolated from other species of nematophagous fungi (3, 13, 21). The function of these proteases is not yet clear, but it has been assumed that they have a role during the fungal penetration of the nematode cuticle and/or during the digestion of the internal tissues of the host.

In this report, the role of *PII* has been studied in more detail by generating several *PII* mutants, using a recently developed transformation system for *A. oligospora* (28). Notably, mutants containing additional copies of the *PII* gene demonstrated an increased pathogenicity. This is the first report showing that genetic engineering can be used to improve the virulence of a nematode-trapping fungus. Furthermore, *PII* was expressed in a heterologous system (*Aspergillus niger*), and it was demonstrated that the recombinant enzyme has a nematotoxic activity in vitro.

MATERIALS AND METHODS

Organisms and plasmids. *A. oligospora* Fres. (ATCC 24927) was maintained on CMA7-containing cornmeal agar (Difco) supplemented with 2 g of KH_2HPO_4 per liter. Trap-containing mycelium was grown in a liquid medium containing 0.01% soya peptone (neutralized; Oxoid) supplemented with 0.05 g of phenylalanine and 0.05 g of valine per liter (27). Two auxotrophic *pyrG* mutant strains of *A. niger* were used for heterologous expression of *PII*: AB1.13pcl3FOA17 (designated AB1.13pclA), which is deficient in both aspergillopepsin A and aspergillopepsin B (due to a regulatory mutation, *prt-13*) (14) and with the *pclA* gene deleted (processing enzyme) (P. J. Punt, unpublished data), and *A. niger* AB1.18, which is mutated in the *pepA* gene encoding aspergillopep-

* Corresponding author. Mailing address: Department of Microbial Ecology, Lund University, Ecology Building, S-223 62 Lund, Sweden. Phone: 46 46 2223757. Fax: 46 46 2224158. E-mail: Anders.Tunlid@mbioekol.lu.se.

† Present address: Department of Ecology and Molecular Biology, The Royal Veterinary and Agricultural University, 1871 Frederiksberg C, Denmark.

TABLE 1. Plasmids used in this study

Plasmid	Description/use	Source
pUBH	Genomic clone of <i>PII</i>	1
pAotubF2	Gene fragment of <i>tubA</i> (from <i>A. oligospora</i>)	This study
pBHE	Gene disruption vector for <i>PII</i>	This study
pCB1003	Transformation vector containing <i>hph</i>	5
pAN7-1	Transformation vector containing <i>hph</i>	19
pCB1003-PII	Transformation vector containing <i>hph</i> and <i>PII</i>	This study
pAFR	PCR clone of <i>PII</i>	This study
pAN52-7Not	<i>glaA</i> promoter (from <i>A. niger</i>)	30
pASP	<i>gluA</i> promoter linked to <i>PII</i>	This study
pAMDSPYRG#1	<i>amdS/pyrG</i> selection marker	(P. J. Punt, unpublished data)

sin A (14). The nematode *Panagrellus redivivus* L. (Goodey) was grown axenically in a soya peptone-liver extract medium (15). The plasmids used in this study are listed in Table 1.

Bioassays. An infection assay using a dialysis membrane technique was employed (20). Briefly, conidia of *A. oligospora* were inoculated onto pieces of dialysis membrane and incubated on agar plates containing a low nutrient mineral salt (LNM) medium. The formation of infection structures (traps) was induced by adding a few specimens of the nematode *P. redivivus* to the hyphae growing on the dialysis membranes. After another 7 to 10 d, the infection experiments were started by adding 20 to 30 nematodes cm⁻². The number of adhered (captured) and killed (captured and not moving) nematodes was counted by using a microscope. The toxic effects of proteases on free-living *P. redivivus* were tested using an assay in microtiter wells (27). The numbers of mobile and immobile (i.e., with arrested movements) nematodes were determined by using a light microscope. The effect of PII was compared with that of proteinase K and trypsin (bovine pancreas) (Sigma). The enzymes were dissolved in water or phosphate-buffered saline (50 mM sodium phosphate buffer [pH 6.5], 0.15 M NaCl). The nematotoxic activities of linoleic acid and ivermectin (from Sigma) were also examined. The compounds were dissolved in 5% methanol (in water). The bioassays were performed with 10 to 15 parallels and repeated at least twice.

Competitive reverse transcription-PCR. Total RNA was isolated from the mycelia growing on the dialysis tubing by acid-guanidinium thiocyanate-phenol-chloroform extraction (20), dissolved in diethyl pyrocarbonate-H₂O, and then stored at -20°C. Traces of genomic DNA were digested by RQ1 DNase (Promega). First-strand synthesis was performed in 10- μ l reaction mixtures by using oligo(dT)₁₂₋₁₈ primer (Gibco/BRL), 2 μ l of total RNA, 30 U of RNasin RNase inhibitor (Promega), and 100 U of Superscript II RNaseH reverse transcriptase (Gibco/BRL) with other conditions according to the manufacturer. After termination, the first-strand product was diluted by adding 20 μ l of 1 \times first-strand buffer (Gibco/BRL). As competitive PCR templates, plasmids containing the *A. oligospora PII* gene (pUBH) and a fragment of the partially characterized *A. oligospora tubulin* gene (*tubA*) were used. Here, the *tubA* template was used as a positive control and for normalizing the data between different measurements and experiments. For cloning of the *tubA* gene fragment, standard PCR was conducted on *A. oligospora* genomic DNA by using the primers Bt2a (5'-GGT AACCAATCGGTGCTGCTTTC) and Bt2b (5'-ACCCTCAGTGTAGTGAC CCTTGGC) (10). A PCR product of the expected size (567 bp) was ligated into the pGEM-T Easy vector (Promega), generating the plasmid pAotubF2. The insert was sequenced and verified as a β -tubulin gene fragment by homology to other fungal tubulin genes (GenBank accession number AY028375).

The primers used for competitive PCR were designed to flank intron regions, resulting in a size difference of the products amplified from genomic DNA (gDNA) and the corresponding cDNA template. For *PII* (GenBank accession number X94121), the primers P106 (5'-TGAGGTCGACTACGTTGAACAAG) and P107 (5'-GGAATCAGTCTTGTCAACAGAGTT) were used, giving PCR products of 320 bp (gDNA) and 259 bp (cDNA), respectively, whereas for *tubA* the primers were P114 (5'-CTCGACGGCTCCGGTGTTA) and P115 (5'-CAGGAAACAGCTATGAC), yielding products of 320 bp (gDNA) and 179 bp (cDNA), respectively. PCR products were separated on a 1.5% agarose gel in 0.5 \times Tris-borate-EDTA and visualized with EtBr and UV light. Equivalence points were calculated by plotting molar ratios of gDNA to cDNA band intensities, and the molar amount of cDNA was determined from the point at the regression line where the ratio between gDNA and cDNA was equal to 1.

