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## Studies on renal progenitor cells and kidney cancer

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# Studies on renal progenitor cells and kidney cancer

Jennifer Hansson



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DOCTORAL DISSERTATION

by due permission of the Faculty of Medicine, Lund University, Sweden.

To be defended at the main lecture hall, Medicon Village, Lund.

Friday 1<sup>st</sup> of April at 09.00 am.

*Faculty opponent*

Associate professor Nigel Mongan,  
Faculty of Medicine & Health Sciences  
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<p>Abstract</p> <p>Kidney cancer and renal injuries affect millions of people in the world resulting in high patient morbidity and mortality, as well as one of the most extensive medical costs for society. Upon renal injury the kidney has an endogenous ability to repair damaged tubules and restore kidney function, provided that the patient receives adequate supportive care in time. The molecular basis for this regeneration process is still not fully elucidated and the cellular origin for regeneration is extensively debated. Kidney cancer is characterized by vague clinical symptoms and pronounced resistance to common cancer therapies, like chemotherapy and radiation. Consequently, nearly 25% of patients present with spread disease already at diagnosis. For these patients a dismal prognosis awaits, with a 5-year survival rate of less than 10%. Thus, there is a desperate need for development of novel targeted therapies and diagnostic methods.</p> <p>In this thesis we describe a new cell type intermingled between the proximal tubular cells of the nephron in normal human kidney. These cells share marker expression with regenerating tubules and display stem cell-like abilities as well as distinct morphological properties, such as low mitochondrial content and extensive structural and anchorage protein expression. By developing a novel human explant model of ATN <i>ex vivo</i> we also show that these cells are more resilient to injury. These cells were detected in normal chimpanzee and pig kidney, but not in mice, not even after induced renal injury. Additionally, the transcriptional profile of these cells is similar to that of papillary renal cell carcinoma (pRCC), and correlate to worse prognosis in clear cell RCC (ccRCC). These results indicate that these cells survive renal insults to a higher extent than bulk proximal tubule cells, and become activated to repopulate the tubule, but also suggest that pRCC might originate from oncogenic transformation of these cells.</p> <p>In the second part of this thesis we show that the dopamine transporter SLC6A3 is highly expressed and functional in ccRCC, while only being expressed at very minute levels in normal kidney and other cancer forms. Additionally, we show that the SLC6A3 expression is affected by hypoxia inducible factor 2 alpha (HIF-2<math>\alpha</math>), an important protein in the cellular oxygen sensing system, which is ubiquitously expressed in the constantly pseudohypoxic ccRCC tumors. We further demonstrate that hypoxia induces SLC6A3 expression in normal kidney cells, but not in cells from other normal tissues like breast or vessels. These results show that SLC6A3 is a highly specific biomarker for ccRCC and that the endogenous features of this dopamine transporter may be utilized for the development of novel treatment modalities in ccRCC.</p>		
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Cover photo by: Jennifer Hansson

The photo shows a histological section of human kidney displaying a proximal tubule with a single proximal tubule progenitor cell (PTPC) positive for CD133 (red) while having low mitochondrial content, compared to the surrounding epithelial cells which are full of mitochondria (green). Nuclei are seen in blue.

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*Till min familj*



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# Abbreviations

AACD	Aromatic L-amino acid decarboxylase	CKI	Chronic kidney injury
ABC	ATP-binding cassette	CNS	Central nervous system
ADHD	Attention deficit hyperactivity disorder	CSC	Cancer stem cell
AKI	Acute kidney injury	DAT	Dopamine transporter
ALDH	Aldehyde dehydrogenase	DT	Distal tubule
AML	Acute myeloid leukemia	EMT	Epithelial to mesenchymal transition
ARF	Acute renal failure	ESC	Embryonic stem cell
ARNT	Aryl hydrocarbon receptor nuclear translocator protein	ESRD	End stage renal disease
ATN	Acute tubular necrosis	FACS	Fluorescence activated cell sorting
ATP	Adenosine triphosphate	FIH	Factor inhibiting HIF
BHD	Birt-Hogg-Dubé	GLUT	Glucose transporter
BMSC	Bone marrow derived stem cells	HIF	Hypoxia inducible factor
CA9/CAIX	Carbonic anhydrase IX	HR	Homologous Recombination
cAMP	cyclic adenosine monophosphate	HRE	Hypoxia response element
ccRCC	clear cell renal cell carcinoma	HSC	Hematopoietic stem cell
CD	Collecting duct	IFN- $\alpha$	Interferon alpha
chRCC	chromophobe renal cell carcinoma	IL-2	Interleukin-2
CKD	Chronic kidney disease	iPS cell	Induced pluripotent stem cell
		IRI	Ischemia reperfusion injury

LOH	Loss of heterozygosity	SPECT	Single photon emission computed tomography
mTOR	mammalian target of rapamycin	TAAR1	Trace amine associated receptor 1
mRNA	messenger RNA	TCA	Tricarboxylic acid
mRCC	metastatic RCC	TCGA	The cancer genome atlas
NOD/SCID	Non-obese-diabetic/severe combined immunodeficiency	TERT	Telomerase reverse transcriptase
NTT	Neurotransmitter transporter	TKI	Tyrosine kinase inhibitor
PD	Parkinson's disease	TLR-2	Toll like receptor 2
PDGF	Platelet derived growth factor	TNM	Tumor-node-metastasis
PD-1/PDL-1	Programmed death receptor 1/ ligand 1	TSC	Tuberous sclerosis complex
PEC	Parietal epithelial cell	UUO	Unilateral urinary obstruction
PHD	Prolyl hydroxylase-domain	VEGF	Vascular endothelial growth factor
PKA	Protein kinase A	VHL	von Hippel-Lindau
PKC	Protein kinase C		
pRCC	papillary renal cell carcinoma		
PT	Proximal tubule		
PTPC	Proximal tubule progenitor cell		
PTRC	Proximal tubular rare cell		
RA	Retinoic acid		
RCC	Renal cell carcinoma		
RNA	Ribonucleic acid		
ROS	Reactive oxygen species		
SLC	Solute carrier		

# List of papers

This thesis is based on the following papers, referred to in the text by their Roman numerals:

- I Isolation and characterization of progenitor-like cells from human renal proximal tubules.  
Lindgren D, Boström AK, Nilsson K, **Hansson J**, Sjölund J, Möller C, Jirstrom K, Nilsson E, Landberg G, Axelson H, Johansson ME.  
*American Journal of Pathology*. vol. 178 (2) pp. 828-37. 2011
- II Evidence for a morphologically distinct and functionally robust cell type in the proximal tubules of human kidney.  
**Hansson J**, Hultenby K, Cramnert C, Pontén F, Jansson H, Lindgren D, Axelson H, Johansson ME.  
*Human Pathology* vol 45 (2) pp 382-93. 2014
- III Species diversity regarding the presence of proximal tubular progenitor cells of the kidney.  
**Hansson J**, Ericsson A, Axelson H and Johansson ME  
*European Journal of Histochemistry* vol 60:2567. 2016
- IV Overexpression of functional SLC6A3 in clear cell renal cell carcinoma.  
**Hansson J**, Lindgren E, Nilsson H, Johansson E, Johansson ME, Gustavsson L and Axelson H  
Manuscript

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# Abstract

Kidney cancer and renal injuries affect millions of people in the world resulting in high patient morbidity and mortality, as well as one of the most extensive medical costs for society. Upon renal injury the kidney has an endogenous ability to repair damaged tubules and restore kidney function, provided that the patient receives adequate supportive care in time. The molecular basis for this regeneration process is still not fully elucidated and the cellular origin for regeneration is extensively debated. Kidney cancer is characterized by vague clinical symptoms and pronounced resistance to common cancer therapies, like chemotherapy and radiation. Consequently, nearly 25% of patients present with spread disease already at diagnosis. For these patients a dismal prognosis awaits, with a 5-year survival rate of less than 10%. Thus, there is a desperate need for development of novel targeted therapies and diagnostic methods.

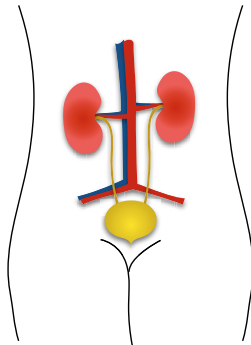
In this thesis we describe a new cell type intermingled between the proximal tubular cells of the nephron in normal human kidney. These cells share marker expression with regenerating tubules and display stem cell-like abilities as well as distinct morphological properties, such as low mitochondrial content and extensive structural and anchorage protein expression. By developing a novel human explant model of ATN *ex vivo* we also show that these cells are more resilient to injury. These cells were detected in normal chimpanzee and pig kidney, but not in mice, not even after induced renal injury. Additionally, the transcriptional profile of these cells is similar to that of papillary renal cell carcinoma (pRCC), and correlate to worse prognosis in clear cell RCC (ccRCC). These results indicate that these cells survive renal insults to a higher extent than bulk proximal tubule cells, and become activated to repopulate the tubule, but also suggest that pRCC might originate from oncogenic transformation of these cells.

In the second part of this thesis we show that the dopamine transporter SLC6A3 is highly expressed and functional in ccRCC, while only being expressed at very minute levels in normal kidney and other cancer forms. Additionally, we show that the *SLC6A3* expression is affected by hypoxia inducible factor 2 alpha (HIF-2 $\alpha$ ), an important protein in the cellular oxygen sensing system, which is ubiquitously expressed in the constantly pseudohypoxic ccRCC tumors. We further demonstrate that hypoxia induces *SLC6A3* expression in normal kidney cells, but not in cells from other normal tissues like breast or vessels. These results show that SLC6A3 is a highly specific biomarker for ccRCC and that the endogenous features of this dopamine

transporter may be utilized for the development of novel treatment modalities in ccRCC.

# The Kidneys

The maintenance of a relatively constant fluid volume and electrolyte composition is essential for homeostasis: the stable environment inside our bodies optimal for survival. Disruption of homeostasis, such as changes in ion concentrations, can have deleterious effects and can rapidly lead to severe illness or death. Homeostasis is maintained by several self-regulating biological systems, where many of the most central ones are localized to the kidneys, such as regulating the balance of electrolytes, blood pressure and acid base equilibrium.



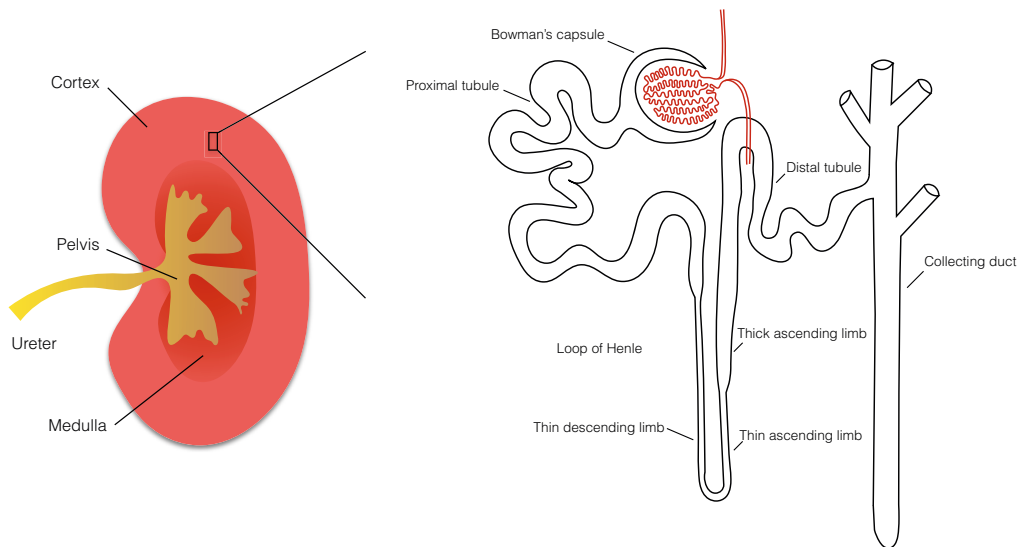
**Figure 1. Anatomical location of the kidneys**

The bean shaped kidneys are located in the retroperitoneal space of the body, on each side of the vertebral column. The kidneys are connected to the bladder via the ureters and to the aorta via the renal arteries.

The kidneys are two bean shaped organs located in the retroperitoneal space of the body, on each side of the vertebral column (Figure 1). For protection, the kidneys are surrounded by a tough fibrous cap, the fascia of Gerota underneath which a perinephric layer of fat resides. Interestingly, under conditions of starvation this fat compartment is the last that remains, reflecting the vital importance of protecting the kidney and thus maintaining kidney function. The kidneys are highly active organs that are estimated to consume 25% of the body's ATP in order to perform their daily tasks, despite their relatively modest weight (1). Apart from the aforementioned functions, the kidneys also serve important endocrine roles via renin and erythropoietin production. Erythropoietin is released in response to low oxygen tension and induce increased production of red blood cells in the bone marrow. The



enzyme renin is released from the juxtaglomerular apparatus of the kidney as a response to low arterial blood pressure, and exerts its function by activating angiotensin I and angiotensin II. The angiotensins induce general vasoconstriction and retention of salt and water, causing increased blood volume and a resulting increase of blood pressure. The kidneys also have a role in calcium homeostasis via the production of calcitriol, the activated form of vitamin D. Calcitriol stimulates the uptake of dietary calcium in the gut as well as inducing augmented calcium reabsorption in the kidneys, resulting in increased  $\text{Ca}^{2+}$  levels in the blood (2). However, the kidneys most apparent function is to filter the blood and eliminate metabolic end products and toxins through the production of urine. Each kidney is connected to the abdominal aorta via the renal arteries, which supply the kidneys with over 20% of cardiac output, by far exceeding the metabolic need of the kidney cells, while reflecting the kidneys essential role in plasma filtration (3). Each day, an astonishing 180 liter of primary filtrate is processed by the kidneys, where about 99,5% are reabsorbed into the body, generating approximately 1,5 liters of urine for excretion (1). This endless process of filtration and reabsorption is performed by the functional units of the kidneys, the nephrons.



**Figure 2. Schematic view of the kidney and the nephron**

The nephrons are the functional units of the kidneys and the majority of nephrons are located in the kidney cortex. The nephrons are composed of the filtering glomerulus followed by the proximal tubule (PT), where the major part of the reabsorption takes place. The PTs leads the filtrate further on into the Loop of Henle continuing into the distal tubule that connects with the collecting duct, which transfers the urine down to the urethra.

# The nephron

The nephrons are the functional units of the kidneys and each kidney contain about one million of these units, each organized into an intricate 3 dimensional architecture. The major part of the nephrons, roughly 90%, is located in the kidney cortex, the outer part of the kidneys (Figure 2). The nephrons are composed of a capillary tuft of the glomerulus, where filtration occurs. This is surrounded by Bowman's capsule where the primary filtrate is collected and guided into a complex tubular system. The renal tubules consist of the proximal tubule (PT), where most of the filtrate is reabsorbed, further leading into the loop of Henle in the renal medulla where it takes a sharp turn to then reach the distal convoluted tubules (DT) where a final modification of the primary filtrate takes place. The distal tubules connect to the collecting ducts (CD) ending in the renal pelvis where the urothelial tract originates and leads to the bladder where the end product urine is collected. The glomerulus is impermeable to larger proteins while salts, ions and all other organic molecules below a certain molecular size are freely filtered through the capillary tuft. The glomerular filtration barrier is composed of three layers: the fenestrated endothelial cell layer, the glomerular basement membrane of negatively charged proteoglycans and collagens, and the epithelial cell layer composed of podocytes with foot like protrusions that surround the capillary loops, allowing the filtrate of solutes and water to finally pass through their slit diaphragm into the Bowman's space (Figure 3).

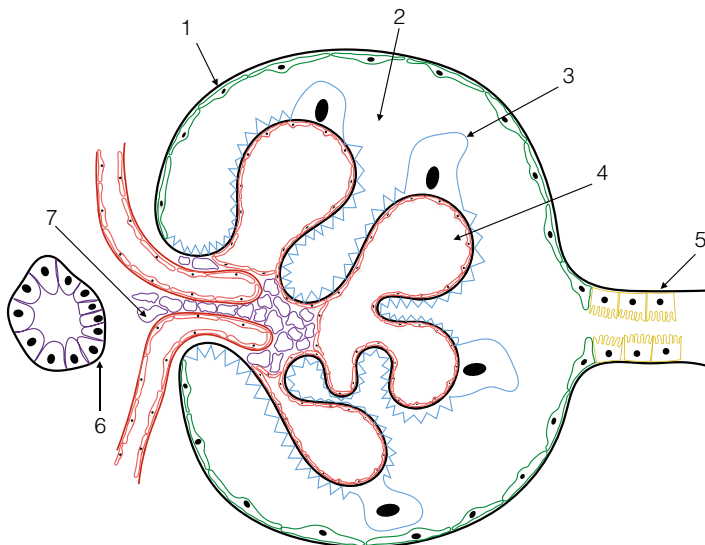


Figure 3. Schematic view of the glomerulus

1. Parietal epithelial cells (PEC) of the Bowman's capsule. 2. Bowman's space. 3. Podocyte. 4. Capillary loops. 5. Proximal tubule. 6. Macula densa. 7. Mesangial cells

The glomerulus also harbors mesangial cells, which besides having a phagocytic function also may regulate glomerular capillary blood flow due to their contractile ability. Following Bowman's space the filtrate enters the first tubular compartment, the PT, where essentially all glucose and amino acids are reabsorbed. The PTs also absorb 65% of the filtered water, sodium, potassium, chloride and bicarbonate. The principle for the reabsorption is vectorial transport and to accomplish this immense task the PT cells are endowed with an apical brush border in order to increase the surface area to the filtrate. Furthermore, numerous transporter protein systems are localized in the apical compartment, facing the tubular lumen, mainly performing secondary active transport of solutes down an electrochemical gradient for sodium. The foundation for this is basally localized  $\text{Na}^+/\text{K}^+$ -ATPases exporting intracellular sodium, thus creating the gradient. In order to fuel these  $\text{Na}^+/\text{K}^+$ -ATPases the PT cells are endowed with a massive abundance of mitochondria, accounting for some 60% of PT cell volume. In addition to reabsorption, the PT cells also actively secrete metabolic waste products, like organic acids and bases into the tubular lumen to ensure rapid excretion of potentially harmful substances. Further down the tubular system the filtrate encounters the loop of Henle that reaches deep into the medulla and have three distinctive segments. The cells of the thin descending segment have low metabolic activity with little active transport while being extensively permeable for water and urea. The water moves through the tubular layer by passive diffusion, due to the hyper osmotic pressure in the medulla caused by the active transport of  $\text{Cl}^-$  and accompanying passive transport of positively charged ions like  $\text{K}^+$  and  $\text{Na}^+$  in the ascending limb. The ascending limb is in turn virtually impermeable to water. Henle's loop in conjunction with the collecting ducts are the foundation for the counter current multiplier system without which the essential property of urine concentration would be impossible. This property allows us to save water by decreasing urinary output during thirst or the reverse if water intake is plentiful (2).

In the thick ascending limb the metabolic activity of the cells increase again, reflected by an increase in mitochondrial content. This segment reabsorbs an additional 25% of sodium chloride and potassium. The loop of Henle leads to the DT, which is highly convoluted and has similar reabsorptive properties to the thick ascending limb. The DT also contains parts of the juxtaglomerular apparatus, which is involved in feedback control of the glomerular filtration rate (GFR) and renal blood flow, and hence blood pressure. The distal tubules then connect to the CD. The late DT and the cortical CD contain two different cell types; the principal cells and the intercalated cells. The principal cells are responsible for  $\text{NaCl}$  reabsorption and  $\text{K}^+$  secretion, while intercalated cells reabsorbs  $\text{K}^+$  and have a role in controlling pH through the secretion of  $\text{H}^+$  or  $\text{HCO}_3^-$ . Worth noting is that the medullary CD stems from the ureteric bud in contrast to the rest of the tubular system, which stems from the metanephric mesenchyme. Finally in the CD, the concentration of the filtrate is fine tuned by a controlled reabsorption of water, via the hormone levels of

vasopressin, before being discharged into the bladder. Ultimately, the formation of the final product, urine, is essential for the body's ability to remove metabolic waste substances like urea, bilirubin, uric acid, hormone metabolites and creatinine (1, 2, 4).



# Stem and Progenitor cells

Stem cells are characterized by their ability to self-renew as well as their capacity to generate various differentiated cells. The stem cell can divide to generate one identical daughter cell as well as a progeny committed to differentiate into a mature specialized cell. Stem cells are classified as either embryonic or adult stem cells, also called tissue specific stem cells. Stem cells have several levels of differentiation potential. In mammals, the totipotent stem cell is found in the morula, the cluster of cells from the first cell divisions following oocyte fertilization. The totipotent cells can mature into all cells of the embryo as well as the extra-embryonic tissue, such as the placenta. The embryonic stem (ES) cells are derived from the inner cell mass of a blastocyst. These cells are pluripotent and can give rise to cells from the three germ layers: ectoderm, mesoderm and endoderm, comprising all the different specialized cell types in the body. Under the right culture conditions ES cells sustain their ability to self-renew indefinitely *in vitro*, while still maintaining their pluripotency (5, 6). This feature makes the cells excellent model systems for studying development, differentiation and function of different tissues

In contrast to the ES cells, the tissue specific stem cells can only give rise to a limited set of specialized cells within a given tissue. These cells can be multipotent or oligopotent, reflecting the potency for the amount of lineages they can give rise to (7). Tissue specific stem cells can be found in several organ systems in the adult body, most typically in tissues that need to continuously replenish their cells, as for instance in the bone marrow, where the hematopoietic stem cells (HSCs) endlessly generate new blood cells, or in the intestinal epithelium where the stem cells of the crypt rapidly replace the shed epithelial cells. Adult stem cells can also remain in a non-dividing state for a very long time until being activated by a need for normal tissue maintenance or by injury or disease, as in the case of stem cells in more quiescent organs like heart, liver and lungs (8-10). The concept of progenitor cells are often confused with stem cells, but in contrast to the latter, progenitor cells are restricted in their self-renewal capacity, that is, they can't divide indefinitely. Additionally, progenitor cells are more specialized than stem cells and they can in some cases be unipotent, that is, give rise to only one kind of mature cell, in which case they can also be termed precursor cells. However, similar to stem cells, some progenitor cells can give rise to several different specialized cell types and thereby be oligopotent (11).

