KCNA5 gene is not confirmed as a systemic sclerosis-related pulmonary arterial hypertension genetic susceptibility factor

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KCNA5 gene is not confirmed as a systemic sclerosis-related pulmonary arterial hypertension genetic susceptibility factor

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Abstract

Introduction: Potassium voltage-gated channel shaker-related subfamily member 5 (KCNA5) is implicated in vascular tone regulation, and its inhibition during hypoxia produces pulmonary vasoconstriction. Recently, a protective association of the KCNA5 locus with systemic sclerosis (SSc) patients with pulmonary arterial hypertension (PAH) was reported. Hence, the aim of this study was to replicate these findings in an independent multicenter Caucasian SSc cohort.

Methods: The 2,343 SSc cases (179 PAH positive, confirmed by right-heart catheterization) and 2,690 matched healthy controls from five European countries were included in this study. Rs10744676 single-nucleotide polymorphism (SNP) was genotyped by using a TaqMan SNP genotyping assay.

Results: Individual population analyses of the selected KCNA5 genetic variant did not show significant association with SSc or any of the defined subsets (for example, limited cutaneous SSc, diffuse cutaneous SSc, anti-centromere autoantibody positive and anti-topoisomerase autoantibody positive). Furthermore, pooled analyses revealed no significant evidence of association with the disease or any of the subsets, not even the PAH-positive group. The comparison of PAH-positive patients with PAH-negative patients showed no significant differences among patients.

Conclusions: Our data do not support an important role of KCNA5 as an SSc-susceptibility factor or as a PAH-development genetic marker for SSc patients.
independently replicated [10]. Wipff et al. [11] recently described the association of potassium voltage-gated channel shaker-related subfamily member 5 (KCNA5) with SSc-related pulmonary arterial hypertension (PAH).

Potassium voltage-gated channels (Kv channels) are homo- or heterotetramers of structural α-subunits and regulatory β-subunits, which control potassium (K) flux. K balance is known to regulate the apoptotic cell shrinkage, a main event in the apoptotic process [12]. Specifically, KCNA5 participates in pulmonary artery smooth muscle cell (PASMC) apoptosis control [13]. It has been reported that overexpression of the KCNA5 gene induces accelerated K efflux and increases caspase-3 proteolytic activity, promoting apoptosis [13].

KCNA5 is also involved in membrane-potential maintenance and vascular-tone regulation [14]. Hypoxic conditions produce a specific inhibition of KCNA5 in the PASMCs, causing pulmonary vasconstriction [15-17]. Moreover, it has been reported that primary pulmonary hypertension patients have an intrinsic reduced level of KCNA5 mRNA in their PASMCs, and this characteristic might play an important role in the pathogenesis of the disease [18].

The role of KCNA5 genetic polymorphisms in pulmonary disease also was explored. Remillard et al. [19] analyzed the influence of KCNA5 genetic variants in IPAH, and showed that different single-nucleotide polymorphisms (SNPs) were related to abnormal function or drug responsiveness. Recently, the importance of KCNA5 variants in SSc-related PAH was analyzed [11]. In the previously mentioned study, the association of KCNA5 with SSc and specifically with PAH-positive (PAH⁺) patients was described [11]. Moreover, the rs10744676 polymorphism was reported as the variant underlying the observed association [11].

In light of the previous evidence, the main goal of this report is to replicate the association of KCNA5 rs10744676 polymorphism with SSc and SSc-related PAH in an independent European population of Caucasian ancestry.

Materials and methods

Subjects

Our study comprised 2,343 SSc cases and 2,690 controls from Spain, The Netherlands, Italy, Sweden, and The United Kingdom. All the individuals included in this report were of Caucasian ancestry. Patients were classified as having limited (lcSSc) or diffuse SSc (dcSSc), according to their skin involvement, as defined by LeRoy et al. [20]. The serologic subgroup stratification of the patients was based on the presence of SSc-associated autoantibodies (anti-centromere antibodies (ACAs) and anti-topoisomerase (ATA)). Pulmonary fibrosis was diagnosed by the presence of interstitial abnormalities in high-resolution computed tomography (HRCT). Patients defined as PAH⁺ showed a mean resting pulmonary artery pressure > 25 mm Hg and a pulmonary capillary wedge pressure ≤15 mm Hg, at the time of a right-heart catheterization [11]. The control population consisted of unrelated healthy individuals recruited in the same geographic regions as the SSc patients and matched by age, sex, and ethnicity with the SSc-patient groups. Approval of local ethical committees was obtained from all the participating centers (Comité de Bioética del Consejo Superior de Investigaciones Científicas, Comitato Etico Azienda Ospedaliera Universitaria Integrata di Verona, Local Ethics Committee of the Radboud University Nijmegen Medical Centre, Medische Ethische Commissie Leids Universitair Medisch Centrum, The regional Ethical Review Board in Lund, Local Research Ethics Committee at Glasgow Royal Infirmary, U.O. Comitato di Etica e Sperimentazione Farmaci Fondazione IRCCS Ca’ Granda-Ospedale Maggiore Policlinico di Milano, Local Research Ethics Committee at Glasgow Royal Infirmary, Royal Free Hospital and Medical School Research Ethics Committee, Manchester University Research Ethics Committee). Written informed consent was required for both patients and controls to be included in the study.