Construction of *PII* mutants. The genomic *PII* clone pUBH was PCR amplified (Expand High-Fidelity PCR system; Roche Molecular Biochemicals) with

two primers designed from the *PII* sequence: a₁ (5'-TGGGAGATGCGAATA CCGGT; position 854 to 873; GenBank accession no. X94121) and a₂ (5'-ACC GTGAATGGTTCATCCCG; position 948 to 967). The *hph* gene under the control of the *trpC* promoter from *Aspergillus nidulans* was excised from the plasmid pCB1003 using *EcoRI* (5). End regions of both the PCR fragment and the excised *hph* gene were filled in with Klenow before ligation, producing the gene disruption vector pBHE. This vector contains the 5' and 3' ends (approximately 1.5 and 1.6 kb, respectively) of the *PII* gene with a displacement of 74 bp (a 61-bp intron and a 13-bp exon region) and a 1.4-kb *hph* insertion, as well as an intact plasmid backbone (see Fig. 2A).

For generating mutants containing additional copies of the *PII* gene, the pUBH vector was cotransformed with the pAN7-1 vector at a molar ratio of 1:1. Multicopy mutants were also generated by transforming the fungus with a vector containing both the *PII* and *hph* genes. The vector designated pCB1003-PII was constructed by excising a *BamHI/HindIII* fragment containing the entire *PII* gene from the plasmid pUBH. The fragment was inserted into the *BamHI/HindIII* sites of pCB1003.

Transformation of *A. oligospora* was performed according to the methods described by Tunlid et al. (28). To confirm the stability of the transformants, they were transferred several times between media with and without selection pressure (hygromycin). Transformants were analyzed by Southern blotting as described below. The *pBHE* gene disruption transformants were also analyzed by PCR amplification. Two primers positioned in the *PII* gene adjacent to the inserted *hph* gene were used: b₁ (5'-TTTGACAAGGCAACTCTCCAGG; position 650 to 671; GenBank/EMBL accession no. X94121) and b₂ (5'-CCACCA GCGCTAAGAACCCTTAAC; position 1092 to 1114) (see Fig 2A). Transformants were screened for extracellular serine protease activity by growing the fungus for 2 to 3 days (130 rpm; 20°C) in a liquid medium containing 0.5% (wt/vol) soya peptone. Transformants were also grown under nitrogen-limited conditions and transferred to an inducing medium containing bovine serum albumin (BSA; 1 mg/ml) as the nitrogen and carbon source (1).

Heterologous expression of *PII*. A gene fragment covering the entire *PII* gene (position 358 to 1682; GenBank accession no. X94121), including regions encoding the signal peptide, propeptide, and the mature enzyme, was obtained by PCR amplifying the genomic clone pUBH using two primers: Aspfw (5'-TCCA TCATGATTACGAACGG) and Aspwr (5'-CTTGGTCAATGATCTTTGGG). Both primers contained *RcaI* sites (underlined). The product was inserted into a pGEM-T vector (Promega), generating the plasmid pAFR. The expression vector pAN52-7Not, containing the *glaA* promoter (30), was cut with *NcoI* and the *RcaI* fragment from pAFR was inserted to produce the expression vector pASP. Junction regions of pASP were sequenced to verify the orientation and cloning sites of the insert. The two auxotrophic *pyrG* mutant strains of *A. niger* (AB1.13pclA and AB1.18) were cotransformed with pASP and the *amdS/pyrG* selection marker (isolated as a *NotI* fragment from pAMDSPYRG#1 [P. J. Punt, unpublished data]) (18). Transformants with strong growth on plates with acetamide-acrylamide as sole nitrogen (and carbon) source were selected (29). To analyze the production of recombinant PII, the transformants of *A. niger* were grown for 48 h at 30°C (200 rpm) in minimal growth medium (2) supplemented with 5% maltodextrin.

Peptide sequencing and protease assays. Proteases were purified from the culture filtrates of *A. oligospora* and *A. niger* by high-performance liquid chromatography (HPLC) (27). Sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) was performed using a MiniProtean II electrophoresis cell (Bio-Rad). Gels were stained with Coomassie brilliant blue. Before peptide sequencing, the protease activity was inhibited by adding phenylmethylsulfonyl fluoride (PMSF; 1 mM). The N-terminal region was analyzed by Edman degra-

dation using an ABI 476A protein sequenator. The hydrolytic activity of PII was measured using the peptide substrate benzoyl (Bz)-Phe-Val-Arg-nitroanilide (NA) and the chromogenic protein substrate Azocoll (Sigma) (27). The substrate specificity, the effects of inhibitors, and the pH dependency of the recombinant PII were examined as previously described (27).

Hydrolysis of nematode cuticle and tissues. The cuticle of *P. redivivus* was isolated according to the protocol of Cox et al. (7). Briefly, nematodes were sonicated in a Tris buffer (10 mM Tris-HCl, pH 7.4), the samples were centrifuged, and the supernatant was collected (intracellular fraction). The pellet was treated with an SDS-containing buffer (7) and, following centrifugation, the supernatant was collected (cuticle-SDS fraction). The pellet was washed with the Tris buffer, lyophilized, and resuspended to give a concentration of 1.0 mg/ml (cuticle-fragment fraction). Before being used in the assays, the intracellular fraction was filtered (0.2- μ m pore size), and the cuticle-SDS fraction was desalted using a PD-10 column (Pharmacia) equilibrated with the Tris buffer. Ten microliters of recombinant PII, proteinase K, or trypsin (1.0×10^{-3} proteolytic units [PU]; 0.3 to 0.8 μ g of protein) was mixed with 300 μ l of the intracellular fraction (total protein content, 18 mg/ml), cuticle-SDS fraction (0.12 mg/ml), and the cuticle-fragments fraction (1.0 mg/ml), and incubated at 37°C for 2 h. Samples were centrifuged and the products of protease hydrolysis were assayed by HPLC using a size-exclusion column (Superdex PC 3.2/30) connected to a SMART system (Pharmacia). The buffer was 10 mM Tris (pH 7.4) containing 0.15 M NaCl. Following the subtraction of a chromatogram (280 nm) obtained from a sample incubated in buffer only, the areas of low-molecular-weight peaks (less than ca. 13 kDa) were integrated using the SMART software. The proteolytic activity is given as area counts per minute.

Southern analysis and RNA dot blots. Genomic DNA was isolated (28), digested, separated on a 0.8% agarose gel, and transferred to a Hybond-N⁺ membrane (Amersham).

RNA was isolated from liquid-grown mycelia using the RNeasy Plant Mini Kit (Qiagen) and quantified by measuring the absorbance at 260 nm. The RNA samples were blotted on a Hybond-N membrane (Amersham) using a Bio-Dot microfiltration apparatus (Bio-Rad). The amount of applied RNA was approximately 1.0 μ g as determined by absorbance at 260 nm.