Hematopoietic stem cells (HSC) provide a good example of a multipotent adult stem cell. The most primitive of the HSC is the long term hematopoietic stem cell (LT-HSC) that has a lifelong ability to self-renew and divide. The LT-HSC can give rise to the short term hematopoietic stem cell (ST-HSC) that has a shorter life span and a more restricted self-renewal. As the ST-HSC matures it gives rise to the multipotent progenitors, MPP, which differentiates into mature cells of the myeloid and lymphoid lineages (12).

### *The stem cell niche*

Adult stem cells reside in a niche, which represents a microenvironment that helps to maintain the stem cells in an undifferentiated and often quiescent state. Stem cell niches have been thoroughly investigated with single cell resolution in invertebrates like *Drosophila* and *C. elegans*, while the study of human niches have been hampered by the vast size of their tissues, as well as by an uncertainty of reliable markers for the true primitive stem cells. In a niche, stem cells can be clustered together like the stem cells of the hair follicle that reside in the bulb, or scattered through the tissue as in the case of satellite muscle stem cells that reside underneath that basal lamina of myofibers. It is postulated that the niche supports the stem cells in several ways; both with supporting cells as the paneth cells in the intestinal crypt, which are required to sustain the Lgr5+ intestinal stem cells (13), and by extracellular signaling as in the case of HSCs, which require the cytokine CXCL12, angiopoitin and stem cell factor (SCF) for their maintenance (9). Also relatively low oxygen pressure (pO<sub>2</sub> 1-8%) has been suggested to play a role for stem cell maintenance of HSCs, mesenchymal stem cells as well as neuronal stem cells (14). In this setting, lower oxygen levels (1%) decrease stem cell proliferation, suggesting a role in maintaining a quiescent state. However, substantial number of reports have ascribed niches to be located in close vicinity of vessels, such as in the bone marrow and spleen where HSC reside in the sinusoidal blood vessels (15). Additionally, what we define as hypoxia in cell culture conditions can be equal to physiologic normoxia, thus the role of oxygen tension in stem cell maintenance is not yet fully clarified.

### *Induced pluripotent stem cells*

In 2006 the Yamanaka group revolutionized the field of stem cell research when reporting that fully differentiated cells could be converted into pluripotent stem cells by only four factors, Oct 3/4, Sox2, Klf4 and c-Myc (16). This suggested that stemness might not be as static as previously thought, but a more plastic state. Induced pluripotent stem (iPS) cells are important tools for the studies of the molecular mechanisms that maintain and control pluripotency. The full potential of iPS cells in human disease is still not clear, but they bring hope to the field of regenerative medicine, with the notion that each patient's own somatic cells could be used for patient specific therapies.

Due to their remarkable regenerating abilities, stem cells have been a hot topic of research in the hope of discovering new ways to treat regenerative and genetic diseases. They also hold enormous potential as candidates for drug screens and toxicity tests. However, deeper knowledge of the genetic pathways that govern the regulation of stem cells is essential in order to use these remarkable cells to their full potential. From a scientific point of view, the presence of adult stem cells in an organ influences the interpretation of the organs regeneration ability and pathophysiology. Furthermore it adds an important aspect to tumor biology, since it has been shown that several cancers and tumors may arise by malignification of organ confined stem cells (17).

## Cancer stem cells

Cancer stem cells (CSC) represent a subpopulation of cells with the ability to self-renew and generate all different cells of a heterogeneous tumor, thereby replenishing the tumor with new cells and maintaining tumor growth. From a superficial point of view, the concept of cancer stem cells stands in contrast to another theory of cancer development, clonal evolution. This latter theory postulates that cancer will arise when a sufficient number of mutations have occurred in any given cell (18). This creates vicious cycle where mutations in the original cancer cell allows the progeny in the growing tumor to accumulate even more mutations, which may be different in individual progeny, thereby giving rise to intra tumor heterogeneity. The clonal evolution theory stipulates that all cells of a tumor harbor the same capacity to maintain and advance tumor growth, while the cancer stem cell theory postulate that only a small subset of cells has this ability However, there are also efforts to combine the two concepts that take into consideration aspects of both theories (19).

The cancer stem cell concept was first experimentally demonstrated in acute myeloid leukemia (AML), where a limited number of an isolated subpopulation of undifferentiated cells transplanted into NOD/SCID mice developed AML with fully differentiated tumor cells, while the remaining subpopulations were unable to give rise to tumor growth (20, 21). Whether or not the CSC originate from transformed normal stem cells or from more mature cells that gain the stem cell properties by mutations, is still not fully elucidated. An adult stem cell would require fewer mutations to gain CSC properties than a mature cell would, and there have been several reports suggesting that tissue stem cells are indeed the origin of many cancers (22-24). Nevertheless, several reports have also shown that CSCs can be created through dedifferentiation or epithelial to mesenchymal transition (EMT) of mature bulk cancer cells (25-27). Much like normal stem cells, CSC have been associated with the expression of multi drug resistance pumps of the ATP-Binding Cassette



(ABC)- family of transporters, contributing to therapy resistance in cancer treatment (28). Additionally, it is believed that conventional therapies like cytostatics affect dividing cells, while leaving the relatively quiescent CSCs unaffected, and that one thus need to specifically kill CSCs with targeted therapies to eradicate the tumor. However, since it has been shown that the bulk tumor cells can transition into CSCs it would be important to target both the bulk and the CSCs simultaneously to get rid of the tumor (18).

## Identification and Isolation

Several techniques have been applied in order to search for, and isolate, stem cells. The use of specific surface markers is one method, like CD34 and Thy-1 that has been ascribed to hematopoietic stem cells or CD133, CD44 and Lgr5 as markers for intestinal crypt cells (29-31). “Label retention” has also been used to identify stem cells. This functional assay is based on the notion that stem cell divide more slowly than their progeny, but also on the “leading strand hypothesis”. This hypothesis states that stem cells divide asymmetrically and that the stem cell always retain the original DNA strand, while the cells bound for differentiation receives the newly synthesized strands. Both of these notions imply that experimentally provided DNA labels, for example a BrdU pulse, would be retained in the stem cells while being washed out of their progeny after a long chase period, thereby allowing for identification of the stem cells by “lable retention”. However, concerns have been raised for whether this technique really labels true primitive stem *in vivo*, partly because some mature cells have been shown to stay non-dividing for a long time, such as paneth cells in the intestinal crypt, (32), but also because not all stem cells divide asymmetrically as described in the leading strand hypothesis (33).

A conceptually different method to isolate stem cells is to use stem cell associated biological functions, as for instance to utilize their inherent and necessary ability to protect themselves against cytotoxic insults. Stem cells have been shown to express ABC-transporters, which allow the cells to efflux substances that can be potentially harmful. This capacity can be used to isolate a so-called side population, by identifying the cells that pump out Hoechst 33 342 dye (34). Another way that stem cells can protect themselves is through an enzyme based detoxification system, based on high aldehyde dehydrogenase (ALDH) activity. ALDH enzymes convert potentially toxic aldehydes to their corresponding weak carboxylic acids (35). An additional feature of ALDH, in association to isolation of stem cells, relates to its enzymatic function to catalyze vitamin A (retinal) into retinoic acid (RA). RA signaling is thought to be essential for cell fate determination and thereby stem cell maintenance. The levels of enzymatic ALDH activity in cells can therefore be used to

isolate subpopulation of cells with stem cell characteristics. Fluorescent activated cell sorting (FACS) based on ALDH activity has successfully been used to sort out stem cells of several origins, in particular HSC, but also breast cancer stem cells, intestinal mucosal stem cells and liver cancer stem cells (36).



# Renal regeneration

## Acute Kidney Injury

Acute kidney injury (AKI) is clinically characterized by a rapid decrease of urine output, oliguria, where between 50-400 ml urine is produced per day in contrast to the normal 1,5 liters of urine, as well as elevated creatinine levels, and/or decreased glomerular filtration rate (GFR) (37). AKI is a life threatening condition that leads to disturbed fluid and electrolyte balance, along with a decreased ability to eliminate metabolic waste products, uremia. AKI represent 3,2- 9,6% of causes for hospital admissions in industrialized countries, and the AKI incidence is estimated to be somewhere around 2000-3000 cases / million individuals / year (38). The most severe form of AKI is acute renal failure and around 6% of all AKI cases require renal replacement therapy and the mortality rates for these patients are as high as 60% (39). Additionally, patients who survive AKI are at higher risk do develop chronic kidney disease (CKD) (38). All in all, it is estimated that over 2 million people die of AKI each year (38, 40).

## Acute Tubular Necrosis

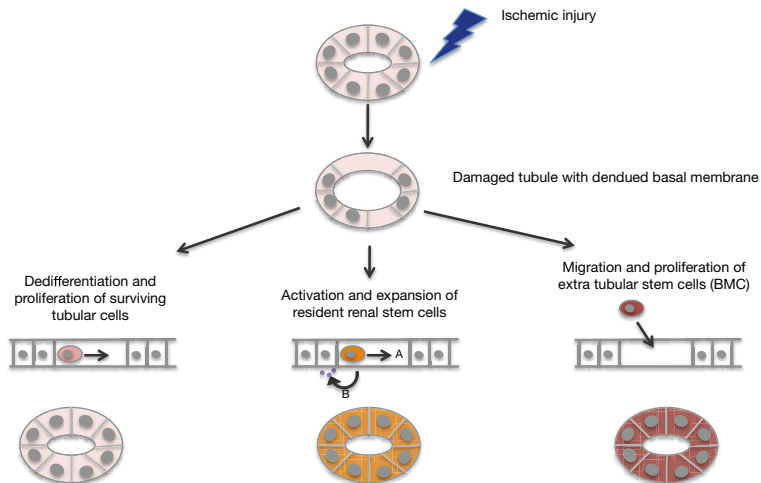
One of the most common causes of acute kidney failure is acute tubular necrosis (ATN) (41). This condition can be caused by a number of insults, for example severe trauma, septicemia and acute pancreatitis (42). ATN can be divided into ischemic ATN and nephrotoxic ATN. Ischemic ATN is associated with hypovolemic shock, but can also be caused by hemolytic crisis, and is due to impaired renal blood flow and thereby diminished oxygen delivery to the tubular cells. Nephrotoxic ATN is less common and caused by intoxication, for example by heavy metals or antibiotics, but can also be caused by ingestion of poisonous mushrooms (41). Renal tubular cells, in particular the proximal tubular cells, are extremely energy dependent and thus very sensitive to disturbances in nutrient and oxygen delivery. The cells of the proximal tubule are packed with mitochondria and are virtually incapable of using non-oxidative glycolysis as their main source of energy (43). Impaired renal blood flow, or toxic renal insult, therefore leads to tubular cell dysfunction. The cells begin to loose their important epithelial polarization, and Na<sup>+</sup>-ATPases shift from their baso-lateral

position to the apical side. This leads to disrupted vectorial transport capacity and consequently that the kidneys essential fluid reabsorption becomes impaired. Less sodium is thus reabsorbed in the proximal tubules, which leads to higher concentrations of  $\text{Na}^+$  reaching the distal tubules where the tubuloglomerular feedback system responds by intrarenal vasoconstriction. This results in decreased glomerular filtration rate (GFR) and oliguria, clinically corresponding to AKI (44). The insult to the renal tubules can also cause the tubular cells to detach, morphologically seen as cells characteristically sloughing into the tubular lumen, leaving behind denuded basal membranes. The shedding of tubular cells can clog up the renal tubular system and in this way contribute to the decreased urine output. The obstructed urine flow can also cause increased intratubular pressure, resulting in leakage from the injured tubular cells into the interstitium, causing edema. However, remarkably, the serious condition of ATN can be reversible through the process of renal tubular regeneration (42).

## Regeneration

Neonephrogenesis denotes the ability to regenerate entire nephrons following injury. This capacity is present in invertebrates like the fruit fly (45), and in lower vertebrates like the zebrafish (46). In contrast, human and other mammals have lost this ability and the renal regeneration capacity is more limited, encompassing tubular, and to some extent, podocyte regeneration. The renal tubules are normally almost mitotically quiescent, in contrast to several other transporting epithelia. Nevertheless, upon injury, the nephrons demonstrate a pronounced capacity to regenerate damaged tubules, and ATN can be reversible if the patient receives supportive care in time. Maintenance of tubular structural integrity is essential for kidney function, and proper clinical management after ATN can thus be the difference between life and death. The regeneration of the renal tubules is based on: critical coverage of denuded basal membranes to ensure structural integrity, cellular proliferation to replace the cells lost by apoptosis or necrosis and finally differentiation to a fully functional tubular cell to regain transporting capacity. In the developing kidney, the cap mesenchyme harbors multipotent progenitors with the ability to give rise to all the cellular components of the nephron. However, the cellular source for renal regeneration in adult kidney is still debated (Figure 4). The classical view is that tubular regeneration is performed by the least damaged tubular cells, which subsequently dedifferentiate to repopulate the tubule (47, 48). This view has in recent years gathered supporting evidence through several animal studies using lineage tracing (49-51). Migrating bone marrow derived stem cells (BMSC) have also been suggested to take part in tubular regeneration (52). Although, animal models has shown that BMSC can ameliorate the symptoms after induced acute renal injury,

they do not seem to directly take part in the regeneration (53). The role of the BMSC might rather be production of protective paracrine factors that can facilitate the epithelial regeneration (54). Interestingly, a phase I clinical trial using bone marrow derived mesenchymal stem cells for therapeutic use, in patient undergoing cardiac surgery with high risk of developing AKI, indeed showed protective effects with regards to kidney injury (55).



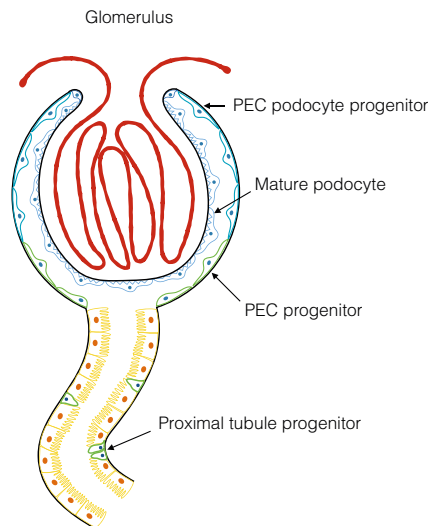
**Figure 4. The hypotheses of the source for renal regeneration**

Schematic image of the different hypotheses concerning renal regeneration. The classical view is that random surviving tubular cells dedifferentiate, divide and repopulate the tubule (left). Another view is that regeneration is performed by resident progenitor cells that become activated in the aftermath of renal injury and divide to repopulate the tubule (A) or that the progenitor cells secrete protective factors that help the bulk tubular cells survive the insult (B) (middle). A contrasting view is that extratubular stem cells like bone marrow derived stem cells (BMC) migrate to the tubule and integrate into the tubular epithelium and divide to repopulate the tubule (right). Modified from Anglani et al. *Frontiers in Bioscience* 2008.

### *Renal progenitor cells*

An alternative hypothesis regarding tubular regeneration is that it is performed by adult resident renal stem cells (56). The localization of these putative progenitor cells have been ascribed to the renal papilla (57), renal interstitium (58, 59), the Bowman's capsule (60, 61), or to the renal tubules (62, 63). These cells may be more resistant to damage and become activated in the wake of renal damages. In human kidney, a progenitor cell population was described in the parietal epithelial cells (PEC) of the Bowman's capsule in 2006 (64). These cells were characterized by CD133 and CD24 expression, markers that have been ascribed to stem cells in several organs (65-68), additionally, CD24 is also expressed in the metanephric mesenchyme during renal development (69). Isolated human CD133+ CD24+ cells were shown to be multipotent and able to develop into several lineages *in vitro*, such as osteogenic cells

and adipocytes, as well as tubular cells. These cells were also able to integrate into renal tubules of SCID mice with glycerol induced renal injury, as well as to reduce the renal damage (64). The PEC progenitor cells were later subdivided into two groups based on their location in Bowman's capsule and on the expression of podocalyxin (PDX). PEC located close to the vascular pole were PDX positive and able to differentiate into podocytes, while PEC at the urinary pole were PDX negative and able to differentiate into both podocytes and tubular cells. The glomerular podocytes are post-mitotic, meaning that they are unable to divide and proliferate. Consequently, if podocytes are injured or lost, glomerular function is dependent on cellular replenishment from an alternative source. PEC of the Bowman's capsule has lately been suggested as the main source of podocyte regeneration (60, 61, 64), but also renin-producing cells in the juxtaglomerular apparatus have been shown to contribute to podocyte regeneration (70).



**Figure 5. Location of the putative renal progenitor cells**

The parietal epithelial cells (PEC) progenitor cells are located in the Bowman's capsule and are positive for CD24 and CD133. PEC located near the vascular pole are thought to give rise to podocytes, and PEC located near the urinary pole is thought to give rise to other PEC as well as to tubular cells. The PT progenitors are scattered among the tubular cells and are often located at plicae positions, where the PT make hairpin turns, and are sometimes found in duplets.

In 2011, our group described a rare population of divergent cells inconspicuously scattered among the differentiated cells of the PT in human kidney (Figure 5) (Discussed in detail in Paper I) (71). These cells were isolated based on high enzymatic activity of ALDH and displayed progenitor features such as clonogenic capacity and anchorage independent growth, while the bulk PT cells (ALDH<sup>low</sup>) did not. Additionally, ALDH<sup>high</sup> cells displayed striking marker similarity to PEC

progenitor cells and expressed CD24, CD133 and vimentin along with many other markers that also were expressed by the PECs. We later showed that the scattered tubular cells have a distinct morphology indicative of increased adherence to the basement membrane, higher mechanical resilience as well as a dramatically lower mitochondrial content compared to the neighboring PT cells. Moreover, we developed a novel human explant culture, simulating ATN *ex vivo*, demonstrating that these cells were more resilient to insult than bulk PT cells (72) (Discussed in Paper II). This finding was strengthened by Angelotti et al. that showed that human renal CD133+ cells indeed were more resistant to death following exposure to nephrotoxic agents compared to other tubular cells. In xenograft experiments, it was also demonstrated that isolated human tubular progenitors, distinguished from the PEC progenitors via the lack of CD106 expression, were able to integrate into renal tubules of SCID mice following renal injury and to generate new epithelial cells that improved the recovery of renal function (73).

Sallustio et al. broadened the view regarding the functional role of these scattered tubular progenitor cells by demonstrating that the cells could protect bulk PT cells against cisplatin induced damage. This protection mechanism was Toll like receptor-2 (TLR2)-dependent and achieved via exosomal shuttling of protective factors from the tubular progenitors to the injured PT cells (74).

There is, however, an alternative interpretation of these scattered tubular cells. Based on the characteristic marker expression of these cells, which other than CD24 and CD133, also express Vimentin, CD44, KIM-1 and annexin-3 (75), some groups have interpreted these markers as, in themselves, signs of dedifferentiation, and that this is in fact an induced phenotype that any differentiated tubular cell could adopt upon injury (76). Thus they interpret the discovery of scattered cells in human kidney samples merely as signs of renal wear and tear, possibly due to old age of the patients from where the renal tissue is derived. However, we have demonstrated that these cells are present also in young healthy kidneys from humans of age 2 years and up (72). Additionally, these cells have a characteristic paucity of mitochondria in comparison to the massive mitochondrial load of bulk PT cells, of which the fully mature PT cells would quickly have to get rid of if this were indeed an induced phenotype. (These matters are discussed in detail in Paper II).



### *Lineage tracing and cell fate labeling in mice*

In order to investigate which cells are responsible for regeneration, and simultaneously try to answer the question whether or not progenitor cells exist in adult kidney, several lineage tracing experiments have been performed. Lineage tracing is performed in transgenic mice that carry a specific promoter, which drives a reporter gene conveying a molecular mark, for example fluorescent proteins such as GFP (green fluorescent protein) or  $\beta$ -galactosidase, to the cell where the promoter is active. Since the activation of the marker gene is irreversible, the marked cells progeny then inherits this molecular mark. In this way, lineage tracing allows identification of all the progeny stemming from a defined single cell. The lineage tracing in kidney have been performed using a variety of promoters, some with the intention to mark a specific cell population, like the proposed renal progenitor cells, and other with a more wide approach intending to label all differentiated tubular cells. I will go through some of them here and also what critique that has been raised concerning the conclusions that has been drawn from the experiments.