Genotyping and statistical analysis

The rs10744676 KCNA5 biallelic variant was analyzed with TaqMan SNP genotyping assay in a 7900HT Real-Time polymerase chain reaction (PCR) system from Applied Biosystems by following the manufacturer’s suggestions (Foster City, CA, USA).

None of the included cohorts showed significant deviation from Hardy-Weinberg equilibrium (HWE) (P value significance threshold, 0.05). Significance for the allelic model in the individual cohort analyses was calculated by 2 × 2 contingency tables and the Fisher Exact test or χ² when necessary. Odds ratios (ORs) were calculated according to the Woolf method. A Breslow-Day test (BD test) was performed to assess the homogeneity of the association among populations, and pooled analysis under a Cochran-Mantel-Haenszel fixed-effect model was used to analyze jointly all the included cohorts. Statistical analyses were performed as implemented in PLINK (v1.07) software package [21]. Power was calculated by using the software Power Calculator for Genetic Studies 2006 and assuming an additive model and previously reported minor allele frequency (MAF) and ORs [22].

Results

The power of our replication study in each stratified analysis is summarized in Table 1. We emphasize that the size of our pooled PAH⁺ patient subgroup represents the largest SSc-related PAH⁺ cohort analyzed to date (n = 179).
The power of the study of this clinical feature in our population to detect an association equivalent to that previously reported by Wipff et al. is 95%, at the 5% significance level. Table 2 shows the results of the comparison of the complete set of SSc and each of the previously defined subgroups with the healthy control group. As observed in the table, the analyzed polymorphism showed no significant association with either the disease or any of the subsets, not even the PAH-positive group. Furthermore, no significant association was observed in the individual population analyses, either in the whole disease comparison or in the different subphenotypes (see Additional file 1, Table S1). The different cohorts showed a high interpopulation combinability, as can be observed in the Breslow-Day test results (Table 2).

The minor allele in the Spanish and Italian control populations included in this study was less frequent than that in the pooled population described by the Wipff et al. (rs10744676*C FrequencySpain = 10.48%; rs10744676*C FrequencyItaly = 10.00%; rs10744676*C Frequencypooled.Wipff = 14.7%). However, these frequencies are in concordance with those reported for the HapMap CEU population (MAF_{HapMap.CEU} = 10.00%). Moreover, MAF differences have been reported in different European populations (MAF_{1000Genomes.CEU} = 17.64%, MAF_{1000Genomes.GRB} = 15.73%, MAF_{1000Genomes.FIN} = 13.98%, MAF_{1000Genomes.TSI} = 8.16%).

In addition, the analysis of PAH+ versus PAH− patients revealed no phenotype-restricted association in this subgroup (P_{MH} = 0.59; OR, 0.91; 95% CI, 0.64 to 1.28).

### Discussion

Despite the previous findings of an association of the rs10744676 KCNA5 genetic variant with PAH+ SSc patients, our data do not corroborate the reported protective effect of the minor allele of this polymorphism on SSc-related PAH.

It is worth mentioning that the prevalence of right-heart catheterization-confirmed PAH in our pooled population (7.64%) is consistent with previous reports [11]. However, SSc is a chronic progressive disease, and some patients classified as PAH− might develop PAH in the future. Therefore, we consider this issue a limitation of our study and similar reports. In this context, we point out that the mean disease duration for the patient group included in this study (mean disease duration, 13.9 ± 14.4 years, as reported in Bossini-Castillo et al. [23]) was higher than that in Wipff et al. [11] (11.9 ± 9.4 years in the discovery.

### Table 1 Overall statistical power of the study for KCNA5 rs10744676 SNP in each analyzed disease subtype at the 5% significance level assuming an additive effect model and a minor allele (rs10744676*C) frequency = 0.10 (MAF_{CEU})

<table>
<thead>
<tr>
<th>Subgroup</th>
<th>Genotype</th>
<th>Allele test</th>
</tr>
</thead>
<tbody>
<tr>
<td>Controls</td>
<td>C/C</td>
<td>C/T</td>
</tr>
<tr>
<td>SS c</td>
<td>100</td>
<td>99</td>
</tr>
<tr>
<td>lcSSc</td>
<td>100</td>
<td>99</td>
</tr>
<tr>
<td>dcSSc</td>
<td>96</td>
<td>91</td>
</tr>
<tr>
<td>ACA+</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>ATA+</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>Fib+</td>
<td>96</td>
<td>91</td>
</tr>
<tr>
<td>PAH+</td>
<td>100</td>
<td>100</td>
</tr>
</tbody>
</table>

* Reference OR, 0.62, except for the PAH+ group, in which OR = 0.47. **Reference OR (lower previously reported 95% CI) = 0.48, except for the PAH+ group, in which OR = 0.32. **Reference OR (upper previously reported 95% CI) = 0.79, except for the PAH+ group, in which OR = 0.71.