The probes were labeled with [α -³²P]dCTP ($\approx 3,000$ Ci mmol⁻¹) using the random-primed DNA method (*rediprime* DNA labeling kit; Amersham).

RESULTS

PII is expressed during the colonization and digestion of the nematode. The *PII* transcript was not detected (<0.001 amole) during the early stages of the infection, including the phase of adhesion (ca. 0 to 12 h of incubation) (Fig. 1). However, the *PII* mRNA was detected 24 h after adding the nematodes. At this time, the fungus had penetrated and killed ca. 70% of the added nematodes, and fungal hyphae were growing inside the infected nematodes. No *PII* transcript was detected after 5 days of incubation, by which time all of the added nematodes had been killed and completely degraded.

Construction of Δ *PII* mutants. Twenty mutants of *A. oligospora* transformed with the gene disruption vector pBHE were examined for proteolytic activity when grown in soya peptone medium. Five of them had significantly reduced proteolytic activity. In Southern analysis, these mutants contained one hybridizing fragment of ca. 4.5 kb (Fig. 2B, lanes 1 to 5) but not the band of 3.2 kb present in the wild type (Fig. 2B, lane 7). This pattern of hybridization is expected after homologous integration of the *hph*-disrupted *PII* gene (Fig. 2A). The other 15 mutants were shown to contain both the 3.2-kb as well as the 4.5-kb fragment (Fig. 2B, lane 6), which show the presence of both the native *PII* gene as well as the disrupted *PII* gene.

The proteolytic activities of two of the replacement transformants (TJ Δ DPII.2 and -3) were further examined and compared with the activity of the wild-type strain by using BSA as the inducing substrate (Fig. 2C and 3A). Under these conditions, the proteolytic activities in TJ Δ DPII.2 and -3 were ca.

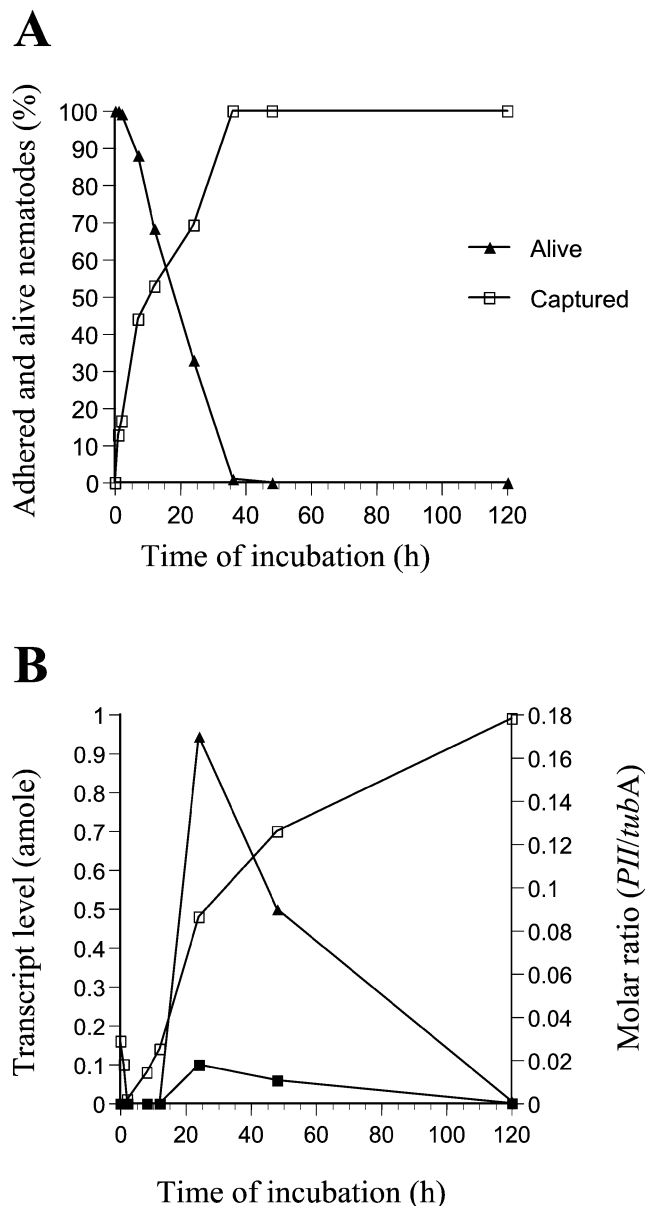


FIG. 1. Expression of the subtilisin PII during the infection of nematodes. (A) Adhesion and killing of the nematode *P. redivivus* during infection by the fungus *A. oligospora*. The numbers of adhered (captured) and killed (captured and not moving) nematodes were counted using a microscope. (B) Results of analysis of transcript levels of *PII* and *tubA*, using competitive reverse transcription-PCR. Tubulin was used as a positive control gene fragment and for normalizing the data. ■, *PII* transcript level; □, *tubA* transcript level; ▲, *PII-tubA* molar ratio.

70% lower than in the wild-type strain (the maximum activity in the wild type corresponding to 100% was 4.2 pmol of NA ml⁻¹ min⁻¹ mg⁻¹). The serine protease inhibitor PMSF (1 mM) completely inhibited the residual activity against Bz-Phe-Val-Arg-NA in both TJ Δ DPII.2 and -3. That the reduction of proteolytic activity in the deletion mutants was due to the lack of PII activity was further verified by partly purifying extracellular proteases expressed by TJ Δ DPII.2 during growth in liquid medium. Under these conditions, the wild-type strain pro-