Humphreys et al. used the *Six2* promoter to mark all cells from the nephron (except collecting duct). *Six2* is expressed during embryogenesis and marks the stem cells of the metanephric mesenchyme that give rise to all differentiated cells of the mature nephron, except for the cells of the collecting duct, which instead are derived from the ureteric bud. Using this model in conjunction with ischemia reperfusion injury (IRI), which simulates human ATN, they could show that the injury-induced regeneration was performed by already labeled tubular cells and could thus exclude extra renal cells like BMSC as a potential cellular source for regeneration (50). However, in this experiment the labeling could not distinguish between fully differentiated tubular cells or a potential progenitor population, and thus no conclusion could be drawn of which intra-renal cell type that achieved the regeneration. The same group tried to resolve this issue using two different subsequent DNA-labeling thymidine analogue injections following IRI, CldU (5-chloro-2-deoxyuridine) 24h after induction of injury and IdU (5-iodo-2-deoxyuridine) 45h after IRI (49). They hypothesized that if a progenitor population indeed existed and were responsible for the regeneration, then the cells of the regenerated tubule would be double labeled. However, only a small fraction of tubular cells were double labeled and the researchers drew the conclusion that the regeneration was a purely stochastic event, performed by surviving differentiated tubular cells and not a progenitor population. These conclusions have been met with some criticism claiming that cells that are labeled with only one of the thymidine analogues not necessarily have completed cell division (77). Lombardi et al. argue that cell cycle labeling only indicate that the cell has entered the cell cycle but not that it has committed to dividing. This notion has been demonstrated in liver where most hepatocytes shows signs of cell cycle entry following partial hepatectomy, but the vast majority of these cells never undergo cell division (78). Additionally, pathological

investigations of injured renal tissue following mercury chloride injections in rats suggest that mitosis is a rare event (79).

In another lineage-tracing experiment Appel et al. used an exogenous promoter composed of a human and rabbit podocalyxin promoter, termed PEC-rtTA, to drive an inducible expression of LacZ (60). Podocalyxin is normally expressed in podocytes, while this model, quite surprisingly, the promoter marked PECs and also scattered cells within the PTs. In mice, nephrogenesis is active also after birth, and using this model on juvenile mice the researchers could show that PEC cells could migrate along the vascular tuft and mature into differentiated podocytes. In a subsequent study by the same lab the researchers investigated the marked scattered tubular cells further (75, 76). The scattered cells expressed several molecular markers common to the ones reported for the putative progenitor population in human such as annexin A3, CD44, KIM1 and src suppressed C-kinase. These cells were then followed after induced IRI. The frequency of marked scattered cells did not change following damage, however, if the mark was induced during injury more cells became positive. The authors thus drew the conclusion that these scattered tubular cells represent a phenotype that can be induced from any differentiated tubular cell following damage, rather than a resident progenitor population. However, no functional studies were performed using these cells, nor any other phenotypic description other than that some markers were shared with the human CD133+/CD24+ cells, although these markers are not the most characteristic for the human cells and they are also expressed in other mature renal cells. It has thus been suggested that the scattered cells marked using the PEC-rtTA promoter is not necessarily the same population that has been described in human kidney (77, 80).

In another study by Langworthy et al. it was demonstrated that mice heterozygous for NFATc1 are more sensitive to HgCl<sub>2</sub> induced renal injury (81). Additionally, NFATc1 driven LacZ expression labeled cells that were more resistant to the HgCl<sub>2</sub> induced renal injury, also the NFATc1 positive cells progeny were responsible for 75% of the regenerated cells. The authors interpreted NFATc1 positive cells as a progenitor population. However, no NFATc1 positivity was detected in the uninjured kidney cortex, nor was NFATc1 expressed during kidney development. Taking these issues into account one may also interpret the findings as that NFATc1 is expressed by dedifferentiated surviving tubular cells that proliferate to restore the tubule.

Kusaba et al. took a different approach and used the promoter of the Slc34a1 (sodium dependent inorganic phosphate transporter) to mark all differentiated PT cells (51). Upon IRI they could not detect any dilution of the marker, suggesting that differentiated PT cells, rather than an unmarked population of stem cells, performed regeneration. However, a putative progenitor population might, to some extent, express the same markers as the differentiated cells that they are destined to become

(82-84). If this is indeed the case regarding Slc34a1, then this labeling tactic would not be able to discern between progenitor based regeneration or regeneration driven by dedifferentiation of mature cells. Additionally, the researchers reported that only 55% of the cells were targeted in the S3 segment of the PTs, which is the most sensitive segment for ARI (85), and also suggested to be the primary site for progenitors (73).

Since there is a lack of a definite progenitor marker in mouse, one cannot with certainty tag progenitor cells using a specific promoter segment. Another approach is to specifically label all cell and follow their clonal expansion following injury. This was recently performed by Rinkevich et al. that used “rainbow mice” where the confetti reporter was driven by the ubiquitous actin promoter. When expressed, the confetti reporter recombines in a stochastic way and then expresses one out of four fluorescent labels in a random manner. In this way, all single cells are individually marked and the expansion of a certain clone is seen as a cluster of unicolored cells, in this otherwise confetti-colored pattern of cells. This method allowed for an unbiased way to follow the clonal expansion of individual cells, both in renal development and following induced renal injury. The study demonstrated that cellular renewal and regeneration in the mouse kidney was performed by clonal expansion of fate-committed precursors, in a segment specific manner (86). This may suggest that unipotent progenitors are located along the different tubular segments of the nephron, and that these cells are responsible for, and restricted to, regeneration of the specific segment in which they reside.

Finally, the discrepancies regarding the cellular source for renal regeneration may be explained by the fact that mouse and human are not one and the same. They differ in longevity, as well as reproductive ability and also biological size and length of nephron. The shortage of convincing evidence for adult tubular progenitors in mice, does not exclude that such progenitors exist in humans, but may suggest that regeneration might be performed in different manners, and possibly by different cells dependent on the severity of the damage, in different species. (This matter is discussed more in Paper III in this thesis.) In the end, deeper knowledge of the mechanisms of regeneration as well as of the features of renal progenitor cells may provide important clues for the development of novel therapeutic strategies for AKI and CKD.

# Kidney Cancer

Kidney cancers comprise all tumors stemming from the kidney, including transitional cell cancer of the renal pelvis and the childhood malignancy Wilms tumor. The most common of kidney cancers, embodying over 90% of all cases, are renal cell carcinomas (RCC). These tumors arise from the tubular epithelium of the kidney and represents 2,4 % of all cancers worldwide. Each year around 338.000 people are newly diagnosed with kidney cancer (87), and 143.000 people succumb to the disease, making it the 16<sup>th</sup> most common cause of cancer related death. RCC is more common in men than women, with a ratio of 2:1, for reasons that are not yet clarified. The disease also represents the ninth most common cancer in men, while it is the 14<sup>th</sup> most common cancer in women. Globally, the incidence of kidney cancer is rising, however, the prevalence varies greatly between countries. Less developed countries display a lower incidence although the mortality rates are similar to those of industrialized countries with more diagnosed cases (88). The growing rates of diagnosed RCC may partly be explained by the increase of accidentally discovered cases due to novel imaging modalities and abdominal scans for unrelated conditions.

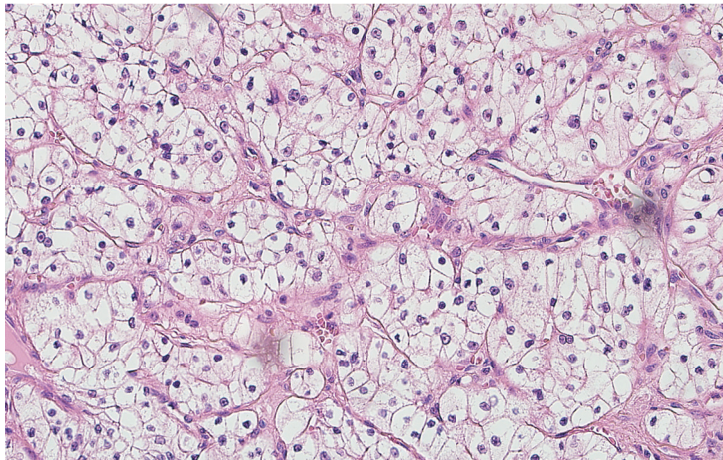
RCC is a slow growing disease, with an estimated growth rate of only 0.28 cm /year (89), and the mean age of diagnosis is at 64 years (90). The causes for kidney cancer are poorly understood, but age, smoking, obesity and hypertension are risk factors (91). For smokers, the relative risk for developing RCC is 1.38 compared to never smokers (92). It has been suggested that this increased risk is due to chronic hypoxia in the renal tissues as well as carbon monoxide exposure and DNA damage induced by tobacco associated N-nitrosamine (93). There is a strong correlation between being overweight and developing RCC, it is estimated that as much as 40% of all RCC cases in USA and 30% of cases in Europe are associated with high body mass index (BMI) (94). The reasons for this increase are not clear, however, it has been suggested that “adipokines” excreted from adipocytes may create a tumor-friendly environment and also induce angiogenesis (95). Additionally, chronic renal diseases increase the risk of developing RCC, possibly due to the persistent inflammatory microenvironment with high levels of inflammatory cytokines and/or due to genetic instability caused by disease associated hypoxia, uremia, and increased growth factor signaling (96). There are also hereditary causes for RCC, such as specific gene mutations, although they represent a minor part, and are estimated to constitute approximately 4% of all cases (97).

## Renal cell carcinoma subtypes

Renal cell carcinoma is a collection of several histologically and phenotypically distinct subtypes that arise along different parts of the nephron. The classification of the subgroups is based on histological and genetic features; accordingly, RCCs are divided into clear cell RCC (ccRCC), papillary RCC (pRCC), chromophobe RCC (chRCC), oncocytomas, collecting duct (Bellini's) RCC or unclassified RCC. These represents the predominant subtypes although there are at least 16 different forms of RCC reported (98).

### *Clear cell renal cell carcinoma*

ccRCC represent the vast majority of all RCC cases, 70-85% (99), and are thought to originate from the PT epithelium of the nephron. This subtype is histologically characterized by fat and glycogen accumulation, and as the name indicates, these features cause the cells to appear “clear” on hematoxylin & eosin (H&E) stained tissue sections (Figure 6).



**Figure 6. Clear cell renal cell carcinoma histology**

Hematoxylin and eosin staining of a histological section of a ccRCC tumor, displaying characteristic “clear” tumor cells and extensive vasculature.

ccRCC are extremely well vascularized tumors, due to high excreted levels of the angiogenesis inducing protein, vascular endothelial growth factor (VEGF). The high VEGF levels are explained by loss of the von Hippel-Lindau (VHL) protein, which occurs in almost all cases of ccRCC. The VHL protein marks the hypoxia inducible factors (HIFs) for degradation at normal oxygen pressure. Loss of VHL causes stabilization of the HIFs and consequently transcription of genes of the *cellular*

*adaptive response to hypoxia*, such as *VEGF*. Additionally, in tumors with wild-type VHL, mutations in *TCEB1* can be found. *TCEB1* encodes the Elongin C protein, which is a part of the VHL-complex responsible for degrading HIF under normoxic conditions. These mutations are mutually exclusive from *VHL* mutations and chromosome 3p loss and occur in 1-5% of all ccRCC cases (100, 101). The fact that the HIF pathway is affected also in wild-type VHL tumors, further stresses the importance of this pathway in ccRCC tumorigenesis. Most tumors tend to rely on aerobic glycolysis, in combination with lactic acid fermentation, instead of oxidative phosphorylation to meet their energy demand, a phenomenon called the Warburg effect (102). By bypassing the tricarboxylic acid (TCA) cycle following glycolysis, and instead using lactate fermentation, significantly lower amounts of ATP is generated. However, this process also allows for the synthesis of membrane components and cellular building blocks that are important for cellular proliferation. Due to the loss of VHL and hence constitutive activation of the HIF, the Warburg phenomenon is especially prominent in ccRCC. Additionally, it has been shown that glucose metabolism is increased with higher grade in ccRCC and that the glutathione pathway is upregulated to inhibit potentially harmful reactive oxygen species (ROS) (103). In this study, an upregulation of tryptophan catabolism was also noted. Metabolites of the essential amino acid tryptophan has been associated with immune suppression in cancer (104).

#### *Papillary renal cell carcinoma*

Papillary RCC is the next most common variant of RCC, embodying 10-15% of all RCC cases and is, similar to ccRCC, thought to emanate from the PT. pRCC has been given its name due to the characteristic papillary growth pattern of the tumor cells. There are two histologically diverse subtypes of pRCC; the more benign type 1, which is characterized by cuboidal epithelial cells and uniform “low grade” nuclei, and the more aggressive type 2 that display a pseudostratified epithelium with “high grade” irregular nuclei (105). pRCC occurs in both hereditary and sporadic forms, which both can be connected to specific genetic mutations. Hereditary, and to a lesser extent, sporadic type 1 pRCC is associated with activating mutations of the proto-oncogene *MET*, a tyrosine kinase receptor, which in these cases becomes constitutively active and thereby increase cellular proliferation. The familial form of type 2 pRCC is associated with loss of the tumor suppressor fumarate hydratase (FH), which causes hereditary leiomyoma renal cell carcinoma (HLRCC). FH converts fumarate to malate in the TCA cycle and loss of this enzyme thus lead to accumulation of fumarate. The increased levels of this metabolite cause VHL independent HIF stabilization by inhibiting the prolyl hydroxylases (PHD), which marks HIFs for VHL recognition. Surprisingly, type 2 pRCC is phenotypically and genetically very different from VHL mutated tumors, highlighting that VHL has more functions than simply targeting the HIFs for destruction. Additionally, it has been shown that the fumarate accumulation lead to HIF independent oncogenic

effects by inhibiting Keap1 and thereby stabilizing the protooncogene Nrf2, a crucial transcription factor in the antioxidant response (106). FH mutations have not been detected in sporadic type 2 pRCC, however, the hereditary and sporadic forms both display upregulation of antioxidant genes such as *AKR1B10*, which may explain their shared phenotype (107). Other rare forms of hereditary pRCC can be attributed to mutations in the tumor suppressors *TSC1* and *TSC2*, encoding hamartin and tuberlin respectively. These mutations cause tuberous sclerosis complex, autosomal dominant disease where patients have a predisposition to develop tumors in several organs such as brain, lung, kidney and skin (108). Additionally, these patients also have an increase risk of developing ccRCC and chRCC.

### *Chromophobe renal cell carcinoma*

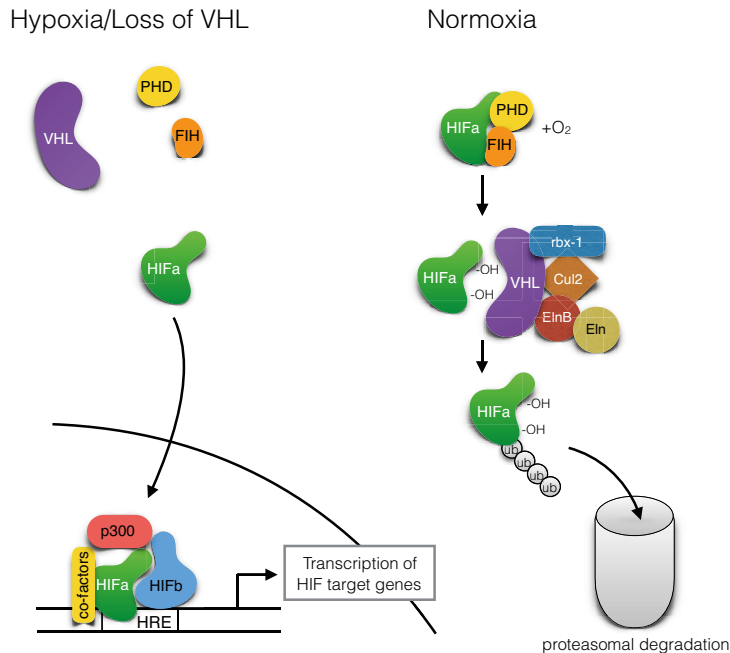
Chromophobe RCCs are low grade tumors, with a considerably better prognosis than the above mentioned subtypes, and represent 4-5% of all RCC cases (109). The cell of origin for chRCC is also suggested to stem from a different, more distal part of the nephron than ccRCC and pRCC does (110, 111). Also in this subgroup the name of the tumor is reflected by its histological appearance. In H&E-stainings chRCC tumor cells thus generally appear pale, but can also sometimes harbor eosinophilic granules in the cytoplasm. Expression of the proto-oncogene *KIT* at well as loss of chromosome 1, 2, 6, 10, 13, 17 and 21 is quite often seen in chRCC (112). A study using data from the cancer genome atlas (TCGA) showed that the most frequently mutated genes in chRCC are *TP53* (in 32% of cases) and in *PTEN* (in 9% of cases). The same study also showed that chRCC display recurrent genomic rearrangements affecting the *TERT* promoter and inducing elevated *TERT* expression (110). Hereditary chRCC is associated with loss of function mutations in the gene coding for follicullin, *BHD*, causing Burt-Hogg-Dube syndrome. Patients with BDH also develop lung cysts and oncocytomas, a benign form of RCC. Follicullin is suggested to play a role in the cells energy sensing mechanism via the AMPK and mTOR pathways (113), and in an animal model of BHD induced kidney cancer, the mice displayed prolonged survival after treatment with the mTOR inhibitor rapamycin (114).

# The genomic landscape of ccRCC

## The von Hippel-Lindau gene and hypoxia inducible factors

ccRCC is characterized by a virtually obligatory loss of VHL. In 80% of all cases of sporadic ccRCC, VHL is mutated with accompanying loss of heterozygosity due to chromosome 3p deletion (115). In an additional 5-10% of cases, VHL is silenced via methylation (116). Hereditary ccRCC represents approximately 2-3% of all cases and are also characterized by a loss of VHL, in this case due to a germline mutation of VHL, causing von Hippel Lindau disease. Apart from ccRCC, patients with VHL-disease also develop pheochromocytomas, renal and pancreatic cysts, neuroendocrine tumors, retinal angiomas and hemangioblastomas (113). These patients also have an earlier onset of ccRCC than sporadic ccRCC, with the mean age of diagnosis 37 years (90). The VHL protein pVHL is the substrate recognition subunit of a E3 ubiquitin ligase complex composed of Elongin B, Elongin C, cul-2 and rbx-1, which is responsible for the degradation of the HIF-1 $\alpha$  and HIF-2 $\alpha$  at normal oxygen levels (Figure 7). When enough oxygen is present PHDs hydroxylate HIF- $\alpha$ s on specific proline residues. This hydroxylation makes the HIF- $\alpha$ s recognizable for VHL that can bind the proteins and mediate ubiquitylation, which in turn signal for proteasomal destruction. Additionally, the HIF- $\alpha$ s are regulated by the enzyme factor inhibiting HIF (FIH) that, at normal oxygen levels, hydroxylate the proteins at an asparagine residue. This modification inhibits the HIF- $\alpha$ s from binding to their transcriptional co-factors, p300/CBP and thereby also impairs their capacity to initiate transcription. At hypoxia, no hydroxylation occurs and the HIF- $\alpha$ s become stabilized and translocate into the nucleus where they form a heterodimer together with HIF- $\beta$ , also known as aryl hydrocarbon receptor nuclear translocator protein (ARNT). This complex subsequently binds specific DNA segments: hypoxia responsive elements (HRE) and recruit co-factors to initiate transcription of genes in the adaptive response to hypoxia. Without a functional pVHL, as in the case of ccRCC, the cells experience pseudohypoxia with transcription of the downstream targets of HIFs even at normal oxygen levels. Consequently, the cells express high levels of genes involved in angiogenesis, glucose uptake, acid-base balance and metabolism, such as *VEGF*, platelet-derived growth factor, (*PDGF*), transforming growth factor alpha (*TGF- $\alpha$* ), glucose transporter (*GLUT1*), carbonic anhydrase 9 (*CA9*) and fibroblast growth factor (*FGF*). At hypoxia, the expression of these transcriptional targets aims to maintain intracellular homeostasis, however, when constitutively expressed, as in the case of VHL loss, they create a tumor promoting setting favoring cellular proliferation and other aspects of tumorigenesis (97).





**Figure 7. Regulation of the hypoxic machinery**

The hypoxia inducible factors (HIFs) are continuously transcribed and translated and are instead regulated on the protein level. The HIF- $\alpha$ s are rapidly degraded in the presence of oxygen in a process mediated by hydroxylation by factor inhibiting HIF (FIH) and prolyl hydroxylases (PHDs) making the HIFs recognizable by the VHL complex that target the HIF- $\alpha$ s for proteasomal degradation. When the oxygen tension is low, the HIF- $\alpha$ s are instead stabilized translocated to the nucleus where they dimerize with HIF- $\beta$  and together with co-factors initiate transcription of genes involved in the adaptive response to hypoxia. The VHL protein is lost in ccRCC causing the HIFs to be stabilized and initiate a hypoxic cellular response even at normoxia.

There are considerable functional similarities between HIF-1 $\alpha$  and HIF-2 $\alpha$  with overlapping transcriptional targets, but they are not functionally redundant and also display distinct exclusive targets. HIF-1 $\alpha$  generally transcribes genes involved in glucose metabolism, while HIF-2 $\alpha$  is more responsible for growth and angiogenesis. The specific HIF-1 $\alpha$  and HIF-2 $\alpha$  targets can even sometimes have opposing cellular effects. For example, HIF-1 $\alpha$  can promote the expression of pro-apoptotic proteins like BNIP3, and it has even been suggested to be a tumor suppressor (117). HIF-1 $\alpha$  is located at chromosome 14q, which is lost in 40% of ccRCC cases (118). In contrast, several reports have suggested that HIF-2 $\alpha$  is the major contributor for the maintenance of tumorigenic features of ccRCC (117). For example, in patients with VHL disease HIF-1 $\alpha$  expression was correlated with neoplastic lesions, while selective expression of HIF-2 $\alpha$  was associated with advanced lesions (119). Furthermore, repression of HIF-2 $\alpha$  in ccRCC has been shown to halt xenograft growth in mice (120). Nevertheless, it may be so that HIF-1 $\alpha$  is important for the initiation of

ccRCC. Quite surprisingly though, so far only HIF-1 $\alpha$  and not HIF-2 $\alpha$  has been shown to promote renal carcinogenesis in conditional mouse models (121-123).