### Table 2 Pooled analysis and stratified analyses of SSc patients and healthy controls for rs10744676 genetic variant, located in the KCNA5 gene

<table>
<thead>
<tr>
<th>Subgroup (n)</th>
<th>Genotype, n (%)</th>
<th>Allele test</th>
</tr>
</thead>
<tbody>
<tr>
<td>Controls (n = 2690)</td>
<td>C/C 37 (1.38)</td>
<td>C/T 597 (22.19)</td>
</tr>
<tr>
<td>SS c (n = 2,343)</td>
<td>C/C 43 (1.84)</td>
<td>C/T 471 (20.10)</td>
</tr>
<tr>
<td>lcSSc (n = 1,642)</td>
<td>C/C 31 (1.89)</td>
<td>C/T 327 (19.91)</td>
</tr>
<tr>
<td>dcSSc (n = 701)</td>
<td>C/C 12 (1.71)</td>
<td>C/T 144 (20.54)</td>
</tr>
<tr>
<td>ACA+ (n = 931)</td>
<td>C/C 18 (1.93)</td>
<td>C/T 197 (21.16)</td>
</tr>
<tr>
<td>ATA+ (n = 568)</td>
<td>C/C 8 (1.41)</td>
<td>C/T 117 (20.60)</td>
</tr>
<tr>
<td>Fib+ (n = 771)</td>
<td>C/C 14 (1.82)</td>
<td>C/T 154 (19.97)</td>
</tr>
<tr>
<td>PAH+ (n = 179)</td>
<td>C/C 2 (1.12)</td>
<td>C/T 37 (20.60)</td>
</tr>
</tbody>
</table>

MAF, minor allele frequency; P_{MH}, Mantel-Haenszel test under fixed-effect P values. Controls are used as reference for all comparisons, and P values have been calculated for the allelic model. OR, odds ratio for the minor allele; 95% CI, 95% confidence interval; SSc, systemic sclerosis; lcSSc, limited cutaneous systemic sclerosis; dcSSc, diffuse cutaneous systemic sclerosis; ACA+, anti-centromere autoantibody-positive patient; ATA+, anti-topoisomerase autoantibody-positive patient; Fib+, lung fibrosis-positive patient (HRCT); PAH+, pulmonary arterial hypertension-positive patients (right-heart catheterization).
cohort and 13.9 ± 14.4 years in the replication), which might influence the observed differences.

The PAH* cohort studied in this report reached a 95% estimated statistical power to detect an association equivalent to that observed by Wipff et al. within this subgroup (OR, 0.47, in the comparison between PAH* SSc patients and controls of the discovery phase and two replication steps [11]). Hence, we consider that the lack of replication observed in our study is unlikely to be caused by a type II error (false negative) because of a reduced statistical power. However, autoimmune-associated variants usually show modest degrees of risk, especially in the case of non-HLA loci [24]. Wipff et al. reported an OR for KCNA5 rs10744676 polymorphism that was remarkably more protective than the SSC genetic-assocation standards for non-HLA loci (that is, modest ORs between 0.70 and 1). Thus, we reflect that if the influence of rs10744676 in the SSC-patient genetic predisposition to PAH is modest, the statistical power to detect a possible association in the PAH* cohort analyzed in the present article might be insufficient, and a possible modest effect of KCNA5 rs10744676 might be overlooked (Table 1).

In addition, it is established that PAH is more frequent in ACA* patients [1], and despite the suggestive association with PAH* patients, no significant association with the ACA* subphenotype was identified in the previous SSC study [11] or in this report. This fact is also consistent with a lack of association of the selected polymorphism with PAH development.

Although rs10744676 might have a functional role in KCNA5 expression because of its location in its putative promoter, no evidence confirms a functional role for this variant. Therefore, we speculate that the previously mentioned association with SSc could be the consequence of a tagged causal variant yet to be discovered. Moreover, because PAH onset time has not been considered for the analyses, it might act as a confounding factor in the discrepant results.

Conclusions

In summary, our data do not support an important role of rs10744676 as a PAH genetic marker in SSc patients.

Additional Material

Additional file 1: Genotype and minor allele frequencies of KCNA5 rs10744676 genetic variant in five European cohorts. This file contains Table S1, showing the genotype and allele distributions of KCNA5 rs10744676 genetic variant in five European cohorts (2,343 SSC cases and 2,690 controls).

Abbreviations

ACA: anti-centromere autoantibodies; ATA: anti-topoisomerase antibody; BD test: Breslow-Day test; CD226: cluster of differentiation 226; CEU: Utah residents with ancestry from northern and western Europe; dcSSC: diffuse cutaneous systemic sclerosis; HRT: High-resolution computed tomography; HWE: Hardy-Weinberg equilibrium; K: potassium; KCNA5: potassium voltage-gated channel shaker-related subfamily member 5, Kv channels: potassium voltage-gated channels; lcSSc: limited cutaneous systemic sclerosis; MAF: minor allele frequency; NOTCH4: Notch (Drosophila) homologue 4; OR: odds ratio; PAH: pulmonary arterial hypertension; PASMC: pulmonary artery smooth muscle cell; PCR: polymerase chain reaction; SNP: single-nucleotide polymorphism; SSc: systemic sclerosis.
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