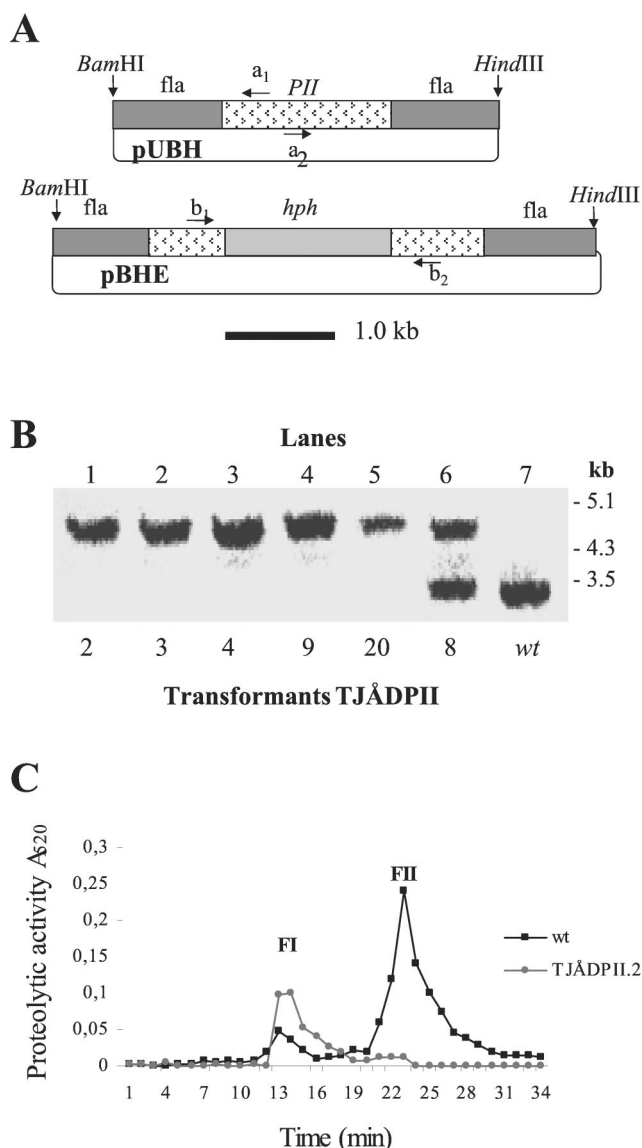


FIG. 2. Construction of ΔPII mutants. (A) Gene disruption vector pBHE constructed by inserting the *hph* gene (*Escherichia coli* hygromycin B resistance gene, under the control of the *trpC* promoter from *A. nidulans*) in the open reading frame of *PII* present in the vector pUBH. *fla*, flanking region. Arrows indicate PCR primers used for cloning (*a*₁ and *a*₂) and for screening (*b*₁ and *b*₂). (B) Southern analysis of gene disruption mutants obtained using the vector pBHE. Genomic DNA was digested with *Bam*HI/*Hind*III and probed with a ³²P-labeled fragment of the *PII* gene (*Bam*HI/*Hind*III fragment excised from pUBH) (30). Lanes 1 to 5 are mutants (designated TJΔDPII.2, -3, -4, -9, and -20) for which the mutated *PII* gene has been homologously integrated into the genome. Lane 6 shows a mutant (TJΔDPII.8) containing nonhomologous integration of the mutated *PII* gene. Lane 7 is the wild type. (C) Partial purification of the extracellular serine proteases produced by the wild type and the deletion mutant TJΔDPII.2 of *A. oligospora* when grown in liquid medium that stimulated the formation of infection structures (traps). The extracts were applied on a Mono Q column equilibrated with a Tris-HCl buffer (pH 7.5) and eluted with a gradient of NaCl using a flow of 1 ml/min (27). One-milliliter fractions were collected and assayed for protease activity by using the chromogenic substrate Azocoll.

duced two fractions of proteolytic activity designated FI and FII, with the latter containing PII (27). In TJΔDPII.2, the proteolytic activity in FII was almost completely absent (Fig. 2C). That *A. oligospora* contains several genes encoding serine proteases of the subtilisin family has been indicated by Southern analyses (1).

Construction of overexpression mutants. In an initial screen, 100 of the mutants obtained after transformation using the pUBH and pAN7-1 vectors and 50 mutants obtained using the pCB1003-PII vector were analyzed for protease activity when grown in soya peptone medium for 2 to 3 days. None of them showed elevated levels of protease activity. However, Southern analysis of randomly selected mutants indicated that several of them contained additional copies of the *PII* gene. The induction of proteases in 45 of these mutants was analyzed in more detail using BSA as carbon and nitrogen source. Notably, two of the mutants (TJΔMPII.1 and TATMPII.D17) started to secrete a protease activity more rapidly than the wild-type strain (Fig. 3A). The integration of DNA in TJΔMPII.1 (transformed with the pUBH and pAN7-1 vectors) was analyzed by Southern blotting. Genomic DNA was digested with *Bam*HI and *Hind*III, and the membranes were hybridized to a *Bam*HI/*Hind*III gene fragment of the *PII* gene excised from pUBH. Two fragments were visualized, one the size of the *PII* band in the wild-type strain and one that was larger (data not shown). As judged from the intensities of these bands, the TJΔMPII.1 mutant contained one additional copy of the *PII* gene. However, the fact that multiple fragments were observed in the Southern blots indicates that there were gene rearrangements of the inserted DNA. Southern analysis of the TATMPII.D17 mutant (transformed with the pCB1003-PII vector) by *Hind*III digestion and hybridization, using the pCB1003 plasmid as probe, showed three DNA fragments, with one the size of the pCB1003-PII vector (8 kb). This suggests integration of a tandemly repeated array of at least two copies of the pCB1003-PII vector (data not shown).

The variations in the level of *PII* mRNA followed approximately the changes in the activity of PII in the wild type, the deletion mutant TJΔDPII.3, and the overexpression mutant TATMPII.D17 (Fig. 3B).

Pathogenicity of PII mutants. The pathogenicities of two ΔPII mutants (TJΔDPII.2 and -3) and two overexpressing mutants (TJΔMPII.1 and TATMPII.D17) were examined using a bioassay with the nematode *P. redivivus*. The ΔPII mutants had a lower number of infection structures (traps) than the wild-type strain (Table 2). However, disruption of *PII* had a limited effect on the capture of nematodes, as significant differences between the ΔPII mutants and the wild type in the capture of nematodes were only observed after 10 h of infection (Fig. 4A). The overexpression mutants had a significantly higher number of traps than the wild-type strain and had an increased speed of capturing nematodes than the wild type (Table 2 and Fig. 4A). The effect of the gene-disrupted and the overexpressing mutants on the immobilization of captured nematodes was also investigated. No significant difference in the immobilization of nematodes was observed between the two ΔPII mutants and the wild-type strain (Fig. 4B). However, the percentage of immobilized nematodes was higher in the overexpressing strains than in the wild type; thus, the overexpressing mutants