## Epigenetic regulators

Intra tumor heterogeneity is described in many cancer forms and seems to be exceptionally common in ccRCC, where the only shared clonal events are VHL loss/inactivation and chromosome 3p loss (124). Additional mutations occurring in ccRCC commonly affects epigenetic regulators such as polybromo 1 (*PBRM1*), SET domain containing 2 (*SETD2*) and BRCA1-associated protein-1 (*BAP1*), which are all located on chromosome 3p (125). *PBRM1* encodes BAF180, a subunit of the SWI/SNIF chromatin remodeling complex. This complex regulates DNA accessibility and thereby indirectly also transcription. Inactivating mutations of *PBRM1* is found in about 40% of ccRCC cases (126) and *PBRM1* suppression has been shown to increase cell proliferation, migration and colony formation, and also to create alterations in pathways controlling chromosome stability (126, 127). *PBRM1* mutations appear to be an early event in ccRCC tumorigenesis and are quite often seen in all cancer cells within a tumor. There are reports suggesting that mutations are correlated to higher grade and worse outcome in ccRCC (128, 129), while other studies report that *PBRM1* mutation frequency is similar in all tumor stages and do not correlate to adverse patient outcome (130, 131). Thus the clinical effect of *PBRM1* mutations in ccRCC patient outcome is not entirely clear, however most studies suggest that *PBRM1* loss stimulate cellular proliferation (125, 126).

*SETD2* is mutated in 10-15% of ccRCC cases and often occurs alongside *PBRM1* mutations. *SETD2* encodes a histone H3 lysine 36 trimethylating (H3K36Me3) enzyme (126). This histone mark is commonly associated with active transcription, but can also take part in alternative splicing and transcriptional repression. In contrast to *PBRM1* mutations, *SETD2* mutations are subclonal, meaning that they are present in some subclones of the tumor, but not in others (132). Additionally, different types of *SETD2* mutations can be found in the same tumor, suggesting a high selective pressure for *SETD2* inactivation (125, 132). *SETD2* is thought to contribute to genomic integrity by promoting DNA repair by homologous recombination and by reducing replication induced cellular stress (101). Mutations in *SETD2* have in some patient cohorts been correlated to low cancer specific survival (130).

Similar to *SETD2*, *BAP1* is mutated in 10-15% of ccRCC cases and also presents as a subclonal event. However, in contrast to *SETD2* mutations, *BAP1* and *PBRM1* mutations are virtually mutually exclusive. *BAP1* is a histone deubiquitinase that is involved in HR repair and protects against chromosomal instability (101). Inactivating mutations of *BAP1* have been associated to tumor aggressiveness and worse prognosis in several studies (125, 126), and loss of *BAP1* protein expression has

been shown to independently predict poor patient outcome in low-risk ccRCC (133). *BAP1* loss has also been associated with mTOR pathway activation (134). The mechanism behind the tumor promoting effects of *BAP1* mutations is still no clear, but mutated tumors have been associated with metastatic spread at diagnosis and higher grade, as well as shorter recurrence free survival, suggesting that *BAP1* mutations might be used for stratifying patients into high risk groups and possibly guide treatment options. ccRCC also harbor additional mutation in genes affecting epigenetic regulation including the histone deacytelases *JARID1C/KDM5A*, which is mutated in 3-7% of tumors and *UTX/KDM6A* which is mutated at a low frequency, around 1% (126).

## Clinical manifestation and prognosis

RCC is characterized by vague clinical symptoms such as blood in urine, persistent flank pain, weight loss, fever, fatigue and possibly also by an abdominal mass. As a consequence of these elusive symptoms, many RCC cases are discovered when the disease has already advanced, and as many as 20-25% of all patients have spread disease at diagnosis. If the disease is locally confined the patients have a very good prognosis, with a 5-year survival of 90% (135). However, approximately 1/3 of these patients later relapse, indicative of micrometastasis that were missed at primary diagnosis. Sadly, patients with metastatic disease await a dismal prognosis with a 5-year survival rate of less than 10%, and a median survival of 18 months (124). The most common metastatic sites are localized to the lung, soft tissue, liver, bone, skin and brain (135).

Great efforts have been made to find prognostic molecular markers in RCC, however the *tumor metastasis node* (TMN) staging system (Table 1) along with Fuhrman nuclear grade are still the most used prognostic indicators. They have together with Eastern Cooperative Oncology Group (ECOG) performance status, also known as the WHO score, been combined into the UCLA integrated staging system UISS (99, 136). The ECOG performance status assesses the general well being and functional impairment of cancer patients. According to the UISS, the 5-year survival rates for patients with localized disease are 92% for the low risk groups 67% for the intermediate risk groups and 44% for the high risk group, while the prognosis for patients with spread disease is more discouraging with 3-year survival rate of 37% for the low risk group, 23% for the intermediate group and 12% for the high risk group (99).

**Table 1**

The TMN staging system defines the localization and spread of the tumor, (T) primary tumor, (M) distant metastasis (N) lymph node involvement

TNM STAGING	
<b>Primary Tumor</b>	
T1a	Organ confined <4 cm
T1b	Organ confined 4-7 cm
T2a	Organ confined 7-10 cm
T2b	Organ confined >10 cm
T3a	Invades into perinephric tissue, renal sinus or renal vein
T3b	Invades into vena cava below the diaphragm
T3c	Invades into vena cava above diaphragm or into wall of vena cava at any level
T4	Invades beyond Gerota's fascia or directly into renal gland
<b>Regional lymph nodes</b>	
N0	No lymph node involvement
N1	Metastasis in one regional lymph node
N2	Metastasis in more than one regional lymph node
<b>Distant metastasis</b>	
M0	No distant metastasis
M1	Distant metastasis
<b>Stage grouping</b>	
Stage I	T1N0M0
Stage II	T2N0M0
Stage III	T1-T2N1 or T3N0-1
Stage IV	T4 (any N or M) or N2 (any T or M) or M1

## Diagnosis and Imaging

If a patient is suspected to have a renal tumor, a computer tomography (CT) scan of the abdomen will be performed. If a mass is indeed detected, the scan will be extended to include the thorax, in order to assess possible metastatic spread (135). The CT scan is performed to evaluate the malignancy of the tumor, as well as the tumor size, possible spread into surrounding tissue and/or adrenal gland, and also to assess lymph node involvement. Magnetic resonance imaging (MRI) investigations are less common, but can also be performed if venous tumor thrombus is suspected, or if the patient cannot tolerate the contrast injections that are included in the CT scan. Most cases of RCC can be correctly assessed following imaging analysis (136). However, if the diagnosis after imaging is uncertain, or if the patient is not suitable for surgery, a biopsy of the primary tumor may be performed and sent to the pathologist for assessment. If the patient presents with multiple metastasis, tumor core biopsies are also taken before starting systemic treatment. Biopsies are also

indicated in cases where the patient previously has had another form of cancer; in this case the biopsy will help to clarify whether the mass is in fact a RCC or a metastasis from the previous malignancy (135). Additional laboratory investigations should also be performed, such as measurements of serum creatinine, hemoglobin, leucocyte and platelet counts, and lactate dehydrogenase levels (136). The results of these tests will also give an estimation of the patient's kidney function, something that is crucial to consider when choosing suitable therapy.

## Treatment of RCC

RCCs are characterized by poor response to common anti-cancer therapies like cytostatic drugs and radiation. At present, surgical resection is the standard treatment for localized RCC and still practically the only curative option for ccRCC patients. In the last couple of years, partial nephrectomy has become increasingly common for smaller tumors. This nephron sparing approach has been proven to be beneficial for patients by maintaining more of the kidney function, whereas radical nephrectomy has been shown to increase the risk of renal failure. Additionally, laparoscopic surgery is associated with decreased morbidity compared to open surgery, however there are no differences in recurrence or survival rates between the surgical approaches (91). There are cases when RCC is not operated on, for instances when the patients are considered to be too frail and/or have other co-morbidities that make them unsuitable for surgery. In these cases the patients can stand under active surveillance, where the disease is monitored closely via continual imaging. As an alternative, elderly patients may also undergo cryo- or radiofrequency ablation, which is considered a less invasive method than conventional surgery (89). For patients with a higher risk of recurrence T2-T4, (Table 1) radical nephrectomy is required, where the whole kidney is removed. For T3-4 tumors additional removal of perinephric fat and Gerota's fascia of along with the adrenal gland and surrounding lymph nodes is performed. These efforts result in a complete cure in 40-60% of RCC cases (89).

### *Cytokine therapy in metastatic RCC*

For advanced disease the situation is different. In these cases nephrectomy is far from sufficient to halt the cancer and, as previously mentioned, metastatic RCC (mRCC) is considered to be one of the most chemotherapy resistant malignancies (95), making therapy exceedingly challenging. However, treatment with radiation and chemotherapy can be used in palliative care for example to reduce pain associated with skeletal metastasis. Traditionally, mRCC has been treated with immune system stimulators interleukin-2 (IL-2) and interferon- $\alpha$  (INF- $\alpha$ ) that rely on the patient's own immune response to eradicate the tumor. However, these treatments have show relatively low response rates (20% and 5-15% respectively) and display severe

systemic toxicity and side effects, such as capillary leakage resulting in dangerously low blood pressure (89). Nevertheless, in some cases high dose IL-2 treatment resulted in complete curative outcome (137). This emphasizes the need for prognostic markers that can predict which patients will respond to a certain treatment. This is a pitfall for many cancer drugs, which may be discarded due to low response rates, while being beneficial for a subset of patients. It has been suggested that positive histological staining against the prototypical HIF target CA9, together with a favorable pathologic features may be predictive for IL-2 responders (138). Finally, retrospective studies have indicated that ccRCC responds better to immune stimulatory therapies, than other histological subgroups of RCC (139).

## Targeted therapy in RCC

### *VEGF- and mTOR- targeting therapies*

In the last 10 years targeted therapies have become increasingly common in the treatment against advanced RCC. These treatments mainly target the biological consequence of VHL loss, and essentially consist of angiogenesis inhibitors, affecting VEGF and mammalian target of rapamycin (mTOR) signaling pathways. The rationale behind angiogenesis inhibitors is based on the notion that tumors cannot grow beyond a few millimeters without a blood supply, and that the tumor cells rely on the very same vessels for oxygen and nutrient delivery (140). Additionally, metastatic spread most often occurs through extravasation across the endothelial cells layer in the tumor vasculature. Thus, it was initially thought that by inhibiting the tumor-induced angiogenesis one would essentially starve the tumor, as well as inhibit metastatic spread (140, 141). However, for most types of cancer, antiangiogenic treatment have shown the best response in combination with for example chemotherapy and have been suggested to normalize the leaky tumor vessels, so that supplementary therapy would actually reach the tumor cells (142).

In 2007, clinical trials for mRCC showed that both a direct VEGF inhibitor, the monoclonal antibody *bevacizumab* and the tyrosine kinase inhibitor (TKI) *sunitinib* with multiple targets (VEGFR, PDGFR and c-Kit) displayed a doubling in progression free survival (10.2–11 vs. 5.4–5 months) compared to INF- $\alpha$  alone (95). Antiangiogenic therapy is now the mainstay of treatment for mRCC. Despite these initial encouraging results, resistance against VEGF targeted therapy is seen in 20–30% of patients trying it for the first time, and a majority of patients with primary response acquire resistance within 1 year (124). As an alternative to TKI inhibitors and treatment specifically targeting VEGF, inhibitors against mTOR have been developed. These drugs target both angiogenesis and tumor proliferation (135). mTOR is a key regulator of cellular homeostasis and is involved in nutrient sensing, energy status and cellular stress (143). The PI3K/Akt/mTOR pathway together

controls multiple cellular features such as proliferation, metabolism and differentiation (144). Additionally, a majority of ccRCC cases display increased mTOR activity, especially apparent in tumors of high grade and with poor prognostic features (143). For mRCC patients with particularly poor prognosis, treatment with the mTOR inhibitor *temsirolimus* was shown to be beneficial over INF- $\alpha$  treatment, or a combinatory treatment of both drugs. Temsirolimus treatment also increased the over all survival for these patients (10.9 vs. 7.3 months). In this study there were no difference in response between ccRCC and non-clear cell RCC (143).

For patients with mRCC that do not respond to treatment or develop resistance, there are alternative so-called second-line agents. *Sorafenib* is a multi kinase inhibitor of Raf1 serine/threonine kinase, b-Raf, VEGFR-2, PDGFR and c-Kit, which in clinical trials have been shown to prolong progression free survival with three months compared to placebo. However, there are no reliable data concerning over all survival, since the study allowed “crossover” from the placebo group arm to the treatment arm and an interim analysis shown that treatment with sorafenib was beneficial (135). As an alternative second-line treatment after failure with VEGF inhibitors, the mTOR inhibitor *everolimus* may be applied. This treatment showed a progression free survival of (4.9 vs. 1.9 months) in a patient cohort of metastasized ccRCC (143).

#### *c-Met inhibitors*

There are limited specific treatment options for metastatic pRCC, possibly owing to a lack of incentive for development of specific pRCC targeted drugs due the fact that pRCC only constitute 5-15% of RCC cases. However, an inhibitor of MET and VEGFR, *foretinib*, has recently been developed and a phase II clinical trial including patients with pRCC, showed an increased progression free survival compared to previous trials using sorafenib or sunitinib (9.3 months vs. 1.6 up to 6 months) (145).

### **The role of surgery in mRCC**

Surgery may not be fruitless for mRCC patients. Radical nephrectomy in patients with metastatic disease, so called cytoreductive nephrectomy (CN) is not a standard procedure, but it has nevertheless been shown that INF- $\alpha$  treatment together with CN significantly prolonged the median overall survival time compared to INF- $\alpha$  treatment alone (13.6 vs. 7.8 months) (146). In line with this, there are now ongoing phase III clinical trials investigating the potential beneficial use of CN together with the standard care of VEGF inhibitors (95). The biology behind this potential beneficial effect is not clear, but it has been suggested that removing the primary tumor might reduce immunosuppressive cytokines, as well as growth promoting signals, creating a systemic effect also on metastasis (89). The possible benefit of surgical removal of metastasis, metastasectomy, is debated since no prospective

randomized studies have been performed and metastasectomy is therefore quite uncommon in routine practice (135). However, it may be employed in cases with solitary lung metastasis or if there has been a long duration between primary tumor and relapse, or between occurring metastasis (136).

## Immune checkpoint inhibitors

RCC is considered to be an immunogenic cancer, based on the reports of spontaneous regressions of RCC in the 80-ties and 90-ties (147, 148), as well as the partial beneficial responses to cytokine therapies mentioned above. Efforts have therefore been made in order to develop novel immunomodulators for treatment of RCC. Programmed death receptor-1 (PD-1) is an immune checkpoint component expressed on T cells, B cells and NK cells, whereas its ligand PD-L1 is expressed on antigen presenting cells as well as tumor cells. When the ligand binds its receptor the cytokine secretion from the immune cell is inhibited. This is an important function in the normal immune tolerance, which prevents us from having an allergic reaction to all exogenous peptides. However, the PD-1/PD-L1 interaction is also suggested to be of major importance for the capacity of tumors to evade the immune response (149). A retrospective study showed that patients with ccRCC tumors staining positive for PD-L1, had a worse prognosis compared to patients with tumors not expressing PD-L1 (41.9% vs. 82.9% in cancer specific survival) (150). Additionally, PD-L1 expression has been correlated to worse outcome also in non-clear cell RCC, suggesting that these patients may also be aided by PD1/PD-L1 targeted therapy (151). Recent clinical trials, for mRCC patients resistant to first-line therapies, using targeted therapy against PD-1/PD-L1 pathways have displayed an overall survival benefit compared to treatment with mTOR inhibitors. Treatment with the PD-1 blocking antibody *nivolumab* prolonged the overall survival with 5 months compared to treatment with everolimus (25 months vs. 19.6 months) (152). Combination treatments with PD1/PD-L1 targeted therapy and other commonly used RCC treatments, such as angiogenesis inhibitors or cytokines, also improved response rates compared to single treatment, while however also displaying higher toxicity (149). These results indicate that treatment with nivolumimab will likely form a new standard of care for patients with mRCC. An additional immune checkpoint receptor, which is being investigated as a possible target for mRCC therapy, is CTLA-4. This receptor which is expressed on T-cells and functions in a similar way to PD-1, have been successfully targeted in metastatic melanoma using the blocking antibody *ipilimumab* (95). Future challenges for immune checkpoint based therapies will be to apprehend which patients that will respond to the therapy. Interestingly, there are contradictory data concerning the use of PD-L1 expression as a biomarker for response, since also negative tumors have been shown to respond to PD1/PD-L1



targeted therapy (149), which may suggest that this therapy has an additional general immune stimulatory effect, regardless of PD-L1 tumor status.

Finally, it may be so that most of the above-mentioned drugs have limited effect as monotherapies, but appear to their full advantage as combination treatments. Avoiding severe adverse side effects in these cases however represents a major challenge. In a manner similar to Darwinian evolution, cancers have a high ability to adapt to their environment, possibly due to the high level of intratumor heterogeneity. Thus, hitting several functionally different targets at once may be necessary to eradicate the tumor. Still, there is a great need for predictive biomarkers in order to correctly guide the most suitable choice of treatment for each patient. The development of resistance remains a huge problem with current treatments and although recent therapies show prolonged survival, virtually no mRCC treatments have been curative. Thus there is a desperate need to develop novel effective targeted therapies for mRCC that may be used complementary to existing treatments. Additionally, there is a need for novel high-resolution diagnostic methods that allows for early detection of the primary tumor, before metastatic traits have been developed, as well as for the identification of early metastasis to ensure that the patients can be treated for mRCC as early as possible, i.e. before the tumor has developed a high level of heterogeneity.

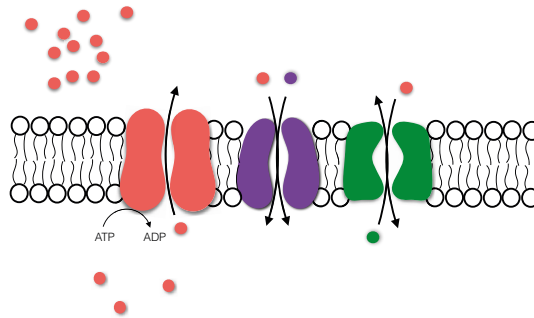
# Transporter proteins

Transporter proteins control vital parts of cellular homeostasis by regulating the import and export of essential substances like sugars, amino acids, solutes and drugs across the plasma membrane. It is estimated that 4% of our DNA codes for transporter proteins and virtually all cells depend on their function (153). However, transporters are deeply understudied, both in proportion to the sheer number of genes and to their importance in normal cellular physiology and disease (154). There are two major families of transporters, the solute carriers (SLC) and the ABC-transporters.

The ABC-transporters represent the second largest transporter family and consist of 48 members divided into 7 subfamilies. The ABC-transporters are intricate pumps that are driven by energy catalyzed from the hydrolysis of ATP in order to efflux molecules across cell membranes (155). The expression of ABC-transporters has been associated with stem and progenitor cells, which might be related to their capacity to eliminate harmful substances in order to protect their DNA and thereby their future progeny (156). Expression of ABC-transporters and have also been coupled to drug resistance in many human diseases, including chemotherapy resistance in cancer (28, 157).

The SLC family is composed of over 400 different members that are organized into 52 subfamilies and is thereby the largest group of cellular transporters. These transporters are mainly involved in molecular uptake (153), and are primarily driven by transmembrane ion gradients (158). The SLC families vary in substrate specificity; some have a very narrow range of substrates while others like the SLCO-family (OATPs), can handle a broad range of chemically diverse substrates (159). The transport of substrates via SLC can be driven in several ways. Passive transporters or facilitated transporters allow substances to diffuse down their concentration gradient. Active transporters on the other hand require energy to transfer substances against their concentration gradient. An example of a primary active transporter is the  $\text{Na}^+/\text{K}^+$  ATPase, which is widely expressed in the kidney.  $\text{Na}^+/\text{K}^+$  ATPase uses the hydrolysis of ATP to pump  $\text{K}^+$  into the cell and  $\text{Na}^+$  out of the cell, against their electrochemical gradients. Secondary active transporter uses the concentration gradients created by the primary active transporters to move a specific substrate, against its own gradient, together with  $\text{Na}^+$  or  $\text{K}^+$  along their electrochemical gradient. Secondary active transporters can be symporters, meaning that the specific substrate and the aid-solute,

for example  $\text{Na}^+$ , moves in the same direction, or antiporters meaning that the substrate and aid-solute move in opposite directions. (Figure 8)



**Figure 8. Secondary active transporters**

Schematic figure of secondary active transporters that are driven by the flow of aid solutes down their electrochemical gradient, such as for example  $\text{Na}^+$  (pink). This gradient is achieved by the active transport of, for example,  $\text{Na}^+$  out of the cell in a process requiring energy in the form of ATP hydrolysis (pink transporter). The secondary active transporter can be symporters (purple transporter or antiporters (green transporter).

Neurotransmitter transporters (NTTs) belong to the SLC6-family, which is one of the largest subgroups of the SLC-family consisting of 20 members. The SLC6-family handles the transport of amino acids or amino acid derivatives into the cells (160). The NTTs transport neurotransmitter substances such as serotonin, dopamine, norepinephrine, GABA and glycine into the cells. These transporters are primarily located to the central nervous system (CNS) in the brain, but can also be found in non-neuronal cells (161). NTTs use the electrochemical gradient of extracellular  $\text{Na}^+$  to transfer their substrates via a symport of  $\text{Na}^+$  into the intracellular compartment.