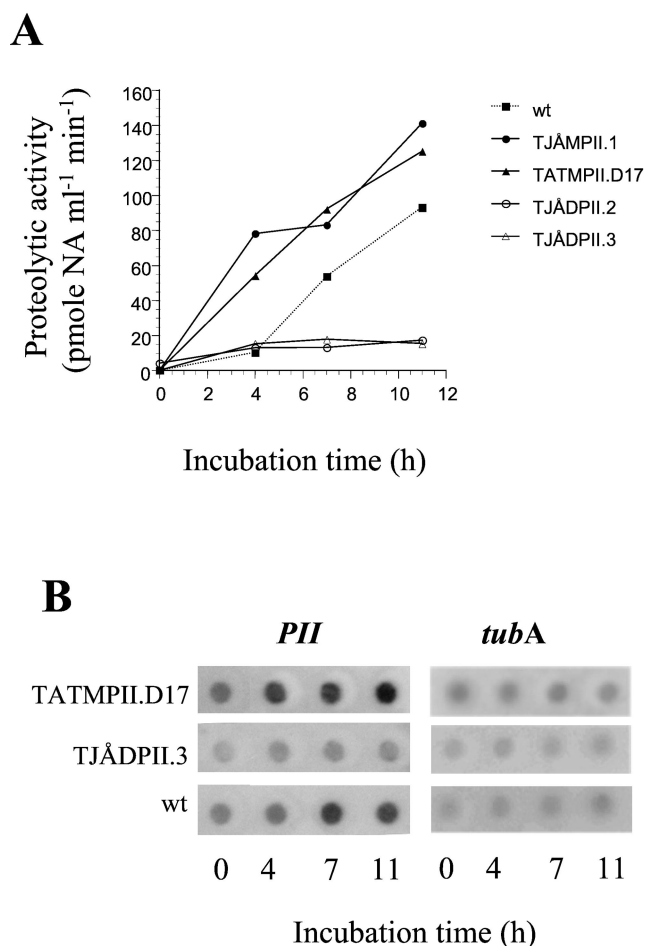


FIG. 3. Proteolytic activity and expression of mRNA in *PII* mutants of *A. oligospora*. Mycelia were grown for 2 days in a nitrogen-limited medium before being transferred to a medium containing albumin as nitrogen and carbon source. (A) Proteolytic activity (in the culture filtrates) as estimated by measuring hydrolysis of the peptide substrate Bz-Phe-Val-Arg-NA. (B) Dot blot of total RNA hybridized with a ³²P-labeled fragment of the *PII* gene (*Bam*HI/*Hind*III fragment excised from pUBH) (1) and *tubA* (*Eco*RI fragment excised from pAotubF2) (cf. Table 1).

showed a more rapid killing of nematodes than the wild-type strain.

Heterologous expression of *PII*. The expression vector pASP was constructed by cloning the open reading frame of *PII* in

TABLE 2. Development of infection structures in various mutants of *A. oligospora*^a

Strain	Traps (cm ⁻²) (mean [SD])
Wild type.....	81 (36)
TJÄDPPII.2.....	51 (20)
TJÄDPPII.3.....	41 (19)
TJÄMPII.1.....	202 (67)
TATMPII.D17.....	229 (72)

^a The fungus was grown on dialysis membranes for 3 d, whereupon nematodes were added to induce trap formation. After another 7 to 10 days, the numbers of traps were counted using a microscope. *n* = 10.

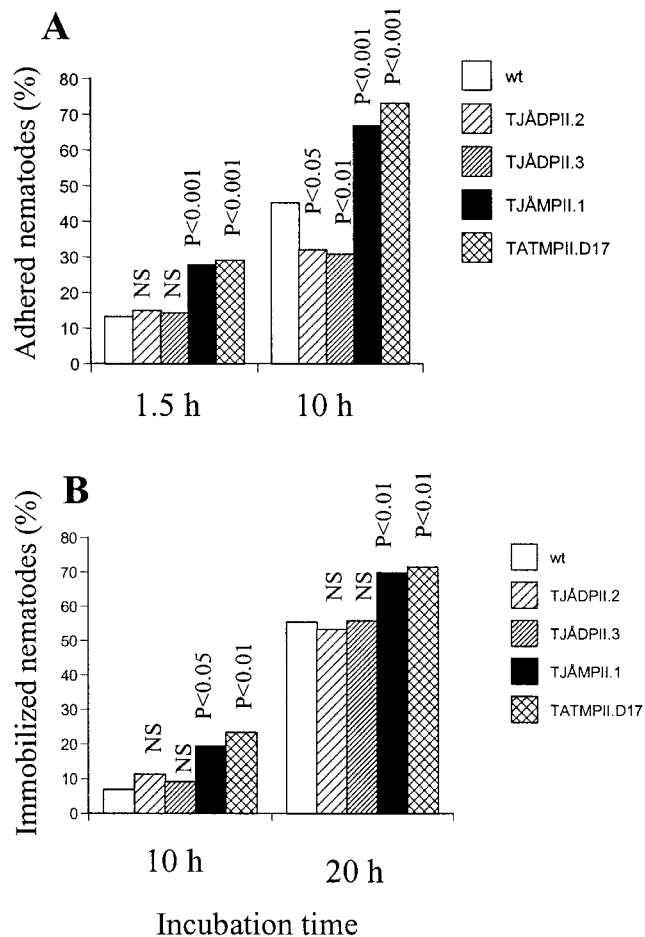


FIG. 4. Infection of nematodes by various mutants of *A. oligospora*. (A) Adhesion of nematodes; (B) immobilization of captured nematodes. Indicated significance levels are from χ^2 tests (*df* = 1) comparing the frequencies of captured and free nematodes (A) or mobile and immobile nematodes (B) in the mutant strains versus the wild type. NS, not significant.

front of the *glaA* (glucoamylase) promoter in *A. niger*. Twenty-eight transformants (19 from strain AB1.13pclA and 9 from AB1.18) of *A. niger* that showed extensive growth on the selective acetamide-acrylamide medium were isolated. When grown in liquid medium after induction of the *glaA* promoter with dextrin, all of these transformants had an increased protease activity compared to the nontransformed control. Southern analysis of genomic DNA from nine different transformants digested with *Bam*HI/*Hind*III showed three or more bands hybridizing to a probe of the *PII* gene (data not shown). Since these enzymes do not cut in the inserted *PII* gene fragment, these data indicate that the transformants contained three or more copies of the *PII* gene.

The proteolytic activities of the 10 AB1.13pclA and 7 AB1.18 transformants showing the highest levels of extracellular protease expression were purified by HPLC. The protease activity of all AB1.13pclA transformants was eluted in one peak by using a size-exclusion column (Fig. 5). Furthermore, the AB1.18 transformants had an additional peak, with activity eluting in a fraction corresponding to protein(s) with a higher

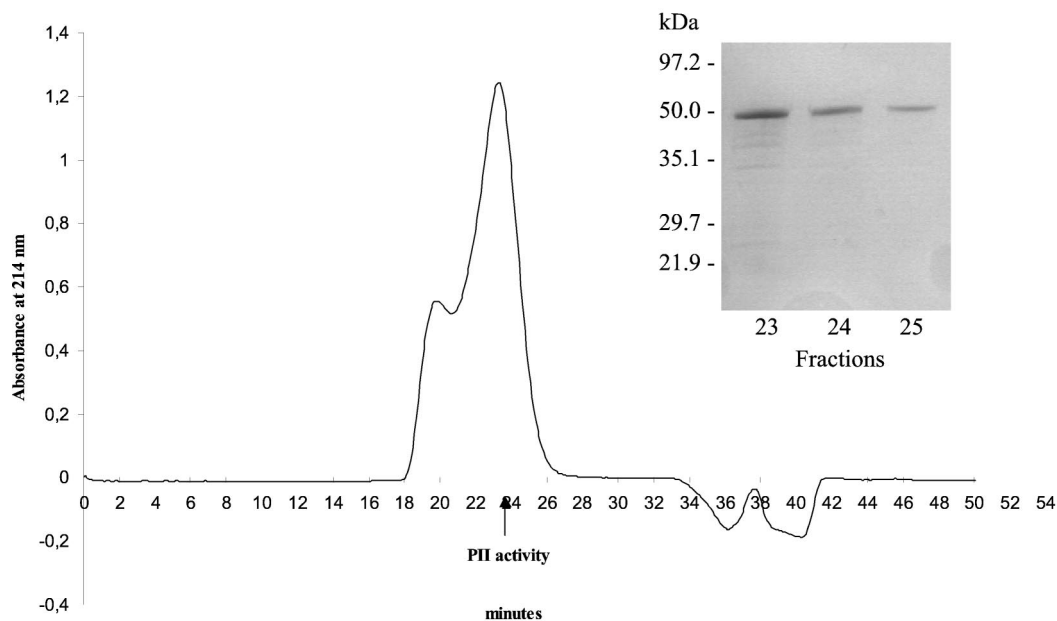


FIG. 5. Heterologous expression of *PII* and purification of heterologous PII from *A. niger* strain AB1.13pclA:3. Extracellular proteases were purified by anion-exchange, hydrophobic-interaction, and size-exclusion chromatography (shown). Protease activities in the different fractions (every minute) were analyzed using the peptide substrate Bz-Phe-Val-Arg-NA. The arrow indicates major PII activity. Inserted is an SDS-PAGE of fraction 23-25.

molecular weight than the protease expressed in the AB1.13pclA transformants. The AB1.13pclA transformant showing the highest proteolytic activity (designated AB1.13pclA:3) was chosen for further studies. SDS-PAGE analysis of the HPLC fractions with the highest activity against Bz-Phe-Val-Arg-NA revealed a polypeptide of approximately 48 kDa in mass (Fig. 5, insert). The N-terminal sequence of this polypeptide was XEQTDSTWGLDRISHEDYSA, with X denoting an unidentified residue. Previous peptide sequencing of PII from *A. oligospora* indicated a blocked N terminus (27). However, based on the alignment of the deduced amino acid sequence of PII with other subtilisin-related fungal serine proteases, it was predicted that the mature PII protease has an N-terminal sequence of TYAEQTDSTWGL (27), which contains two additional residues compared to that of the recombinant PII. The difference observed between the molecular mass of the recombinant PII (48 kDa when using SDS-PAGE) and the native enzyme (40 kDa) (27) could be explained by differences in glycosylation. An alteration in the pattern of glycosylation has been observed for many heterologously produced proteins in *Aspergillus*, including a *Rhizomucor miehi* aspartic proteinase (6).

The yield of PII obtained from AB1.13pclA:3 was 120 $\mu\text{g/liter}$, which is significantly higher than the production by *A. oligospora* (0.7 $\mu\text{g/liter}$) (27). The specific activity of the heterologous PII was 0.8 PU/mg of protein, which is lower than that of PII from *A. oligospora* (1.9 PU/mg) (27). There were no significant differences between the recombinant and native PII with regard to the effects of various inhibitors, pH optimum, and hydrolysis of synthetic peptide substrates (data not shown). Thus, both enzymes were inhibited by PMSF, chymostatin, and antipain, both had a broad pH optimum in the

alkaline range (approximately 7 to 9), and their preferred peptide substrate was Bz-Phe-Val-Arg-NA (27).

Hydrolytic and nematotoxic activity of recombinant PII. Recombinant PII showed high hydrolytic activity against an intracellular fraction of proteins isolated from the nematode *P. redivivus* (Fig. 6). Some activity was observed against proteins present in the cuticle-SDS fraction, which is supposed to contain muscle fragments and the tissue remains of the cuticle wall

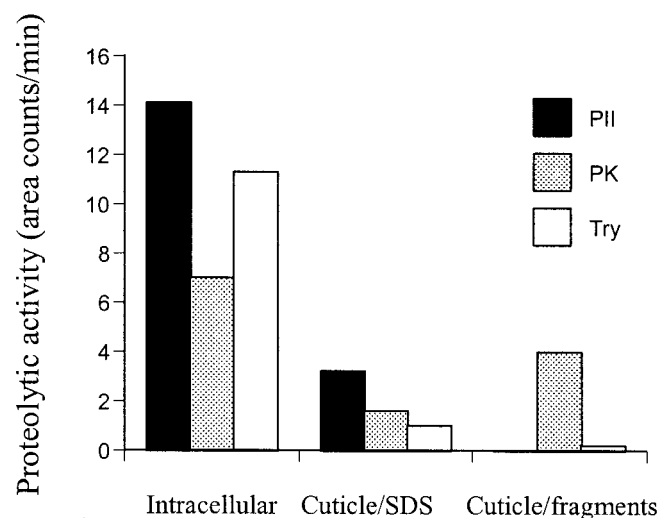


FIG. 6. Hydrolysis of nematode cuticle and tissues by various serine proteases. The nematode *P. redivivus* was sonicated and separated into three fractions. Heterologous-produced PII, proteinase K (PK), or trypsin (Try) were added to the fractions and incubated at 37°C for 2 h. The amount of released peptides was assayed by using HPLC.

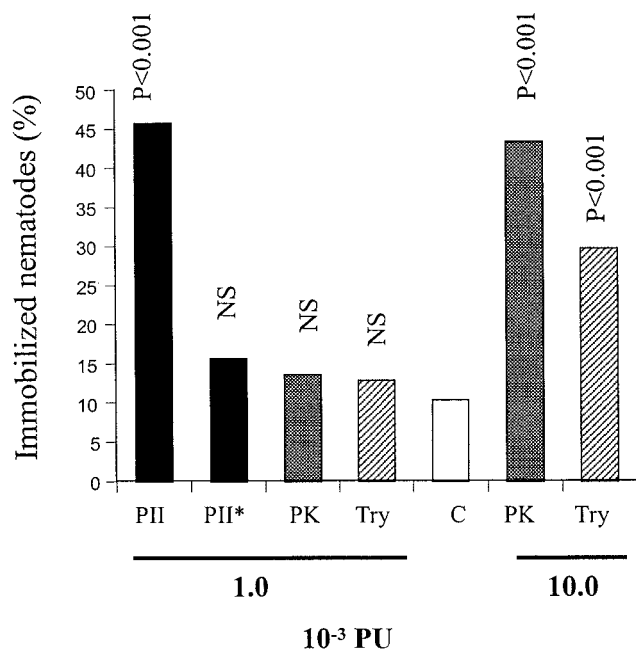


FIG. 7. Nematotoxic activity of heterologous PII. The proteases were incubated with the nematode *P. redivivus* in microtiter wells. After 20 to 22 h the number of mobile and immobile (i.e., with arrested movements) nematodes were counted in a light microscope. Indicated significance levels are from χ^2 tests ($df = 1$) comparing the frequencies of mobilized and immobilized nematodes in treated versus control samples (without enzymes). NS, not significant. PII, PII produced in *A. niger*; PII*, boiled PII (10 min); PK, proteinase K; Try, trypsin; C, control (buffer). The proteolytic activities of the added enzymes, as measured by using the substrate Azocoll, are indicated.