## The dopamine transporter SLC6A3

Dopamine was first described as a neurotransmitter in the late 1950's by the Nobel Prize laureate Arvid Carlsson. The function of dopamine in the brain mainly involves control of voluntary movement via the basal ganglia. Degeneration of the dopaminergic neurons in the nigrostriatal pathway causes Parkinson's disease (PD) where patients have a decreased ability to execute smooth controlled movements. The contrary is seen in disorders associated with elevated dopamine levels such as Huntington's disease and Tourette's syndrome where patients display jerking involuntary movements (162, 163). Other than motor deficiencies, PD patients also present with other symptoms such as depression, hallucinations, cardiovascular symptoms and even cognitive impairment and dementia in later stages (164). These

symptoms may be due to other changes occurring in PD, affecting noradrenergic, serotonergic and cholinergic pathways (165) but may also reflect dopamine's non-motor functions. For example, in nucleus accumbens and striatum dopamine plays a part in the reward system, and is released upon pleasure stimuli like food or sex (166). Dopamine also has a role in the frontal lobe where it is involved in managing the information flow from other parts of the brain. Disorders affecting this system cause cognition deficiency and impaired attention, memory and problem solving (167, 168).

Dopamine release into the synaptic cleft results in binding to G-protein coupled dopamine receptors that convey excitatory or inhibitory signals through the post synaptic neuron, depending on which receptor that is activated. There are five dopamine receptors D<sub>1</sub>-D<sub>5</sub> divided into either D<sub>1</sub>-like family (D<sub>1</sub> and D<sub>5</sub>) or D<sub>2</sub>-like family (D<sub>2</sub>, D<sub>3</sub> and D<sub>4</sub>) (169). The D<sub>1</sub>-like family is coupled to the stimulatory G-protein G<sub>s</sub> that activates the enzyme adenylyl cyclase and thereby increases the levels of the common second messenger cyclic adenosine monophosphate (cAMP), while the D<sub>2</sub>-like family is coupled to inhibitory G<sub>i</sub>, which inhibits adenylyl cyclase and thus the formation of cAMP (170). The potency of the dopamine signaling also requires careful control of its levels at the synapse; this control is primarily mediated by the clearance of dopamine from the synaptic cleft via the dopamine reuptake transporter SLC6A3. The *SLC6A3* or *DAT1* gene is located on chromosome 5p15 and encodes a transporter protein with 12 transmembrane domains that is dependent on a symport of sodium and chloride to execute its function (160). At the synapse, the extracellular concentration of Na<sup>+</sup> and Cl<sup>-</sup> by far exceeds their intracellular concentration, the ions thus move through SLC6A3 down their electrochemical gradient, bringing dopamine along.

### Pharmacological interaction with SLC6A3

SLC6A3 have been extensively studied from a pharmacological perspective, since drugs like cocaine and amphetamine primarily exert their function through SLC6A3 inhibition, thereby prolonging the dopaminergic signaling with molecular and behavioral effects as a result. SLC6A3s normal function of dopamine influx can in some instances be reversed, at least in some *in vitro* experimental settings. For example, if the electrochemical gradients of ions are reversed, with high intracellular concentration of Na<sup>+</sup> and Cl<sup>-</sup>, the transporter can efflux dopamine out of the cells (171, 172). SLC6A3 can also be modified by phosphorylation via phosphokinase C (PKC) and phosphokinase A (PKA). Phosphorylated SLC6A3 causes decreased dopamine uptake due to either internalization of the transporter via PKC phosphorylation or an actual efflux of dopamine through the transporter via PKA phosphorylation (173). The PKA and PKC activities are mediated via the trace amine

associated receptor 1 (TAAR1) that can be activated by methamphetamine (174). Amphetamine is taken up via SLC6A3, inhibiting dopamine reuptake in several ways: in a competitive manner, through internalization (175), and also through activation of TAAR1 and subsequent induction of SLC6A3 dopamine efflux (176). Additionally, the action of amphetamine can cause calcium-induced depolarization, which activates Ca<sup>2+</sup>/calmodulin-dependent protein kinase II (CAMII) that also can cause dopamine efflux by SLC6A3 phosphorylation (177, 178). Cocaine directly binds to SLC6A3 and inhibits the dopamine reuptake (179). This inhibition has been suggested to be caused by a stabilization of the transporter in an outward facing conformation (180, 181), however the exact binding sites of cocaine have not been exactly established (181).

### **SLC6A3 in disease**

Mutations in SLC6A3 have been associated with infantile parkinsonism-dystonia, a rare autosomal recessive disorder that causes PD like symptoms and muscle contractions during infancy (160). The mutations cause either loss of function resulting in reduced levels of operative SLC6A3 (182), or severely decrease the binding affinity of dopamine to SLC6A3 (183). The symptoms may be explained by impaired replenishment of dopamine into synaptic vesicles. SLC6A3 harbors a 40 base pairs variable number tandem repeat VNTR section in its untranslated 3' end. The VNTR has been suggested to affect SLC6A3 availability on the cell surface and variants of this VNTR have been linked to attention deficit hyperactive disorder (ADHD) susceptibility (184, 185). In humans, the most common forms are 10 or 9 repeats where the 9 repeat form has been associated with higher SLC6A3 activity (186). Other disorders that have been linked to malfunctioning SLC6A3 include ADHD, bipolar disorder, Tourette's syndrome and alcoholism (160, 187-189). Additionally, SLC6A3 knock-out mice display neurological deficiencies including hyperactivity resembling the symptoms seen in patients with ADHD (190).

#### *Imaging using SLC6A3*

As previously mentioned, PD is a progressive neurodegenerative disease where the dopaminergic cells die causing a variety of symptoms (191). The diagnosis of this disease can be aided using imaging modalities based on the presence of SLC6A3. Radioligands, such as <sup>123</sup>I-β-CIT or <sup>123</sup>I-Ioflupane also known as DaTSCAN are cocaine analogues that selectively bind to SLC6A3, which subsequently can be imaged using single photon emission computed tomography (SPECT) (192). This imaging allows for a quantitative measure of the dopaminergic neurons in striatum. In patients with parkinsonian symptoms these imaging techniques make it possible to differentiate PD from other neurodegenerative disorders such as essential tremor

(193) and additionally distinguish between Lewy body dementia and Alzheimer's disease (194).

## Dopamine system in the kidneys

As previously mentioned, dopamine exerts several important functions in the brain in its role as a neurotransmitter. This vital role of dopamine is emphasized by the fact that disturbances in its signaling causes several severe disorders. However, in the last 30 years it has become apparent that dopamine signaling also has various important physiological functions in non-neuronal tissues (195). In the kidney, dopamine has a natriuretic effect, resulting in decreased blood pressure and more sodium excreted with the urine. Disturbances in the renal dopamine system have therefore been associated with the development of hypertension (196). The renal tubular cells are equipped with amino acid transporters located on the apical and basal side that take up circulating or filtered L-dopa (197). Inside the cells, L-dopa is converted to dopamine via the enzyme aromatic L-amino acid decarboxylase (AADC). The kidneys express one of the highest levels of AADC in the body (198), and the activity of the enzyme is highly affected by sodium intake where it is upregulated by high levels of sodium and conversely downregulated by low sodium intake (199). The dopamine generated from the AADC conversion is subsequently secreted into the tubular lumen where it can bind dopamine receptors in an autocrine or paracrine manner. The D<sub>1</sub>-D<sub>5</sub> receptors are all expressed in the kidney, although some of them at very low levels (200). Binding of D<sub>1</sub> receptors in the proximal tubules results in inhibition of Na<sup>+</sup>/K<sup>+</sup>-ATPase as well as the Na<sup>+</sup>/H<sup>+</sup>-exchanger, causing decreased sodium reabsorption. Animal studies have estimated that as much as 50% of sodium excretion is dependent on renal dopamine signaling (200). Apart from its role in blood pressure regulation dopamine is also suggested to be important in redox balance in the kidney. The dopamine receptors D<sub>1</sub>, D<sub>2</sub> and D<sub>5</sub> can have antioxidant effects, for example by inhibiting NADPH oxidase and stimulation of superoxide dismutase (SOD) (201, 202). Additionally, dopamine has been shown to have a dampening role on inflammation by inhibiting immune cells from releasing interferon  $\gamma$  (INF $\gamma$ ), IL-12 and IL-4 (203, 204). Also, mice with malfunctioning renal dopamine system displayed increased infiltration of immune cells and increased oxidative stress (203). The transfer of dopamine through the apical and basal membranes of proximal tubule cells, both inwards and outwards, have been shown to be unaffected by cocaine or the specific SLC6A3 inhibitor GBR-12909 (198). Additionally, there are hardly any *SLC6A3* mRNA transcripts detected in the kidney (205), suggesting that SLC6A3 take very little, if any, part in the dopamine system in the kidney.



# The present investigation

## Overall Aim

The overall aim of this thesis was to shed light on kidney regeneration and the connection between different cell types of the kidney and renal malignancies.

## Paper I-III

### Aims and Results

*Paper I: Isolation and characterization of progenitor-like cells from human renal proximal tubules*

#### *Specific Aims*

The aim of this paper was to define and characterize a potential progenitor cell population in the kidney using an assay based on the stem cell associated aldehyde dehydrogenase (ALDH) activity.

#### *Key Findings*

In this paper we used the FACS-based ALDH assay to isolate human renal cortical cells with stem cell-like features. Transcriptional analysis of the two cell fractions ALDH<sup>high</sup> and ALDH<sup>low</sup> showed that the ALDH<sup>high</sup> cells were associated with a stem cell signature and cytoskeletal-, adhesion-, leading edge- profiles as well as with negative regulation of apoptosis. The ALDH<sup>low</sup> cells on the other hand, expressed genes involved in normal renal tubular functions, such as transmembrane transportation, solute carrier function and metabolic processes. We could also show that the ALDH<sup>high</sup> cells displayed stem cell like features like clonogenic capacity and anchorage independent growth while the ALDH<sup>low</sup> cells did not. Immunohistochemical examination of human kidney cortex showed that the genes upregulated in the ALDH<sup>high</sup> cells delineated a population of cells positive for vimentin, CD133, CD24, KRT7, KRT19 and BCL2 that were scattered within the



PTs. These cells were also negative for several characteristic PT markers such as XPNPEP and SUSD2. They were also immunophenotypically very similar to the parietal epithelial cells (PEC) of the Bowman's capsule, which previously has been described as a progenitor cell population. Additionally, regenerating tubules in kidneys with ATN, showed long stretches of cells positive for the markers of the ALDH<sup>high</sup> cells. Furthermore, we demonstrated that the transcriptional profile of the ALDH<sup>high</sup> cells showed a substantial overlap with that of pRCC. Collectively the findings of Paper I, suggest that the scattered progenitor-like cells may be more resistant to apoptotic stimuli associated with renal insult and that they may also be involved in renal tubular regeneration. Finally, we propose that pRCC might be derived from oncogenic transformation these cells.

*Paper II: Evidence for a morphologically distinct and functionally robust cell type in the proximal tubules of human kidney*

#### *Specific Aims*

The aim was to further characterize the progenitor-like cell discovered in Paper I both at an ultrastructural and functional level.

#### *Key Findings*

In this paper we explored the ultrastructural features of the cells previously described in Paper I, using conventional electron microscopy (EM) and vimentin-immuno-EM. Structurally, we discovered that the cells were flask shaped, had a dense nucleus and were smaller than surrounding PT cells. The cells were also endowed with an extensive filamentous mat and had drastically fewer mitochondria than surrounding bulk cells. The mitochondrial paucity was also confirmed using immunofluorescence co-stainings with the characteristic markers described in Paper I and the mitochondrial markers MTCO2 or VDAC1. To assess the functionality of the cells we also developed an *ex vivo* model of human ATN, where kidney pieces were subjected to ischemia in conjunction with nephrectomy and subsequently reoxygenated in cell culture conditions. Immunofluorescent analysis showed that the progenitor-like cells largely remained attached to the membrane in contrast to the bulk PT cells where 80% were lost. Additionally, we investigated the presence of these cells in renal injuries ranging from acute to chronic and demonstrated that they were present in regenerating tubules during all phases. We also showed that the cells could be detected in renal wedge biopsies from young healthy kidney donors. Collectively, these results show that the progenitor-like cells are morphologically distinct and indicate that these cells are more resilient to renal insult than bulk PT cells.

### *Paper III: Species diversity regarding the presence of proximal tubular progenitor cells of the kidney*

#### *Specific Aims*

The aim was to investigate the potential presence of renal progenitor cells as defined in human kidney, in different species.

#### *Key Findings*

In this paper we investigated the presence of cells similar to human proximal tubular progenitors (PTPC) in kidneys from chimpanzee, pig, rat and mouse, using immunohistochemical markers that characterize the human PTPCs such as Vimentin, CD133, KRT7, KRT19, CLDN1 and low mitochondrial content (MTCO2<sup>low</sup>). PTPCs could be detected in kidneys from chimpanzee and pig, however we were unable to find these cells in mouse kidneys. In rat we could find occasional, very rare cells with vimentin positivity and lower mitochondrial content. In an attempt to provoke the appearance of these cells in murine kidneys, we subjected mice to kidney injury in the form of unilateral urinary obstruction (UUO). Despite clear kidney damage we could not detect any vimentin positive cells within mouse kidney tubules. These findings have implications for the interpretation of the apparently contradicting results that prevail in the field of kidney regeneration.

## **Discussion**

The kidneys are highly energy demanding organs and are consequently very sensitive to disturbances in renal blood flow causing hypoxia and decreased nutrient delivery. Ischemic or toxic insults can lead to acute tubular necrosis where the tubular cells detach and die. Normally, the cells of the kidney rarely divide, but following injury the renal tubules possess a remarkable regenerative ability. The cells or factors that control this regeneration process are still largely unknown.

#### *Do renal progenitors exist?*

In the field of regeneration there is a vivid discussion of whether renal progenitor cells exist, and if so, whether they have a role in the regeneration process. In this thesis we describe a previously overlooked cell population inconspicuously scattered within the epithelial cells of the proximal sections of the nephron. These cells displayed progenitor cell characteristics, such as clonogenicity and anchorage independent growth and were, as demonstrated by a novel explant model of human ATN, more resilient to renal insult than surrounding bulk PT cells. The cells also express a characteristic set of markers, among others, the stem cell associated CD24 and CD133, the antiapoptotic BCL-2 protein, and markers associated with cellular

structural integrity and attachment, which may contribute with mechanical resilience. These markers are virtually all shared with the population of PEC progenitors of Bowman's capsule. Additionally, we could find long stretches of cells positive for these characteristic markers in regenerating tubules of clinical samples of acute as well as more prolonged renal injury. The most striking morphological difference between the scattered cells and the bulk PT cells were the mitochondrial content. PT cells are packed with mitochondria, reflecting their normal function of endless active reabsorption and hence pronounced energy demand. In contrast, the scattered cells display remarkable mitochondrial paucity. Low mitochondrial content have been associated with stem cells and may contribute to DNA protection from reactive oxygen species (ROS) linked to mitochondrial respiration (206). Based on these findings we suggest that these proximal tubular rare cells, also called PTPCs, (which I will refer them to from now on) are better equipped to survive renal insults and may become activated to proliferate and repopulate the tubules in the wake of renal injury, thereby providing a cellular source for regeneration. On the contrary, the results from most lineage tracing studies performed in mice support the classical theory, that regeneration is not performed by a specialized progenitor cell population, but through dedifferentiation of randomly surviving cells. However, as we show in Paper III, human kidneys and mice kidneys are not mirror images of each other, at least with regard to the presence of PTPCs. The lack of PTPCs in mice does not exclude that progenitors exist, but may suggest that they have other characteristics and express other markers in mice. Recently, two papers applying lineage tracing techniques in renal epithelia have supported the notion of progenitor cells also in mice. Kang et al. demonstrated that Sox9 positive cells proliferated following damage and contributed to regeneration to all nephron segments except for Bowman's capsule, glomerular cells and collecting duct. Additionally, conditionally knocking out *sox9* resulted in fibrosis (207). Rinkevich et al. revealed that Wnt responsive precursors gave rise to segment specific renal regeneration (86). Interestingly, we noted that *SOX9* is upregulated in the ALDH<sup>high</sup> fraction described in Paper I.

There are also a few alternative notions regarding the source for cellular regeneration. The group of Marcus Moeller has described a specific population of cells appearing *de novo* after damage. These cells are morphologically distinct and similar to the cells described by our group, most strikingly so with the reported lack of mitochondria and vimentin expression (75). This group reports a massive increase of these, so called scattered tubular cells (STC), upon AKI in mice and show that the STCs contribute to tubular regeneration. The STCs express KIM-1 and vimentin, which have been associated with kidney injury, this group therefore interpret the presence of STCs in normal human kidneys as signs of micro injury that possibly occur with age. However, we have found PTPCs in healthy human kidneys from ages 2 (which was the lowest age that we could get hold of) and up. Additionally, the PTPCs are immunophenotypically very similar to the PEC of Bowman's capsule, which are

widely accepted as a progenitor cell population and also express vimentin. It seems unlikely that these cells reside in a state of permanent injury. We thus propose that the PTPCs /STCs are indeed a resident progenitor population rather than an induced phenotype that could be adopted by any mature tubular cell, as suggested by the Moeller group (208). An alternative perhaps complementary hypothesis to the progenitor cell theory is presented by Sallustio et al. They propose that resident tubular progenitor cells aid in the regeneration process by protective paracrine signaling. They show that resident tubular progenitor cells secrete of protective factors such as MCP-1, C3 and IL-6 in response to TLR-2 activation signaled from the injured bulk cells (209), and that progenitor cell-secreted FGF2 and inhibin-A, as well as micro vesicle shuttled decorin and cyclin D1, could protect bulk epithelial cells from cisplatin induced damage via inhibition of apoptosis (74). Together these factors thus act on remaining bulk cells and protect them against injury and also induce proliferation, in this way minimizing the tissue damage. Thus resident renal progenitor cells may have several functions in AKI: to protect bulk PT cells from apoptosis and possibly stimulate their proliferation, as well as to regenerate tubules by proliferating to cover denuded basal membranes and subsequently differentiate to restore tubular function.

As indicated here, there is still confusion regarding the cellular origin of tubular regeneration. It may be so that the two main theories of renal regeneration, progenitors vs. dedifferentiation are not mutually exclusive in mice, but work in concert to achieve tubular integrity after damage. In Paper III we did not see an induction of PTPCs even after damage. Our findings may shed some light regarding the discrepant results and conclusions concerning tubular regeneration in the field and may also suggest that tubular regeneration in mice and men are not orchestrated in exactly the same way. However, lineage tracing remains the current dogma for investigating cellular sources for biological processed *in vivo*, and this is for natural reasons not feasible in humans. Hence, the exact process of renal regeneration in humans is currently difficult to fully elucidate. Importantly though, one has to be careful when translating results attained in mice directly onto the human setting.

#### *Anatomical location of PTPCs, a stem cell niche?*

Adult stem cells are most often described to be located in a niche, which supports the optimal environment for the maintenance of these cells (9). In the process of characterizing the PTPCs we noted that they were often located in plicae positions, where the proximal tubules make hairpin turn. It is tempting to speculate that this localization may serve as a niche for the PTPCs. The plicae position could entail specific environmental features that might somehow support the PTPCs. One feature that may be specific for this anatomical location is the possible currents created from the flow of the filtrate, which may affect the PTPCs from the luminal side. Another specific feature may be the distance to the capillaries at this position. Low oxygen

tension has been suggested to be an important factor in maintaining quiescence in other adult stem cells (14). However the possible importance of the anatomical location of the PTPCs requires further investigation.

#### *Connection to cancer and clinical impact?*

In Paper I we found that the transcriptional profile of PTPC showed significant resemblance to that of pRCC. Additionally, two of the most characteristic PTPC markers KRT7 and KRT19, are used in the clinic to diagnose pRCC and are also expressed in renal adenomas, which are hypothesized to be a precursor lesion to pRCC. We thus propose that pRCC may be caused by a stepwise oncogenic transformation of PTPCs. This also relates to the hypothesis concerning adult stem cells as a plausible cell of origin in some cancers, as discussed in the stem and progenitor cell chapter of this thesis. Interestingly, the incidence of pRCC is higher in kidney transplant patients, both in native kidneys and in the allograft (210). Higher incidence of malignancies after organ transplants are often ascribed to the immunosuppressive treatment required upon transplantation (211) However, since the pRCC incidence is especially elevated, it is tempting to envision that this could be associated with PTPCs. In a retrospective study by Tillou et al. it was found that pRCC represented over 50% of RCC cases in comparison to around 10-15% in non-transplanted cases. Additionally, the proportion of pRCC in native kidneys of transplant patients has also been reported to be elevated, around 30% (210, 212). In the case of pRCC in allografts, could the transient hypoxic challenge in conjunction with the transplantation procedure cause erroneous activation of PTPCs, which in the long run could cause malignant transformation? Additionally, the incidence of pRCC is also elevated in patients with chronic kidney injury (96)), where PTPCs are also activated in regenerating tubules, as showed in Paper II. This may suggest a link between PTPCs, CKD and pRCC development.