(7). The activity against the purified cuticle fragments was very low. Trypsin had a pattern of hydrolytic activity against the various cellular fractions similar to that of PII. Proteinase K degraded proteins present in all three fractions, including the cuticle fragments.

When added to free-living nematodes, the heterologous-produced PII had a nematotoxic activity, i.e., the nematodes were arrested in their movements (Fig. 7). The effect was not observed after boiling the preparation, indicating that it was due to the presence of protease activity in PII. No toxic effect was observed after adding a similar level of proteolytic activity of two other commercially available serine proteases, the fungal subtilisin proteinase K and trypsin. At least 10 times higher proteolytic activity was needed for these enzymes to immobilize nematodes at a level corresponding to that of PII (Fig. 7).

The concentration of heterologous PII that immobilized ca. 50% of the nematodes in the microwell assay (Fig. 7) was calculated to be 13 $\mu\text{g/ml}$ (2.7×10^{-8} M). Using the same microwell plate assay, linoleic acid and ivermectin immobilized ca. 50% of the nematodes at 100 $\mu\text{g/ml}$ (3.6×10^{-4} M) and 0.5 $\mu\text{g/ml}$ (5.7×10^{-7} M), respectively. Linoleic acid is the only nematotoxic compound that so far has been isolated from *A. oligospora* (22). Ivermectin is a macrocyclic lactone and belongs to the avermectin group of antihelmintics and insecticides (4).

DISCUSSION

A deletion of the *PII* gene in *A. oligospora* by homologous recombination had only a limited effect on the capture and infection of nematodes. This could be explained by the presence of a significant residual proteolytic activity in the ΔPII mutants. However, the role of PII in virulence was demonstrated by analyzing mutants containing additional copies of *PII*. Two of these mutants showed an increased speed in capturing nematodes compared to the wild type.

That the activity of PII has a role in the initiation or development of traps in *A. oligospora* was indicated by the fact that the deletion mutants had a lower number and the multicopy mutants a larger number of traps compared to the wild type. It is well documented that the formation of traps in *A. oligospora* and several other nematode-trapping fungi is stimulated by peptides released from the nematode host (8). Proteases secreted by the fungi could have a role in the generation of such peptides by degrading and solubilizing proteins of the host cuticle. The chymoelastase serine protease Pr1 in *Metarhizium anisopliae* is induced by peptides from the insect cuticle produced by the basal activity of Pr1 and Pr2 (trypsin-like serine protease) (17). An alternative explanation for the stimulation of trap formation by the *PII* mutants is that they more rapidly digested the tissues of the infected nematodes, thus making nutrients available for the development of infection structures. Whether the activity of PII could affect the adhesion of nematodes to the traps of *A. oligospora* is difficult to evaluate, since the ratio of added nematodes to trap numbers varied between the various strains tested. However, that the *PII* transcript was not detected during adhesion (Fig. 1) and that treatment of *A. oligospora* with inhibitors against serine proteases did not affect the capture of nematodes (26) make it unlikely that PII has a role in adhesion.

The factor(s) that contributes to the comparable rapid killing of nematodes captured by nematophagous fungi has intrigued scientists for a long time. Early studies indicated that toxic fungal metabolites might be involved in the immobilization of nematodes by *A. oligospora* (16). Based on experiments demonstrating that only extracts from nematodes infected by the fungus, but not extracts from the fungus or nematodes alone, were toxic to nematodes, it was proposed that *A. oligospora* secretes a nematotoxic substance which paralyzes nematodes following the capture and penetration of them (16). This earlier paper stimulated a number of studies attempting to isolate and identify nematotoxic compounds from nematophagous fungi. Despite extensive efforts from a number of different laboratories, the results have been confusing (11). The only nematotoxic compounds identified so far from nematophagous fungi are fatty acids, including linoleic acid (12, 22). The observations that recombinant PII immobilized free-living nematodes in vitro indicate that PII can function as a nematotoxic compound. In agreement with the observations by Olthof and Estey (16), this toxic activity is produced after the nematodes are caught by the adhesive trapping nets (Fig. 1). Notably, the toxic activity of PII was significantly higher than that of two other commercially available serine proteases. The difference in nematotoxic activity between the enzymes was apparently not related to their hydrolytic activity on purified cuticle fragments, since proteinase K had a significantly higher

enzymatic activity on cuticle fragments but lower nematocidal activity than PII. In combination with the observation that PII is mainly degrading proteins in the intracellular fraction, we propose that the added recombinant PII may be transported inside the nematode and display an intracellular, not-yet-characterized cytotoxic effect.

This is the first report demonstrating that genetic engineering can be used to improve the virulence of a nematophagous fungus. In contrast to some other fungi, including the mycoparasite *Trichoderma harzianum* (9), it appeared to be difficult to insert a large number of copies of PII into the genome of *A. oligospora*. Previous studies of integration patterns of transformants generated by using the pAN7-1 vector also indicated a low frequency of multiple and tandem-repeated integrations in *A. oligospora* (28). Furthermore, the Southern analysis of generated mutants indicated a high frequency of rearrangements of the inserted DNA. Alternatively, the expression levels of PII could be modified or elevated by using a constitutive promoter. This approach was recently successfully used in the entomopathogen *M. anisopliae* (25). In the future it should be possible to express recombinant subtilisins with nematocidal activity in other organisms that are present in the habitat of parasitic nematodes (e.g., host plant).

ACKNOWLEDGMENTS

This study was supported by grants from the Swedish Natural Science Research Council, the Swedish Council for Forestry and Agricultural Research Council, and an EMBO short fellowship (to J.Å.). We thank Bo Ek for peptide sequencing.