A deeper knowledge of the process of regeneration and also the features of the putative renal progenitor cells may aid in the understanding of how AKI is best supported and possibly also how to exogenously induce the activation of the patients own progenitor cells. However, too much proliferation may lead to fibrosis and also have implications in the development of CKD, which is more prevalent in patients that previously have experienced AKI. So far, the molecular mechanisms of PTPC activation are unclear. Future studies are required to investigate which factors that activate the PTPCs and also what makes them differentiate. Knowledge of how to control this balance could be of clinical importance for both AKI and CKD, and possibly also hold clues to pRCC development.

# Paper IV

## Aims and Results

*Paper IV: Overexpression of functional SLC6A3 in clear cell renal cell carcinoma*

### *Specific Aims*

The aim was to investigate transporter proteins differentially expressed in renal cell carcinoma compared to normal kidney.

### *Key Findings*

In this paper we show that the dopamine transporter *SLC6A3* is highly expressed in ccRCC, while hardly being detected in normal kidney or other tumor types and thus bear the hallmarks of a highly specific biomarker ccRCC. We could show that ccRCC cells actively transported [<sup>3</sup>H]-dopamine into the cells, an uptake that could be repressed using a specific SLC6A3-inhibitor, GBR-12909. Additionally, by knocking down HIFs using siRNA, we could see that the *SLC6A3* expression was maintained by HIF-2 $\alpha$  expression. Furthermore, we could also show that *SLC6A3* expression could be induced in normal kidney cells grown in hypoxia. This effect was not seen in normal epithelial cells from breast (MCF10A), nor in normal endothelial cells (HUVEC). Altogether, this study shows that the high expression of *SLC6A3* in ccRCC is accompanied by an enhanced uptake of dopamine and that the elevated expression level of the dopamine transporter can be linked to the main hallmark of ccRCC, i.e. a pseudohypoxic phenotype as a consequence of VHL-loss.

## Discussion

Transporter proteins are understudied in the field of cancer, despite their important role in homeostasis and disease. Moreover, transporter proteins also hold great promise as potential targets for drug delivery. In this paper we performed an unbiased screen of transporter genes in several tumor types, compared to their corresponding normal tissues, including RCC and normal kidney. We found that RCC display a unique expression pattern of transporter genes compared to other tumor types, which most likely reflect their origin from the transporter-rich kidney. In general the tumor types reflected their tissue of origin and clustered together in the unsupervised hierarchical clustering analyses. However, there were examples of transporter genes that were present in the normal tissue but lost in the tumor samples. This may be due to dedifferentiation associated with tumorigenesis, but could also reflect the cell of origin. This is particularly evident when comparing normal kidney and the different

RCC subtypes. The kidneys are complex organs with advanced 3D structure and hence each normal biopsy contains many different cell types. Samples from the medulla and samples from the cortex will display different transcriptional profiles, reflecting their cellular composition. For example, the major part of the cortical samples will be composed of PTs. In the analysis of transporter gene expression, chRCC clustered away from the other RCC subtypes, and also away from the normal kidney samples. This may be explained by the notion that pRCC and ccRCC stem from the PT, while the cell of origin for chRCC is suggested to be located in the more distal part of the nephron, and that distal tubules represent a minor proportion of the cells in the normal samples. This highlights the importance of understanding the cellularity of normal samples, when comparing to tumors that originate from a distinct subset of cells within a given normal tissue.

When comparing RCC and normal kidney the most distinct differentially expressed transporter gene was the dopamine transporter *SLC6A3*. *SLC6A3* was highly expressed in ccRCC, while other tumor types, including pRCC and chRCC, barely express noticeable levels of this transporter. In normal tissue, *SLC6A3* expression is only seen in dopaminergic neurons in the brain, where it normally exerts its function of transporting released dopamine back into the neuron for later use. *SLC6A3* plays a vital role for the dopamine signaling in the brain, and disturbances of its function cause marked neurological and behavioral effects. However, *SLC6A3* has not been reported to be expressed at appreciable levels elsewhere. The exact reason for the upregulation of *SLC6A3* in ccRCC is not clear, but we could demonstrate that the expression is affected by HIF, in particular HIF-2 $\alpha$ . We also show that *SLC6A3* is specifically induced by hypoxia in primary normal renal cells. These primary cultures are predominantly composed of PT cells, which are also suggested to be the cells of origin for ccRCC. As previously mentioned ccRCC has a very strong hypoxic drive due to the functional loss of VHL, and whereas HIF-1 $\alpha$  has been reported to be lost in 40% of ccRCC cases, HIF-2 $\alpha$  is continuously expressed (118). Collectively, this may suggest that the *SLC6A3* locus is specifically accessible in renal proximal tubule cells and that HIF-2 $\alpha$  is responsible for its transcription, causing high expression in ccRCC exclusively.

Importantly, we also demonstrate that ccRCC cells can actively take up dopamine and that this is mediated by *SLC6A3*, providing evidence for functional *SLC6A3* in ccRCC. The effect of the active dopamine uptake on the tumor cells is however not clear. In preliminary viability assays inhibition of *SLC6A3* has not shown any significant effects, (unpublished data). However, long term treatment effects of *SLC6A3* inhibition has not been studied. Additionally, the consequence of *SLC6A3* expression may be different *in vivo* and *in vitro*. The environment of cell culture conditions is indisputably altered from the microenvironment for ccRCC tumor cells *in vivo*. As mentioned in the transporter chapter of this thesis, dopamine has been reported to have immune suppressive properties. One of the prerequisites for tumor

development is to evade the immune system, perhaps this could be an advantageous trait of the *SLC6A3* expression? Dopamine is also involved in antioxidant protection, which may be beneficial in a tumor environment. Antioxidants have been implicated in promotion of metastasis in melanoma (213, 214), lung cancer (215) and breast cancer (216). Additionally, it may also protect the tumor cells from free radicals similar to the effect of antioxidants in normal cells. Antioxidant supplement in combination with radiation or chemotherapy has shown to have adverse effects on tumor progression in several cancer forms (217). Another possible usage for the dopamine uptake might be for metabolic purposes, perhaps for energy or building blocks. Dopamine can for example be catabolized to homovanillic acid (HVA) by the enzymes monoamine oxidase (MAO) and catechol-O-methyl transferase (COMT) (218). Dopamine is also the precursor for norepinephrine and epinephrine. Epinephrine has been shown to protect breast- and prostate cancer cells from apoptosis via indirect inhibition of the pro-apoptotic protein BAD (219). However, the metabolic pathways that could affect dopamine in ccRCC, and the possible consequence of the metabolites, require further investigation.

Interestingly, primary ccRCC cells, but only two of the tested ccRCC cell lines in Paper IV display elevated *SLC6A3* expression. The KMRC3 and SNU-349 cell lines show high levels of *SLC6A3* mRNA and are the only cell lines (apart from cultured primary ccRCC cells) that specifically take up dopamine mediated by *SLC6A3*. The growth characteristic as well as the morphology of these cell lines are also more in line with primary ccRCC cells than conventionally used cell lines, such as 786-O. This observation emphasizes the importance of using representative model systems for studying human diseases *in vitro*.

Just recently a paper was published on *SLC6A3* in ccRCC. Schrödter et al. report that *SLC6A3* mRNA levels are extremely high in ccRCC, but also that the protein levels are very low and conversely, that normal kidney samples have low mRNA levels but that the PTs express high protein levels of *SLC6A3* (220). The authors also claim that the ccRCC cell lines Caki-1 and A-498 can be killed using an inhibitor of *SLC6A3*. However, they use *sertraline*, which is a selective serotonin reuptake inhibitor (SSRI). Moreover, this inhibitor was used at high concentrations, and appropriate controls using cells lacking *SLC6A3* expression are missing. In light of our detailed transcriptional profiling showing exceedingly low levels of *SLC6A3* in normal kidney cortex, the results presented by Schrödter et al., are difficult to comprehend. We have also tried to detect the *SLC6A3* protein in kidney tissue, both tumor and normal, using immunohistochemistry, but we have so far been unsuccessful. There are antibodies that work well on neuronal tissue, while no staining could be seen in kidney tissue. There could be several reasons for this discrepancy. Kidney tissue and neuronal tissue are of very distinctive natures and are usually handled differently prior to staining. Additionally, there might be differences in the post-translational modifications of *SLC6A3* in ccRCC compared to dopaminergic neurons. Also,



kidney cells, and PT cells in particular, are crowded with transporter proteins reflecting their normal function, opening up for the possibility that the antibody used by Schrödter et al. not only detects SLC6A3 but also another transporter, since the structural homology between transporter proteins can be high (221, 222). This putative cross-reaction might not have been seen when validating the antibody in neuronal tissue, since dopaminergic cells and kidney cells do not in general share transporter expression.

ccRCC is exceptionally resistant to conventional cancer treatments such as radiation and chemotherapy, and patients with metastatic ccRCC await a dismal prognosis. Despite recent treatment advances using targeted therapy, virtually all patients with mRCC will succumb to the disease. There is thus a desperate need for novel targeted therapies and diagnostic methods. The observation that SLC6A3 is highly expressed and functional in ccRCC, may have noticeable clinical impact for the development of novel targeted therapies and diagnostics in ccRCC. It is tempting to envision a therapeutic approach that allows for SLC6A3-specific uptake of small toxic molecules into the ccRCC cells. Alternatively, one could use cocaine-analogues for delivery of targeted radionuclide therapy, which would specifically bind SLC6A3 and selectively kill the ccRCC cells. These approaches would target ccRCC features distinct from the pathways targeted with current therapies, and could perhaps be used in combination to achieve synergistic effects. Additionally, there are already diagnostic techniques that utilize the presence for SLC6A3 for detecting and quantifying dopaminergic neurons using imaging modalities. These are based on radiolabelled cocaine analogues and are currently in clinical use for the diagnosis of PD (223). We propose that these compounds could be used also for the diagnosis of ccRCC, and possibly allow for high resolution of lymph node involvement and early metastatic spread. We are currently in the process of initiating such a clinical trial. Since the prognosis of ccRCC is extremely different depending on whether the disease is spread or not, the proposed alternative diagnostic approach could possibly help to guide treatment options and initiate targeted therapies as early as possible.

## Conclusion and Future perspective

In this thesis I have studied renal progenitor cells and kidney cancer and the findings of the included papers may lead to elevated insight to renal regeneration as well as to renal tumor development.

We herein present evidence for a novel progenitor cell type scattered among the tubular cells of the proximal part of the nephron. We propose that this cell type, PTPC, is involved in renal regeneration, and that it may represent the cell of origin for pRCC. Increased knowledge of the process of renal regeneration and progenitor cells has implications for treatment development for AKI and CKD, conditions associated with extensive morbidity and mortality as well as enormous medical cost for society. Further investigations of the regulation of PTPCs could perhaps be utilized for future cell based- or molecular therapies, to improve outcome following acute kidney damage as well as progressive kidney disease. Additionally, our findings regarding the species distribution of PTPCs could shed some light on the discrepant findings of the cellular source for regeneration within the field.

In this thesis we also present evidence for HIF driven upregulation of functional dopamine transporter *SLC6A3* in ccRCC. The consequences of the *SLC6A3* overexpression in ccRCC are yet to be revealed, however the finding has implications for the development of novel specific drugs targeting ccRCC, which currently is essentially therapy resistant. Finally, since there are already diagnostic methods based on radiolabelled *SLC6A3* ligands, I have high hopes for the usage of the same, or similar, imaging modalities for detection of ccRCC tumors and metastasis in the near future.



# Populärvetenskaplig sammanfattning

Njurarna har en mängd viktiga funktioner i kroppen men har som främsta uppgift att filtrera blodet och föra ut gifter ur kroppen genom bildandet av urin. Njurarna filtrerar uppskattningsvis 180 liter primärurin varje dygn och 99,5% av all vätska pumpas tillbaka till kroppen genom njurarnas rörsystem, vilket slutligen resulterar i endast 1,5 liter urin. Detta konstanta pumpande är extremt energikrävande och medför att njurcellerna är mycket känsliga för störningar i sin energi- och syretillförsel från blodet. Om njurarna får syrebrist, till exempel vid en massiv blödning efter en olycka, kan cellerna i njurens rörsystem dö, vilket kan leda till njursvikt som är direkt livshotande. Om patienten snabbt får hjälp att rena blodet på konstgjord väg, via dialys, kan njuren dock laga sig själv via så kallad regenerering. Hur det går till och vilka celler som utför hoplappandet av rörsystemen i njuren är fortfarande inte helt klarlagt. Njurarna kan också, som så många andra organ i kroppen, drabbas av cancer, där tumörcellerna delar sig okontrollerat på grund av mutationer, felaktigheter i cellens DNA. Njurcancer drabbar cirka 1000 personer om året i Sverige. Symptomen för njurcancer är vaga och hos en stor del av patienterna så har sjukdomen redan spridit sig utanför njuren då man upptäcker sjukdomen. För dessa patienter är prognosen mycket dystert då njurcancer inte svarar på vanliga cancerbehandlingar såsom cellgifter och strålning och knappt 10 % av dessa patienter förväntas överleva i fem år. Behovet av nya behandlingsformer så väl som nya diagnostiska metoder är därför mycket stort.

I den här avhandlingen beskrivs en helt ny celltyp som ligger vilande mellan de vanliga cellagren i njurens rörsystem. Celltypen är stamcellslik och är utrustad med förankringsprotein som troligtvis bidrar till att den inte lossnar lika lätt vid skada. Vi visar även att celler i regenererande rör delar markörer med den nya celltypen. I en ny metod för att simulera njurskada utanför kroppen, i en cellodlingsskål, såg vi att den nya celltypen är mycket mer motståndskraftig mot celldöd än de andra cellerna i njurens rörsystem. Vissa har hävdade att ytmarkörerna som uttrycks av den nya celltypen endast är tecken på skada efter ett långt liv av blodfiltrering. Här visar vi däremot visat att cellerna finns i njurar från unga friska personer från 2 år och uppåt. Till vår förvåning så fann vi att de nya cellerna inte kunde hittas i njurar från möss, men att de fanns i njurar från schimpans och gris. Detta skulle kunna förklara den förvirring som råder inom njurregenerations-fältet om huruvida cellerna som ansvar för reparation är stamceller som finns på plats innan skadan eller om skadan repareras

av vanliga njurceller som omprogrammeras. Många av de rapporter som hävdar att njurregenerationen inte utförs av specialiserade stamceller grundar sig nämligen på studier i möss. Vi föreslår alltså att det finns en skillnad i hur njurregenereringen går till i olika arter. Baserat på de fynd som gjorts i denna avhandling så tror vi att de nya cellerna kan aktiveras efter skada och då börjar dela sig för att ersätta det förlorade cellagret i den mänskliga njuren. Men det finns även en helt annan sida av celltypen. Celltypens genuttryck var nämligen mycket likt en typ av njurcancer, så kallad papilläer njurcancer. Det skulle alltså kunna vara just i dessa celler som denna typ av njurcancer uppkommer. Nästa steg är att ta reda på vad det är som aktiverar cellen och bidrar till läkning efter njurskada, men också vad som händer när cellen blir elakartad och ger upphov till cancer. På så sätt skulle denna upptäckt möjligtvis kunna bidra med idéer till nya behandlingar vid både njurskador och njurcancer.

I en annan del av avhandlingen har vi upptäckt att klarcellig njurcancer uttrycker mycket höga nivåer av dopamintransportören SLC6A3. I normala fall uttrycks endast denna transportör i dopaminproducerande celler i hjärnan, samma celler som för övrigt förloras vid Parkinsons sjukdom. Vi fann att andra tumörformer, och normal njure, uttrycker synnerligen låga nivåer av SLC6A3 och att det därför är en mycket specifik biomarkör för just klarcellig njurcancer. Vi visade också att njurcancer cellerna aktivt kan ta upp dopamin och att detta upptag kan hämmas genom att sätta till en specifik SLC6A3-hämmare. Det verkade även vara så att SLC6A3-uttrycket i njure styrs av ett protein, HIF-2a, som normalt sett uttrycks specifikt när cellerna har syrebrist, så kallad hypoxi. När vi behandlade normala njurceller med lågt syretryck så inducerades uttrycket av SLC6A3, men detta skedde inte med celler från andra normala vävnader så som bröst och blodkärl. I klarcellig njurcancer förekommer en speciell mutation som gör att cellerna alltid befinner sig i ett tillstånd som om det rådde hypoxi, även vid normalt syretryck. Denna pseudohypoxi skulle kunna vara en av förklaringarna till det höga SLC6A3-uttrycket i klarcellig njurcancer. Genom att använda sig av dopamin transportörens egenskaper tror vi att dessa fynd skulle kunna leda till utveckling av nya riktade behandlingsformer och ny diagnostik för klarcellig njurcancer.

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*“Forskning är lätt när man har rätt”*

Håkan Axelsson

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# References

1. Lote CJ. Principles of renal physiology. 5th ed. New York: Springer; 2012.
2. Guyton AC, Hall JE. Textbook of medical physiology. 11th ed. Philadelphia: Elsevier Saunders; 2006.
3. McDonuogh A, Thomson S, Brebbber B, Rector F. Metabolic basis of solute transport. *The Kidney: Physiology and Pathophysiology*. 2011:138-57.
4. Deshmukh SR, Wong NWK. *The renal system explained : an illustrated core text*. Nottingham: Nottingham University Press; 2009.
5. Slack JM. Stem cells in epithelial tissues. *Science*. 2000;287(5457):1431-3.
6. Ludwig TE, Levenstein ME, Jones JM, Berggren WT, Mitchen ER, Frane JL, et al. Derivation of human embryonic stem cells in defined conditions. *Nat Biotechnol*. 2006;24(2):185-7.
7. Holtzer H. Cell lineages, stem cells and the 'quantal' cell cycle concept. B.I. Lord CSP, and R.J. Cole. , editor. New York: Cambridge University Press; 1978.
8. Goichberg P, Chang J, Liao R, Leri A. Cardiac stem cells: biology and clinical applications. *Antioxid Redox Signal*. 2014;21(14):2002-17.
9. Morrison SJ, Spradling AC. Stem cells and niches: mechanisms that promote stem cell maintenance throughout life. *Cell*. 2008;132(4):598-611.
10. Simons BD, Clevers H. Strategies for homeostatic stem cell self-renewal in adult tissues. *Cell*. 2011;145(6):851-62.
11. Seaberg RM, van der Kooy D. Stem and progenitor cells: the premature desertion of rigorous definitions. *Trends Neurosci*. 2003;26(3):125-31.
12. Morrison SJ, Weissman IL. The long-term repopulating subset of hematopoietic stem cells is deterministic and isolatable by phenotype. *Immunity*. 1994;1(8):661-73.
13. Sato T, van Es JH, Snippert HJ, Stange DE, Vries RG, van den Born M, et al. Paneth cells constitute the niche for Lgr5 stem cells in intestinal crypts. *Nature*. 2011;469(7330):415-8.
14. Mohyeldin A, Garzon-Muvdi T, Quinones-Hinojosa A. Oxygen in stem cell biology: a critical component of the stem cell niche. *Cell Stem Cell*. 2010;7(2):150-61.
15. Oh M, Nor JE. The Perivascular Niche and Self-Renewal of Stem Cells. *Front Physiol*. 2015;6:367.
16. Takahashi K, Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell*. 2006;126(4):663-76.

17. Alison MR, Guppy NJ, Lim SM, Nicholson LJ. Finding cancer stem cells: are aldehyde dehydrogenases fit for purpose? *J Pathol.* 2010;222(4):335-44.
18. Pietras A. Cancer stem cells in tumor heterogeneity. *Adv Cancer Res.* 2011;112:255-81.
19. Greaves M. Evolutionary determinants of cancer. *Cancer Discov.* 2015;5(8):806-20.
20. Beachy PA, Karhadkar SS, Berman DM. Tissue repair and stem cell renewal in carcinogenesis. *Nature.* 2004;432(7015):324-31.
21. Lapidot T, Sirard C, Vormoor J, Murdoch B, Hoang T, Caceres-Cortes J, et al. A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. *Nature.* 1994;367(6464):645-8.
22. Visvader JE. Cells of origin in cancer. *Nature.* 2011;469(7330):314-22.
23. Waters D, Newman B, Levy ML. Stem cell origin of brain tumors. *Adv Exp Med Biol.* 2010;671:58-66.
24. White AC, Lowry WE. Refining the role for adult stem cells as cancer cells of origin. *Trends Cell Biol.* 2015;25(1):11-20.
25. DiMeo TA, Anderson K, Phadke P, Fan C, Perou CM, Naber S, et al. A novel lung metastasis signature links Wnt signaling with cancer cell self-renewal and epithelial-mesenchymal transition in basal-like breast cancer. *Cancer Res.* 2009;69(13):5364-73.
26. Giannoni E, Bianchini F, Masieri L, Serni S, Torre E, Calorini L, et al. Reciprocal activation of prostate cancer cells and cancer-associated fibroblasts stimulates epithelial-mesenchymal transition and cancer stemness. *Cancer Res.* 2010;70(17):6945-56.
27. Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, et al. The epithelial-mesenchymal transition generates cells with properties of stem cells. *Cell.* 2008;133(4):704-15.
28. Moitra K, Lou H, Dean M. Multidrug efflux pumps and cancer stem cells: insights into multidrug resistance and therapeutic development. *Clin Pharmacol Ther.* 2011;89(4):491-502.
29. Baum CM, Weissman IL, Tsukamoto AS, Buckle AM, Peault B. Isolation of a candidate human hematopoietic stem-cell population. *Proc Natl Acad Sci U S A.* 1992;89(7):2804-8.
30. Hou NY, Yang K, Chen T, Chen XZ, Zhang B, Mo XM, et al. CD133+ CD44+ subgroups may be human small intestinal stem cells. *Mol Biol Rep.* 2011;38(2):997-1004.
31. Huang EH, Hynes MJ, Zhang T, Ginestier C, Dontu G, Appelman H, et al. Aldehyde dehydrogenase 1 is a marker for normal and malignant human colonic stem cells (SC) and tracks SC overpopulation during colon tumorigenesis. *Cancer Res.* 2009;69(8):3382-9.
32. Barker N, van Es JH, Kuipers J, Kujala P, van den Born M, Cozijnsen M, et al. Identification of stem cells in small intestine and colon by marker gene *Lgr5*. *Nature.* 2007;449(7165):1003-7.
33. Goldman B. Magic marker myths. *Nature Reports Stem Cells.* 2008.

34. Goodell MA, Brose K, Paradis G, Conner AS, Mulligan RC. Isolation and functional properties of murine hematopoietic stem cells that are replicating in vivo. *J Exp Med.* 1996;183(4):1797-806.
35. Sladek NE. Human aldehyde dehydrogenases: potential pathological, pharmacological, and toxicological impact. *J Biochem Mol Toxicol.* 2003;17(1):7-23.
36. Douville J, Beaulieu R, Balicki D. ALDH1 as a functional marker of cancer stem and progenitor cells. *Stem Cells Dev.* 2009;18(1):17-25.
37. Bellomo R, Ronco C, Kellum JA, Mehta RL, Palevsky P, Acute Dialysis Quality Initiative w. Acute renal failure - definition, outcome measures, animal models, fluid therapy and information technology needs: the Second International Consensus Conference of the Acute Dialysis Quality Initiative (ADQI) Group. *Crit Care.* 2004;8(4):R204-12.
38. Tao Li PK, Burdmann EA, Mehta RL. Acute kidney injury: global health alert. *Int J Organ Transplant Med.* 2013;4(1):1-8.
39. Uchino S, Kellum JA, Bellomo R, Doig GS, Morimatsu H, Morgera S, et al. Acute renal failure in critically ill patients: a multinational, multicenter study. *JAMA.* 2005;294(7):813-8.
40. Murugan R, Kellum JA. Acute kidney injury: what's the prognosis? *Nat Rev Nephrol.* 2011;7(4):209-17.
41. Vinay Kumar, Abdul K. Abbas, Nelson Fausto, Mitchell R. Robbins Basic Pathology. 8th ed. Philadelphia: W. B. Saunders Company; 2007. p. 541-77.
42. Jennette JC, Heptinstall RH. Heptinstall's pathology of the kidney. 6th ed. Philadelphia: Lippincott Williams & Wilkins; 2007.
43. Weinberg JM, Venkatachalam MA, Roeser NF, Saikumar P, Dong Z, Senter RA, et al. Anaerobic and aerobic pathways for salvage of proximal tubules from hypoxia-induced mitochondrial injury. *Am J Physiol Renal Physiol.* 2000;279(5):F927-43.
44. Stevens A, Lowe JS. Pathology. 2nd ed. Edinburgh ; New York: Mosby; 2000. p. xv, 652 p.
45. Singh SR, Liu W, Hou SX. The adult *Drosophila* malpighian tubules are maintained by multipotent stem cells. *Cell Stem Cell.* 2007;1(2):191-203.
46. Diep CQ, Ma D, Deo RC, Holm TM, Naylor RW, Arora N, et al. Identification of adult nephron progenitors capable of kidney regeneration in zebrafish. *Nature.* 2011;470(7332):95-100.
47. Vogetseder A, Picard N, Gaspert A, Walch M, Kaissling B, Le Hir M. Proliferation capacity of the renal proximal tubule involves the bulk of differentiated epithelial cells. *Am J Physiol Cell Physiol.* 2008;294(1):C22-8.
48. Witzgall R, Brown D, Schwarz C, Bonventre JV. Localization of proliferating cell nuclear antigen, vimentin, c-Fos, and clusterin in the postischemic kidney. Evidence for a heterogenous genetic response among nephron segments, and a large pool of mitotically active and dedifferentiated cells. *J Clin Invest.* 1994;93(5):2175-88.

49. Humphreys BD, Czerniak S, DiRocco DP, Hasnain W, Cheema R, Bonventre JV. Repair of injured proximal tubule does not involve specialized progenitors. *Proc Natl Acad Sci U S A*. 2011;108(22):9226-31.
50. Humphreys BD, Valerius MT, Kobayashi A, Mugford JW, Soeung S, Duffield JS, et al. Intrinsic epithelial cells repair the kidney after injury. *Cell Stem Cell*. 2008;2(3):284-91.
51. Kusaba T, Lalli M, Kramann R, Kobayashi A, Humphreys BD. Differentiated kidney epithelial cells repair injured proximal tubule. *Proc Natl Acad Sci U S A*. 2014;111(4):1527-32.
52. De Broe ME. Tubular regeneration and the role of bone marrow cells: 'stem cell therapy'--a panacea? *Nephrol Dial Transplant*. 2005;20(11):2318-20.
53. Kale S, Karihaloo A, Clark PR, Kashgarian M, Krause DS, Cantley LG. Bone marrow stem cells contribute to repair of the ischemically injured renal tubule. *J Clin Invest*. 2003;112(1):42-9.
54. Humphreys BD, Duffield JS, Bonventre JV. Renal stem cells in recovery from acute kidney injury. *Minerva Urol Nefrol*. 2006;58(4):329-37.
55. Togel FE, Westenfelder C. Kidney protection and regeneration following acute injury: progress through stem cell therapy. *Am J Kidney Dis*. 2012;60(6):1012-22.
56. Sagrinati C, Ronconi E, Lazzeri E, Lasagni L, Romagnani P. Stem-cell approaches for kidney repair: choosing the right cells. *Trends Mol Med*. 2008;14(7):277-85.
57. Oliver JA, Maarouf O, Cheema FH, Martens TP, Al-Awqati Q. The renal papilla is a niche for adult kidney stem cells. *J Clin Invest*. 2004;114(6):795-804.
58. Bussolati B, Bruno S, Grange C, Buttiglieri S, Deregibus MC, Cantino D, et al. Isolation of renal progenitor cells from adult human kidney. *Am J Pathol*. 2005;166(2):545-55.
59. Hishikawa K, Marumo T, Miura S, Nakanishi A, Matsuzaki Y, Shibata K, et al. Musculin/MyoR is expressed in kidney side population cells and can regulate their function. *J Cell Biol*. 2005;169(6):921-8.
60. Appel D, Kershaw DB, Smeets B, Yuan G, Fuss A, Frye B, et al. Recruitment of podocytes from glomerular parietal epithelial cells. *J Am Soc Nephrol*. 2009;20(2):333-43.
61. Ronconi E, Sagrinati C, Angelotti ML, Lazzeri E, Mazzinghi B, Ballerini L, et al. Regeneration of glomerular podocytes by human renal progenitors. *J Am Soc Nephrol*. 2009;20(2):322-32.
62. Gupta S, Verfaillie C, Chmielewski D, Kren S, Eidman K, Connaire J, et al. Isolation and characterization of kidney-derived stem cells. *J Am Soc Nephrol*. 2006;17(11):3028-40.
63. Maeshima A, Sakurai H, Nigam SK. Adult kidney tubular cell population showing phenotypic plasticity, tubulogenic capacity, and integration capability into developing kidney. *J Am Soc Nephrol*. 2006;17(1):188-98.
64. Sagrinati C, Netti GS, Mazzinghi B, Lazzeri E, Liotta F, Frosali F, et al. Isolation and characterization of multipotent progenitor cells from the Bowman's capsule of adult human kidneys. *J Am Soc Nephrol*. 2006;17(9):2443-56.

65. Jiang Y, Jahagirdar BN, Reinhardt RL, Schwartz RE, Keene CD, Ortiz-Gonzalez XR, et al. Pluripotency of mesenchymal stem cells derived from adult marrow. *Nature*. 2002;418(6893):41-9.
66. Kubota H, Avarbock MR, Brinster RL. Spermatogonial stem cells share some, but not all, phenotypic and functional characteristics with other stem cells. *Proc Natl Acad Sci U S A*. 2003;100(11):6487-92.
67. Richardson GD, Robson CN, Lang SH, Neal DE, Maitland NJ, Collins AT. CD133, a novel marker for human prostatic epithelial stem cells. *J Cell Sci*. 2004;117(Pt 16):3539-45.
68. Shackleton M, Vaillant F, Simpson KJ, Stingl J, Smyth GK, Asselin-Labat ML, et al. Generation of a functional mammary gland from a single stem cell. *Nature*. 2006;439(7072):84-8.
69. Platt JL, LeBien TW, Michael AF. Stages of renal ontogenesis identified by monoclonal antibodies reactive with lymphohemopoietic differentiation antigens. *J Exp Med*. 1983;157(1):155-72.
70. Pippin JW, Sparks MA, Glenn ST, Buitrago S, Coffman TM, Duffield JS, et al. Cells of renin lineage are progenitors of podocytes and parietal epithelial cells in experimental glomerular disease. *Am J Pathol*. 2013;183(2):542-57.
71. Lindgren D, Bostrom AK, Nilsson K, Hansson J, Sjolund J, Moller C, et al. Isolation and characterization of progenitor-like cells from human renal proximal tubules. *Am J Pathol*. 2011;178(2):828-37.
72. Hansson J, Hultenby K, Cramnert C, Ponten F, Jansson H, Lindgren D, et al. Evidence for a morphologically distinct and functionally robust cell type in the proximal tubules of human kidney. *Hum Pathol*. 2014;45(2):382-93.
73. Angelotti ML, Ronconi E, Ballerini L, Peired A, Mazzinghi B, Sagrinati C, et al. Characterization of renal progenitors committed toward tubular lineage and their regenerative potential in renal tubular injury. *Stem Cells*. 2012;30(8):1714-25.
74. Sallustio F, Costantino V, Cox SN, Loverre A, Divella C, Rizzi M, et al. Human renal stem/progenitor cells repair tubular epithelial cell injury through TLR2-driven inhibin-A and microvesicle-shuttled decorin. *Kidney Int*. 2013;83(3):392-403.
75. Smeets B, Boor P, Dijkman H, Sharma SV, Jirak P, Mooren F, et al. Proximal tubular cells contain a phenotypically distinct, scattered cell population involved in tubular regeneration. *J Pathol*. 2013;229(5):645-59.
76. Berger K, Bangen JM, Hammerich L, Liedtke C, Floege J, Smeets B, et al. Origin of regenerating tubular cells after acute kidney injury. *Proc Natl Acad Sci U S A*. 2014;111(4):1533-8.
77. Lombardi D, Becherucci F, Romagnani P. How much can the tubule regenerate and who does it? An open question. *Nephrol Dial Transplant*. 2015.
78. Miyaoka Y, Ebato K, Kato H, Arakawa S, Shimizu S, Miyajima A. Hypertrophy and unconventional cell division of hepatocytes underlie liver regeneration. *Curr Biol*. 2012;22(13):1166-75.

79. Cuppage FE, Tate A. Repair of the nephron following injury with mercuric chloride. *Am J Pathol.* 1967;51(3):405-29.
80. Romagnani P, Rinkevich Y, Dekel B. The use of lineage tracing to study kidney injury and regeneration. *Nat Rev Nephrol.* 2015;11(7):420-31.
81. Langworthy M, Zhou B, de Caestecker M, Moeckel G, Baldwin HS. NFATc1 identifies a population of proximal tubule cell progenitors. *J Am Soc Nephrol.* 2009;20(2):311-21.
82. Ousset M, Van Keymeulen A, Bouvencourt G, Sharma N, Achouri Y, Simons BD, et al. Multipotent and unipotent progenitors contribute to prostate postnatal development. *Nat Cell Biol.* 2012;14(11):1131-8.
83. Mascré G, Dekoninck S, Drogat B, Youssef KK, Brohee S, Sotiropoulou PA, et al. Distinct contribution of stem and progenitor cells to epidermal maintenance. *Nature.* 2012;489(7415):257-62.
84. Hsu YC, Li L, Fuchs E. Transit-amplifying cells orchestrate stem cell activity and tissue regeneration. *Cell.* 2014;157(4):935-49.
85. Basile DP, Anderson MD, Sutton TA. Pathophysiology of acute kidney injury. *Compr Physiol.* 2012;2(2):1303-53.
86. Rinkevich Y, Montoro DT, Contreras-Trujillo H, Harari-Steinberg O, Newman AM, Tsai JM, et al. In vivo clonal analysis reveals lineage-restricted progenitor characteristics in mammalian kidney development, maintenance, and regeneration. *Cell Rep.* 2014;7(4):1270-83.
87. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer.* 2015;136(5):E359-86.
88. Znaor A, Lortet-Tieulent J, Laversanne M, Jemal A, Bray F. International variations and trends in renal cell carcinoma incidence and mortality. *Eur Urol.* 2015;67(3):519-30.
89. Rini BI, Atkins MB. Resistance to targeted therapy in renal-cell carcinoma. *Lancet Oncol.* 2009;10(10):992-1000.
90. Shuch B, Vourganti S, Ricketts CJ, Middleton L, Peterson J, Merino MJ, et al. Defining early-onset kidney cancer: implications for germline and somatic mutation testing and clinical management. *J Clin Oncol.* 2014;32(5):431-7.
91. Ljungberg B, Cowan NC, Hanbury DC, Hora M, Kuczyk MA, Merseburger AS, et al. EAU guidelines on renal cell carcinoma: the 2010 update. *Eur Urol.* 2010;58(3):398-406.
92. Hunt JD, van der Hel OL, McMillan GP, Boffetta P, Brennan P. Renal cell carcinoma in relation to cigarette smoking: meta-analysis of 24 studies. *Int J Cancer.* 2005;114(1):101-8.
93. Chow WH, Dong LM, Devesa SS. Epidemiology and risk factors for kidney cancer. *Nat Rev Urol.* 2010;7(5):245-57.
94. Gati A, Kouidhi S, Marrakchi R, El Gaaied A, Kourda N, Derouiche A, et al. Obesity and renal cancer: Role of adipokines in the tumor-immune system conflict. *Oncoimmunology.* 2014;3(1):e27810.

95. Penticuff JC, Kyprianou N. Therapeutic challenges in renal cell carcinoma. *Am J Clin Exp Urol.* 2015;3(2):77-90.
96. Nagy A, Walter E, Zubakov D, Kovacs G. High risk of development of renal cell tumor in end-stage kidney disease: the role of microenvironment. *Tumour Biol.* 2016.
97. Cohen HT, McGovern FJ. Renal-cell carcinoma. *N Engl J Med.* 2005;353(23):2477-90.
98. Ng KL, Rajandram R, Morais C, Yap NY, Samaratunga H, Gobe GC, et al. Differentiation of oncocytoma from chromophobe renal cell carcinoma (RCC): can novel molecular biomarkers help solve an old problem? *J Clin Pathol.* 2014;67(2):97-104.
99. Patard JJ, Leray E, Rioux-Leclercq N, Cindolo L, Ficarra V, Zisman A, et al. Prognostic value of histologic subtypes in renal cell carcinoma: a multicenter experience. *J Clin Oncol.* 2005;23(12):2763-71.
100. Sato Y, Yoshizato T, Shiraishi Y, Maekawa S, Okuno Y, Kamura T, et al. Integrated molecular analysis of clear-cell renal cell carcinoma. *Nat Genet.* 2013;45(8):860-7.
101. Turajlic S, Larkin J, Swanton C. SnapShot: Renal Cell Carcinoma. *Cell.* 2015;163(6):1556- e1.
102. Ward PS, Thompson CB. Metabolic reprogramming: a cancer hallmark even warburg did not anticipate. *Cancer Cell.* 2012;21(3):297-308.
103. Wettersten HI, Hakimi AA, Morin D, Bianchi C, Johnstone ME, Donohoe DR, et al. Grade-Dependent Metabolic Reprogramming in Kidney Cancer Revealed by Combined Proteomics and Metabolomics Analysis. *Cancer Res.* 2015;75(12):2541-52.
104. Chung KT, Gadupudi GS. Possible roles of excess tryptophan metabolites in cancer. *Environ Mol Mutagen.* 2011;52(2):81-104.
105. Courthod G, Tucci M, Di Maio M, Scagliotti GV. Papillary renal cell carcinoma: A review of the current therapeutic landscape. *Crit Rev Oncol Hematol.* 2015;96(1):100-12.
106. Kinch L, Grishin NV, Brugarolas J. Succination of Keap1 and activation of Nrf2-dependent antioxidant pathways in FH-deficient papillary renal cell carcinoma type 2. *Cancer Cell.* 2011;20(4):418-20.
107. Ooi A, Wong JC, Petillo D, Roossien D, Perrier-Trudova V, Whitten D, et al. An antioxidant response phenotype shared between hereditary and sporadic type 2 papillary renal cell carcinoma. *Cancer Cell.* 2011;20(4):511-23.
108. Cheadle JP, Reeve MP, Sampson JR, Kwiatkowski DJ. Molecular genetic advances in tuberous sclerosis. *Hum Genet.* 2000;107(2):97-114.
109. Storkel S, Eble JN, Adlakhia K, Amin M, Blute ML, Bostwick DG, et al. Classification of renal cell carcinoma: Workgroup No. 1. Union Internationale Contre le Cancer (UICC) and the American Joint Committee on Cancer (AJCC). *Cancer.* 1997;80(5):987-9.
110. Davis CF, Ricketts CJ, Wang M, Yang L, Cherniack AD, Shen H, et al. The somatic genomic landscape of chromophobe renal cell carcinoma. *Cancer Cell.* 2014;26(3):319-30.



111. Prasad SR, Narra VR, Shah R, Humphrey PA, Jagirdar J, Catena JR, et al. Segmental disorders of the nephron: histopathological and imaging perspective. *Br J Radiol.* 2007;80(956):593-602.
112. Stec R, Grala B, Maczewski M, Bodnar L, Szczylik C. Chromophobe renal cell cancer--review of the literature and potential methods of treating metastatic disease. *J Exp Clin Cancer Res.* 2009;28:134.
113. Linehan WM, Pinto PA, Bratslavsky G, Pfaffenroth E, Merino M, Vocke CD, et al. Hereditary kidney cancer: unique opportunity for disease-based therapy. *Cancer.* 2009;115(10 Suppl):2252-61.
114. Baba M, Furihata M, Hong SB, Tessarollo L, Haines DC, Southon E, et al. Kidney-targeted Birt-Hogg-Dube gene inactivation in a mouse model: Erk1/2 and Akt-mTOR activation, cell hyperproliferation, and polycystic kidneys. *J Natl Cancer Inst.* 2008;100(2):140-54.
115. Nickerson ML, Jaeger E, Shi Y, Durocher JA, Mahurkar S, Zaridze D, et al. Improved identification of von Hippel-Lindau gene alterations in clear cell renal tumors. *Clin Cancer Res.* 2008;14(15):4726-34.
116. Herman JG, Latif F, Weng Y, Lerman MI, Zbar B, Liu S, et al. Silencing of the VHL tumor-suppressor gene by DNA methylation in renal carcinoma. *Proc Natl Acad Sci U S A.* 1994;91(21):9700-4.
117. Gossage L, Eisen T, Maher ER. VHL, the story of a tumour suppressor gene. *Nat Rev Cancer.* 2015;15(1):55-64.
118. Shen C, Beroukhi R, Schumacher SE, Zhou J, Chang M, Signoretti S, et al. Genetic and functional studies implicate HIF1alpha as a 14q kidney cancer suppressor gene. *Cancer Discov.* 2011;1(3):222-35.
119. Mandriota SJ, Turner KJ, Davies DR, Murray PG, Morgan NV, Sowter HM, et al. HIF activation identifies early lesions in VHL kidneys: evidence for site-specific tumor suppressor function in the nephron. *Cancer Cell.* 2002;1(5):459-68.
120. Kondo K, Kim WY, Lechpammer M, Kaelin WG, Jr. Inhibition of HIF2alpha is sufficient to suppress pVHL-defective tumor growth. *PLoS Biol.* 2003;1(3):E83.
121. Minton DR, Fu L, Mongan NP, Shevchuk MM, Nanus DM, Gudas LJ. Role of NADH Dehydrogenase (Ubiquinone) 1 alpha subcomplex 4-like 2 in clear cell renal cell carcinoma. *Clin Cancer Res.* 2016.
122. Fu L, Wang G, Shevchuk MM, Nanus DM, Gudas LJ. Generation of a mouse model of Von Hippel-Lindau kidney disease leading to renal cancers by expression of a constitutively active mutant of HIF1alpha. *Cancer Res.* 2011;71(21):6848-56.
123. Fu L, Wang G, Shevchuk MM, Nanus DM, Gudas LJ. Activation of HIF2alpha in kidney proximal tubule cells causes abnormal glycogen deposition but not tumorigenesis. *Cancer Res.* 2013;73(9):2916-25.
124. Soultati A, Stares M, Swanton C, Larkin J, Turajlic S. How should clinicians address intratumour heterogeneity in clear cell renal cell carcinoma? *Curr Opin Urol.* 2015;25(5):358-66.