REFERENCES

- Åhman, J., B. Ek, L. Rask, and A. Tunlid. 1996. Sequence analysis and regulation of a gene encoding a cuticle-degrading serine protease from the nematophagous fungus *Arthrobotrys oligospora*. *Microbiology* **142**:1605–1616.
- Bennet, J. W., and L. L. Lasure. 1991. Growth media, p. 444–445. In J. W. Bennet and L. L. Lasure (ed.), *More gene manipulations in fungi*. Academic Press, San Diego, Calif.
- Bonants, P. J. M., P. F. L. Fitters, H. Thijs, E. Den-Belder, C. Waalwijk, and J. W. D. M. Henfling. 1995. A basic serine protease from *Paecilomyces lilacinus* with biological activity against *Meloidogyne hapla* eggs. *Microbiology* **141**:775–784.
- Campbell, W. C. 1982. Progress and prospects in the chemotherapy of nematode infections of man and other animals. *J. Nematol.* **15**:608–615.
- Carrol, A. M., J. A. Sweigard, and B. Valent. 1995. Improved vectors for selecting resistance to hygromycin. *Fungal Gen. News* **41**:22.
- Christensen, T., H. Woeldike, E. Boel, S. B. Mortensen, L. T. Hjortshøj, and M. T. Hansen. 1988. High level expression of recombinant genes in *Aspergillus oryzae*. *Bio/Technology* **6**:1419–1422.
- Cox, G. N., M. Kusch, and R. S. Edgar. 1981. Cuticle of *Caenorhabditis elegans*: its isolation and partial characterization. *J. Cell Biol.* **90**:7–17.
- Dijksterhuis, J., M. Veenhuis, W. Harder, and B. Nordbring-Hertz. 1994. Nematophagous fungi: physiological aspects and structure-function relationships. *Adv. Microb. Physiol.* **36**:111–143.
- Flores, A., I. Chet, and A. Herrera-Estrella. 1997. Improved biocontrol activity of *Trichoderma harzianum* by overexpression of the proteinase-encoding gene *prb1*. *Curr. Genet.* **31**:30–37.
- Glass, N. L., and C. G. Donaldson. 1995. Development of primer sets designed for use with PCR to amplify conserved genes from filamentous ascomycetes. *Appl. Environ. Microbiol.* **61**:1323–1330.
- Jansson, H.-B., A. Tunlid, and B. Nordbring-Hertz. 1997. Biological control of nematodes, p. 38–50. In T. Anke (ed.), *Fungal bio/technology*. Chapman and Hall, Weinheim, Germany.
- Kwok, O. C. H., R. Plattner, D. Weisleder, and D. T. Wicklow. 1992. A nematocidal toxin from *Pleurotus ostreatus* NRRL 3526. *J. Chem. Ecol.* **18**:127–136.
- Lopez-Llorca, L. V., and W. M. Robertson. 1992. Immunocytochemical localization of a 32-kDa protease from the nematophagous fungus *Verticillium suchlasporium* in infected nematode eggs. *Exp. Mycol.* **16**:261–267.
- Mattern, I. E., J. M. Van Noort, P. Van den Berg, D. B. Archer, I. N. Roberts, and C. A. M. J. J. Van den Hondel. 1992. Isolation and characterization of mutants of *Aspergillus niger* deficient in extracellular proteases. *Mol. Gen. Genet.* **234**:332–336.
- Nordbring-Hertz, B. 1977. Nematode induced morphogenesis in the predacious fungus *Arthrobotrys oligospora*. *Nematologica* **23**:443–451.
- Olthof, T. H. A., and R. H. Estey. 1963. A nematotoxin produced by the nematophagous fungus *Arthrobotrys oligospora*. *Nature* **197**:514–525.
- Paterson, I. C., A. K. Charnley, R. M. Cooper, and J. M. Clarkson. 1994. Partial characterization of specific inducers of a cuticle-degrading protease from the insect pathogenic fungus *Metarhizium anisopliae*. *Microbiology* **140**:3153–3159.
- Punt, P. J., and C. A. M. J. J. Van den Hondel. 1992. Transformation of filamentous fungi based on hygromycin B and phleomycin resistance marker. *Methods Enzymol.* **216**:447–457.
- Punt, P. J., R. P. Oliver, M. A. Dingemans, P. H. Pouwels, and C. A. M. J. J. Van den Hondel. 1987. Transformation of *Aspergillus* based on the hygromycin B resistance marker from *Escherichia coli*. *Gene* **56**:117–124.
- Rosén, S., K. Sjollem, M. Veenhuis, and A. Tunlid. 1997. A cytoplasmic lectin produced by the fungus *Arthrobotrys oligospora* functions as a storage protein during saprophytic and parasitic growth. *Microbiology* **143**:2593–2604.
- Segers, R., T. M. Butt, B. R. Kerry, and J. F. Peberdy. 1994. The nematophagous fungus *Verticillium chlamyosporium* Goddard produces a chymoelastase-like protease which hydrolyses host nematode proteins *in situ*. *Microbiology* **140**:2715–2723.
- Stadler, M., H. Anke, and O. Sterner. 1993. Linoleic acid—the nematocidal principle of several nematophagous fungi and its production in trap-forming submerged cultures. *Arch. Microbiol.* **160**:401–405.
- Stirling, G. R. 1991. Biological control of plant parasitic nematodes: progress, problems and prospects. CAB International, Wallington, United Kingdom.
- Stirling, G. R., K. A. Licastro, L. M. West, and L. J. Smith. 1998. Development of commercially acceptable formulations of the nematophagous fungus *Verticillium chlamyosporium*. *Biol. Control* **11**:217–223.
- St. Leger, R. J., L. Joshi, M. J. Bidochka, and D. W. Roberts. 1996. Construction of an improved mycoinsecticide overexpressing a toxic protease. *Proc. Natl. Acad. Sci. USA* **93**:6349–6354.
- Tunlid, A., and S. Jansson. 1991. Proteases and their involvement in the infection and immobilization of nematodes by the nematophagous fungus *Arthrobotrys oligospora*. *Appl. Environ. Microbiol.* **57**:2868–2872.
- Tunlid, A., S. Rosén, B. Ek, and L. Rask. 1994. Purification and characterization of an extracellular serine protease from the nematode-trapping fungus *Arthrobotrys oligospora*. *Microbiology* **140**:1687–1695.
- Tunlid, A., J. Åhman, and R. P. Oliver. 1999. Transformation of the nematode-trapping fungus *Arthrobotrys oligospora*. *FEMS Microbiol. Lett.* **173**:111–116.
- Verdoes, J. C., P. J. Punt, J. M. Schrickx, H. W. Van-Verseveld, A. H. Stouthamer, and C. A. M. J. J. Van den Hondel. 1993. Glucoamylase overexpression in *Aspergillus niger*: molecular genetic analysis of strains containing multiple copies of the *gluA* gene. *Transgen. Res.* **2**:84–92.
- Verdoes, J. C., P. J. Punt, A. H. Stouthamer, and C. A. M. J. J. Van den Hondel. 1994. The effect of multiple copies of the upstream region on expression of the *Aspergillus niger* glucoamylase-encoding gene. *Gene* **145**:179–187.
- Wolstrup, J., P. Nansen, J. Grönvold, S. A. Henriksen, and A. Zorn. 1996. Toward practical biological control of parasitic nematodes in domestic animals. *J. Nematol.* **28**:129–132.