125. Brugarolas J. Molecular genetics of clear-cell renal cell carcinoma. *J Clin Oncol.* 2014;32(18):1968-76.
126. Liao L, Testa JR, Yang H. The roles of chromatin-remodelers and epigenetic modifiers in kidney cancer. *Cancer Genet.* 2015;208(5):206-14.
127. Varela I, Tarpey P, Raine K, Huang D, Ong CK, Stephens P, et al. Exome sequencing identifies frequent mutation of the SWI/SNF complex gene PBRM1 in renal carcinoma. *Nature.* 2011;469(7331):539-42.
128. da Costa WH, Rezende M, Carneiro FC, Rocha RM, da Cunha IW, Carraro DM, et al. Polybromo-1 (PBRM1), a SWI/SNF complex subunit is a prognostic marker in clear cell renal cell carcinoma. *BJU Int.* 2014;113(5b):E157-63.
129. Pawlowski R, Muhl SM, Sulser T, Krek W, Moch H, Schraml P. Loss of PBRM1 expression is associated with renal cell carcinoma progression. *Int J Cancer.* 2013;132(2):E11-7.
130. Hakimi AA, Ostrovskaya I, Reva B, Schultz N, Chen YB, Gonen M, et al. Adverse outcomes in clear cell renal cell carcinoma with mutations of 3p21 epigenetic regulators BAP1 and SETD2: a report by MSKCC and the KIRC TCGA research network. *Clin Cancer Res.* 2013;19(12):3259-67.
131. Kapur P, Pena-Llopis S, Christie A, Zhrebker L, Pavia-Jimenez A, Rathmell WK, et al. Effects on survival of BAP1 and PBRM1 mutations in sporadic clear-cell renal-cell carcinoma: a retrospective analysis with independent validation. *Lancet Oncol.* 2013;14(2):159-67.
132. Gerlinger M, Rowan AJ, Horswell S, Larkin J, Endesfelder D, Gronroos E, et al. Intratumor heterogeneity and branched evolution revealed by multiregion sequencing. *N Engl J Med.* 2012;366(10):883-92.
133. Joseph RW, Kapur P, Serie DJ, Eckel-Passow JE, Parasramka M, Ho T, et al. Loss of BAP1 protein expression is an independent marker of poor prognosis in patients with low-risk clear cell renal cell carcinoma. *Cancer.* 2014;120(7):1059-67.
134. Pena-Llopis S, Vega-Rubin-de-Celis S, Liao A, Leng N, Pavia-Jimenez A, Wang S, et al. BAP1 loss defines a new class of renal cell carcinoma. *Nat Genet.* 2012;44(7):751-9.
135. Nationellt vårdprogram för Njurcancer. Regionalt cancercentrum Stockholm Gotland: 2013 2013-03-26. Report No.: Contract No.: ISBN: 978-91-85947-37-9.
136. Escudier B, Eisen T, Porta C, Patard JJ, Khoo V, Algaba F, et al. Renal cell carcinoma: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol.* 2012;23 Suppl 7:vii65-71.
137. McDermott DF, Atkins MB. Immunotherapy of metastatic renal cell carcinoma. *Cancer J.* 2008;14(5):320-4.
138. Atkins M, Regan M, McDermott D, Mier J, Stanbridge E, Youmans A, et al. Carbonic anhydrase IX expression predicts outcome of interleukin 2 therapy for renal cancer. *Clin Cancer Res.* 2005;11(10):3714-21.
139. Atkins MB, Choueiri TK, Cho D, Regan M, Signoretti S. Treatment selection for patients with metastatic renal cell carcinoma. *Cancer.* 2009;115(10 Suppl):2327-33.

140. Folkman J. Tumor angiogenesis: therapeutic implications. *N Engl J Med.* 1971;285(21):1182-6.
141. Al-Husein B, Abdalla M, Trepte M, Deremer DL, Somanath PR. Antiangiogenic therapy for cancer: an update. *Pharmacotherapy.* 2012;32(12):1095-111.
142. Carmeliet P, Jain RK. Principles and mechanisms of vessel normalization for cancer and other angiogenic diseases. *Nat Rev Drug Discov.* 2011;10(6):417-27.
143. Hudes GR. Targeting mTOR in renal cell carcinoma. *Cancer.* 2009;115(10 Suppl):2313-20.
144. Polivka J, Jr., Janku F. Molecular targets for cancer therapy in the PI3K/AKT/mTOR pathway. *Pharmacol Ther.* 2014;142(2):164-75.
145. Choueiri TK, Vaishampayan U, Rosenberg JE, Logan TF, Harzstark AL, Bukowski RM, et al. Phase II and biomarker study of the dual MET/VEGFR2 inhibitor foretinib in patients with papillary renal cell carcinoma. *J Clin Oncol.* 2013;31(2):181-6.
146. Flanigan RC, Mickisch G, Sylvester R, Tangen C, Van Poppel H, Crawford ED. Cytoreductive nephrectomy in patients with metastatic renal cancer: a combined analysis. *J Urol.* 2004;171(3):1071-6.
147. Fairlamb DJ. Spontaneous regression of metastases of renal cancer: A report of two cases including the first recorded regression following irradiation of a dominant metastasis and review of the world literature. *Cancer.* 1981;47(8):2102-6.
148. Vogelzang NJ, Priest ER, Borden L. Spontaneous regression of histologically proved pulmonary metastases from renal cell carcinoma: a case with 5-year followup. *J Urol.* 1992;148(4):1247-8.
149. Weinstock M, McDermott D. Targeting PD-1/PD-L1 in the treatment of metastatic renal cell carcinoma. *Ther Adv Urol.* 2015;7(6):365-77.
150. Thompson RH, Kuntz SM, Leibovich BC, Dong H, Lohse CM, Webster WS, et al. Tumor B7-H1 is associated with poor prognosis in renal cell carcinoma patients with long-term follow-up. *Cancer Res.* 2006;66(7):3381-5.
151. Choueiri TK, Fay AP, Gray KP, Callea M, Ho TH, Albiges L, et al. PD-L1 expression in nonclear-cell renal cell carcinoma. *Ann Oncol.* 2014;25(11):2178-84.
152. Motzer RJ, Escudier B, McDermott DF, George S, Hammers HJ, Srinivas S, et al. Nivolumab versus Everolimus in Advanced Renal-Cell Carcinoma. *N Engl J Med.* 2015;373(19):1803-13.
153. Hediger MA, Clemençon B, Burrier RE, Bruford EA. The ABCs of membrane transporters in health and disease (SLC series): introduction. *Mol Aspects Med.* 2013;34(2-3):95-107.
154. Cesar-Razquin A, Snijder B, Frappier-Brinton T, Isserlin R, Gyimesi G, Bai X, et al. A Call for Systematic Research on Solute Carriers. *Cell.* 2015;162(3):478-87.
155. Dean MC. The human ATP-binding cassette (ABC) transporter superfamily. Bethesda, MD: NCBI; 2002. Available from: <http://www.ncbi.nlm.nih.gov/books/NBK3>.

156. Tang L, Bergevoet SM, Gilissen C, de Witte T, Jansen JH, van der Reijden BA, et al. Hematopoietic stem cells exhibit a specific ABC transporter gene expression profile clearly distinct from other stem cells. *BMC Pharmacol.* 2010;10:12.
157. Choudhuri S, Klaassen CD. Structure, function, expression, genomic organization, and single nucleotide polymorphisms of human ABCB1 (MDR1), ABCC (MRP), and ABCG2 (BCRP) efflux transporters. *Int J Toxicol.* 2006;25(4):231-59.
158. Nakanishi T. Drug transporters as targets for cancer chemotherapy. *Cancer Genomics Proteomics.* 2007;4(3):241-54.
159. Hagenbuch B, Stieger B. The SLCO (former SLC21) superfamily of transporters. *Mol Aspects Med.* 2013;34(2-3):396-412.
160. Kristensen AS, Andersen J, Jorgensen TN, Sorensen L, Eriksen J, Loland CJ, et al. SLC6 neurotransmitter transporters: structure, function, and regulation. *Pharmacol Rev.* 2011;63(3):585-640.
161. Penmatsa A, Wang KH, Gouaux E. X-ray structure of dopamine transporter elucidates antidepressant mechanism. *Nature.* 2013;503(7474):85-90.
162. Marsden CD. The neuropharmacology of abnormal involuntary movement disorders (the dyskinesias). *Mod Trends Neurol.* 1975;6:141-66.
163. Albin RL, Mink JW. Recent advances in Tourette syndrome research. *Trends Neurosci.* 2006;29(3):175-82.
164. Chaudhuri KR, Healy DG, Schapira AH, National Institute for Clinical E. Non-motor symptoms of Parkinson's disease: diagnosis and management. *Lancet Neurol.* 2006;5(3):235-45.
165. Scatton B, Javoy-Agid F, Rouquier L, Dubois B, Agid Y. Reduction of cortical dopamine, noradrenaline, serotonin and their metabolites in Parkinson's disease. *Brain Res.* 1983;275(2):321-8.
166. Wise RA, Bozarth MA. Brain mechanisms of drug reward and euphoria. *Psychiatr Med.* 1985;3(4):445-60.
167. Bach ME, Barad M, Son H, Zhuo M, Lu YF, Shih R, et al. Age-related defects in spatial memory are correlated with defects in the late phase of hippocampal long-term potentiation in vitro and are attenuated by drugs that enhance the cAMP signaling pathway. *Proc Natl Acad Sci U S A.* 1999;96(9):5280-5.
168. Goldberg TE, Weinberger DR. The effects of clozapine on neurocognition: an overview. *J Clin Psychiatry.* 1994;55 Suppl B:88-90.
169. Kebabian JW, Calne DB. Multiple receptors for dopamine. *Nature.* 1979;277(5692):93-6.
170. Beaulieu JM, Gainetdinov RR. The physiology, signaling, and pharmacology of dopamine receptors. *Pharmacol Rev.* 2011;63(1):182-217.
171. Pifl C, Agneter E, Drobny H, Reither H, Singer EA. Induction by low Na<sup>+</sup> or Cl<sup>-</sup> of cocaine sensitive carrier-mediated efflux of amines from cells transfected with the cloned human catecholamine transporters. *Br J Pharmacol.* 1997;121(2):205-12.

172. Uhl GR. Dopamine transporter: basic science and human variation of a key molecule for dopaminergic function, locomotion, and parkinsonism. *Mov Disord.* 2003;18 Suppl 7:S71-80.
173. Miller GM. The emerging role of trace amine-associated receptor 1 in the functional regulation of monoamine transporters and dopaminergic activity. *J Neurochem.* 2011;116(2):164-76.
174. Xie Z, Miller GM. A receptor mechanism for methamphetamine action in dopamine transporter regulation in brain. *J Pharmacol Exp Ther.* 2009;330(1):316-25.
175. Saunders C, Ferrer JV, Shi L, Chen J, Merrill G, Lamb ME, et al. Amphetamine-induced loss of human dopamine transporter activity: an internalization-dependent and cocaine-sensitive mechanism. *Proc Natl Acad Sci U S A.* 2000;97(12):6850-5.
176. Fleckenstein AE, Volz TJ, Riddle EL, Gibb JW, Hanson GR. New insights into the mechanism of action of amphetamines. *Annu Rev Pharmacol Toxicol.* 2007;47:681-98.
177. Cameron KN, Solis E, Jr., Ruchala I, De Felice LJ, Eltit JM. Amphetamine activates calcium channels through dopamine transporter-mediated depolarization. *Cell Calcium.* 2015;58(5):457-66.
178. Steinkellner T, Yang JW, Montgomery TR, Chen WQ, Winkler MT, Sucic S, et al. Ca(2+)/calmodulin-dependent protein kinase IIalpha (alphaCaMKII) controls the activity of the dopamine transporter: implications for Angelman syndrome. *J Biol Chem.* 2012;287(35):29627-35.
179. Kuhar MJ, Ritz MC, Boja JW. The dopamine hypothesis of the reinforcing properties of cocaine. *Trends Neurosci.* 1991;14(7):299-302.
180. Beuming T, Kniazeff J, Bergmann ML, Shi L, Gracia L, Raniszewska K, et al. The binding sites for cocaine and dopamine in the dopamine transporter overlap. *Nat Neurosci.* 2008;11(7):780-9.
181. Verma V. Classic Studies on the Interaction of Cocaine and the Dopamine Transporter. *Clin Psychopharmacol Neurosci.* 2015;13(3):227-38.
182. Blackstone C. Infantile parkinsonism-dystonia: a dopamine "transportopathy". *J Clin Invest.* 2009;119(6):1455-8.
183. Kurian MA, Zhen J, Cheng SY, Li Y, Mordekar SR, Jardine P, et al. Homozygous loss-of-function mutations in the gene encoding the dopamine transporter are associated with infantile parkinsonism-dystonia. *J Clin Invest.* 2009;119(6):1595-603.
184. Gizer IR, Ficks C, Waldman ID. Candidate gene studies of ADHD: a meta-analytic review. *Hum Genet.* 2009;126(1):51-90.
185. Yang B, Chan RC, Jing J, Li T, Sham P, Chen RY. A meta-analysis of association studies between the 10-repeat allele of a VNTR polymorphism in the 3'-UTR of dopamine transporter gene and attention deficit hyperactivity disorder. *Am J Med Genet B Neuropsychiatr Genet.* 2007;144B(4):541-50.
186. Faraone SV, Spencer TJ, Madras BK, Zhang-James Y, Biederman J. Functional effects of dopamine transporter gene genotypes on in vivo dopamine transporter functioning: a meta-analysis. *Mol Psychiatry.* 2014;19(8):880-9.

187. Du Y, Nie Y, Li Y, Wan YJ. The association between the SLC6A3 VNTR 9-repeat allele and alcoholism-a meta-analysis. *Alcohol Clin Exp Res.* 2011;35(9):1625-34.
188. Grunhage F, Schulze TG, Muller DJ, Lanczik M, Franzek E, Albus M, et al. Systematic screening for DNA sequence variation in the coding region of the human dopamine transporter gene (DAT1). *Mol Psychiatry.* 2000;5(3):275-82.
189. Tarnok Z, Ronai Z, Gervai J, Kereszturi E, Gadoros J, Sasvari-Szekely M, et al. Dopaminergic candidate genes in Tourette syndrome: association between tic severity and 3' UTR polymorphism of the dopamine transporter gene. *Am J Med Genet B Neuropsychiatr Genet.* 2007;144B(7):900-5.
190. Gainetdinov RR. Dopamine transporter mutant mice in experimental neuropharmacology. *Naunyn Schmiedebergs Arch Pharmacol.* 2008;377(4-6):301-13.
191. Dickson DW, Braak H, Duda JE, Duyckaerts C, Gasser T, Halliday GM, et al. Neuropathological assessment of Parkinson's disease: refining the diagnostic criteria. *Lancet Neurol.* 2009;8(12):1150-7.
192. Tissingh G, Bergmans P, Booij J, Winogrodzka A, van Royen EA, Stoof JC, et al. Drug-naive patients with Parkinson's disease in Hoehn and Yahr stages I and II show a bilateral decrease in striatal dopamine transporters as revealed by [123I]beta-CIT SPECT. *J Neurol.* 1998;245(1):14-20.
193. Bajaj N, Hauser RA, Grachev ID. Clinical utility of dopamine transporter single photon emission CT (DaT-SPECT) with (123I) ioflupane in diagnosis of parkinsonian syndromes. *J Neurol Neurosurg Psychiatry.* 2013;84(11):1288-95.
194. Walker Z, Costa DC, Walker RW, Lee L, Livingston G, Jaros E, et al. Striatal dopamine transporter in dementia with Lewy bodies and Parkinson disease: a comparison. *Neurology.* 2004;62(9):1568-72.
195. Chen NH, Reith ME, Quick MW. Synaptic uptake and beyond: the sodium- and chloride-dependent neurotransmitter transporter family SLC6. *Pflugers Arch.* 2004;447(5):519-31.
196. Harris RC, Zhang MZ. Dopamine, the kidney, and hypertension. *Curr Hypertens Rep.* 2012;14(2):138-43.
197. Soares-da-Silva P, Fernandes MH, Pinto-do OP. Cell inward transport of L-DOPA and 3-O-methyl-L-DOPA in rat renal tubules. *Br J Pharmacol.* 1994;112(2):611-5.
198. Soares-Da-Silva P, Serrao MP, Vieira-Coelho MA. Apical and basolateral uptake and intracellular fate of dopamine precursor L-dopa in LLC-PK1 cells. *Am J Physiol.* 1998;274(2 Pt 2):F243-51.
199. Seri I, Kone BC, Gullans SR, Aperia A, Brenner BM, Ballermann BJ. Influence of Na<sup>+</sup> intake on dopamine-induced inhibition of renal cortical Na<sup>(+)</sup>-K<sup>(+)</sup>-ATPase. *Am J Physiol.* 1990;258(1 Pt 2):F52-60.
200. Carey RM. Theodore Cooper Lecture: Renal dopamine system: paracrine regulator of sodium homeostasis and blood pressure. *Hypertension.* 2001;38(3):297-302.
201. Cuevas S, Villar VA, Jose PA, Armando I. Renal dopamine receptors, oxidative stress, and hypertension. *Int J Mol Sci.* 2013;14(9):17553-72.

202. Yang Y, Zhang Y, Cuevas S, Villar VA, Escano C, L DA, et al. Paraoxonase 2 decreases renal reactive oxygen species production, lowers blood pressure, and mediates dopamine D2 receptor-induced inhibition of NADPH oxidase. *Free Radic Biol Med.* 2012;53(3):437-46.
203. Choi MR, Kouyoumdzian NM, Rukavina Mikusic NL, Kravetz MC, Roson MI, Rodriguez Fermepin M, et al. Renal dopaminergic system: Pathophysiological implications and clinical perspectives. *World J Nephrol.* 2015;4(2):196-212.
204. Hasko G, Szabo C, Nemeth ZH, Deitch EA. Dopamine suppresses IL-12 p40 production by lipopolysaccharide-stimulated macrophages via a beta-adrenoceptor-mediated mechanism. *J Neuroimmunol.* 2002;122(1-2):34-9.
205. Alpern RJ, Herbert SC, Seldin DW, Giebisch GH. Seldin and Giebisch's the kidney : physiology & pathophysiology. 4th ed. Amsterdam ; Boston: Elsevier Academic Press; 2008.
206. Qian W, Nishikawa M, Haque AM, Hirose M, Mashimo M, Sato E, et al. Mitochondrial density determines the cellular sensitivity to cisplatin-induced cell death. *Am J Physiol Cell Physiol.* 2005;289(6):C1466-75.
207. Kang HM, Huang S, Reidy K, Han SH, Chinga F, Susztak K. Sox9-Positive Progenitor Cells Play a Key Role in Renal Tubule Epithelial Regeneration in Mice. *Cell Rep.* 2016;14(4):861-71.
208. Berger K, Moeller MJ. Mechanisms of epithelial repair and regeneration after acute kidney injury. *Semin Nephrol.* 2014;34(4):394-403.
209. Sallustio F, De Benedictis L, Castellano G, Zaza G, Loverre A, Costantino V, et al. TLR2 plays a role in the activation of human resident renal stem/progenitor cells. *FASEB J.* 2010;24(2):514-25.
210. Tillou X, Doerfler A, Collon S, Kleinclaus F, Patard JJ, Badet L, et al. De novo kidney graft tumors: results from a multicentric retrospective national study. *Am J Transplant.* 2012;12(12):3308-15.
211. Sherston SN, Carroll RP, Harden PN, Wood KJ. Predictors of cancer risk in the long-term solid-organ transplant recipient. *Transplantation.* 2014;97(6):605-11.
212. Doublet JD, Peraldi MN, Gattegno B, Thibault P, Sraer JD. Renal cell carcinoma of native kidneys: prospective study of 129 renal transplant patients. *J Urol.* 1997;158(1):42-4.
213. Le Gal K, Ibrahim MX, Wiel C, Sayin VI, Akula MK, Karlsson C, et al. Antioxidants can increase melanoma metastasis in mice. *Sci Transl Med.* 2015;7(308):308re8.
214. Piskounova E, Agathocleous M, Murphy MM, Hu Z, Huddlestun SE, Zhao Z, et al. Oxidative stress inhibits distant metastasis by human melanoma cells. *Nature.* 2015;527(7577):186-91.
215. Sayin VI, Ibrahim MX, Larsson E, Nilsson JA, Lindahl P, Bergo MO. Antioxidants accelerate lung cancer progression in mice. *Sci Transl Med.* 2014;6(221):221ra15.
216. Davison CA, Durbin SM, Thau MR, Zellmer VR, Chapman SE, Diener J, et al. Antioxidant enzymes mediate survival of breast cancer cells deprived of extracellular matrix. *Cancer Res.* 2013;73(12):3704-15.

217. Lawenda BD, Kelly KM, Ladas EJ, Sagar SM, Vickers A, Blumberg JB. Should supplemental antioxidant administration be avoided during chemotherapy and radiation therapy? *J Natl Cancer Inst.* 2008;100(11):773-83.
218. Meiser J, Weindl D, Hiller K. Complexity of dopamine metabolism. *Cell Commun Signal.* 2013;11(1):34.
219. Sastry KS, Karpova Y, Prokopovich S, Smith AJ, Essau B, Gersappe A, et al. Epinephrine protects cancer cells from apoptosis via activation of cAMP-dependent protein kinase and BAD phosphorylation. *J Biol Chem.* 2007;282(19):14094-100.
220. Schrodter S, Braun M, Syring I, Klumper N, Deng M, Schmidt D, et al. Identification of the dopamine transporter SLC6A3 as a biomarker for patients with renal cell carcinoma. *Mol Cancer.* 2016;15(1):10.
221. Dahl SG, Sylte I, Ravna AW. Structures and models of transporter proteins. *J Pharmacol Exp Ther.* 2004;309(3):853-60.
222. Manavalan P, Smith AE, McPherson JM. Sequence and structural homology among membrane-associated domains of CFTR and certain transporter proteins. *J Protein Chem.* 1993;12(3):279-90.
223. Wang L, Zhang Q, Li H, Zhang H. SPECT molecular imaging in Parkinson's disease. *J Biomed Biotechnol.* 2012;2012:412486.



